Supplemental Table S1: Amino acid replacements causing defects in allosteric control in Hsp70s

Activities and conformational changes of amino acid replacement variants relative to the wild type protein.

			Crystal structure contacts ^a		ATPase	Effects on allostery					
Position in DnaK	replace- ment	protein analyzed ^b	1HPM	4B9Q	rel. k _{cat} c	Substrate stimulated ATPase ^d	Dna J- stimulated ATPase ^e	DnaJ- Substrate stimulation ^f	ATP- stimulated substrate release ^g	ATP-induced conformational change ^h	Reference
11	T37G	CgBiP	T13/OG1-Pi/O4	T13/OG1-ATP/O1G	0.19	-			-	- (proteolysis)	(1)
	T13G	BtHsc70	T13/ N-Pi/O3		0.002					no ∆Rg (SAXS) no blueshift	(2)
	T13S	BtHsc70			0.09					red. ΔRg (SAXS)	(2)
	T13V	BtHsc70			0.02					no ∆Rg (SAXS) no blueshift	(2)
70	K71M	BtHsc70	K71/NZ-E175/OE2 K71/NZ-Pi/O3	K70/NZ-ATP/O1G K70/NZ-E171/OE2	0.00					no ΔRg (SAXS)	(3)
	K70A	EcDnaK	K71/CE-P147/CG K71/CE-P147/CD	K70/CE-P143/CG K70/CE-P143/CD	0.04				-	no blueshift	(4)
143	P143S	EcDnaK	P147/CG-K71/CE	P143/CG-K70/CE	0.06	+	+				(5)
	B 4404		P147/CD-K71/CE	P143/CD-K70/CE	0.07	4.40	0.00	0.40	0.40		(0)
	P143A	EcDnaK	P147/CG-F150/CE1	P143/CG-F146/CE1	0.27	1.43	0.66	0.46	0.16	red. blueshift	(6)
	P143G	EcDnaK	F 147/CD-F 150/CE2	F 143/CD-F 140/CE2	0.12	0.49	0.39	0.24	0.67	red. blueshift	(6)
145,147,148	YND>AAA	EcDnaK			3.49	0.92	0.15	0.125	0.34	red. blueshift	(7)
145	Y145A	EcDnaK	Y149/OH-H2O-D206/OD2		16.10	0.85	0.30	0.03	0.27	red. blueshift	(8)
	Y145F	EcDnaK	Y149/OH-H2O-PiO1		2.20	1.27	2.88	0.60	0.61	wt blueshift	(8)
	Y145H	EcDnaK			2.40	3.13	7.34	1.00	0.58	wt blueshift	(8)
146	F146A	EcDnaK	F150/CZ-K70/CG F150/CE1-P143/CG F150/CE2-P143/CD	F146/CZ-K70/CG F146/CE1-P143/CG F146/CE2-P143/CD	5.17	0.66	1.33	0.34	0.06	red. blueshift	(8)
148	D148A	EcDnaK		D148/OD1-L484/N	3.41	0.55	1.05	0.07	2.4	wt blueshift	(8)
151	R151A	EcDnaK	R155/NH1-N170/OD1 R155/NH2-F146/O	R151/NH1-N170/OD1 R151/NH2-F146/O	4.5	0.12	0.37	0.03	0	no blueshift	(6)
	R151K	EcDnaK	R155/NH2-A144/O R155/NE-F146/O	R151/NH2-A144/O R151/NE-D481/O	17.6	0.26	0.29	0.02	0.06	red. blueshift	(6)
155	K155A	EcDnaK			0.86	0.82	1.79	0.16	0.03	red. blueshift	(9)
	K155D	EcDnaK			1.44	1.46	0.73	0.11	0.00	red. blueshift	(9)

