

## **Supplementary information, Materials and Methods**

### **Generation of dsDNA targets**

Short dsDNA targets were prepared through annealing of two complementary oligonucleotides (see Table S2). Paired oligonucleotides (0.8 µM) were annealed in 1×PCR buffer (Transgen Biotech) in a total volume of 50 µL, and then subjected to the annealing program: initial denaturation at 95 °C for 5 min and then cool down from 95 °C to 20 °C with 1 °C decrease per min using a thermocycler.

### **Transcription of crRNAs**

First, the transcription templates were prepared through annealing of the synthesized oligonucleotides with T7-crRNA-F (Table S3), following the same procedures as previously described<sup>1-2</sup>. crRNAs were synthesised using the T7 High Yield Transcription Kit (ThermoFisher Scientific) and the reaction was performed at 37 °C overnight (about 16 h). RNA was purified using the RNA Clean & Concentrator<sup>TM-5</sup> (Zymo Research) and quantified with NanoDrop 2000C (ThermoFisher Scientific).

### **Purification of Cas12a proteins**

Ten Cas12a coding sequences were codon-optimised and synthesized by Tolo Biotech (Shanghai, China) and then cloned into pET28a (Novagen) with N-terminal 6×His tagging. Oligonucleotides for the construction of site-directed Cas12a mutants are listed in Table S4. For example, to obtain FnCas12a mutants, plasmid pET28a-FnCas12a was amplified by a pair of primers, the products of which were digested with DpnI (1 h, 37 °C) before being transformed into DH10B competent cells. The whole *Cas12a* gene was verified by DNA sequencing before being used for protein expression and purification. Other mutants were obtained following the same procedure.

Cas12a expression plasmids were transformed into *E. coli* BL21 (DE3) (Table S5). For protein expression, a single clone was first cultured overnight in 5-mL liquid LB tubes and then 1% (v/v) inoculated into 3 L of fresh liquid LB. Cells were grown with shaking at 220 rpm and 37 °C until the OD<sub>600</sub> reached 0.8, and IPTG was then added to a final concentration of 0.2 mM followed by further culture of the cells at 16 °C for about 16 h before the cell harvesting.

Cells were resuspended in 50 mL of lysis buffer [50 mM Tris-HCl (pH 8.0), 1.5 M NaCl, 1 mM DTT and 5% glycerol] with 1 mM PMSF as the protease inhibitor and lysed by high pressure. The obtained lysate was then centrifuged twice at 15,000 rpm for 30 min. After centrifuging, the supernatant was mixed with 5 mL of Ni-NTA beads (GE Healthcare) and softly shaken for 1 h at 4 °C before being loaded onto a 30-mL column. The packing was then washed with wash buffer (lysis buffer supplemented with 30 mM imidazole) and eluted with elution buffer (lysis buffer supplemented with 600 mM imidazole). The elution was dialysed with dialysis buffer 1 [50 mM Tris-HCl (pH 8.0), 200 mM NaCl, 1 mM DTT and 5% glycerol]. Before the protein solution was loaded onto an anion exchange column (HiTrapTM Q HP, GE Healthcare), it was diluted until the final NaCl concentration reached below 80 mM. After that, the column was washed and then eluted with a gradient concentration of NaCl (AKTA Pure, GE Healthcare). Fractions containing Cas12a proteins were verified by SDS-PAGE and then pooled for dialysis with dialysis buffer 2 [20 mM Tris-HCl (pH 8.0), 600 mM NaCl, 2 mM DTT, 0.2 mM EDTA] overnight. Finally, the protein was collected and diluted to a final concentration of 6 µM and mixed with an equal volume of 100% cold glycerol prior to being frozen at –80 °C.

#### ***In vitro* cleavage assay**

Unless mentioned otherwise, FnCas12a was used for the cleavage assays in this study. For the cleavage of target plasmid, the *in vitro* cleavage system contained 500 ng of target DNA, 250 nM Cas12a and 500 nM synthesised crRNA in a 20-µL reaction system. The reaction was performed at 37 °C in NEB buffer 3 for 1 h. For cleavage of M13mp18 by Cas12a, the reaction was performed at 37 °C in NEB buffer 3 for 30 min, employing 100 nM Cas12a, 250 nM synthesised crRNA, 1 µg M13mp18 DNA, 40 nM *trans* ssDNA and 10 U RNase inhibitor (Takara) in a 20-µL volume. For cleavage of M13mp18 by exonuclease T (NEB), the reaction was incubated at 25 °C for 30 min in NEB buffer 4.

Cleavage of FAM-labelled single-stranded oligonucleotides or double-stranded oligonucleotides was performed at 37 °C in NEB buffer 3 for 1 h, employing 250 nM Cas12a, 500 nM synthesised crRNA, 40 nM target DNA or/and 40 nM *trans* ssDNA and 10 U RNase inhibitor (Takara) in a 20-µL volume. Reactions were stopped by heating at 98 °C for 10 min, followed by immediately chilling on ice before further analysis through urea PAGE. For the time-course experiments in Supplementary information Figure S2a, the final

concentration of crRNA and Cas12a was reduced to 100 nM.

For LC-MS detection, cleavage of samples was performed in NEB buffer 3 at 37 °C for 2 h, containing 1 μM FnCas12a, 1 μM synthesised crRNA-T1, 1 μM target DNA (target-T1-18), 25 μM *trans* ssDNA (10T-FAM-5' or 10T-FAM-3') and 10 U RNase inhibitor.

#### **Analysis of Cas12a-digested products by denaturing urea PAGE or LC-MS**

To analyze the products by urea PAGE, FAM-labelled DNA was first digested by Cas12a and then heated at 98 °C for 10 min after the addition of loading buffer to stop the reactions. Heated samples were immediately chilled on ice, followed by being loaded on 10%–15% denaturing polyacrylamide gels containing 7 M urea. Electrophoresis was performed by running at 1800 V (about 40 V/cm) for about 70–90 min (using the Sequi-Gen GT Sequencing Cell system, Bio-Rad) or at 200 V (about 40 V/cm) for about 25 min (using the Mini-PROTEAN Tetra Cell system, Bio-Rad) in 1× TBE buffer. Gels were scanned using a FLA-9000 phosphoimager (FujiFilm Corporation, Japan).

For LC-MS analysis, FnCas12a-digested samples were first de-salted using a Novatia Oligo HTCS trap column (C18 300A, 1×10 mm), and were then analyzed with a Thermo LCQ ion trap with an ESI source (negative ion mode), where the electrospray gas was heated to about 300 °C. The liquid phase conditions are HFIPA-based aqueous reagents, and the system was operated with a flow of 40% ACN mixed with 40% MeOH and 20% water.

## Supplementary information, Table S1 Sequences of Cas12a proteins used in this study

Name	GI number	Species
FnCas12a	489130501	<i>Francisella tularensis</i>
AsCas12a	545612232	<i>Acidaminococcus</i> sp. BV3L6
LbCas12a	917059416	<i>Lachnospiraceae bacterium</i> ND2006
Lb5Cas12a	652820612	<i>Lachnospiraceae bacterium</i> NC2008
HkCas12a	491540987	<i>Helcococcus kunzii</i> ATCC 51366
OsCas12a	909652572	<i>Oribacterium</i> sp. NK2B42
TsCas12a	972924080	<i>Thiamicrospira</i> sp. XS5
BbCas12a	987324269	<i>Bacteroidales bacterium</i> KA00251
BoCas12a	496509559	<i>Bacteroidetes oral taxon</i> 274 str. F0058
Lb4Cas12a	769130406	<i>Lachnospiraceae bacterium</i> MC2017

**Supplementary information, Table S2 Oligonucleotides used for preparation of cleavage templates in this study**

**Supplementary information, Table S3 Oligonucleotides used for preparation of transcription templates in this study**

Oligo names	Sequences (5'-3')
T7-crRNA-F	GAAATTAATACGACTCACTATAGGG
T7-T1-24-R	gaattcgttagaaagtgcgataaATCTACAACAGTAGAAATTCCCTATAGT
	GAGTCGTATTAATTTC
T7-T1-8-R	tgcgataaATCTACAACAGTAGAAATTCCCTATAGTGAGTCGTATT
AATTTC	
T7-T1-10-R	gttgcgataaATCTACAACAGTAGAAATTCCCTATAGTGAGTCGT
ATTAATTTC	
T7-T1-12-R	aagtgcgataaATCTACAACAGTAGAAATTCCCTATAGTGAGTCGT
ATTAATTTC	
T7-T1-14-R	gaaagttgcgataaATCTACAACAGTAGAAATTCCCTATAGTGAGT
CGTATTAAATTTC	
T7-T1-15-R	agaaaagttgcgataaATCTACAACAGTAGAAATTCCCTATAGTGAGT
CGTATTAAATTTC	
T7-T1-16-R	tagaaagttgcgataaATCTACAACAGTAGAAATTCCCTATAGTGAG
TCGTATTAAATTTC	
T7-T1-17-R	gtagaaagttgcgataaATCTACAACAGTAGAAATTCCCTATAGTGAG
TCGTATTAAATTTC	
T7-T1-18-R	agtagaaagttgcgataaATCTACAACAGTAGAAATTCCCTATAGTGAG
GTCGTATTAAATTTC	
T7-T1-20-R	tcaagtagaaagttgcgataaATCTACAACAGTAGAAATTCCCTATAGTGAG
AGTCGTATTAAATTTC	
T7-T1-22-R	attcagtagaaagttgcgataaATCTACAACAGTAGAAATTCCCTATAGTGAG
GAGTCGTATTAAATTTC	
T7-T1-30-R	aagcttgaatttcgttagaaagtgcgataaATCTACAACAGTAGAAATTCCCT
ATAGTGAGTCGTATTAAATTTC	
T7-crRNA-DNMT-23nt-R	GAGTAACAGACATGGACCACATCAGATCTACAACAGTAGAAATT
	CCCTATAGTGAGTCGTATTAAATTTC
T7-crRNA-DNMT-(-8)-R	gacatggaccatcggaaacattATCTACAACAGTAGAAATTCCCTATAGTGAGTCGTATTAAATTTC
T7-crRNA-DNMT-(+4)-R	aggcgagtaacagacatggaccaATCTACAACAGTAGAAATTCCCTATAGTGAGTCGTATTAAATTTC
T7-crRNA-DNMT-(+8)-R	tgacaggcgagtaacagacatggATCTACAACAGTAGAAATTCCCTATAGTGAGTCGTATTAAATTTC
T7-crRNA-DNMT-16nt-R	agacatggaccatcgtAGTACAACAGTAGAAATTCCCTATAGTGAGT
	CGTATTAAATTTC
T7-crRNA-DNMT-18nt-R	acagacatggaccatcgtAGTACAACAGTAGAAATTCCCTATAGTGAGT
	GTCGTATTAAATTTC
T7-crRNA-DNMT-20nt-R	taacagacatggaccatcgtAGTACAACAGTAGAAATTCCCTATAGTGAGT
	AGTCGTATTAAATTTC
T7-T2-R	ctagagtaaaggctgttgtcgttagtaATCTACAACAGTAGAAATTCCCTATAGTGAGTCGTATTAAATTTC
T7-T3-R	ggatccactgtctgttgtcgatAGTACAACAGTAGAAATTCCCTATAGTGAGTCGTATTAAATTTC
T7-T4-R	ttcaaggagaaactgcgactgttgtcgatAGTACAACAGTAGAAATTCCCTATAGTGAGTCGTATTAAATTTC

