## SUPPLEMENTARY FILE

## Membrane depolarization and aberrant lipid distributions in the neonatal rat brain following hypoxic-ischaemic insult

Dominika Luptakova<sup>1,2</sup>, Ladislav Baciak<sup>2,3</sup>, Tomas Pluhacek<sup>1,4</sup>, Anton Skriba<sup>1</sup>, Blanka Sediva<sup>1</sup>, Vladimir Havlicek<sup>1,4\*</sup>, Ivo Juranek<sup>2</sup>

<sup>1</sup>Institute of Microbiology of the Czech Academy of Sciences, Prague 4, 142 20, Czech Republic

<sup>2</sup>Institute of Experimental Pharmacology and Toxicology, CEM of the SAS, Bratislava, 841 04, Slovakia

<sup>3</sup>Slovak University of Technology, Central Laboratories, Bratislava, 812 37, Slovakia

<sup>4</sup>Regional Centre of Advanced Technologies and Materials, Department of Analytical Chemistry, Olomouc, 771 47, Czech Republic

\*vlhavlic@biomed.cas.cz

## **Table of Contents:**

**Figure S1**: Acute (cytotoxic) brain ooedema evolution in 7-day old rat pup (**panel A**; *in vivo* DW-MRI image taken at 0 h after HI insult of pup #16). **Panel B** – Volume of acute eodema in the ipsilateral (ipsi, dotted line) and contralateral (contra, full line) hemisphere (mean  $\pm$  SEM, n = 7). **Panel C** – Evolution of the total lesion size relative to the whole brain volume in time; two rats (out of nine) that died around 36-hour interval were excluded from the graph. Rats # 3, 5, 7, 10 and 18 recovered in time (green lines) while rats # 9 and 16 developed progressive aggravation of brain lesion (red lines). **Panel D** – Representative coronary brain MRI consecutive scans (N=8) of pup #9 taken at acute (0 h), subacute (36 h) and late phase (144 h) of HI injury; cytotoxic oedema is indicated by red arrows (0 h); vasogenic oedema/early necrosis reflected by hypointensive MRI signal are indicated by white arrows (36 h, 144 h).

**Figure S2:** Representative averaged MALDI mass spectra of affected rat pup brain tissue acquired in positive ion mode (**A**) and negative ion mode (**B**).

**Figure S3:** MALDI MSI analysis showing the spatial distribution of Na<sup>+</sup> and K<sup>+</sup> adducts (upper and lower panel, respectively) of phosphocholine PC (14:0/16:0) within the representative HI-insulted (HII) and control (sham) brain at striatal level; the respective characteristic fragmentation spectra are depicted on the left.

**Figure S4:** Sodium and potassium calibration plots in LA-ICP-MSI. Two line scans were ablated along the whole diameter of each filter paper disks (no tissue matrix was present) in the following order: 0, 10, 100, 500, 1000 and 4000  $\mu$ g.g<sup>-1</sup> for both sodium and potassium. The background-subtracted signal intensity data were averaged over the two line scans per each concentration level and plotted against the spiked concentrations. For both calibration curves, the linear relationship was found with the coefficient of determination value over 0.994. Sodium and potassium signals were divided by the

corresponding <sup>12</sup>C isotope signal. This standardization allowed for signal compensation arising from different interactions of the laser with the sample surface as well as local differences in the tissue thickness.

**Figure S5**: The fragmentation spectrum of adenosine diphosphate (ADP) in a negative ion mode (MALDI).

**Figure S6**: The fragmentation spectrum of adenosine monophosphate (AMP) in a negative ion mode (MALDI).

**Figure S7**: MALDI MSI analysis of the subacute molecular changes in the HI-affected rat pup brain (36 h after the insult); molecules discussed in the main text were: phosphocholines (PC), adenosine monophosphate (AMP), adenosine diphosphate (ADP), *N*-acylphosphatidylethanolamine (NAPE), gangliosides (GM2 and GM3), phosphoserines (PS), phosphatidylinositols (PI) and phosphoethanolamines (PE).

**Figure S8**: The fragmentation spectrum of monosialoganglioside GM2 in a negative ion mode (MALDI). All product ions arose from the dissociation in saccharidic part of the molecule.