167	R167A	EcDnak		R167/NE-D/81/OD1	2.8	1.0/	0.56	0.11	0.03	red blueshift	(0.10)
107		EcDnak			2.0	0.42	0.50	0.11	0.00	no bluochift	(0)
		EcDitar			1.5	0.42	0.51	0.03	0	no bluesnin	(9)
170		EcDilar						reduced			(10)
170	N170A			N170/ND2-D393/OD2	0.04			reduced			(10)
171	E175S	BtHsc/0	E175/0E1-PI/01	E1/1/0E2-K/0/NZ	0.01					red ΔRg (SAXS)	(3)
	E171D	EcDnaK	E175/0E2-R71/NZ E175/0E2-H20-Ma	E171/0E2-1120-1vig E171/0E1-H20-	0.02	1.15	0.5	0.25	0.18	red. blueshift	(6)
	E171Q	EcDnaK		D201/OD2	0.01	1.83	0.4	0.18	0.11	red. blueshift	(6)
173	T173A	EcDnaK		T173/OG1-D393/OD2 T173/CG2-L391/CD2				reduced			(10)
194	D199S	BtHsc70	D199/OD1-H2O-Mg D199/OD2-H2O-Mg D199/N-D206/O D199/O-D206/N	D194/OD1-H2O-Mg D194/OD2-H2O-Mg D194/N-D201/O D194/O-D201/N	0.32					no ∆Rg (SAXS)	(3)
196	G226D	CgBiP			0.33				-	- (proteolysis)	(1)
197	G227D	CgBiP	G202/N-ADP/O1B	G197/N-ATP/O3B	0.29				-	- (proteolysis)	(1)
199	T199A	EcDnaK	T204/OG1-Pi/O4	-	0.030				+	wt blueshift	(4)
	T204E	BtHsc70			7.500						(11)
	T204V	BtHsc70			0.350						(11)
	T229G	CgBiP			-				+	yes	(1)
201	D201N	EcDnaK	D206/OD2-H2O-Y149/OH	D201/OD2-H2O-	10.3	0.1-0.2	-			red. (proteolysis)	(12)
	D206S	BtHsc70	D206/OD2-H2O-Pi/O1	E171/OE1	0.28					red ΔRg (SAXS)	(3)
217,218	EV>AA	EcDnaK			4.29	0.85	0.11	0.07	0.10	red. blueshift	(7)
326	D326V	EcDnaK		D326/OD2-K414/NZ D326/OD2-N415/ND2	ca. 2	0.43				red. blueshift	(13)
388	D388R	EcDnaK			1.33	0.81	2.3	0.35	0.96	wt blueshift	(9)
389-392	VLLL>AAAA	EcDnaK				-	-	-	-		(14,15)
389-392	VLLL>VDDL	EcDnaK				-	-	-	-		(14,15)
389	V389A	EcDnaK		V389/CG1-L177/CD2 V389/CG1-V281/CG2		-	+	+	reduced		(16)
	V389D	EcDnaK		V389/CG1-F216/CD1		-	-	reduced	-		(16)
390	L390A	EcDnaK				reduced	reduced	reduced			(16)
	L390D	EcDnaK				reduced	-	-			(16)
391	L391A	EcDnaK		L391/CD1-L177/CD2 L391/CD1-F216/CB		-	-	reduced	reduced		(16)
	L391D	EcDnaK		L391/CD2-T173/CG2		-	-	-	-		(16)

392	L392A	EcDnaK			reduced	-	-			(16)
	L392D	EcDnaK			reduced	-	-	reduced		(16)
393	D393A	EcDnaK	D393/OE2-N170/ND2	2.40	0.44	0.51	0.02	0.01	no blueshift	(9)
	D393R	EcDnaK	D393/OE2-T173/OG1	1.36	0.36	0.51	0.02	0.00	no blueshift	(9)
414	K414I	EcDnaK	K414/NZ-D326/OD2	25.9				0.002	no blueshift	(8,17)
415	N415G	EcDnaK	N415/ND2-D326/OD2 N415/OD1-T221/N	ca. 1.6	0.5				red. blueshift	(13)
440	V440A	EcDnaK	V440/CG1-L484/CD2	1.97	0.42	1.43	0.06	2.4	red. blueshift	(8)
481	D481A	EcDnaK	D481/OD1-I168/N D481/OD1-R167/NE	84	0.56	0.89	0.03	0.002	no blueshift	(8)
	D481K	EcDnaK	D481/O-R151/NE	70	0.64	0.57	0.02	0.001	no blueshift	(8)
484	L484A	EcDnaK	L484/CD2-V440/CG1	1.72	0.5	0.98	0.06	2.9	red. blueshift	(8)

a: Crystal structure contacts in the ADP·Pi bound NBD (1HPM, DnaK numbering) and the ATP bound DnaK (4B9Q) in pdb nomenclature. Distances between H-bond donor and acceptor <3.5 Å; hydrophobic contacts <4.2Å; b: Cg, Cricetulus griseus; Bt, Bos taurus; Ec, Escherichia coli; c: V_{max}(mut)/V_{max}(wt) or k_{cat}(mut)/k_{cat}(wt); d: Substrate-stimulated ATPase rate; -, not detectable; +, proficient; numbers= [k_{cat}(mut+substrate)/k_{cat}(wt)]/[k_{cat}(wt+substrate)/k_{cat}(wt)]; e: DnaJ-stimulated ATPase rate; -, not detectable; +, proficient; numbers= [k_{cat}(mut+DnaJ)/k_{cat}(wt)]/[k_{cat}(wt+DnaJ)/k_{cat}(wt)]; f: synergistic stimulation by DnaJ+substrate; -, not detectable; +, proficient; numbers= [k_{cat}(mut+DnaJ+substrate)/k_{cat}(wt)]; g: ATP-stimulated substrate release; -, not detectable; +, proficient; numbers= [k_{cat}(mut+ATP)/k_{off}(mut)]/[k_{off}(wt+ATP)/k_{off}(wt)]; h: ATP-induced conformational changes as determined by proteolysis, tryptophane fluorescence (blueshift), or small angle X-ray scattering (SAXS); red., reduced; wt, wild type-like; ΔRg, change in radius of gyration.