**Supplementary information, Table S4 Oligonucleotides used for plasmids construction in this study**

Oligo names	Sequences (5'-3')
FnCas12a-K869A-F	gctgtctgcgtcgatcgatgtactcgaaacacg
FnCas12a-K869A-R	atctgatcgccgacaaaggcggttaccggaggataag
FnCas12a-K852A-F	tgccttatttgcgtggcgatggcctccttggc
FnCas12a-K852A-R	atcgccaacgcaaataaggacaatctaagaagg
FnCas12a-H843A-F	ttggctgggtgtgtatctttagggatggac
FnCas12a-H843A-R	agaagatcacagcaccagccaaggaggccatcg

FnCas12a-R1218A-F	ggcttgcttggccatctgcaggattgtttc
FnCas12a-R1218A-R	tcctgcagatggccaacagaacaaggccccacagagc
FnCas12a-E1006A-F	agttcagatctgcgaacaccacgatggcatttgc
FnCas12a-E1006A-R	tctgttgttcgcagatctgaacttcggcttaagatcg
FnCas12a-D917A-F	tctcgccccgtgcgatctcaggatgtgcacatcg
FnCas12a-D917A-R	tcctgagcatgcacgggggagagacacctgg
FnCas12a-D1255A-F	gcccgttgctgcggcatctgaggatattcttgg
FnCas12a-D1255A-R	ctcaggatgccgcagccaacgggcctatcacatggcctg

**Supplementary information, Table S5 Plasmids and strains used in this study**

Plasmids or Strains	Relevant properties or genotypes	Sources
<b>Plasmids</b>		
pET28a-TEV	pET28a with the thrombin cleavage site changed to the TEV protease cleavage site	<sup>3</sup>
pET28a-TEV-FnCas12a	pET28a-TEV carrying FnCas12a	<sup>1</sup>
pET28a-TEV-AsCas12a	pET28a-TEV carrying AsCas12a	<sup>2</sup>
pET28a-TEV-LbCas12a	pET28a-TEV carrying LbCas12a	<sup>2</sup>
pET28a-TEV-Lb5Cas12a	pET28a-TEV carrying Lb5Cas12a	This study
pET28a-TEV-HkCas12a	pET28a-TEV carrying HkCas12a	This study
pET28a-TEV-OsCas12a	pET28a-TEV carrying OsCas12a	This study
pET28a-TEV-TsCas12a	pET28a-TEV carrying TsCas12a	This study
pET28a-TEV-BbCas12a	pET28a-TEV carrying BbCas12a	This study
pET28a-TEV-BoCas12a	pET28a-TEV carrying BoCas12a	This study
pET28a-TEV-Lb4Cas12a	pET28a-TEV carrying Lb4Cas12a	This study
pET28a-TEV-FnCas12a-K869A	pET28a-TEV carrying FnCas12a-K869A	This study
pET28a-TEV-FnCas12a-K852A	pET28a-TEV carrying FnCas12a-K852A	This study
pET28a-TEV-FnCas12a-H843A	pET28a-TEV carrying FnCas12a-H843A	This study
pET28a-TEV-FnCas12a-R1218A	pET28a-TEV carrying FnCas12a-R1218A	This study
pET28a-TEV-FnCas12a-E1006A	pET28a-TEV carrying FnCas12a-E1006A	This study
pET28a-TEV-FnCas12a-D917A	pET28a-TEV carrying FnCas12a-D917A	This study
pET28a-TEV-FnCas12a-D1255A	pET28a-TEV carrying FnCas12a-D1255A	This study
M13mp18 single-stranded DNA		NEB
pCB1A2_2		<sup>2</sup>
<b>Strains</b>		
<i>E. coli</i> DH10B	F <sup>-</sup> endA1 deoR <sup>+</sup> recA1 galE15 galK16 nupG rpsL Δ(lac)X74 φ80lacZΔM15 araD139 Δ(ara,leu)7697 mcrA Δ(mrr-hsdRMS-mcrBC) Str <sup>R</sup> λ <sup>-</sup>	Invitrogen
<i>E. coli</i> BL21(DE3)	F <sup>-</sup> ompT gal dcm lon hsdSB(rB- mB-) λ(DE3 [lacI lacUV5-T7 gene 1 indI sam7 nin5])	Invitrogen

## **Supplementary information, nucleotide sequences used in this study**

>FnCas12a (codon-optimised DNA sequence)

ataggcatctatcaggagttcgtaataagtacagccgtcaagaccctgcgggtagctgatccccaggcaagacactggagaacatcaaggccagggg  
cctgatcctggacgatgagaagcgcgccaaggactataagaaggcaagcagatcatcgataagtaccaccagtttatcgaggagatcctgagcagcgtgt  
catctctgaggatctgctcagaattacagcgtgttcaagactgaagaactgtcgatgacaacctcgagaaggactcaagagcgcaggacaccatc  
aagaagcagatcagcgtatataaggactccgagaagttcaaccagaatctgatcgatgcaagaaggccaggagtcgcacctgatcctgt  
ggctgaagcagcttaaggacaatggcatcgagctgtcaaggccaactctgatcaccgatatcgacgaggccctggagatcatcaagagcttaaggctgga  
ccacatacttaaggcctcacgagaacaggaaacctgtacagcagcaacgacatccctacaaggcatcatctaccgcacgtggatgacaatctgc当地  
cctggagaacaaggcaagttatgagtcctgaaggacaaggccccgaggccatcaattacgagcagatcaagaaggatctggccgaggagtcacccat  
atcgactataagacatccgagggtgaaccaggcgggtttctctggacgagggtttgagatcgcaattcaacaattacctgatcgaccatccggcatccaagtt  
aatacaatcatcgccggcaagttgtgaacggcagaatccaagagaaaggcatcaacgagatcatcaatctgtatagccagcagatcaacgacaagacc  
gaagaagtacaagatgagcgtgtcaagcagatccgtccgatacagactaagagcttgatcgataagctggaggatgactctgacgtgggacc  
atgcagagctttatgagcagatcgccctcaagaccgtggaggagaagtcataaggagacactgagccgtctgatgacctaaggccagaagctg  
gacctgtctaagatctactcaagaacgataagtcctgaccgcacccgtctcagcaggtttgatgactatagcgtgatcgccaccgcgtctggagatcatc  
cagcagatcgcggcaagaacctggataatccctctaagaaggaggcaggagctgatcgccagaagaccgagaaggccagatctgacccggag  
agetggccctggaggagttcaataagcaccggatatcgacaaggcagttgaggagatctggcaacttcgcgeccatccatgatcttgatgag  
cgeccagaaacaaggacaatctggccagatctccatcaagtaccagaaccaggcagaaggaccctgctgceggcctctggaggatgacgtgaaggcc  
caaggatctgacccggagacaacaatctgctgacaagacttccacatctccagttgaggataaggccaatatctggataaggacgacactt  
atctgggttgcaggagttacttcgagctggcaacatctgcccgttacaacaagatcagaaattatatcacacagaaggctactccgacgagaagtt  
tgaacttcgagaacagcaccctggcaacggctgggataagaataaggagcctgacaacacagccatctgtcatcaaggatgacaagttactatctgg  
tgaataagaacaataagatctcgatgacaaggccatcaaggagaacaaggccgaggctacaagaagatcgatgatgatgactctgcccggccaata  
gatgctgcctaagggtttccgccaagtcataaggcttccatcaaccatccgaggacatctgcggatcagaatctccaccacacaagaacggcttcc  
ccagaaggctatgagaagttgagttcaatatcgaggattgccggaaagttatcgacttctacaaggcagacatctccaggccctgagttgaaagg  
caggttttagcgcaccccgccgtacaactccatcgacgaggcttctacagagaggtggagaatcagggtataagctgacatttggataacatct  
gacagcgtggtaatcaggcaagctgtacctgtccagatctataacaaggacttcagcgcctattccaaggccggcaacccctgtactgg  
gcccgttgcagagagaaatctgcaggacgtgggtataagctgatcgacggccgaggcttacaggaaggactgatccatccataagaagat  
cccaaggaggccatcgccacaagaataaggacaatccataagaaggagagcgtttcgagatctgatcaaggacaaggccgttaccggaggata  
ttccactgtccatcacaatcaactcaagtcctctggcccaacaagttatgacgagatcatctgctgtcaaggagaaggccacatgtgc  
catcgaccggggcgagagacaccctggctactataccctgggtggatggcaaggcaatcatcaaggcaggatccatcaacatcatcg  
agacaaactaccacgataagctggcccatcgagaaggatagggactccgcccgaaggactggaaagaagatcaacaatatcaagg  
gatgacggggct  
atctgtctcagggtggtcacgagatcgccaaagctgtcatcgactacaatgcctatcgatgggttcaaggatctgatcgatgact  
gagaaggcaggtgtatcagaagctggagaagatgtcgatcgagaagactgatgatggatcggatcttgcgatcggatcttgc  
ataccagctgaccggccctttagacatcaagaagatggcaaggcagacaggcatctactatgtgc  
ggcttgcgatcggatcttgcgatcggatcttgcgatcggatcttgcgatcggatcttgcgatcggatcttgcgatcggatcttgcgatc  
ttccctcgattataagaacttggcgacaaggcccaaggcaagtggaccatcgccctttcgccagccgctgatcaactttagaaattccgata  
aaccaca  
attgggacccggggatgtfacccaacaaaggagctggagaagctgtcgatcgaggactacagcatcgatgtggccacggcagatgc  
gtggcgagagcataagaagtttcgccaagctgacccgtctgatcaataatccctcgatcgatgcggaaacagcaagaccggc  
cagagactggactacccatctgc  
ccccgtggccatgtgaacggcaacttctcgacagcagacaggcccccaagaatatgcctcaggatgccc  
ggcctgtatcgatcggcaggatcaagaacaatcaggaggcaagaagactgatgacccatctggatc  
aattga

>AsCas12a (codon-optimised DNA sequence)

>LbCas12a (codon-optimised DNA sequence)

atgctgaagaacgtggcattcgacagactggatgtggagaaggcagaagaacatgagcaagctggagaagttccacaactgtcacagcctgagaacc  
ctgagattcaaggccatccccgtggaaaaacccaggagaacatcgacaacaagagactgtggtgaggacgaaaagagagccgaggactacaagggcgt

>Lb5Cas12a (codon-optimised DNA sequence)

atggagaactactacgcacagecgtgaccagacaatacccggtgaccaagaaggccatcagacaggagctgaagcccgaaaacctggagaacatcaagac  
gccgagatcatcgagggcacaaggcagaagaaggaggctacgtgaaggtgaaggagctgtggacgagttccacaagagcatcatcgagaagagcctggt  
ggccattaaactggacggcctgagcgaatttggagaagctgttacaagaatcaagaccacccgacgaggacaagaacagaatcagcggactgttctactatcg  
gaaaggcagatcgccgacgcctgaagaacagcagagactacggctacgtggacaacaaggacctgtcgagaagatcctgcccggagagactgtgaaggacgag  
aacagccctgaacgcctgagctgtcaaggcttaccacacttccaccgactactacaagaacaggagaacatctacagcgcacgaggagaagcatagcac

