Supporting Information: Gluco-1*H***-imidazole: a new class of azole-type βglucosidase inhibitor**

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Experimental procedures and characterization data

General: Chemicals were purchased from Acros, Sigma Aldrich, Biosolve, VWR, Fluka, Merck and Fisher Scientific and used as received unless stated otherwise. Tetrahydrofuran (THF), *N,N*dimethylformamide (DMF) and toluene were stored over molecular sieves before use. Traces of water from reagents were removed by co-evaporation with toluene in reactions that required anhydrous conditions. All reactions were performed under an argon atmosphere unless stated otherwise. TLC analysis was conducted using Merck aluminum sheets (Silica gel 60 F₂₅₄) with detection by UV absorption (254 nm), by spraying with a solution of $(NH_4)_6MO_7O_{24}$ -4H₂O (25 g/L) and $(NH_4)_4$ Ce(SO₄)₄·2H₂O (10 g/L) in 10% sulfuric acid or a solution of KMnO₄ (20 g/L) and K₂CO₃ (10 g/L) in water, followed by charring at ~150 °C. Column chromatography was performed using Screening Device b.v. silica gel (particle size of 40 – 63 µm, pore diameter of 60 Å) with the indicated eluents. ¹H NMR and 13 C NMR spectra were recorded on a Brüker AV-400 (400 and 101 MHz respectively) or a Brüker AV-500 (500 and 125 MHz respectively) spectrometer in the given deuterated solvent. Chemical shifts are given in ppm (δ) relative to the residual solvent peak or tetramethylsilane (0 ppm) as internal standard. Coupling constants are given in Hz. High-resolution mass spectrometry (HRMS) analysis was performed with a LTQ Orbitrap mass spectrometer (Thermo Finnigan), equipped with an electronspray ion source in positive mode (source voltage 3.5 kV, sheath gas flow 10 mL/min, capillary temperature 250 °C) with resolution R = 60000 at m/z 400 (mass range m/z = 150 – 2000) and dioctyl phthalate (m/z = 391.28428) as a "lock mass", or with a Synapt G2-Si (Waters) , equipped with an electronspray ion source in positive mode (ESI-TOF), injection via NanoEquity system (Waters), with LeuEnk (m/z = 556.2771) as "lock mass". Eluents used: MeCN:H2O (1:1 v/v) supplemented with 0.1% formic acid. The high-resolution mass spectrometers were calibrated prior to measurements with a calibration mixture (Thermo Finnigan). Glucoimidazole **5** was prepared according to the procedure by Vasella *et al*.^[1] NMR spectra of this compound are provided herein and are in agreement with those previously reported.

General procedure 1 (GP1): Bis-azidation

The diol starting material was dissolved in dry CHCl₃ (0.2 M), then Et₃N (3 equiv.) and *N*-methyl imidazole (10 equiv.) were added and the mixture was cooled to 0 °C. MsCl (4 equiv.) was added and the mixture was stirred 16 h at rt. The mixture was quenched with water at 0 °C, diluted with EtOAc, washed with aq. 1M HCl (2 x), H₂O and brine. The organic layer was dried over MgSO₄, filtered and concentrated. After co-evaporation with toluene (2 x), the crude intermediate product was dissolved in dry DMF (0.1 M). NaN₃ (10 equiv.) was added and the mixture was stirred 16 h at 100 °C. Then, the mixture was diluted with H₂O and extracted with Et₂O (3 x). The combined organic layers were washed with H₂O and brine, dried over MgSO₄, filtered and concentrated. The product was purified by flash column chromatography using the indicated eluent.

General procedure 2 (GP2): Azide reduction

The bisazido starting material was dissolved in THF (0.05 M) under N_2 atmosphere. PtO₂ (30 mol%) was added, the reaction mixture was purged with H_2 with a balloon, and the mixture was stirred vigorously for 16 h. Then, the mixture was filtered over a small Celite pad and concentrated. The product was purified by flash column chromatography using the indicated eluent.

General procedure 3 (GP3): Imidazoline formation

The diamino starting material was dissolved in HFIP (0.1 M), the appropriate trimethyl orthoester (3 equiv.) was added and the mixture was stirred for 16 h at rt. The mixture was diluted with Et₂O and washed with aq. 1M NaOH (3 x), H₂O and brine, dried over MgSO₄, filtered and concentrated. The product was purified by flash column chromatography using the indicated eluent.