**Figure S9:** Typical histological picture of the rat pup brain at the late phase (144 h) of HI injury (Panels A, C, D) in comparison to the sham brain (panel B). In somatosensory cortex, morphologically deformed neurons (red circles) were observed in the HI-insulted (ipsilateral) hemisphere (A) while intact neurons (green circle) prevailed in the sham brain (B). At striatal level, necrotic areas (marked in brown) were observed (C); individual necrotic cells (white circles) and reactive astrocyte (black circle) seen in enlarged image (D).

**Figures S10-S16:** Graphical representation of differences in means and medians between HI-insulted (HII) group and control group using standard box-and-whisker diagrams.

**Figures S17-S23**: Graphical representation of the individual results for rats of the HII group compared with the 95% prediction intervals of expected value based on control group (red area – prediction intervals based on control group)

**Table S1**: Lesion size related to cytotoxic ooedema expressed as hemisphere(ipsilateral/contralateral) volume in seven experimental rats. SD and SEM stand for StandardDeviation and Standard Error of the Mean, respectively.

Table S2: Characterization of the observed Na<sup>+</sup> and K<sup>+</sup> adducts of phosphocholines (PCs).

**Table S3**: *N*-acylphosphatidylethanolamines (NAPEs) detected in the HI-affected brains at 36 h after the HI insult.

**Table S4:** Statistical analysis of differences between the HII group and control group performed using nonparametric two-tailed Mann–Whitney U test. *P* values indicating statistical significance are in red color.

**Table S5**: Statistical analysis of differences between the HII group and control group performed using parametric two-tailed t-tests. *P* values indicating statistical significance are in red color.

Table S6: Tissue reduction at 144 hours after the HI insult recorded by MRI.

**Table S7:** Multiple parametric testing of the time effect based on one-way ANOVA. *P* values indicating statistical significance are in red color.

**Table S8**: Multiple nonparametric testing of the time effect based on Kruskal-Wallis method. *P* values indicating statistical significance are in red color.

**Table S9:** Technical variability expressed as Coefficients of Variation (CV) for each group of technical replications.

Table S10: Biological variability expressed as Coefficient of Variation (CV).

Table S11: Percentage of the biological variability from the total variability for each dataset.

**Table S12**: *P*-values of the Lilliefors corrected Kolmogorov-Smirnov test of the normality. Values indicating statistical significance (if the *P*-value rejected normal distribution) are in red.

**Table S13**: Values (mean  $\pm$  SEM) of the relative abundance of the NAPE representatives, GM2 and GM3 of the 0, 36 and 144 time intervals.



**Figure S1**: Acute (cytotoxic) brain oedema evolution in 7-day old rat pup (**panel A**; *in vivo* DW-MRI image taken at 0 h after HI insult of pup #16). **Panel B** – Volume of acute oedema in the ipsilateral (ipsi, dotted line) and contralateral (contra, full line) hemisphere (mean  $\pm$  SEM, n = 7). **Panel C** – Evolution of the total lesion size relative to the whole brain volume in time; two rats (out of nine) that died around 36-hour interval were excluded from the graph. Pups # 3, 5, 7, 10 and 18 recovered in time (green lines) while pups # 9 and 16 developed progressive aggravation of brain lesion (red lines). **Panel D** – Representative coronary brain MRI consecutive scans (N=8) of pup #9 taken at acute (0 h), subacute (36 h) and late phase (144 h) of HI injury; cytotoxic oedema is indicated by red arrows (0 h); vasogenic oedema/early necrosis reflected by hypointensive MRI signal are indicated by white arrows (36 h, 144 h).



**Figure S2:** Representative averaged MALDI mass spectra of affected rat pup brain tissue acquired in positive ion mode (**A**) and negative ion mode (**B**).

![](_page_4_Figure_0.jpeg)

**Figure S3:** MALDI MSI analysis showing the spatial distribution of Na<sup>+</sup> and K<sup>+</sup> adducts (upper and lower panel, respectively) of phosphocholine PC (14:0/16:0) within the representative HI-insulted (HII) and control (sham) brain at striatal level; the respective characteristic fragmentation spectra are depicted on the left.