>HkCas12a (codon-optimised DNA sequence)

>OsCas12a (codon-optimised DNA sequence)

atggagaccgagatcctgaagtacgacttgcagagagaggcaagtacatgtactacgcacggcctgaccaaacaatacgccctgagcaagaccatcaga  
aa  
cgagctggccatcgaaaaaccctggacaacatcaagaagaacagaatcctggaggccgacatcaagagaagagcgcactacgagcacgtgaagaact  
gatggacatgtaccacaagaagatcatcaacgaggccctggacaactcaagctgagcgtgtggaaagatgccgcccacatctacttaacaac  
a  
agagagacatcgacgccttcgtaaagatccaggacaagactgagaaaggagatcgtggagcagcgtgaagggacacaccgactacagcaagg  
tggccaacaag  
gactttctggcctgctgaaagctgctgtagcaccgaggaagacagaatcctgatcgagagctcgtacaacttctacaccacttcacc  
agctacaacaaggtgagaa  
gcaacctgtacagcgcgaggataagagcagcaccgtggctacagactgatcaacgagaacctgccaagttctcgacaacatcaagg  
cctacagaaccgt  
agaaacgcggagtgtacagcggagacatgagcatcgtggagcaggcagactgtttgagggtggacacccctgacc  
cctgac  
cataaccacatgatcgccagctgaacagcgcacatcaccgtacaacc  
a  
cagaatgatcgtggccggcagttaaaagctgccaagatgaagg  
gact  
tacaaggcagctgctgaccgagagagaggaggatcatcgaggag  
tac  
acccgacgcacgagggtgtctgattacc  
cgcgtacaactac  
tgcgtgac  
actacctgaacagcgcacaaggfggagagcttctcgacacc  
ctgagaa  
a  
agacgcacggcaaggagggtgttcatcaagaac  
ac  
gcgtgagcaagacc  
c  
accatg  
caacatctgttcgacaacttggagcaccatcgacgcac  
ctgatcaacc  
ac  
cgactgac  
c  
gagtc  
ac  
gcac  
ct  
cc  
gaga  
act  
gt  
ga  
aga  
a  
gac  
ca  
gg  
ac  
c  
acc  
at  
tc

>TsCas12a (codon-optimised DNA sequence)

atgacccaagacccctcgacagcgagttctcaacctgtacagcctgcagaagaccgtgagattcgagctaaaccctggggagaaacagacttagctcgaggagac  
ttcaagaacgagggcctgaagagagtggtgagcgaggacgaaagaagagccgtggactaccagaaggatcatcgacgactaccacagagactt  
atcgaggagagcctgaactactccccgagcaggtgagcaaagacgetctggagcaggccttacactgtaccagaagetgaaggccctaagggtggaggaaa  
gagagaaggccctgaaggatggaaagccctgcagaagaagctgagagagaaggggtgtgaagtgttcagcgcacagcaacaaggccagattcagcagaatc  
gacaagaaggagctgatcaaggaggacctgatcaactggctggccagaatagagaggacgcacatccctaccgtggagaccccaacaacttaccaccta  
cttcaccggctccacgagaacagaaacatctacagcaaggacgaccacgcctaccgcacatcagcttgcactgtatccacgagaacctggcccaagtcttgc  
caacgtgatcagctcaacaagctgaaggagggttcccggactgtaagttcagacaaggtaaggaggactggaggtggactacatctgaagcgcctcg  
agatcgagacttcgtgacttcgtgaccgcaggccggaaatcgaccactacaactacccctggccggaaacactggaggatggcacaagaagcaggcat  
gaacgagcagatcaacctgttcaagcagcagcagcagaccagagacaaggccagacagatccccaaactgtatccctgttcaagcagatctgagcagagaacc  
gagagccagagcttcatccccaaagcagttcgagagcgcaccaggagcttgcacgcctgcagaagctgcacaacaactgccaggacaagttcacccgtgtca  
gcaggctattctggactggctgaggctgatctgaagaagggtgttcatcagaaccgcgcacccgttgcacgcctgcagaacaccatctggcaactacgcgttgc  
agcgcacgcctgaacctgtacaaggagagcctgaagaccaagaagcccgaggaggccctcgaaaaactgcccgcctatgcacccatgcacccgttgc  
tggagcaggatcaacagcagcgcggacgtgaaaagcagcagcagacacccgtgtgaacttcatcaagaccgacgagctgtacagcagattcatcaag  
agcaccagcggaggcttccaccaagtgcacccctgtttgagctggaaagccctgagcagcagaaaagaagaccccccggagagcgaagatgaaggccctaagg  
caagaaggctcgagcagatcaagagaatcaaggcctacccgtggacacacttatggaggccgtgcactttgtcaagccctgtatctggfagggcagaagatg

atcgaggggcctggacaaggaccagagttctacgaggcctcgagatggcctaccaggaaactggagagcctgtatccccatctacaacaaggccagaagcta  
cctgagcagaaagccctcaaggccgacaagttcaagatcaacttcgacaacaacccctgtctggatggacccaacaaggaaacgcacgcggccagg  
atcctgttcaagaaggacggcctgtactacctggcatcatgcccaaggcaagacccctgtcgactacttcgtgagcagcggaggacagcgagaagctgaag  
cagagaagacagaagaccgcccggaggaagcttgctcaagacggcgaagacttgcgagaagatcagatacagactgtgtccggcgcttagaaaaatgtcg  
ccaagggtttcagcaacaagaacatcggttctacaaccccagcgcacgcacatcctgagaatcagaaaacaccgcacacccaaatggcaccccccag  
aagggacatagcaagggtggatcaacctgaacgactgcccacaagatgatcgactttcaagagcagcatccagaacgcaccccgaaatgggaagctcggtt  
cacattcagcgcacaccaggcacttcgaagacatgagcgcctctacagagaggtggagaaccagggttacgtgtacgatcagttcgacaagatcaaggagacactaca  
tccagagccagggtggagcaggaaacctgttccatctacaacaaggacttcgacccctactctaaggcagccaaatgtgcacaccctgtactgg  
aggccctgttcaagaggccaaacctgaataacgtggggcaagctgttccatctacaacaaggacttcgacccctactctaaggcagccaaatgtgcacaccctgtactgg  
gggtgcacctgtccaatcaagccatcgacaacaagaaccccccacaccggagaaaacccagagcaccttcgagtacgcacctgttgaaggacaagagatcaccca  
ggacaagtttctccactgtggccatcagctgaacttaaggcccagggttgcgatcaacgcacaagggttacgggttccatcggaaaccccgacgt  
gaacatcatcgccatcgacagaggcgaagacaccctgttgcacttcaccgttgcgatcaaggcagatccctgttgcaggagacccgttgcacaccctgtatgt  
gctgacaaggccacgttacgcactaccaggcagaagctggacaagaaggagcagggaaagacgcggccagaaaatctggaccaccgttgcgatcaa  
ggagctgaaggaggctatctggccacgttgcataagctggccacccgtatcatcaagtacaacgcacgttgcgttgcctggaggacccgttgcacttc  
gagaggcagattcaagggtggagaagcaggttgcacttgcgagaaggccctgtatcgacaagctgttgcacttgcgatccctgttgcaggagaaggagctggcga  
atgtggacatttctggccatccaacttgcacgcactcccttcgaggttgcacttgcgatccctgttgcaggagaaggctgttgcacttgcgatccctgttgc  
tctaagatcgaccccaacccggcttcgttgcacttgcgatccctgttgcaggataccaggcgttgcaggagaaggctgttgcacttgcgatccctgttgc  
aacagcgttgcagaactacttcgatccgttgcacttgcgatccctgttgcaggataccaggcgttgcaggataccaggcgttgcacttgcgatccctgttgc  
acgttgcaggataccaggcgttgcacttgcgatccctgttgcaggataccaggcgttgcaggataccaggcgttgcaggataccaggcgttgcacttgcgatcc  
acaaccaccgttgcacttgcgatccctgttgcaggataccaggcgttgcaggataccaggcgttgcaggataccaggcgttgcaggataccaggcgttgc  
gaccatgaccctgttgcaggataccaggcgttgcaggataccaggcgttgcaggataccaggcgttgcaggataccaggcgttgcaggataccaggcgttgc  
agaagtgtggccaaagatgtggccatgttgcacttgcgatccctgttgcaggataccaggcgttgcaggataccaggcgttgcaggataccaggcgttgc  
ctgaacccgttgcacttgcgatccctgttgcaggataccaggcgttgcaggataccaggcgttgcaggataccaggcgttgcaggataccaggcgttgc

>BbCas12a (codon-optimised DNA sequence)

atgaagaagtccacaaccgtaccccgtagcagaagaccctgagattcggacttaccccaaggcataacctctaagcacctgtcaagatcatccaggaggac  
gaggcagatcgtgaggatagccaggaggtaagaagctgctggacagataccacaaggagttcatgccatgcctgagcagcttcataagccccctggc  
caaagagatcatcccaagctgaaggagtcccccagatcagagctacaggcagccaaaggcataatgcgtacggactgagatccatcaggacgagctgagagatcttggtg  
aaggcgttaagggagagggcgagcaggagagaagatacaagatccgtatggcgtaagggcaaccctaatgcgtacgagctgttcaacaccggagctgtatca  
acttcctgaaggaccctgtgaacaggctcttgtaagaagttccagaagcacaccggctacttcctggcttcaacgagaacagaaacatgtacagcgcca  
aggctcagagcacagccattgcctacagactgtatccacgagaacccgtatccctggacaacatcaccacccatcggagaaggtaagacactacccgttcaaggag  
gagatccccagctggagaaggaactggtagagccggagctctctggtagccatgtggacagcgtgttaccatcgacttcttcctggagggttccatccaaa  
gcggcatcgaccaatacaacgcctgtatggcaagatcgtgaaccacagcaggccggaggtaaaaggcctgaacgagagaatcaacccgttacaaccaggc  
acaaggcaggaggctaagctgcctgttcaagccctgtacaagcagatcctgagcagacagagcaacttagctggctggccgaagcttacaacaggcacaag  
gaccctgtggacagcatccagaagtactaccagctgtatcgacaccagatctcgagagaatccctggactgtgcacacactggagaaagccccctgg  
caagatctggatcacctacgacaccctgaccagcatcagaacaccctgtacggcagctggagactgttggagaggccctggcagaatgccttacaag  
gagaaggagagaaagagcagccagaagaaggccctgaactacacgcctggagagcatcaatcaggccatgccttacaaggccatgcctggcagcatgg  
atccagaagtacttcatccctggagaagcaaccctgaccaagaagacgcgtacccggcactctggataaggtgagaagcgttacaaggccctgg  
gacatccattacaaccctgaccataccggcaagaagctgtatccaggacaagaaggcaggctggacactgttggacaccctgtatccctgcag  
attcatcaagccctgtgtacagcaacaacgcgagaacgagaccacaaggacgaggcttacaccggactgttgcaccatcgacccatcatgg  
ggccctgtacaacaaggtaagaaactaccgtaccagaageccctacagcaccgagaagttcaagatcaactcaagagcagcagccctgtggatgg  
agaaacaaggagaaggacaaccctggccgtatccatgttcaagagagagagagaacttacccatcatggacaaggctcacaacgcacccttcaagaacaag

