Supplemental Information

Dynamic Role of SETD7 as a Transcriptional Activator of Cardiac Lineage Commitment

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0	0	•	Calcium signaling pathway	•	٥	٥	•	Developmental Biology		
			Dilated cardiomyonathy	•	0	0	۰	FGFR1b ligand binding and activation		
				•	0	0	۰	Loss of Function of FBXW7 in Cancer and NOTCH1 Signali	ng	
٠	٠	•	Selenocompound metabolism	•	0	0	۰	Activation of NOXA and translocation to mitochondria		
٠	٠	•	Melanogenesis	٥	•	0	٠	HATs acetylate histones		
	0	0	Systemic lupus erythematosus	0	•	0	0	Chromatin modifying enzymes		
•	٥	0	Glycine, serine and threonine metab	olism [.]	•	0	٠	Chromatin organization		
۰		۰	Notch signaling pathway	0	•		٠	SIRT1 negatively regulates rRNA Expression		
			Taurine and hypotaurine metabolism			0	٠	PRC2 methylates histones and DNA		
	•					0	٠	Condensation of Prophase Chromosomes		
0	•	0	whit signaling pathway	٠		0	0	Cellular Senescence		
0	•	٠	beta-Alanine metabolism	۰		0	۰	Meiotic recombination		
٠	•	•	Butanoate metabolism	۰		0	٠	Amyloids		
0	•	0	Alanine, aspartate and glutamate metabolisi			0	٠	formation of the beta-catenin:TCF transactivating complex		
0	•	0	Type I diabetes mellitus	0	0		۰	GABA synthesis		
0	•	0	Adherens junction	•	0			Constitutive Signaling by NOTCH1		
	, in the second s		Lunertrenkie eerdiemuenethy (LCM	•	0			Signaling by NOTCH3		
ũ	۰				0			Signaling by NOTCH4		
0	0		IGF-beta signaling pathway	•	٠			Receptor-ligand binding initiates the second proteolytic cleavage of Notch receptor		
٠	0		Cardiac muscle contraction Hedgehog signaling pathway		٠			Constitutive Signaling by NOTCH1 HD Domain Mutants		
0	٠				0		٠	negative regulation of ICF-dependent signaling by WNT ligand antagonists		
٠	٠		Endocytosis	۵	0		•	GABA synthesis, release, reuptake and degradation		
0	•	•	Cytokine-cytokine receptor interaction	'n	0	•	٠	RNA Polymerase III Chain Elongation		
			Optopologi differentiation		•		•	RNA Polymerase III Transcription Termination		
				۰	0	0				
0	0	•	Light junction -Log(P	· value)	0	0		Muscle contraction	-Log(P-value)	
0	٥	٠	Pathways in cancer	0.0	•	0		Molecules associated with elastic fibres	• 0.0	
			•	2.0	•	•			• 2.0 • 4.0	
				6.0	v			Signaling by BMP	6.0	
			_ ≥	8.0	•	•		Socium/Calcium exchangers	○ ≥ 8.0	
				•		•				
			Min	Max	0	÷		Extracellular matrix organization	Min Max	

Figure S1 (Related to Figure 1). Dynamic enrichment pattern of SETD7 during CM differentiation.

(A) Representative microscopy images of H7 hESCs and H7 hESC-CMs at differentiation day 10. Scale bar, 100 uM

(B) Representative fluorescence microscopy images of H7 derived CMs. Green color represents cardiac Troponin T and blue color represents DAPI. Scale bar, 10 uM.

(C) Representative line-scan images and spontaneous calcium signaling of H7 hESC-CMs showed regular calcium transients.

(D) The mRNA expression of MYH6, MYL4, TNNT2, TNNT1, BMP2, GATA4, MEF2C, TBX5, and NKX2-5 during CM differentiation was measured by RT-qPCR. Blue bar represents cardiac structure genes and red bar is cardiac transcription factors.

(E-H) The average profiles of SETD7 enrichment in gene body regions of target genes from each cluster.

(I-J) KEGG and Reactome pathway analysis of each cluster. Color code indicates minus log transformed multiple testing adjusted P value of category enrichment.





Figure S2 (Related to Figure 2 and Figure 3). SETD7 is required for early cardiac lineage commitment.

(A) Real-time PCR analysis of cardiac genes and NANOG expression levels in in hESCs (H7) and hESC-CMs expressing doxycycline-inducible SETD7 shRNA or Scramble shRNA. Doxycycline was treated at differentiation day 0. The total RNA was extracted at differentiation day 10. One way ANOVA method and expressed as *P < 0.05; **P < 0.01; ***P < 0.005; (mean \pm SEM; n=4).

(B) Immunoblot analysis of SETD7 protein levels in control and SETD7-overexpressed stable lines at day 14 of CM differentiation. The stable cell lines were generated using a lentiviral overexpression with puromycin selection step. The total lysates were extracted from 3 different experiments.

(C) Flow cytometry analysis of TNNT2 positive cells of control and SETD7-overexpressed stable lines at day 14 of CM differentiation (mean ± SEM; n=5).

(D) Heat-map of differentially expressed genes in mesoderm lineage cells derived from SETD7^{+/+} or SETD7^{-/-} (n=2). GO enrichment analysis of the differentially expressed genes. Color bar indicates multiple testing adjusted q value.

(E) Time line of EB formation and random differentiation.

(F) Immunoblot analysis of cell lysate from hESCs or EBs (day 14) treated with or without doxycycline. Alpha tubulin was used as a loading control.

(G) mRNA level of endomesoderm or ectoderm maker genes in hESCs or EBs (day 14) treated with or without doxycycline.