![](_page_5_Figure_0.jpeg)

**Figure S4:** Sodium and potassium calibration plots in LA-ICP-MSI. Two line scans were ablated along the whole diameter of each filter paper disks (no tissue matrix was present) in the following order: 0, 10, 100, 500, 1000 and 4000  $\mu$ g.g<sup>-1</sup> for both sodium and potassium. The background-subtracted signal intensity data were averaged over the two line scans per each concentration level and plotted against the spiked concentrations. For both calibration curves, the linear relationship was found with the coefficient of determination value over 0.994. Sodium and potassium signals were divided by the corresponding <sup>12</sup>C isotope signal. This standardization allowed for signal compensation arising from different interactions of the laser with the sample surface as well as local differences in the tissue thickness.

![](_page_6_Figure_0.jpeg)

**Figure S5**: The fragmentation spectrum of adenosine diphosphate (ADP) in a negative ion mode (MALDI).

![](_page_6_Figure_2.jpeg)

**Figure S6**: The fragmentation spectrum of adenosine monophosphate (AMP) in a negative ion mode (MALDI).

![](_page_7_Figure_0.jpeg)

**Figure S7**: MALDI MSI analysis of the subacute molecular changes in the HI-affected rat pup brain (36 h after the insult); molecules discussed in the main text were: phosphocholines (PC), adenosine monophosphate (AMP), adenosine diphosphate (ADP), *N*-acylphosphatidylethanolamine (NAPE), gangliosides (GM2 and GM3), phosphoserines (PS), phosphatidylinositols (PI) and phosphoethanolamines (PE).

![](_page_8_Figure_0.jpeg)

**Figure S8**: The fragmentation spectrum of monosialoganglioside GM2 in a negative ion mode (MALDI). All product ions arose from the dissociation in saccharidic part of the molecule.

![](_page_8_Figure_2.jpeg)

**Figure S9**: Typical histological picture of the rat pup brain at the late phase (144 h) of HI injury (**Panels A, C, D**) in comparison to the sham brain (**panel B**). In somatosensory cortex, morphologically deformed neurons (red circles) were observed in the HI-insulted (ipsilateral) hemisphere (**A**) while intact neurons (green circle) prevailed in the sham brain (**B**). At striatal level, necrotic areas (marked in brown) were observed (**C**); individual necrotic cells (white circles) and reactive astrocyte (black circle) seen in enlarged image (**D**).

![](_page_9_Figure_0.jpeg)

**Supplemental Figures S10-S16:** Graphical representation of differences in means and medians between HI-insulted (HII) group and control group using standard box-and-whisker diagrams.

![](_page_10_Figure_0.jpeg)

![](_page_11_Figure_0.jpeg)

![](_page_12_Figure_0.jpeg)

![](_page_13_Figure_0.jpeg)

![](_page_14_Figure_0.jpeg)

![](_page_15_Figure_0.jpeg)

**Supplemental Fig S17-S23**: Graphical representation of the individual results for rats of the HII group compared with the 95% prediction intervals of expected value based on control group (red area – prediction intervals based on control group)

![](_page_16_Figure_1.jpeg)

0

0 h

36 h

144 h

**NAPE 52:4** 

![](_page_17_Figure_0.jpeg)

NAPE 58:6

![](_page_17_Figure_2.jpeg)

![](_page_18_Figure_0.jpeg)

![](_page_18_Figure_1.jpeg)

![](_page_19_Figure_0.jpeg)

Table S1: Lesion size related to cytotoxic ooedema expressed as hemisphere
(ipsilateral/contralateral) volume in seven experimental rats. SD and SEM stand for Standard
Deviation and Standard Error of the Mean, respectively.