agcctgcccacacaaggagagtctacgagaagatggagtacaagctgcccggccaaataaatgtgcccaggtgtacatcaccagcaagaaggc  
atcgagagttccatcccagcgaagagctgcagaagaagtacaagctggccacccacaagaaggagccagcttcaacctgagcgacatgagagccctgatc  
gactacttcaaggagagcctggagaagcatgaggagcacagccaattcggttccacttcagcgacaccagcacctacgaagacatcagcggcttacagaga  
ggtgagcagcaggcctacaagatcacccctcagaaggtgagctggaggtacatcgaccagctgtgaacgggcaagctgtacccatgttccagatctacaaca  
aggacttcagccctacagaaggcaccctaatctgcacaccctgtacttgaagatgttgcacccgcaatctgcaggacatgttacaagctgaac  
gagaggccgagggttctcagaagaaagagcctgcagtacgacagacccacacccctaagggcaaccatcaacaagaagagcctgtgaac  
agaccagcctttgactacgacactgtatcaaggacagaagattcaccgtggacaagtccagttccacgtgcccacatcaccatgaacttcaaggccaccaggc  
ccaaagtgaaccagatggcaggaggaggtgaagaagagaaggcgttccacctgtatccacgttgcgaatcgacagaggcggagaaaacctgttacatcgfttgc  
caacgagagaggcggagatcatcgagcgttgcagcttgcacaagatctgttgcacccatcaccaggagaaggacacccgttgcactataaggccctgttgc  
gagaagccagagcagacttggaggagagaagagactggcagaccatcgagaacatcaaggagcttgcaggggcggttccatcagaaccagaaagaagttc  
gagttcagctgttgcacccatcaccaggagaacactgttgcacccatcaccaggagaacactgttgcacccatcaccaggagaac  
gaagaagctgatcgacaagctggcttgcgttgcacccatcaccaggagaacactgttgcacccatcaccaggagaacactgttgcacccatcaccaggaga  
gcttgcacccatcaccaggagaacactgttgcacccatcaccaggagaacactgttgcacccatcaccaggagaacactgttgcacccatcaccaggaga  
atcgagagcatcgcaagaccaggactgtatcgacccatcaccaggagaacactgttgcacccatcaccaggagaacactgttgcacccatcaccaggaga  
ttcggcaacagagactaaggcagcagaagcaagtggagactgttgcacccatcaccaggagaacactgttgcacccatcaccaggagaacactgttgcacccatcaccaggaga  
aaagcgacagcatgggttctgaccggaggccttcaaggacgttgcacccatcaccaggagaacactgttgcacccatcaccaggagaacactgttgcacccatcaccaggaga  
gacaaggcccttctcgacttgcacccatcaccaggagaacactgttgcacccatcaccaggagaacactgttgcacccatcaccaggagaacactgttgcacccatcaccaggaga  
gatgaaaacggcgagtttcgcacccatcaccaggagaacactgttgcacccatcaccaggagaacactgttgcacccatcaccaggagaacactgttgcacccatcaccaggaga  
tgctggacaagatcagaagaaagaccgagaaggtaactggcttgcacccatcaccaggagaacactgttgcacccatcaccaggagaacactgttgcacccatcaccaggaga  
tga

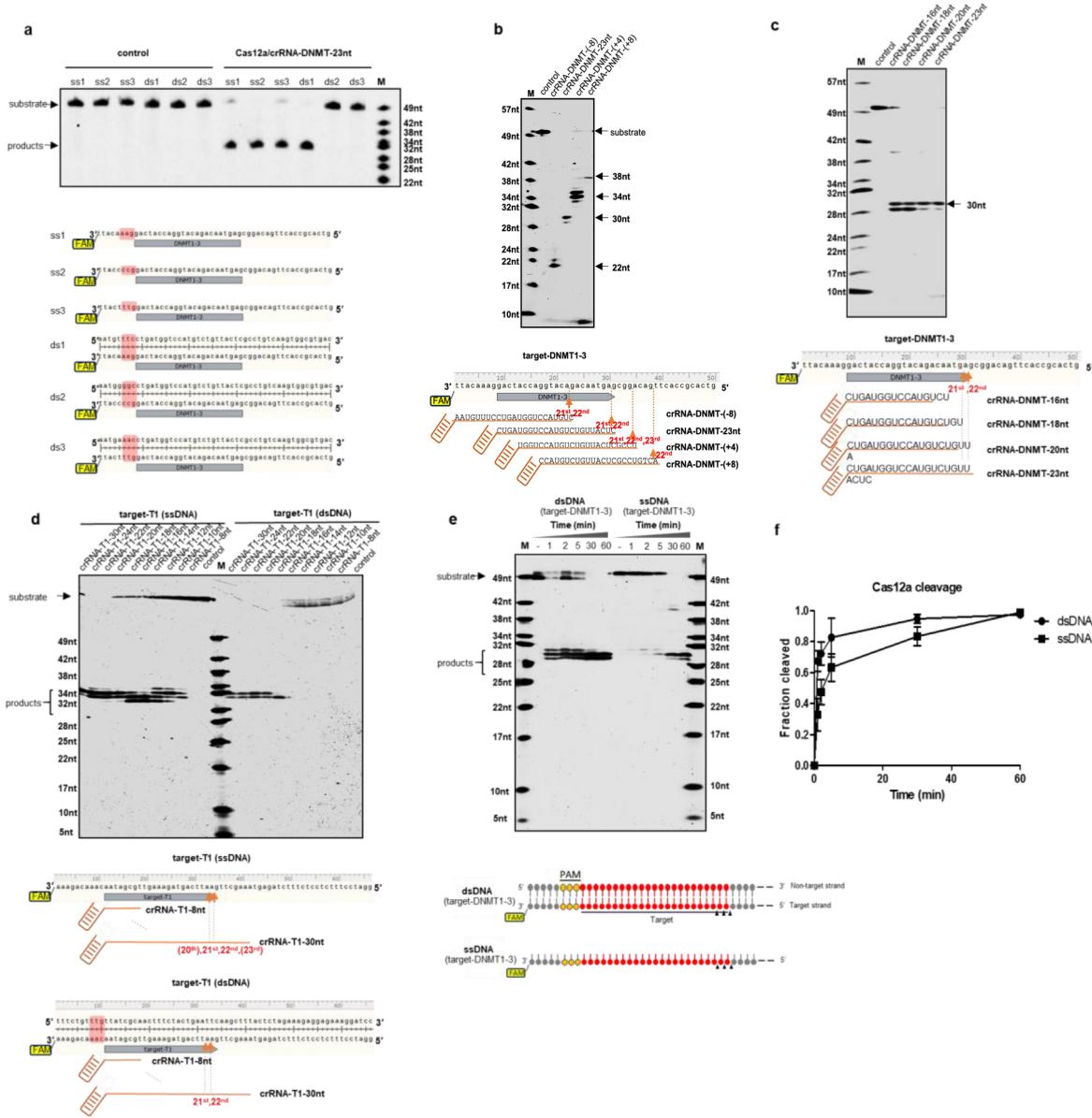
>BoCas12a (codon-optimised DNA sequence)

atagaggaagtcaacgagttcgccccctgttccatcgaaagacccttgcgttccatcgaaagatgttgcacccatcaccaggagaacactgttgcacccatcaccaggaga  
gcttgcgttccatcgaaagacccttgcgttccatcgaaagatgttgcacccatcaccaggagaacactgttgcacccatcaccaggagaacactgttgcacccatcaccaggaga  
atttgcacccatcaccaggagaacactgttgcacccatcaccaggagaacactgttgcacccatcaccaggagaacactgttgcacccatcaccaggagaacactgttgcacccatcaccaggaga  
accatccaaacactgttccatcgaaagacccttgcgttccatcgaaagatgttgcacccatcaccaggagaacactgttgcacccatcaccaggagaacactgttgcacccatcaccaggaga  
gacttgcacccatcaccaggagaacactgttgcacccatcaccaggagaacactgttgcacccatcaccaggagaacactgttgcacccatcaccaggagaacactgttgcacccatcaccaggaga  
ggaggagaagacccttgcgttccatcgaaagacccttgcgttccatcgaaagatgttgcacccatcaccaggagaacactgttgcacccatcaccaggagaacactgttgcacccatcaccaggaga  
ccgaaaaatcaacgcctgtacggacttgcacccatcaccaggagaacactgttgcacccatcaccaggagaacactgttgcacccatcaccaggagaacactgttgcacccatcaccaggaga  
aagcagatcgacccatcaccaggagaacactgttgcacccatcaccaggagaacactgttgcacccatcaccaggagaacactgttgcacccatcaccaggagaacactgttgcacccatcaccaggaga  
cagcagaccgacagaagcaacactgttgcacccatcaccaggagaacactgttgcacccatcaccaggagaacactgttgcacccatcaccaggagaacactgttgcacccatcaccaggaga  
gcgacaagaacactgttgcacccatcaccaggagaacactgttgcacccatcaccaggagaacactgttgcacccatcaccaggagaacactgttgcacccatcaccaggagaacactgttgcacccatcaccaggaga  
ctacgacccatcaccaggagaacactgttgcacccatcaccaggagaacactgttgcacccatcaccaggagaacactgttgcacccatcaccaggagaacactgttgcacccatcaccaggaga  
gacttgcacccatcaccaggagaacactgttgcacccatcaccaggagaacactgttgcacccatcaccaggagaacactgttgcacccatcaccaggagaacactgttgcacccatcaccaggaga  
cgacgttgcacccatcaccaggagaacactgttgcacccatcaccaggagaacactgttgcacccatcaccaggagaacactgttgcacccatcaccaggagaacactgttgcacccatcaccaggaga  
gataccaccatcaccaggagaacactgttgcacccatcaccaggagaacactgttgcacccatcaccaggagaacactgttgcacccatcaccaggagaacactgttgcacccatcaccaggaga  
aggcacaacactgttgcacccatcaccaggagaacactgttgcacccatcaccaggagaacactgttgcacccatcaccaggagaacactgttgcacccatcaccaggagaacactgttgcacccatcaccaggaga  
gctgttgcacccatcaccaggagaacactgttgcacccatcaccaggagaacactgttgcacccatcaccaggagaacactgttgcacccatcaccaggagaacactgttgcacccatcaccaggaga  
agaagatcagaactgttgcacccatcaccaggagaacactgttgcacccatcaccaggagaacactgttgcacccatcaccaggagaacactgttgcacccatcaccaggagaacactgttgcacccatcaccaggaga  
gctgttgcacccatcaccaggagaacactgttgcacccatcaccaggagaacactgttgcacccatcaccaggagaacactgttgcacccatcaccaggagaacactgttgcacccatcaccaggaga  
tacccatcaccaggagaacactgttgcacccatcaccaggagaacactgttgcacccatcaccaggagaacactgttgcacccatcaccaggagaacactgttgcacccatcaccaggagaacactgttgcacccatcaccaggaga  
ggcccaataaaatgttgcacccatcaccaggagaacactgttgcacccatcaccaggagaacactgttgcacccatcaccaggagaacactgttgcacccatcaccaggagaacactgttgcacccatcaccaggaga  
gaaggggagccggatttgcacccatcaccaggagaacactgttgcacccatcaccaggagaacactgttgcacccatcaccaggagaacactgttgcacccatcaccaggagaacactgttgcacccatcaccaggaga