It should be noted that for both glucose and conduritol configurations, oxidation of the 2-butylimidazolines to the 2-butyl-imidazoles proceeded in only moderate yields when IBX/DMSO was employed. In contrast, we found that oxidation proceeded more smoothly under Swern conditions.[2,3]

General procedure 4 (GP4): Oxidation to the imidazole (IBX, DMSO)

The imidazoline starting material was dissolved in DMSO (0.1 M), IBX $^{[4]}$ (1.5 equiv.) was added and the mixture was stirred 16 h at 45 °C. Next, the mixture was cooled to rt, quenched with aq. 10% Na₂S₂O₃ and aq. 1M NaOH. The mixture was stirred for 15 min, diluted with Et₂O, washed with H₂O (3 x) and brine, dried over $MgSO_4$, filtered and concentrated. The product was purified by flash column chromatography using the indicated eluent.

General procedure 5 (GP5): Oxidation to the imidazole (Swern conditions)

To dry DCM (0.1 M based on starting material) was added DMSO (7 equiv.) and the mixture was cooled to -60 °C. Then, oxalyl chloride (5 equiv.) was added slowly and the mixture was stirred for 30 min. The imidazoline starting material was co-evaporated with toluene (2 x), dissolved in dry DCM (1 mL) and added dropwise. The mixture was stirred for 1 h at -60 °C and subsequently quenched with Et3N (7 equiv.). The cooling bath was removed and the mixture was allowed to reach rt. After stirring 1 h at rt, the mixture was diluted with EtOAc, washed with $H₂O$ (3 x) and brine. The organic layer was dried with MgSO₄, filtered and concentrated. The product was purified by flash column chromatography using the indicated eluent.

General procedure 6 (GP6): Hydrogenation

The imidazole starting material was dissolved in MeOH (0.03 M) under $N₂$ atmosphere, then HCl (1.25M in MeOH, 10 equiv.) and Pd(OH)₂/C (20 wt%) were added and the mixture was purged with H₂ with a balloon. The mixture was stirred vigorously for 16 h, filtered over a small Celite pad and finally concentrated which afforded the pure product.

Compound 11b and **compound 11a**

Cyclohexene 10^{5} (1.28 g, 2.46 mmol) was dissolved in EtOAc (15 mL) and MeCN (15 mL) and cooled to 0 °C. A solution of RuCl $_3.3H_2O$ (36 mg, 0.17 mmol) and NaIO₄ (789 mg, 3.69 mmol) in H₂O (4.9 mL) was added and the mixture was stirred vigorously at 0 °C

for 90 min. The mixture was quenched by addition of aq. 10% $Na₂S₂O₃$ (20 mL) and the mixture was stirred for 15 min. Then the mixture was diluted with H₂O (100 mL) and extracted with EtOAc (3 x 60 mL). The combined organic layers were washed with brine, dried over MgSO₄, filtrated and concentrated. The product was purified by flash column chromatography (pentane/EtOAc, 4:1 \rightarrow 2:1) affording compound **11b** (432 mg, 32%) and **11a** (539 mg, 40%) as white solids. *Analytical data for 11b:* ¹H-NMR (400 MHz, CDCl3) δ 7.69 – 6.77 (m, 20H), 5.00 – 4.85 (m, 4H), 4.79 (d, *J* = 11.1 Hz, 1H), 4.59 – 4.39 (m, 3H), 4.25 (s, 1H), 3.89 (m, 3H), 3.73 (dd, *J* = 8.9, 2.9 Hz, 1H), 3.61 – 3.45 (m, 2H), 3.34 (s, OH), 2.41 (d, *J* = 4.8 Hz, OH), 1.74 (dq, *J* = 8.0, 2.3 Hz, 1H). ¹³C-NMR (101 MHz, CDCl3) δ 138.7, 138.7, 138.4, 137.6, 128.7, 128.7, 128.6, 128.6, 128.1, 128.1, 127.9, 127.9, 127.9, 127.8, 127.7, 86.7, 82.4, 77.4, 75.8, 75.7, 75.6, 74.6, 73.7, 71.0, 68.9, 43.5. IR (neat, cm⁻¹): ν 3441, 2866, 1452, 1058. HRMS (ESI) m/z: [M+H]⁺ calc for C₃₅H₃₉O₆ 555.27412, found 555.27411. Analytical data for 11a: ¹H-NMR (400 MHz, CDCl3) δ 7.38 – 7.15 (m, 20H), 4.94 (d, *J* = 10.8 Hz, 1H), 4.87 (d, *J* = 10.8 Hz, 1H), 4.82 (d, *J* = 10.8 Hz, 1H), 4.71 (s, 2H), 4.55 – 4.38 (m, 3H), 4.14 (s, 1H), 3.96 (t, *J* = 9.4 Hz, 1H), 3.84 (dd, *J* = 9.0, 2.5 Hz, 1H), 3.68 – 3.65 (m, 2H), 3.46 – 3.30 (m, 2H), 3.05 (d, *J* = 6.2 Hz, OH), 2.62 (s, OH), 2.18 (tdd, *J* = 10.9, 5.0, 2.5 Hz, 1H). ¹³C-NMR (101 MHz, CDCl3) δ 138.8, 138.5, 138.0, 138.0, 128.5, 128.5, 128.4, 128.4, 128.0, 128.0, 127.9, 127.7, 127.7, 127.7, 127.6, 82.9, 80.0, 77.7, 75.7, 75.3, 73.3, 72.5, 70.3, 69.3, 67.7, 43.2. IR (neat, cm⁻¹): v 3441, 2868, 1452, 1064. HRMS (ESI) m/z: [M+H]⁺ calc for $C_{35}H_{39}O_6$ 555.27412, found 555.27374. These data are in agreement with those previously reported.^[6]