Lesion si	ze/ hemisphere vol	ume (%)
rat No.	ipsi	contra
rat3	7.1	0.0
rat5	8.0	0.0
rat7	13.8	0.0
rat9	13.7	0.0
rat10	13.9	0.0
rat16	15.7	4.5
rat18	11.3	7.5
mean (%)	11.9	1.7
SD (%)	3.3	3.1
SEM (%)	1.2	1.2

**Table S2:** Characterization of the observed Na<sup>+</sup> and K<sup>+</sup> adducts of phosphocholines (PCs).

Theoretical <i>m/z</i>	Measured m/z	Error (ppm)	Formula	Phosphocholine	Adduct
728.5201	728.5197	-0.6		DC(14.0/16.0)	Na⁺
744.4940	744.4937	-0.4	C38H76NO8P	PC(14:0/16:0) =	K+
754.5357	754.5352	-0.7		DC(16.0/16.1)	Na⁺
770.5097	770.5102	0.7	C40H78NU8P	PC (10:0/10:1) -	K+
756.5514	756.5505	-1.2		DC(16.0/16.0)	Na <sup>+</sup>
772.5253	772.5249	-0.6	C40H80NU8P	PC (10:0/10:0) -	K+
782.5670	782.5673	0.4		DC(16.0/10.1)	Na⁺
798.5410	798.5411	0.2	C42H82NO8P	PC (10:0/18:1) -	K+
784.5827	784.5793	4.31		DC(16.0/10.0)	Na⁺
800.5566	800.5512	5.26	C42H84NO8P	PC (10:0/18:0) -	K+

Theoretical <i>m/z</i>	Measured m/z	Error (ppm)	Formula	Lipid compound	Relative intensity (%)
974.7219	974.7201	1.90	$C_{57}H_{102}NO_9P$	NAPE (52:5)	10
976.7376	976.7356	2.07	$C_{57}H_{104}NO_9P$	NAPE (52:4)	51
998.7219	998.7210	0.93	$C_{59}H_{102}NO_9P$	NAPE (54:7)	7
1000.7376	1000.7366	0.94	$C_{59}H_{104}NO_9P$	NAPE (54:6)	49
1002.7532	1002.7496	3.68	$C_{59}H_{106}NO_9P$	NAPE (54:5)	52
1004.7689	1004.7282	0.68	$C_{59}H_{108}NO_9P$	NAPE (54:4)	100
1006.7845	1006.7875	3.00	$C_{59}H_{110}NO_9P$	NAPE (54:3)	22
1024.7376	1024.7375	0.11	$C_{61}H_{104}NO_9P$	NAPE (56:8)	6
1026.7532	1026.7493	3.86	$C_{61}H_{106}NO_9P$	NAPE (56:7)	25
1030.7845	1030.7822	2.30	$C_{61}H_{110}NO_9P$	NAPE (56:5)	46
1032.8002	1032.7994	0.81	$C_{61}H_{112}NO_9P$	NAPE (56:4)	53
1034.8158	1034.8155	0.35	$C_{61}H_{114}NO_9P$	NAPE (56:3)	11
1052.7689	1052.7663	2.46	$C_{63}H_{108}NO_9P$	NAPE (58:8)	7
1054.7845	1054.7826	1.88	$C_{63}H_{110}NO_9P$	NAPE (58:7)	18
1056.8002	1056.7990	1.09	$C_{63}H_{112}NO_9P$	NAPE (58:6)	37
1060.8315	1060.8323	0.82	$C_{63}H_{116}NO_9P$	NAPE (58:4)	9

**Table S3**: *N*-acylphosphatidylethanolamines (NAPEs) detected in the HI-affected brains at 36 h after the HI insult.

**Table S4:** Statistical analysis of differences between the HII group and control group performed using nonparametric two-tailed Mann–Whitney U test. *P* values indicating statistical significance are in red color.

	NAPE	NAPE	NAPE	NAPE	Sum of		
	52:4	54:4	56:4	58:6	NAPEs	GM2	GM3
Time 0 h	0.182	0.180	0.061	0.061	0.180	0.132	0.240
Time 36 h	0.015	0.026	0.061	0.015	0.026	0.026	0.004
Time 144 h	1.000	0.486	1.000	1.000	0.486	0.486	0.200

**Table S5**: Statistical analysis of differences between the HII group and control group performed using parametric two-tailed t-tests. *P* values indicating statistical significance are in red color.