cgacaccaggcacctaccaggacatcagcgaggcttacagagaggtggaggcagcagggttacaagaatgagcttcagaaagggtggactacatcaagag  
cctgggtggaggaggaaagctgtacctgttccagatctacaacaaggacttcagcgcctactctaaggaaacacccaacatgcacaccctgtactggagatgt  
gttcgacgaggagaacctgaaggacgtggtgtacaagctgaacggagaggccgagggtttcagaaagagcagcatcaccgtcagagccctacacatct  
gccaacagccccatcaagaacaagaaccagaagaaggagagcaagtgcagtgatcaaggacagaagatacaccgtggacaagttc  
ctgttccacgtgcccattaccatgaacttcaagagcgtggccggcagcaacattaaccagctggtaagagacacatcagaagcgcaccgacattc  
ggcatcgacagaggcgaagacacctgtgttccacgttccatcaaggacatcaggcagttcagcgcaccatccagaacatcag  
acggcaacacccatcagaaccgactaccacgagctgtggacacaagagagggcgaaagaaccggaggccagaagaaaactggcagaccatccagaacatcag  
agagctgaaggaggctatctgagccagggtatccacaaagatcagcggcactcaaggatcagcgcaccatccagaacatcag  
gagaagcagacagaaggtggagaagcaggtgtaccagaagttcggagaagatgctgatcacaagctgatccatctggcttcaacttccatc  
aacaggaggactgctgagacgcctatcaactgaccggcagttcggagactttaagaccctggcaagcaaagcggcatctgttctacgtgcccgttca  
ccagcaagatcgtccctgtgaccggcttgcacaccactacgagaacatcgagaaggccaaagggttctcgacaagttcaagacgcata  
caacagcgacaaggactgttgcagttcgtggacgactacaccagattcagcccaaggccgaaggaaccagaagagactggaccatctgcaccagg  
caagagaatccagatctgcagaaaccaccagagaaacaacgagttggaggggcaagaaatcgaccctgaccacggccatcaaggacacttcgagg  
cgtggacatcagcaaggacactgagagagcagatcaacacccagaacaagaaggagtttgcaggagctgctgagactctgagactgaccctgc  
aagatgaaacagcatgcccgatcgactacctgtatcagccctgtggctatgcacacaggcttctcgacagcagaagcggccagactgaagg  
ccgtgtccatgtatgcgatgccaacggagttacaacatcgccagaaggccatcggccatcggatcagacggagaacgcacagcgcca  
aaatcagectggccatcagcaacaaggagtggctgaagttcgtcagaccaagccctatctggaggactgt

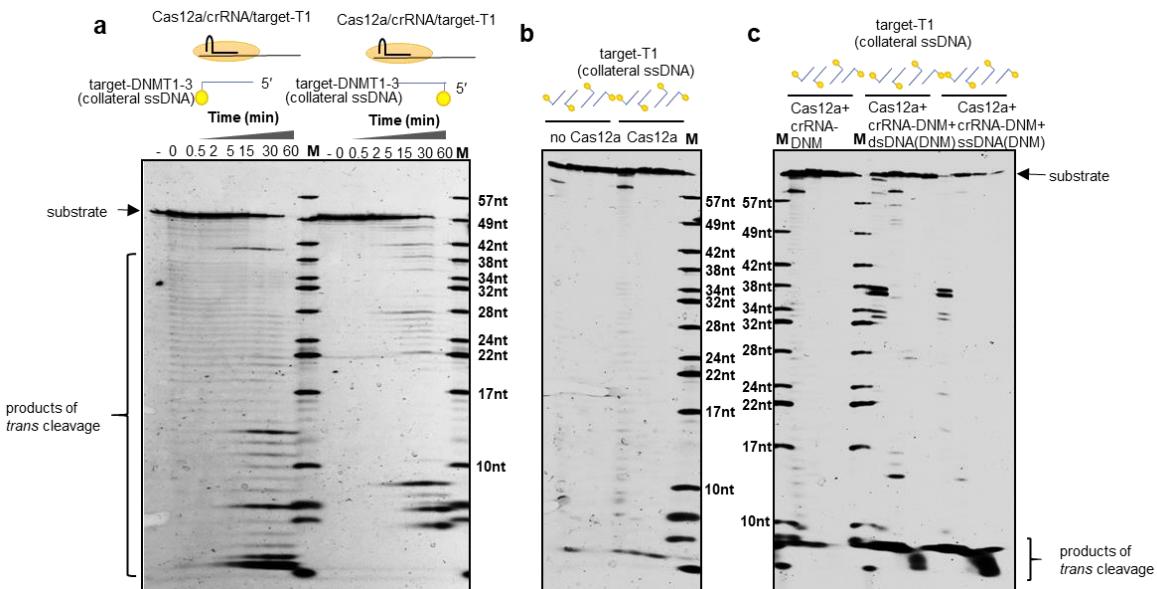
>Lb4Cas12a (codon-optimised DNA sequence)

aagatacacccgtggacaagttccagttcacgtgcccctgaagatgaactcaaggccgacgagaagaatcaacgacgtatcgaggccatcaga  
agcaacaaggcatccacgtatcgaaatcgacagaggcgagagaaacctgttacctgatcaacgaggaggcagaatcatcgacgagaagc  
ctgaacatcatcgacagcggcgaaggccataccagaactacagagacactgtggacagcagagagaaggacagagagaaggccagagagaactggcagg  
agatccaggagatcaaggacctaagaccggatctgagccaagccatccacaccatccaagtggatgaaggagtacaacgcctatcgatcgagg  
cctgaacgacagattccaacccggcagaagaaggatggagaaggcaggttaccagaagttcgagaagatgtatcgacaaactactatcgacaaag  
gacgaggagttcgacagaatggggcggcacacatagactctgcaactgaccgagaagttcgagagcttcagaagctggagacagaccggatcttca  
tgtcccccttggaaacacatctaagctggatcccacaaccggattctgtggacactgttacccaaacttacaagagcgtggacgccaccaaggacttcatcaagaa  
gttcgacttcatcgattcaacacgacgagaactacttcgagttccggctgcactacagcaacttcaccggatcgacatccaaggcagctgtggacacttgc  
tgcaactacggcaacagaatctgtgaacttcagaaacccggccaaagaacaacacagctggactacaaggagatcgacatccaaggcagctgtggacacttgc  
agaagaacggcatcgacgtgaagcaggagaacctgtatcgacacatctgcgagatgaaggacaagccctttcaagggcctgtatcgccaaacatcaagatcg  
ctgcagatcagaacacagcgctagccggacatcgactacatgtatcgccccccatgaatgacagaggcgagtttcgacaccagaaggccctgcaac  
aactgcctctggacgctgtatcgtaacggccctataacatcgccaaagaagggcctgtggatcgaccatcagaacaccaccggcaacaacgtgaagatg  
gccatgaccaacagagatggatgcacttcggccaggaaagcagacttgcttga

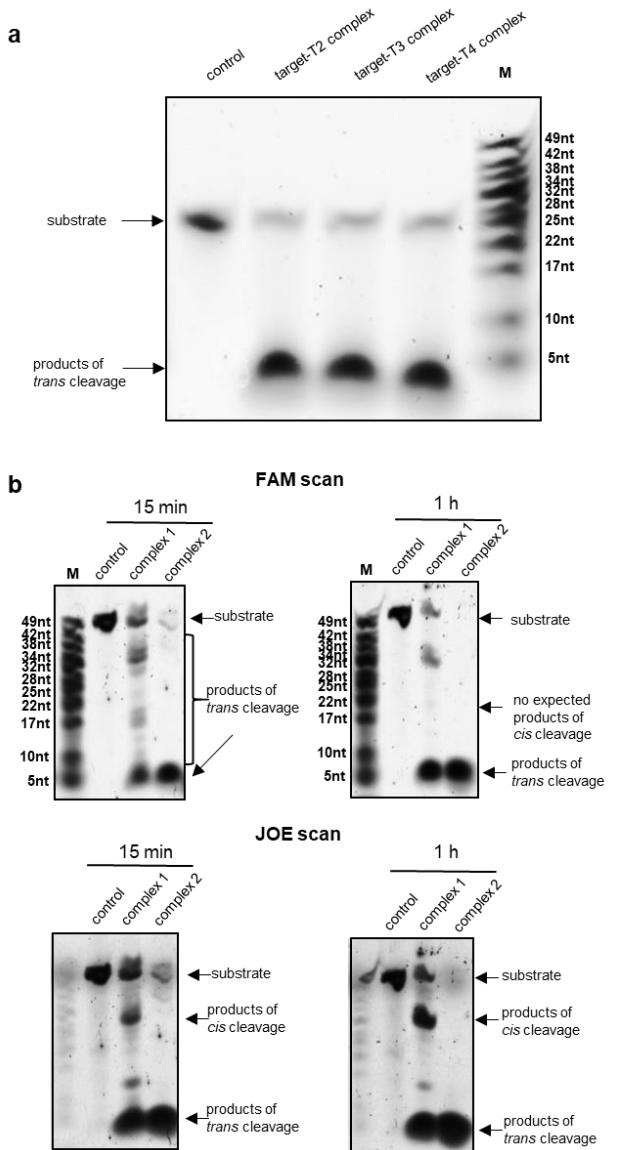


**Figure S1** Cleavage of ssDNA by Cas12a with a programmable and PAM-independent mechanism. **(a)** Cleavage of ssDNA and dsDNA by Cas12a. 3'-FAM-labelled ssDNA or dsDNA containing different PAMs were cleaved by Cas12a and crRNA-DNMT-23nt. For dsDNA, only ds1 with the PAM sequence of “TTC” was successfully cleaved; however, for ssDNAs, all targets with or without PAM sequences could be cleaved. Schematic of the substrates was shown in the lower panel. **(b)** Identification of the Cas12a cleavage sites on ssDNA of target-DNMT1-3 using urea PAGE. The cleavage sites were near the 22<sup>nd</sup> base downstream of the first crRNA-paired base with different crRNAs. Schematic of the ssDNA substrate and crRNAs was shown in the lower panel. Paired guide sequences were shown by orange lines and the cleavage sites were indicated by both arrows and numbers. **(c)** Determination of the cleavage sites of Cas12a on target-DNMT1-3 ssDNA. crRNAs had short guide sequences of 16 nts, 18 nts and 20 nts, respectively. As there were 8 nts before the paired site in the target ssDNA, the 30-nt product indicated the cleavage of target ssDNA after the 22<sup>nd</sup> base by Cas12a. Schematic of the ssDNA substrate and crRNAs was shown in the lower panel. **(d)** crRNAs with different lengths of guide sequences were tested on dsDNA and ssDNA. At least 16-nt guide sequences were required by Cas12a for cleavage of target dsDNA;