Compound 12a

Starting from **11a** (55 mg, 0.1 mmol) and following **GP1**, the product was purified by flash column chromatography (pentane/EtOAc, 15:1) affording compound **12a** as a white solid (45 mg, 74%). ¹H-NMR (400 MHz, CDCl₃) δ 7.44 – 7.11 (m, 20H), 4.96 – 4.74 (m, 5H), 4.55 – 4.37 (m, 3H), 4.16 (t, *J* = 2.9 Hz, 1H), 3.86 – 3.75 (t, *J* = 9.6 Hz, 1H), 3.73 (dd, *J* = 8.9, 4.3 Hz, 1H), 3.58 –

3.49 (m, 2H), 3.49 – 3.41 (m, 1H), 3.38 (dd, *J* = 10.2, 9.2 Hz, 1H), 2.08 – 1.95 (m, 1H). ¹³C-NMR (101 MHz, CDCl3) δ 138.4, 137.9, 137.8, 137.8, 128.6, 128.6, 128.6, 128.4, 128.2, 128.1, 128.1, 128.0, 127.8, 127.6, 87.0, 81.1, 78.0, 76.1, 75.9, 75.6, 73.6, 67.5, 65.9, 61.1, 43.8. IR (neat, cm-1): ν 2858, 2102, 1359, 1066. HRMS (ESI) m/z: $[M+Na]^+$ calc for $C_{35}H_{37}N_6O_4$ 605.28708, found 605.33734.

Compound 12b

Starting from **11b** (55 mg, 0.1 mmol) and following **GP1**, the product was purified by flash column chromatography (pentane/EtOAc, 15:1) affording compound **12b** as a white solid (35 mg, 58%). ¹H-NMR (400 MHz, CDCl₃) δ 7.39 – 7.13 (m, 20H), 4.95 – 4.67 (m, 5H), 4.48 (d, *J* = 4.3 Hz, 1H), 4.45 (d, *J* = 3.5 Hz, 1H), 4.34 (d, *J* = 11.5 Hz, 1H), 4.05 (t, *J* = 2.9 Hz, 1H), 3.86 (t, *J* = 9.5 Hz,

1H), 3.82 (d, *J* = 9.2 Hz, 1H), 3.56 (ddd, *J* = 9.6, 6.5, 4.1 Hz, 2H), 3.51 (dd, *J* = 10.6, 2.4 Hz, 2H), 2.02 (t, *J* $= 11.2$ Hz, 1H). ¹³C-NMR (101 MHz, CDCl₃) δ 138.6, 138.4, 138.0, 137.6, 128.7, 128.5, 128.2, 128.1, 128.1, 127.9, 127.8, 127.8, 83.1, 80.4, 77.7, 76.0, 75.7, 73.3, 73.3, 65.0, 63.7, 57.7, 42.5. IR (neat, cm-¹): v 2858, 2098, 1359, 1082. HRMS (ESI) m/z: $[M+Na]^+$ calc for C₃₅H₃₆N₆O₄Na 627.26902, found 627.26849.

Compound 13a

Starting from **12a** (367 mg, 0.61 mmol) and following **GP2**, the product was purified by flash column chromatography (DCM/MeOH, 99:1 \rightarrow 49:1) affording compound 13a as a colorless oil (269 mg, 80%). ¹H-NMR (400 MHz, CDCl3) δ 7.42 – 7.11 (m, 20H), 4.99 (d, *J* = 11.1 Hz, 1H), 4.96 – 4.83 (m, 3H), 4.66 (d, *J* = 11.1 Hz, 1H), 4.55 – 4.41 (m, 3H), 3.91 (dd, *J* = 11.0, 9.3 Hz, 1H),