	NAPE	NAPE	NAPE	NAPE	Sum of		
	52:4	54:4	56:4	58:6	NAPEs	GM2	GM3
Time 0h	0.115	0.080	0.048	0.036	0.059	0.220	0.177
Time 36h	0.113	0.124	0.120	0.121	0.120	0.012	0.016
Time 144h	NaN	0.882	NaN	NaN	0.882	0.528	0.125

Н	II	SHA	AM
rat No.	144h	rat No.	144h
rat3	8.9%	rat12S	0.7%
rat5	11.2%	rat13S	2.7%
rat7	15.6%	rat14S	2.4%
rat9	18.1%	rat15S	0.1%
rat10	9.3%	rat17S	3.3%
rat16	17.1%	rat19S	4.1%
rat18	11.3%		
mean (%)	13.1	mean (%)	2.2
<b>SD</b> (%)	3.8	<b>SD</b> (%)	1.5
<b>SEM</b> (%)	1.4	<b>SEM (%)</b>	0.6

Table S6: Tissue reduction at 144 hours after the HI insult recorded by MRI.

**Table S7**: Multiple parametric testing of the time effect based on one-way ANOVA. *P* values indicating statistical significance are in red color.

	NAPE	NAPE	NAPE	NAPE	Sum of		
	52:4	54:4	56:4	58:6	NAPEs	GM2	GM3
Time 0h ver. 36h	0.188	0.228	0.276	0.269	0.235	0.003	0.013
Time 0h ver. 144h	1.000	1.000	1.000	1.000	1.000	1.000	1.000
Time 36h ver.144h	0.219	0.233	0.240	0.241	0.233	0.010	0.107

**Table S8**: Multiple nonparametric testing of the time effect based on Kruskal-Wallis method. *P* values indicating statistical significance are in red color.

	NAPE	NAPE	NAPE	NAPE	Sum of		
	52:4	54:4	56:4	58:6	NAPEs	GM2	GM3
Time 0h ver. 36h	0.521	0.825	1.000	1.000	0.825	0.046	0.039
Time 0h ver. 144h	0.639	0.278	0.353	0.268	0.278	1.000	0.667
Time 36h ver.144h	0.041	0.024	0.096	0.052	0.024	0.248	0.947

**Table S9:** Technical variability expressed as Coefficients of Variation (CV) for each group of technical replications.

	NAPE	NAPE	NAPE	NAPE	Sum of		
HII group	52:4	54:4	56:4	58:6	NAPEs	GM2	GM3
Time 0h	0.018±0.010	0.035±0.007	0.033±0.013	0.042±0.021	0.043±0.008	0.094±0.012	0.114±0.014
Time 36h	0.077±0.031	0.068±0.012	0.034±0.012	0.057±0.017	0.072±0.015	0.097±0.026	0.078±0.014

Time 144h	0	0.034±0.007	0	0	0.034±0.007	0.068±0.017	0.098±0.025
	NAPE	NAPE	NAPE	NAPE	Sum of		
Control group	52:4	54:4	56:4	58:6	NAPEs	GM2	GM3
Time 0h	0	0.043±0.007	0	0	0.043±0.007	0.093±0.012	0.135±0.020
Time 36h	0	0.048±0.013	0	0	0.048±0.013	0.082±0.020	0.121±0.040
Time 144h	0	0.043±0.009	0	0	0.043±0.009	0.053±0.013	0.055±0.024

Table S10: Biological variability expressed as Coefficient of Variation (CV).

	NAPE	NAPE	NAPE	NAPE	Sum of		
HII group	52:4	54:4	56:4	58:6	NAPEs	GM2	GM3
Time 0h	1.286	0.677	0.942	0.859	0.817	0.228	0.182
Time 36h	1.278	1.233	1.309	1.313	1.272	0.465	0.335
Time 144h	0.000	0.218	0.000	0.000	0.218	0.262	0.140
	NAPE	NAPE	NAPE	NAPE	Sum of		
Control group	NAPE 52:4	NAPE 54:4	NAPE 56:4	NAPE 58:6	Sum of NAPEs	GM2	GM3
<b>Control group</b> Time 0h	NAPE 52:4 0.000	<b>NAPE 54:4</b> 0.146	NAPE 56:4 0.000	NAPE 58:6 0.000	Sum of NAPEs 0.146	<b>GM2</b> 0.189	<b>GM3</b> 0.130
<b>Control group</b> Time Oh Time 36h	NAPE 52:4 0.000 0.000	NAPE 54:4 0.146 0.170	NAPE 56:4 0.000 0.000	NAPE 58:6 0.000 0.000	Sum of NAPEs 0.146 0.170	<b>GM2</b> 0.189 0.259	<b>GM3</b> 0.130 0.121