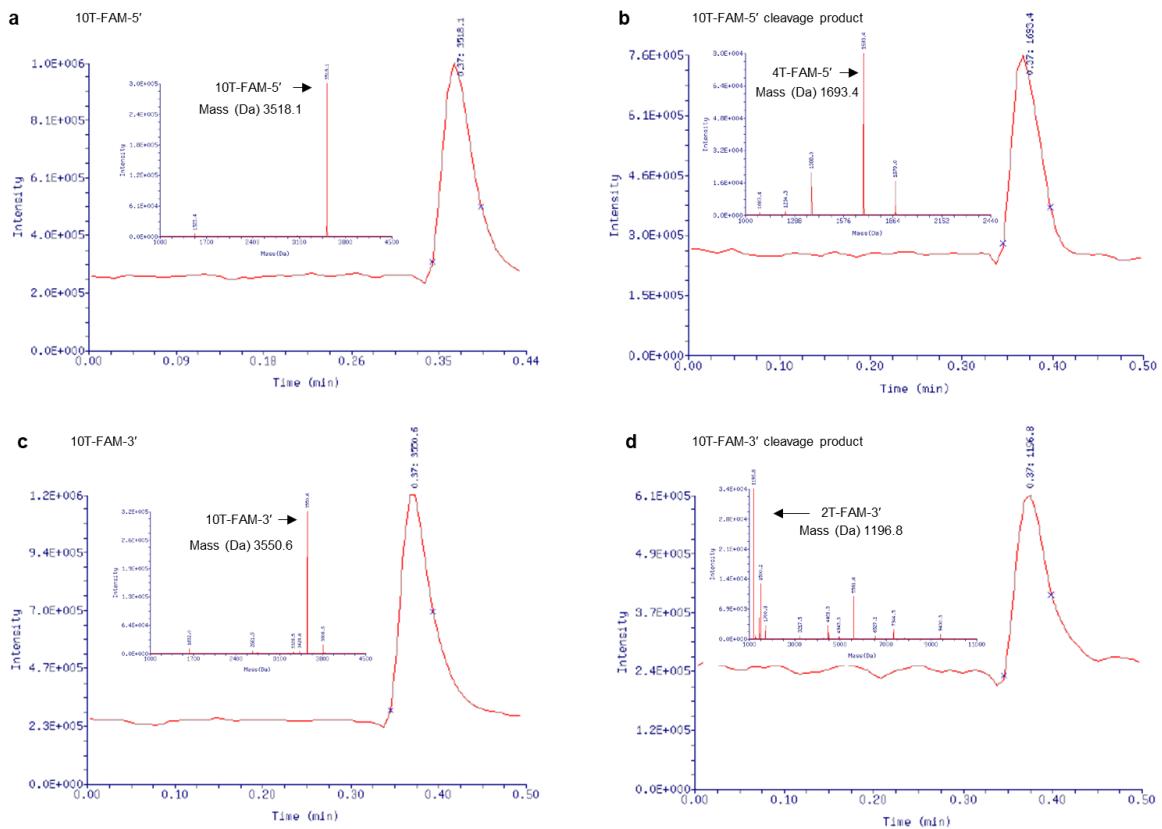
however, even 10-nt guide sequence of crRNA enabled the cleavage of ssDNA by Cas12a. Schematic of the target DNA (target-T1) and the crRNAs was shown in the lower panel. **(e)** Time-course cleavage experiment. FAM was labelled on the 3'-end of the T-strand of dsDNA or target ssDNA, with the schematic of DNA substrates shown in the lower panel. Target sequence was shown in red and the PAM site of “TTC” was shown in yellow. **(f)** Quantification of the Cas12a DNA cleavage activity on dsDNA and ssDNA substrates. Cleavage assays were conducted in triplicate and data were represented as mean  $\pm$  SEM.



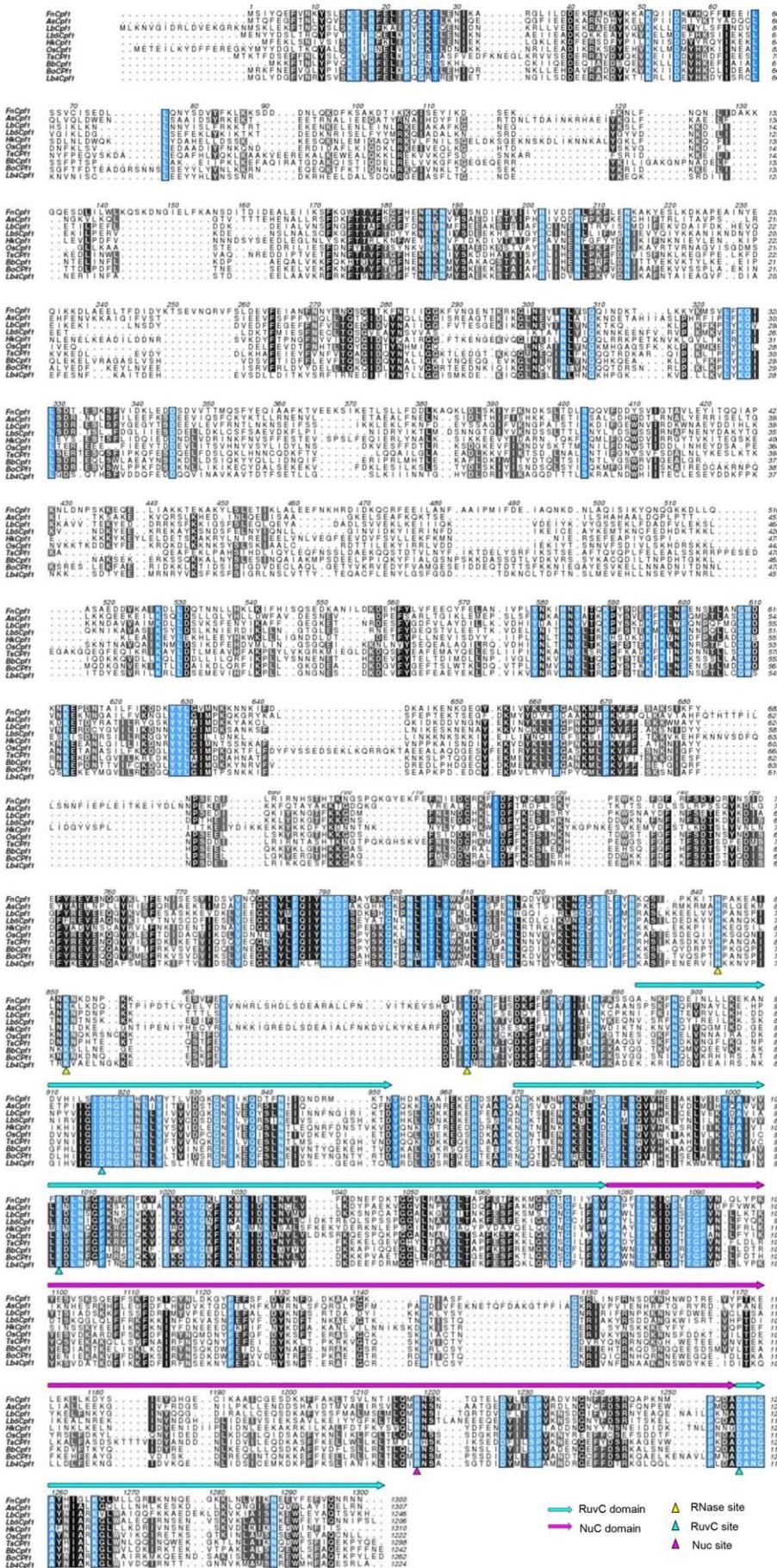
**Figure S2** The Cas12a *trans* cleavage activity on ssDNA. **(a)** Time-course cleavage of DNMT1-3 ssDNA in the presence of Cas12a/crRNA-T1-24nt/target-T1 complex. Collateral ssDNAs (DNMT1-3) labelled with FAM at either the 3'-end (left part) or 5'-end (right part) were promiscuously cleaved. The concentration of crRNA and Cas12a used here was reduced to 100 nM each. **(b and c)** Cleavage of FAM-labelled collateral ssDNAs (i.e. target-T1-F-FAM, target-T1-R-FAM, target-T1-FAM-3'-F and target-T1-FAM-5'-R) by Cas12a **(b)** or its complex **(c)**. Digestion reactions with no Cas12a or only Cas12a were employed as negative controls **(b)**. When only Cas12a and crRNA-DNMT-23nt were added, collateral ssDNAs (target-T1, FAM-labelled) could not be cleaved. However, upon the formation of the ternary complex of Cas12a/crRNA-DNMT-23nt/target-DNMT1-3 (dsDNA or ssDNA) **(c)**, collateral ssDNAs (target-T1, FAM-labelled) were *trans* cleaved.



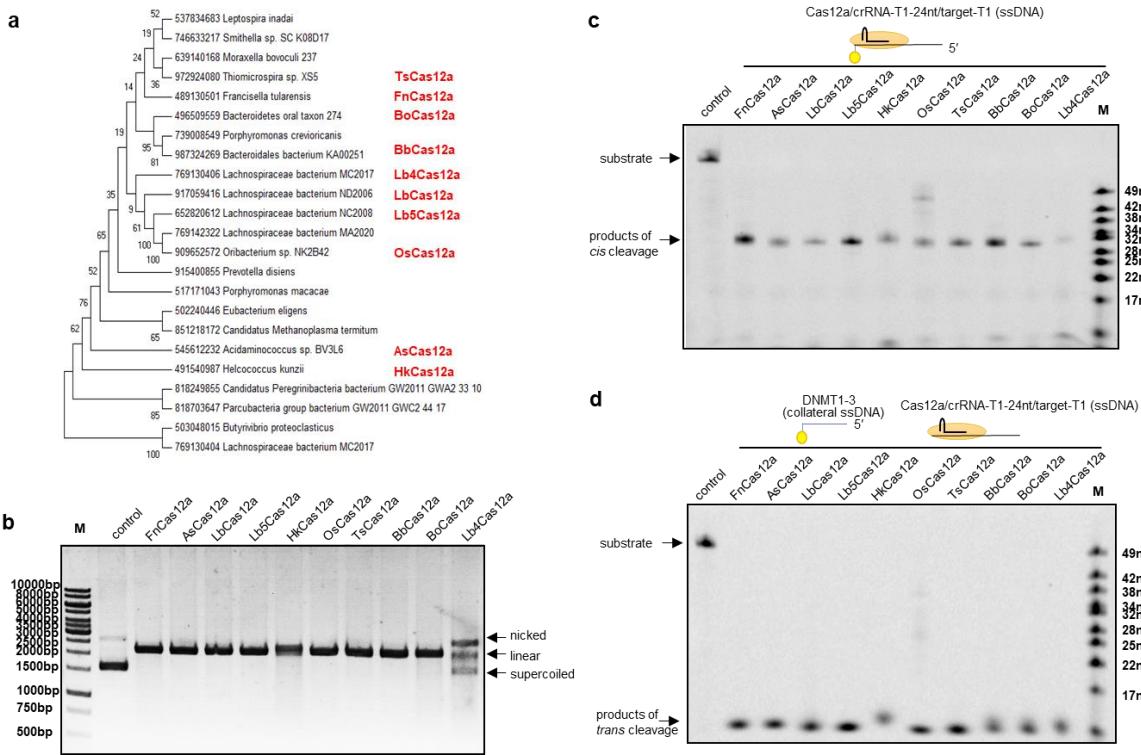
**Figure S3** Cas12a-mediated *cis*- and *trans*-cleavage of ssDNA. **(a)** Collateral ssDNA substrate N25-3'FAM was *trans*-cleaved by the Cas12a complexes. All complexes, including Cas12a/crRNA-T2/target-T2, Cas12a/crRNA-T3/target-T3 and Cas12a/crRNA-T4/target-T4, had the ssDNA *trans*-cleavage activity. **(b)** Cleavage of double-end labelled ssDNA substrate (target-DNMT1-3-R-FAM-5'-JOE-3'), which was labelled with 5'-FAM and 3'-JOE. Double-end labelled ssDNA was cleaved for 15 min and 1 h by complex 1 (Cas12a/crRNA-DNMT-23nt/target-DNMT1-3-R-FAM-5'-JOE-3') or complex 2 (Cas12a/crRNA-T1/target-T1-24-R). 3'-JOE-labelled *cis*-cleavage product (about 30 nt) only appeared with JOE scan, and no 5'-FAM-labelled *cis*-cleavage product (expected at 20 nt) could be observed.