3.77 – 3.63 (m, 3H), 3.59 (t, *J* = 9.2 Hz, 1H), 3.38 (t, *J* = 3.0 Hz, 1H), 2.80 (dd, *J* = 10.0, 3.4 Hz, 1H), 1.87 (ddt, J = 11.0, 7.4, 3.2 Hz, 1H), 1.53 (s, 4H, 2 x NH₂). ¹³C-NMR (101 MHz, CDCl₃) δ 138.9, 138.7, 138.6, 138.2, 128.6, 128.5, 128.5, 128.1, 128.0, 127.8, 127.8, 127.7, 127.5, 88.3, 82.3, 78.5, 75.8, 75.4, 75.3, 73.3, 68.8, 56.6, 51.7, 44.8. IR (neat, cm⁻¹): ν 2856, 1361, 1066. HRMS (ESI) m/z: [M+H]⁺ calc for C₃₅H₄₁N₂O₄ 553.30608, found 553.30585.

Compound 13b

Starting from **12b** (858 mg, 1.42 mmol) and following **GP2**, the product was purified by flash column chromatography (DCM/MeOH, 99:1 \rightarrow 7:3) affording compound 13b as a colorless oil (750 mg, 96%). ¹H-NMR (400 MHz, CDCl3) δ 7.27 (m, 20H), 4.91 (t, *J* = 11.9 Hz, 2H), 4.79 (d, *J* = 10.7 Hz, 1H), 4.69 (d, *J* = 11.7 Hz, 1H), 4.64 (d, *J* = 11.6 Hz, 1H), 4.46 (d, *J* = 27.6 Hz, 3H), 3.90 (t,

J = 9.2 Hz, 1H), 3.77 (d, *J* = 7.6 Hz, 1H), 3.64 (d, *J* = 7.8 Hz, 1H), 3.50 (t, *J* = 9.6 Hz, 1H), 3.49 - 3.43 (m, 2H), 2.91 (d, J = 10.9 Hz, 1H), 1.97 (t, J = 10.9 Hz, 1H), 1.87 (s, 4H, 2 x NH₂). ¹³C-NMR (101 MHz, CDCl₃) δ 139.1, 138.9, 138.5, 128.6, 128.5, 128.1, 128.0, 127.9, 127.9, 127.8, 127.6, 127.6, 83.1, 81.6, 78.9, 75.7, 75.4, 73.2, 72.2, 66.1, 53.3, 49.6, 43.4. IR (neat, cm⁻¹): v 2860, 1602, 1496, 1452, 1359, 1066. HRMS (ESI) m/z: [M+Na]⁺ calc for C₃₅H₄₀N₂O₄Na 575.28803, found 575.28741.

Compound 14a

Starting from **13a** (55 mg, 0.1 mmol) and following **GP3** using trimethyl orthoformate, the product was purified by flash column chromatography (DCM/MeOH, 99:1 \rightarrow 7:3) affording compound **14a** as a colorless oil (43 mg, 76%). ¹H-NMR (400 MHz, CD₃CN) δ 7.41 – 7.18 (m, 20H), 7.07 (s, 1H), 4.84 – 4.64 (m, 5H), 4.51 – 4.42 (m, 3H), 4.01 (dd, *J* = 9.5, 4.4 Hz, 1H), 3.86 (dd, *J* =

9.4, 6.2 Hz, 1H), 3.77 (dd, *J* = 9.2, 4.1 Hz, 1H), 3.68 (t, *J* = 8.8 Hz, 1H), 3.58 – 3.51 (m, 1H), 3.50 – 3.40 (m, 2H), 2.25 (ddt, J = 12.3, 8.4, 4.2 Hz, 1H). ¹³C-NMR (101 MHz, CD₃CN) δ 156.2, 140.0, 139.9, 139.8, 139.7, 129.3, 129.2, 129.2, 128.9, 128.8, 128.7, 128.5, 128.4, 128.4, 85.1, 83.5, 78.9, 74.7, 74.5, 74.1, 73.7, 69.9, 65.6, 60.7, 41.7. IR (neat, cm⁻¹): v 3278, 3030, 2862, 1654, 1543, 1359, 1066. HRMS (ESI) m/z: $[M+H]^+$ calc for $C_{36}H_{38}N_2O_4$ 563.29043, found 563.29022.

