Supplemental Information

Confocal Imaging:

Cells from each stages of microglial differentiation were grown on pre-treated German glass cover slips (GG-18-1.5-pre, NeuVitro, Vancouver, USA). Prior to fixing, they were gently washed 3X with PBS-T (PBS containing 0.1% Triton-X100) for 10 min, followed by two hours of incubation in blocking solution (PBS-T with 5% goat serum). Both conjugated and unconjugated primary antibodies were prepared in the above blocking solution. Following the blocking step, coverslips were incubated overnight 4°C with the primary antibodies. After the overnight incubation, the glass coverslips were washed again 3X in PBS-T for 15 min, and further incubated with respective secondary antibodies for 2 hours at room temperature (RT). The coverslips were washed then 3X for 10 min in PBS-T following incubation at RT and counterstained with ProLongTM Diamond Antifade Mountant containing DAPI for 15 min at RT. Finally the coverslips were washed for 2X in PBS and kept in dark prior to imaging. The confocal images were acquired in Zeiss LSM 710 and analysed using ZEN 2012 SP1 software (Zeiss, Thornwood, NY). Please see the antibody list below for reference (Supplementary table 1).

Flow cytometry analysis

Cultured cells from different stages of microglia induction and differentiation were enzymatically harvested by Accutase treatment for 5 min at 37°C and then gently scrapped with cell scraper (Sigma-Aldrich). Cells were then washed twice using a customized staining buffer that contained 1x PBS with 0.2% bovine serum albumin and 0.09% sodium azide. Cells were pelleted in 5810R Eppendorf centrifuge at a speed of 1600 rpm for 5 mins. These cell pellets were re-suspended in 200 µl of their respective medium containing the required ratio of fluorescence-conjugated antibodies as per the manufacturer's instructions and were incubated on ice for 45 min in a dark place. Following incubation with unconjugated primary antibodies, the cells were washed twice using the above staining buffer and again pelleted using similar centrifugation step. Cell pellets were further resuspended with florochrome-conjugated secondary antibodies and incubated on ice for 1 hour. Prior to analysis, cells were washed, repelleted and finally resuspended in the staining buffer. Isotype controls or secondary antibodies only were used to quantitate the baseline background. Data was acquired using a BD Biosciences ARIA-III workstation and analyzed using BD FACS DivaTM software.

Supplementary Table 1. List of Antibodies used for flow cytometry and immunofluorescence analyses.

Name	Host	Vendor	Cat. No.
CD184-APC	Mouse	BD Pharmingen	555976
CD24-PE	Mouse	BD Pharmingen	555428
SPHK1	Mouse	Santa Cruz Biotechnology, Inc.	sc-365401

Oct3/4-PE	Mouse	BD Pharmingen	560186
PTX3	Mouse	R&D Systems	PP-PPJ0069-00
IBA1	Rabbit	Wako	019-19741
Anti-mouse IgM-RRX	Goat	Jackson Immunoresearch	115-296-020
Anti-rabbit IgG-Alexa-633	Goat	Invitrogen	A21070
Anti-mouse IgG-Alexa-594	Goat	Invitrogen	A11012

Supplementary Table 2. The mapped GO ID to corresponding GO Molecular function /Biological process names.

GO ID	GO process	Adj_pvalue
GO:0004872	receptor activity	3.70E-08
GO:0004871	signaling transducer activity	5.82E-08
GO:0010644	cell communication by electrical coupling	1.02E-07
GO:0038023	signaling receptor activity	1.22E-07
GO:0051270	regulation of cellular component movement	4.12E-07
GO:0008284	positive regulation of cell proliferation	5.92E-07
GO:0034220	ion transmembrane transport	6.32E-07
GO:0044057	regulation of system process	6.70E-07
GO:0009605	GO:0009605 response to external stimulus	
GO:0071310	cellular response to organic substance	1.30E-06
GO:0051050	positive regulation of transport	1.50E-06

GO:0098660	inorganic ion transmembrane transport	1.51E-06
GO:0086065	cell communication involved in cardiac conduction	1.73E-06
GO:0010959	regulation of metal ion transport	1.77E-06
GO:0009719	response to endogenous stimulus	2.29E-06
GO:0055085	transmembrane transport	2.34E-06
GO:0042127	regulation of cell proliferation	2.88E-06
GO:0002376	Immune system process	4.30E-06