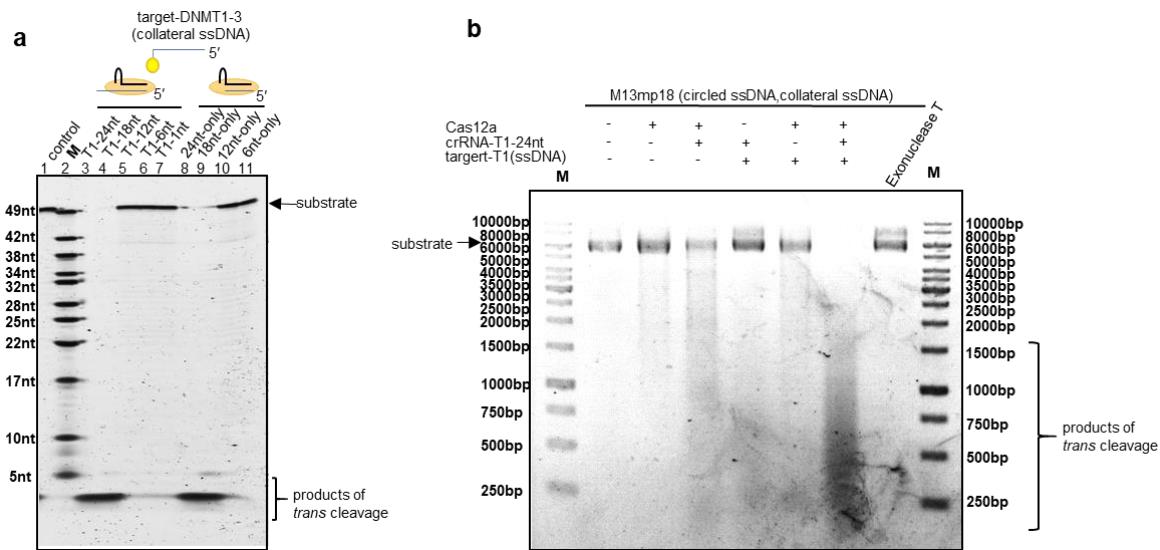
**Figure S4** Analysis of ssDNA by LC-MS. Analysed ssDNAs included 10T-FAM-5' (**a**), 10T-FAM-3' (**c**) and their corresponding products (**b** and **d**) *trans*-cleaved by complex of Cas12a/crRNA-T1/target-T1-18-R. The main cleavage product of substrate 10T-FAM-5' was 4T-FAM-5' with the molecular weight of 1693.4 (**b**), and main cleavage product of 10T-FAM-3' was 2T-FAM-3' with the molecular weight of 1196.8 (**d**).



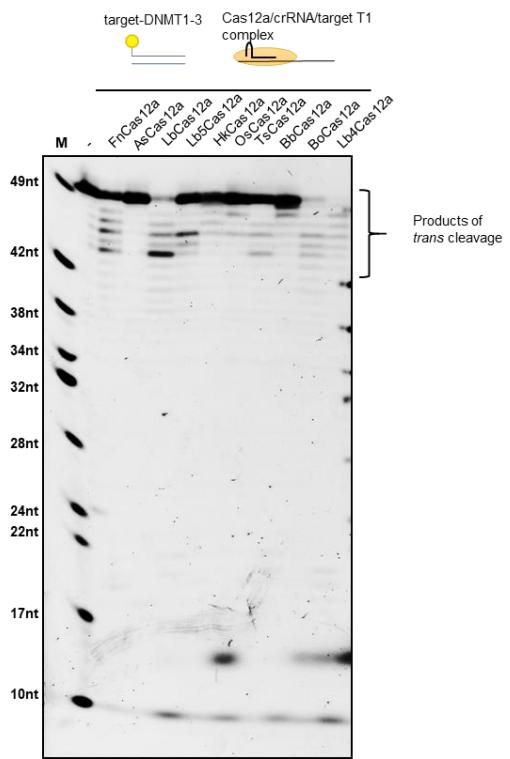
**Figure S5** Multiple sequence alignment of Cas12a proteins. The RuvC and Nuc domains were labelled above the amino acid sequences, and key residues were indicated by coloured triangles. Fn, *Francisella tularensis*; As, *Acidaminococcus* sp. BV3L6; Lb, *Lachnospiraceae bacterium* ND2006; Lb5, *Lachnospiraceae bacterium* NC2008; Hk, *Helcoccusc kunzii* ATCC 51366; Os, *Oribacterium* sp. NK2B42; Ts, *Thiomicrospira* sp. XS5; Bb, *Bacteroidales bacterium* KA00251; Bo, *Bacteroidetes oral taxon* 274 str. F0058; Lb4, *Lachnospiraceae bacterium* MC2017.



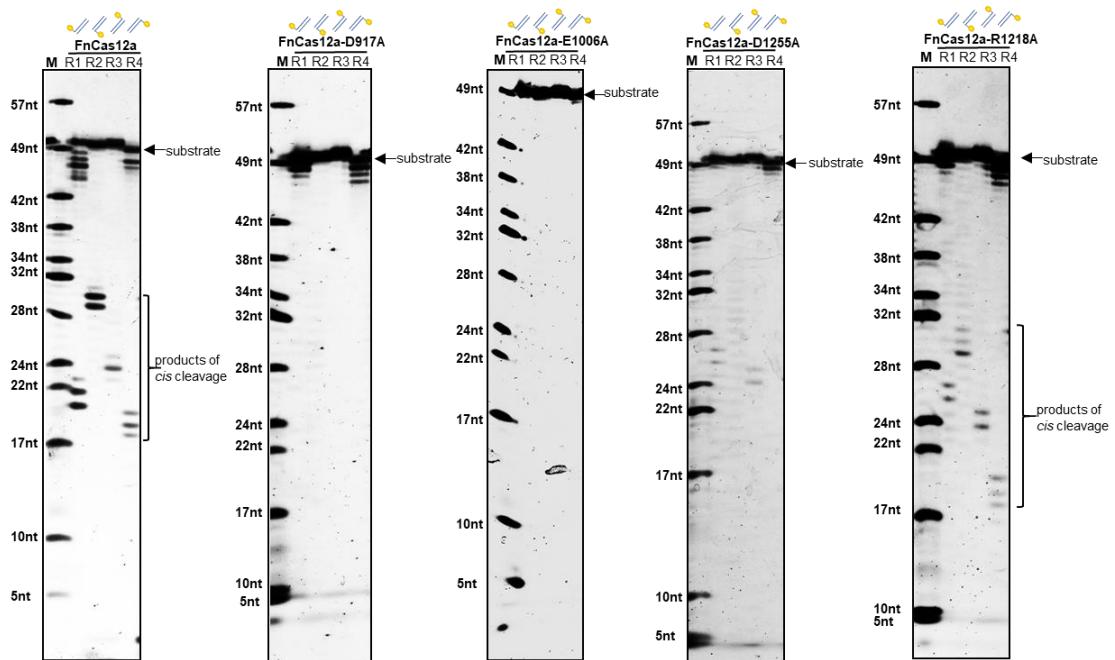
**Figure S6** Characterization of the *cis* and *trans* cleavage activities of ten Cas12a proteins from different species. **(a)** A phylogenetic tree of partial Cas12a-family proteins, among which ten were chosen for further biochemical analyses and were indicated in red. **(b)** Cleavage of plasmid pCB1A2\_2 by ten different Cas12a proteins with crRNA-T1-24nt. **(c)** *Cis* cleavage of the ssDNA target-T1 by ten different Cas12a proteins with crRNA-T1-24nt. All Cas12a proteins showed *cis* cleavage activity. **(d)** *Trans* cleavage of the ssDNA target-DNMT1-3 by the complex of Cas12a/crRNA-T1-24nt/target-T1 (ssDNA). All Cas12a complexes showed *trans* cleavage activity.



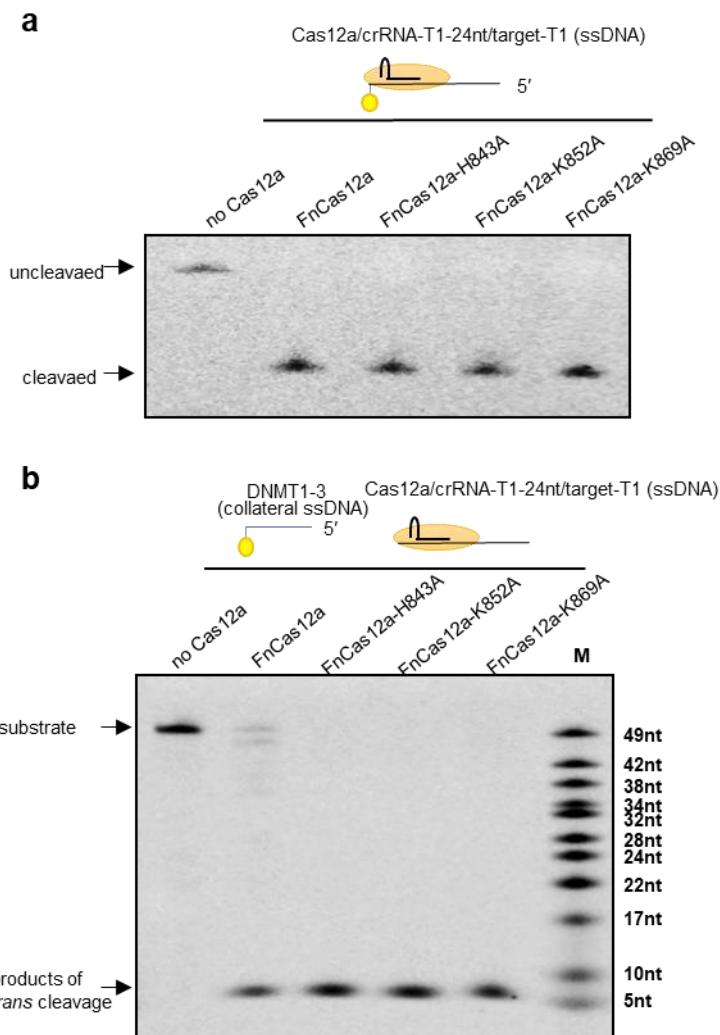
**Figure S7** The Cas12a *trans* cleavage activity on ssDNA. **(a)** *Trans* cleavage of collateral ssDNA (target-DNMT1-3) by the Cas12a complex with different lengths of target ssDNA. In lanes 3-7, 3'-extended sequences were designed on the target ssDNA, which was indicated in the schematic; lanes 8-11, there were no extended sequences beyond the 3' terminal of the target ssDNA. The crRNA-T1-24nt was used in the assay, and detailed information of the target ssDNAs used could be found in Extended Data Table 1. **(b)** Cas12a *trans* cleavage of circular target ssDNA of M13mp18. Once the ternary complex of Cas12a/crRNA/target DNA formed, circular ssDNA M13mp18 was promiscuously cleaved. Exonuclease T (Exo T), which is a single-stranded RNA- or DNA-specific nuclease that requires a free 3' terminus and removes nucleotides in the 3' to 5' direction, showed no cleavage activity on circular M13mp18 and was therefore employed as a negative control.