**Table S11**: Percentage of the biological variability from the total variability for each dataset.

	NAPE	NAPE	NAPE	NAPE	Sum of		
HII group	52:4	54:4	56:4	58:6	NAPEs	GM2	GM3
Time 0h	99.78%	99.85%	99.74%	99.00%	99.77%	86.23%	75.67%
Time 36h	99.50%	99.62%	99.68%	99.66%	99.62%	95.28%	93.20%
Time 144h		97.09%			97.09%	94.72%	67.26%
	NAPE	NAPE	NAPE	NAPE	Sum of		
Control group	NAPE 52:4	NAPE 54:4	NAPE 56:4	NAPE 58:6	Sum of NAPEs	GM2	GM3
<b>Control group</b> Time 0h	NAPE 52:4	<b>NAPE 54:4</b> 91.72%	NAPE 56:4	NAPE 58:6	Sum of NAPEs 91.72%	<b>GM2</b> 80.91%	<b>GM3</b> 52.66%
<b>Control group</b> Time Oh Time 36h	NAPE 52:4	NAPE 54:4 91.72% 90.22%	NAPE 56:4	NAPE 58:6	Sum of NAPEs 91.72% 90.22%	<b>GM2</b> 80.91% 88.41%	<b>GM3</b> 52.66% 39.05%

**Table S12**: *P*-values of the Lilliefors corrected Kolmogorov-Smirnov test of the normality. Values indicating statistical significance (if the *P*-value rejected normal distribution) are in red.

	NAPE	NAPE	NAPE	NAPE	Sum of		
HII group	52:4	54:4	56:4	58:6	NAPEs	GM2	GM3
Time 0h	0.148	0.500	0.500	0.500	0.500	0.500	0.450
Time 36h	0.243	0.246	0.304	0.210	0.267	0.034	0.500
Time 144h	<0.001	0.202	<0.001	<0.001	0.202	0.500	0.500
	NAPE	NAPE	NAPE	NAPE	Sum of		
Control group	NAPE 52:4	NAPE 54:4	NAPE 56:4	NAPE 58:6	Sum of NAPEs	GM2	GM3
<b>Control group</b> Time 0h	NAPE 52:4 < <mark>0.001</mark>	<b>NAPE</b> <b>54:4</b> 0.182	NAPE 56:4 < <mark>0.001</mark>	NAPE 58:6 < <mark>0.001</mark>	Sum of NAPEs 0.182	<b>GM2</b> 0.026	<b>GM3</b> 0.095
<b>Control group</b> Time Oh Time 36h	NAPE 52:4 <0.001 <0.001	NAPE 54:4 0.182 0.500	NAPE 56:4 <0.001 <0.001	NAPE 58:6 <0.001 <0.001	Sum of NAPEs 0.182 0.500	<b>GM2</b> 0.026 0.070	<b>GM3</b> 0.095 0.126

**Table S13**: Values (mean ± SEM) of the relative abundance of the NAPE representatives, GM2 and GM3 of the 0, 36 and 144 time intervals.

	NAPE	NAPE	NAPE	NAPE	Sum of		
HII group	52:4	54:4	56:4	58:6	NAPEs	GM2	GM3
Time 0h	0.194±0.096	0.847±0.220	0.483±0.175	0.261±0.086	1.784±0.560	0.386±0.036	0.497±0.040
Time 36h	2.910±1.430	5.789±2.743	3.375±1.697	1.936±0.977	14.010±6.84 5	1.300±0.237	0.888±0.118
Time 144h	0	0.300±0.030	0	0	0.300±0.030	0.411±0.050	0.590±0.046
	NAPE	NAPE	NAPE	NAPE	Sum of		
Control group	52:4	54:4	56:4	58:6	NAPEs	GM2	GM3
Time 0h	0	0.334±0.020	0	0	0.334±0.020	0.328±0.026	0.433±0.030
Time 36h	0	0.403±0.028	0	0	0.403±0.028	0.350±0.037	0.459±0.034
Time 144h	0	0.307±0.034	0	0	0.307±0.034	0.367±0.033	0.478±0.044