References

- 1. Wei J, Gaut JR, Hendershot LM. In vitro dissociation of BiP-peptide complexes requires a conformational change in BiP after ATP binding but does not require ATP hydrolysis. J Biol Chem. 1995 Nov 3;270(44):26677–82.
- 2. Sousa MC, McKay DB. The hydroxyl of threonine 13 of the bovine 70-kDa heat shock cognate protein is essential for transducing the ATP-induced conformational change. Biochemistry. 1998 Nov 3;37(44):15392–9.
- 3. Johnson ER, McKay DB. Mapping the role of active site residues for transducing an ATP-induced conformational change in the bovine 70-kDa heat shock cognate protein. Biochemistry. 1999 Aug 17;38(33):10823–30.
- 4. Barthel TK, Zhang J, Walker GC. ATPase-defective derivatives of Escherichia coli DnaK that behave differently with respect to ATP-induced conformational change and peptide release. Journal of Bacteriology. 2001 Oct 1;183(19):5482–90.
- 5. Burkholder WF, Panagiotidis CA, Silverstein SJ, Cegielska A, Gottesman ME, Gaitanaris GA. Isolation and characterization of an Escherichia coli DnaK mutant with impaired ATPase activity. Journal of Molecular Biology. 1994 Sep 30;242(4):364–77.
- 6. Vogel M, Bukau B, Mayer MP. Allosteric regulation of Hsp70 chaperones by a proline switch. Molecular Cell. 2006 Feb 3;21(3):359–67.

- 7. Gässler CS, Buchberger A, Laufen T, Mayer MP, Schröder H, Valencia A, et al. Mutations in the DnaK chaperone affecting interaction with the DnaJ cochaperone. Proc Natl Acad Sci USA. 1998 Dec 22;95(26):15229–34.
- 8. Kityk R, Vogel M, Schlecht R, Bukau B, Mayer MP. Pathways of allosteric regulation in Hsp70 chaperones. Nat Commun. 2015;6:8308.
- 9. Vogel M, Mayer MP, Bukau B. Allosteric regulation of Hsp70 chaperones involves a conserved interdomain linker. J Biol Chem. 2006 Dec 15;281(50):38705–11.
- 10. Suh WC, Burkholder WF, Lu CZ, Zhao X, Gottesman ME, Gross CA. Interaction of the Hsp70 molecular chaperone, DnaK, with its cochaperone DnaJ. Proc Natl Acad Sci USA. 1998 Dec 22;95(26):15223–8.
- 11. O'Brien MC, McKay DB. Threonine 204 of the chaperone protein Hsc70 influences the structure of the active site, but is not essential for ATP hydrolysis. J Biol Chem. 1993 Nov 15;268(32):24323-9.
- 12. Kamath-Loeb AS, Lu CZ, Suh WC, Lonetto MA, Gross CA. Analysis of three DnaK mutant proteins suggests that progression through the ATPase cycle requires conformational changes. J Biol Chem. 1995 Dec 15;270(50):30051–9.
- 13. Smock RG, Rivoire O, Russ WP, Swain JF, Leibler S, Ranganathan R, et al. An interdomain sector mediating allostery in Hsp70 molecular chaperones. Mol Syst Biol. 2010 Sep 21;6:414.
- 14. Laufen T, Mayer MP, Beisel C, Klostermeier D, Mogk A, Reinstein J, et al. Mechanism of regulation of hsp70 chaperones by DnaJ cochaperones. Proc Natl Acad Sci USA. 1999 May 11;96(10):5452–7.
- 15. Mayer MP, Laufen T, Paal K, McCarty JS, Bukau B. Investigation of the interaction between DnaK and DnaJ by surface plasmon resonance spectroscopy. Journal of Molecular Biology. 1999 Jun 18;289(4):1131–44.
- 16. Kumar DP, Vorvis C, Sarbeng EB, Cabra Ledesma VC, Willis JE, Liu Q. The four hydrophobic residues on the Hsp70 inter-domain linker have two distinct roles. Journal of Molecular Biology. 2011 Sep 2;411(5):1099–113.
- 17. Montgomery DL, Morimoto RI, Gierasch LM. Mutations in the substrate binding domain of the Escherichia coli 70 kDa molecular chaperone, DnaK, which alter substrate affinity or interdomain coupling. Journal of Molecular Biology. 1999 Feb 26;286(3):915–32.