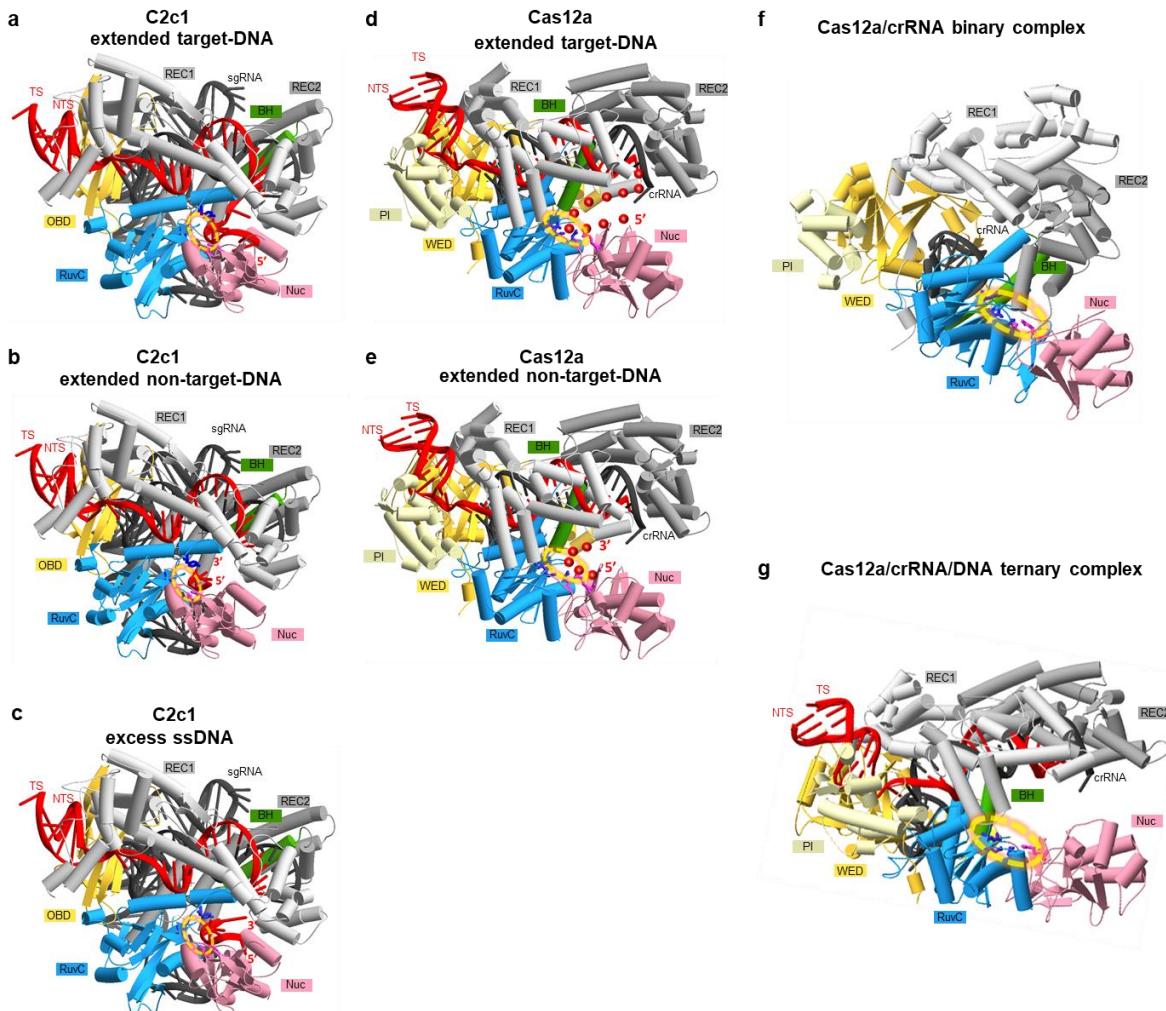
**Figure S8** *Trans* cleavage of dsDNA substrate by ten different Cas12a complexes. Upon the formation of Cas12a/crRNA-T1/target-T1 (ssDNA), most Cas12a complexes (except AsCas12a) had *trans* cleavage activity on the ends of dsDNA substrate of target-DNMT1-3, among which the activity of LbCas12a, BoCas12a and Lb4Cas12a complexes was much higher.



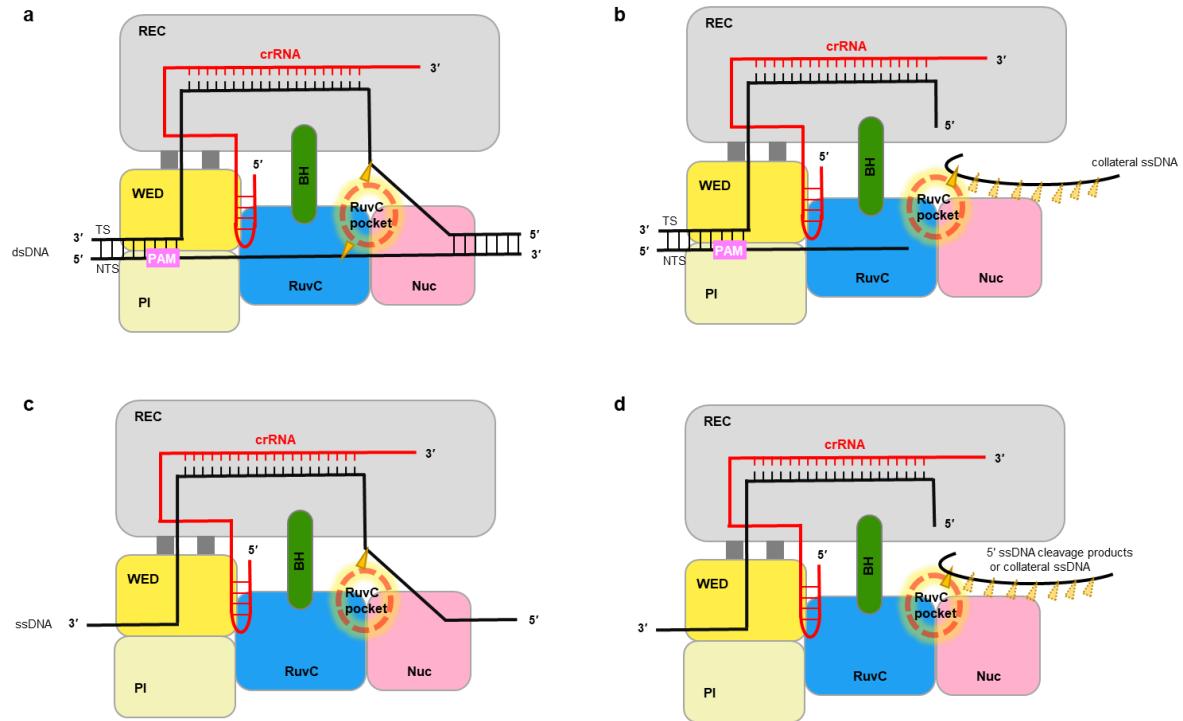
**Figure S9** Identification of the cleavage sites in double-stranded DNMT1-3 by FnCas12a and its mutants (D917A, E1006A and D1255A mutations in the RuvC domain; R1218A mutation in the Nuc domain), employing crRNA-DNMT-23nt and FAM-labelled dsDNA. Labelling of targets was indicated above each lane, i.e. R1: 5'-labelled non-target strand, R2: 3'-labelled target strand, R3: 3'-labelled non-target strand, and R4: 5'-labelled target strand. All mutants showed either weakened or completely lost activity on dsDNA cleavage.



**Figure S10** Identification of key residues in Cas12a that were involved in the *cis* and *trans* ssDNA cleavage. **(a)** *cis* cleavage of target ssDNA (target-T1-R-FAM) by FnCas12a and three FnCas12a mutants with single amino acid replacement in H843, K852 and K869, which are associated with the RNase activity. No significant difference was found in the *cis* cleavage activity among the tested Cas12a proteins. **(b)** *Trans* cleavage of the 3'-FAM-labelled collateral ssDNA of target-DNMT1-3-R by the ternary complexes of wild-type FnCas12a or its mutants. The three tested residues (H843, K852 and K869 in FnCas12a) are associated with the RNase activity. No significant difference was found in the *trans* cleavage activities among the tested Cas12a proteins.



**Figure S11** The RuvC catalytic pocket model for C2c1 and Cas12a. **(a-c)** C2c1 complex: the ternary complex C2c1 with sgRNA and extended target DNA (PDB: 5U30 in Figure S11a), extended non-target DNA (PDB: 5U33 in Figure S11b) and excess ssDNA (PDB: 5U31 in Figure S11c). All above substrate DNAs were positioned in the RuvC catalytic pocket (labelled in dashed yellow circle). **(d-e)** Cas12a complex (PDB: 5B43): the ternary complex of Cas12a with crRNA and proposed extended target DNA (Figure S11d) and extended non-target DNA (Figure S11e). Red dots represented the proposed positions of extended target-DNA and non-target-DNA. **(f-g)** Figure S11f represented the binary complex Cas12a/crRNA with a triangle-shaped structure (PDB: 1WJX) and Figure S11g represented the ternary complex Cas12a/crRNA/DNA with a bilobed architecture (PDB: 5B43). Molecular graphic images were prepared using CueMol (<http://www.cuemol.org>). DNA was colored in Red, RNA was colored in black and the RuvC catalytic pocket was indicated by dashed yellow circles.



**Figure S12** The Cas12a cleavage models. Substrates included target dsDNA **(a)**, collateral ssDNA *trans*-cleaved by the ternary complex of Cas12a/crRNA/target dsDNA **(b)**, target ssDNA *cis*-cleaved by the Cas12a complex **(c)** and *cis*-cleaved ssDNA which was then *trans*-cleaved by the Cas12a complex **(d)**. All substrate DNAs were proposed to be cleaved by the active sites in the Cas12a RuvC pocket.

## References

1. Li, S. Y., Zhao, G. P. & Wang, J. C-Brick: A New Standard for Assembly of Biological Parts Using Cpf1. *ACS Synth. Biol.* **5**, 1383-1388 (2016).
2. Lei, C. *et al.* The CCTL (Cpf1-assisted Cutting and Taq DNA ligase-assisted Ligation) method for efficient editing of large DNA constructs *in vitro*. *Nucleic Acids Res.* **45**, e74 (2017).
3. Carneiro, F. R. *et al.* Spectroscopic characterization of the tumor antigen NY-REN-21 and identification of heterodimer formation with SCAND1. *Biochemical and biophysical research communications* **343**, 260-268 (2006).