(H) Timeline of DOX treatment during CM differentiation.

(I-J) Immunoblot analysis of SETD7, GAPDH, H3K36me3, and H3 levels in hESCs, hESC-CMs, and different tissues of human fetus. (Fetus-1: 16.4 week male; Fetus-2: 17.0 week female).



Figure S3 (Related to Figure 3). Effects of SETD7 on lineage-specific gene expression of noncardiomyocyte cell types.

(A) Immunoblot analysis of SETD7, NES, PAX6, GAPDH, H3K36me3, H3Kme1, and H3 level at different time points of NPC differentiation. The total lysates were extracted from 2~3 different experiments. NPC differentiation was performed according to the manufacturer's protocol.

(B) Relative signal intensities of NES and PAX protein level in Figure A.

(C) Real-time PCR analysis of NES, PAX6, SOX2, SOX1, and NEUROG1 expression during NPC differentiation. Statistical significance was analyzed using the one way ANOVA. *P < 0.05; **P < 0.01; ***P < 0.005; (mean ± SEM; n=3).

(D) Immunostaining of NES (green) and DAPI (blue) in SETD7^{+/+} and SETD7^{-/-}-derived NPCs.

(E) Immunoblot analysis of SETD7, AFP, ALB, GAPDH, H3K36me3, and H3 level at different time points of hepatocyte differentiation. The total lysates were extracted from 2 different experiments. Hepatocyte differentiation was performed according to previously published protocol (Song et al., 2009).

(F) Real-time PCR analysis of SOX17, NHE-4, AFP, and ALB expression during hepatocyte differentiation. Statistical significance was analyzed using the one way ANOVA. *P < 0.05; **P < 0.01; ***P < 0.005; (mean ± SEM; n=3).

(G) Immunostaining of AFP (red) and ALB (green) in SETD7^{+/+} and SETD7^{-/-}-derived hepatocyte-like cells.
(H) Representative images of bona fide ECs derived from SETD7^{+/+} and SETD7^{-/-} lines. EC differentiation

was performed according to previous papers (Gu et al., 2017; Hu et al., 2016).

(I) Cell count before (total) and after (CD144⁺) magnetic-activated cell sorting (MACS).

0.05; ***P* < 0.01; ****P* < 0.005; (mean ± SEM; n=3).

Figure S4



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Figure S4 (Related to Figure 4). Identification of co-factors of SETD7 during CM differentiation.

(A) Immunoblot analysis of SETD7 immunoprecipitates (IP line) and cell lysates (Input line) from mesoderm lineage and CM lineage cells derived from hESCs (H7).

(B) Heatmaps showing relative enrichment levels of SETD7 (in human) and Srf, Mef2a, Gata4, and Tbx5 (in mouse), on TSS of ortholog coding genes in CMs. The heatmaps were ranked by the enrichment of SETD7. ChIP-seq data were adapted from a previous study (He et al., 2011).

Figure S5



160

H3K4me³





Figure S5 (Related to Figure 5). Correlation between the SETD7 enrichment and several histone markers at each stage of CM differentiation.

(A) Immunoblot analysis of SETD7, H3K4me1, and H3K36me3. *In vitro* methylation assay was performed with histone 3 and active SETD7-FLAG protein. PFI-2 and DMSO were treated as indicated.

(B-C) Corrgram package analysis was used to calculate correlation/r-values. The piechart where the intersection represents the correlation value between the two factors/marks. The size of the shaded region within the pie reflects correlation/r-values (bigger shaded region represents higher correlation or r-value), whereas the color of the shaded region represents the direction of correlation (blue reflects positive correlation and red reflects negative correlation). The lower panel represents the confidence ellipses and smoothed lines.

(D-F) ChIP-qPCR assay of SETD7, H3K4me3, and H3K27me3 enrichment at promotor region of several lineage specific genes during CM differentiation.





Figure S6 (Related to Figure 5 and Figure 6). SETD7 associates with H3K36me3 on its target genes during CM differentiation.

(A) Peptide array of 384 different histone modification with SETD7 proteins conjugated with two different tags. Red box showed the position of unmethylated H3K36, H3K36me, H3K36me2, H3K36me3, and H3K36Ac.

(B) Hierarchical clustering analysis based on the enrichment of histone marks and SETD7 of genebodies of coding genes.

(C-F) ChIP-qPCR assay of SETD7 and H3K36me3 enrichment at several target genes during CM differentiation. Primer targets gene-body region of each gene.

(G-I) ChIP-qPCR assay of SETD7 and H3K36me3 enrichment at tissue specific genes in different organs of human fetus (Fetus-1: 16.4 week male).

Figure S7



Figure S7 (Related to Figure 6). SETD7 is not necessary for methylation of H3K36, but required for Pol II-mediated gene transcription.

(A) Immunoblot analysis of SETD7 and H3K36me3. *In vitro* methylation assay was performed with histone peptides (biotin-conjugated; H3 a.a. 22-44) and active SETD7-FLAG protein. H3K36me3 peptide were used as positive control.

(B) ChIP-qPCR analysis of H3K36 enrichment at gene-body regions of T and MIXL1 genes at mesoderm stage. Statistical significance was analyzed using the one way ANOVA. *P<0.05; **P < 0.01; (mean ± SEM; n=3).

(C) Genome browser screenshots of SETD7, H3K36me3, Pol II, and PolII-S2P ChIP-seq profiles on MIXL1 gene at mesoderm lineage of SETD7^{+/+} and SETD^{-/-} lines.

(D) The genome-wide average profiles of Pol II enrichment in gene body region.

(E) The genome-wide average profiles of Pol II-S2P enrichment in gene body region of each cluster showed in Figure 5J.