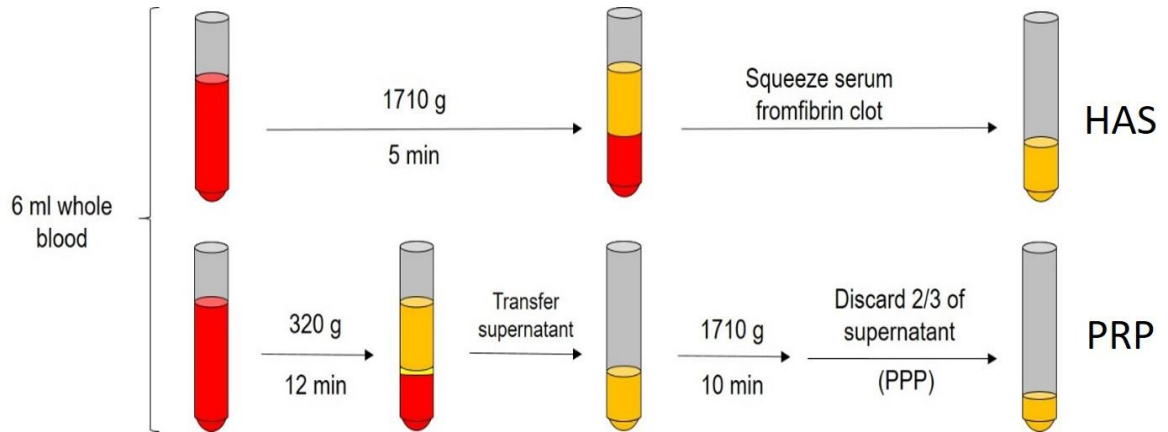


## Supplementary data



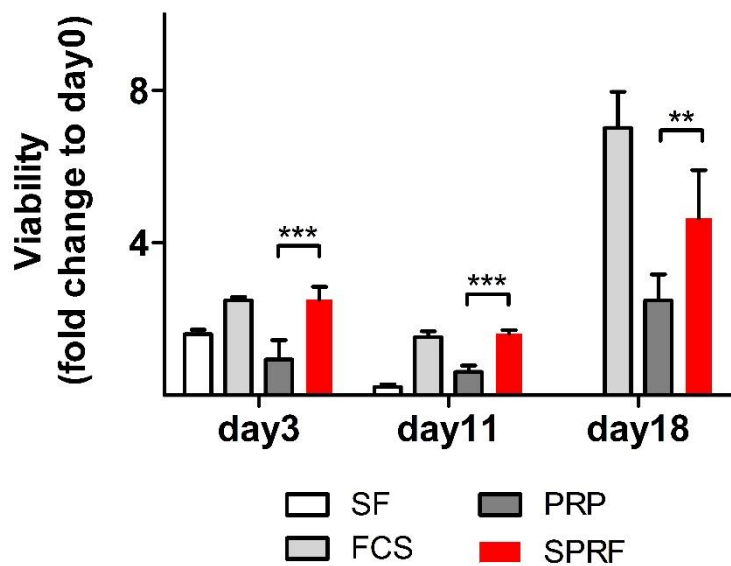
*Supplementary figure 1. Isolation method of HAS and PRP.*

		<b>GENE NAME</b>	<b>ASSAY ID</b>	<b>GENE SYMBOL</b>	<b>ENTREZ GENE ID</b>
1.	Mesenchymal stem cell markers	activated leukocyte cell adhesion molecule	Hs00977641_m1	ALCAM	214
2.		integrin subunit beta 1	Hs01127536_m1	ITGB1	3688
3.		alanyl aminopeptidase	Hs00174265_m1	ANPEP	290
4.		endoglin	Hs00923996_m1	ENG	2022
5.	Osteoblast markers	collagen type I.	Hs00164004_m1	COL1A1	1277
6.		runt related transcription factor 2	Hs01047973_m1	RUNX2	860
7.		alkaline phosphatase, liver/bone/kidney	Hs01029144_m1	ALPL	249
9.	Adipocyte markers	peroxisome proliferator activated receptor gamma	Hs00234592_m1	PPARG	5468
10.		fatty acid binding protein 4	Hs01086177_m1	FABP4	2167
11.		adiponectin	Hs00605917_m1	ADIPOQ	9370
12.	Hematopoietic stem cell markers	CD14	Hs02621496_s1	CD14	929
13.		CD34	Hs02576480_m1	CD34	947
14.		protein tyrosine phosphatase, receptor type C	Hs04189704_m1		5788
15.	Osteocyte markers	podoplanin	Hs00366766_m1	PDPN	10630
16.		dentin matrix acidic phosphoprotein 1	Hs01009390_m1	DMP1	1758
17.		matrix extracellular phosphoglycoprotein	Hs00220237_m1	MEPE	56955
18.	Apoptotic marker	BCL2 associated X	Hs00180269_m1	BAX	581
19.	Antiapoptotic marker	BCL2	Hs00608023_m1	BCL2	596
20.	Houskeeping control gene	actin beta	Hs01060665_g1	ACTB	60

*Supplementary figure 2. Real-time qPCR primer collection.*

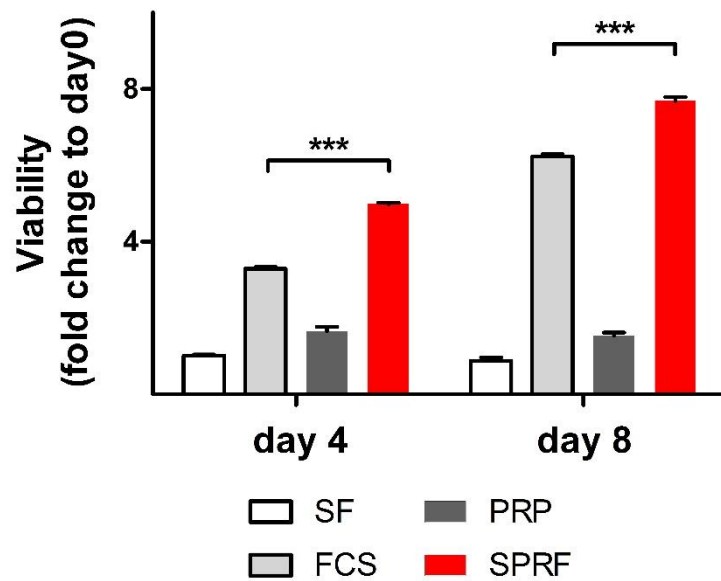
<b>Positive Marker Cocktail</b>	CD105 PerCP-Cy5.5/CD73 APC/CD90 FITC
<b>Additional Positive Drop-In Marker</b>	CD44 PE
<b>Negative Marker Cocktail</b>	CD45/CD34/CD11b/CD19/HLA-DR PE
<b>Isotype Controls</b>	mlgG1, κ PerCP-Cy5.5/ mlgG1, κ APC / mlgG1, κ FITC (for positive cocktail)
	mlgG1, κ/mlgG2a, κ PE (for negative cocktail)
	mlgG2b,κ PE (for CD44 drop in)

*Supplementary figure 3. Flow cytometry analysis antibody cocktails.*



*Supplementary figure 4. Time-course effect of serum supplements on osteoblasts.*

Subconfluent osteoblasts (LONZA, Walkersville, USA) were cultured in high glucose DMEM with pyruvate (Gibco, Paisley, Scotland) and with antibiotics (penicillin 200 U/mL; streptomycin 0.2 mg/mL, and amphotericin B 2.5 µg/mL) (Sigma-Aldrich, St. Louis, USA), and l-ascorbic acid (50 µg/mL; Sigma-Aldrich, St. Louis, USA). in the absence of supplement (□), 10 (v/v)% of FCS (▤), 10 (v/v)% FCS + 1 ng/mL bFGF (▥), 10 (v/v)% PRP (▦) or 10 (v/v)% SPRF (■). XTT viability assay was performed on the third, 11<sup>th</sup> and 18<sup>th</sup> day. Results are presented as means of triplicate samples in three experiments ± SD.  $p < 0.0001$  \*\*\*.



*Supplementary figure 5. Time-course effect of serum supplements on osteoarthritic chondrocytes.* Cells were cultured in DMEM/F12 (Gibco, Paisley, Scotland) with antibiotics (penicillin 200 U/mL; streptomycin 0.2 mg/mL, and amphotericin B 2.5 µg/mL) (Sigma-Aldrich, St. Louis, USA), and l-ascorbic acid (50 µg/mL; Sigma-Aldrich, St. Louis, USA), in the absence of supplement (□), 10 (v/v)% of FCS (▤), 10 (v/v)% FCS + 1 ng/mL bFGF (▥), 10 (v/v)% PRP (▦) or 10 (v/v)% SPRF (■). XTT viability assay was performed on the 4<sup>th</sup> and 8<sup>th</sup> day. Results are presented as means of triplicate samples in three experiments ± SEM.  $p < 0,0001$  \*\*\*.