Compound 14b

Starting from **13b** (110 mg, 0.2 mmol) and following **GP3** using trimethyl orthoformate, the product was purified by flash column chromatography (DCM/MeOH, 99:1 \rightarrow 4:1) affording compound **14b** as a colorless oil (98 mg, 87%). ¹H-NMR (400 MHz, CD₃CN) δ 7.38 – 7.20 (m, 20H), 7.03 (s, 1H), 4.78 – 4.62 (m, 5H), 4.53 – 4.44 (m, 3H), 3.97 (dd, *J* = 9.3, 4.3 Hz, 1H), 3.86 – 3.81

(m, 1H), 3.81 – 3.78 (m, 1H), 3.70 (t, *J* = 7.0 Hz, 1H), 3.67 – 3.63 (m, 2H), 3.38 (dd, *J* = 11.3, 7.2 Hz, 1H), 1.74 - 1.66 (m, 1H). ¹³C-NMR (101 MHz, CD₃CN) δ 155.7, 140.1, 140.0, 140.0, 139.8, 129.3, 129.2, 129.2, 128.9, 128.8, 128.7, 128.7, 128.5, 128.4, 128.4, 83.4, 79.6, 79.1, 74.5, 74.4, 73.8, 73.3, 69.2, 61.7, 60.5, 46.2. IR (neat, cm⁻¹): v 3030, 2868, 1681, 1595, 1454, 1087. HRMS (ESI) m/z: [M+H]⁺ calc for $C_{36}H_{39}N_2O_4$ 563.29043, found 563.29010.

Compound 16a

Starting from **13a** (119 mg, 0.21 mmol) and following **GP3** using trimethyl orthovalerate, the product was purified by flash column chromatography (DCM/MeOH, 99:1 \rightarrow 8:2) affording compound 16a as a colorless oil (99 mg, 74%). ¹H-NMR (400 MHz, CD₃CN) δ 7.39 – 7.18 (m, 20H), 4.73 (m, 5H), 4.51 – 4.43 (m, 3H), 3.99 (dd, *J* = 9.0, 4.4 Hz, 1H), 3.77 (dt, *J* = 9.2, 4.6 Hz, 2H), 3.68 (t, *J* = 8.8 Hz, 1H), 3.53 (t, *J* = 7.5 Hz, 1H), 3.46 – 3.39 (m, 2H), 2.19 (dq, *J* = 12.8, 4.3 Hz, 1H), 2.07 (td, *J* = 7.4, 3.6

Hz, 2H), 1.51 – 1.42 (m, 2H), 1.31 (dq, *J* = 14.2, 7.2 Hz, 2H), 0.88 (t, *J* = 7.3 Hz, 3H). ¹³C-NMR (101 MHz, CD3CN) δ 168.4, 140.2, 140.0, 139.9, 139.8, 129.3, 129.3, 129.2, 129.2, 128.9, 128.8, 128.7, 128.7, 128.4, 128.4, 128.3, 85.5, 83.7, 79.1, 74.8, 74.6, 73.9, 73.5, 70.0, 66.8, 61.6, 42.2, 29.7, 29.4, 23.1, 14.1. IR (neat, cm⁻¹): v 2868, 1600, 1454, 1363, 1066. HRMS (ESI) m/z: [M+H]⁺ calc for C₄₀H₄₇N₂O₄ 619.35303, found 619.35266.

Compound 16b

Starting from **13b** (110 mg, 0.2 mmol) and following **GP3** using trimethyl orthovalerate, the product was purified by flash column chromatography (DCM/MeOH, 99:1 \rightarrow 4:1) affording compound **16b** as a colorless oil (96 mg, 78%). ¹H-NMR (400 MHz, CD₃CN) δ 7.40 – 7.19 (m, 20H), 4.76 (dd, J = 11.2, 6.3 Hz, 2H), 4.71 – 4.62 (m, 3H), 4.53 – 4.43 (m, 3H), 4.00 (dd, *J* = 9.0, 3.5 Hz, 1H), 3.83 (t, *J* = 9.4 Hz, 1H), 3.77 – 3.63 (m, 4H), 3.39 (dd, *J* = 11.2, 6.5 Hz, 1H), 2.18 (t, *J* = 7.6 Hz, 2H), 1.73 (t, *J* = 10.4 Hz, 1H), 1.52 (m,

2H), 1.31 (m, 2H), 0.87 (t, J = 7.3 Hz, 3H). ¹³C-NMR (101 MHz, CD₃CN) δ 168.5, 140.1, 140.0, 139.8, 129.3, 129.2, 129.2, 129.1, 128.8, 128.7, 128.7, 128.6, 128.4, 128.4, 128.4, 128.3, 83.6, 80.0, 79.0, 74.6, 74.5, 73.8, 73.3, 69.1, 62.5, 61.0, 46.4, 29.7, 29.5, 23.1, 14.2. IR (neat, cm-1): ν 2862, 1608, 1454, 1359, 1091. HRMS (ESI) m/z: $[M+H]^+$ calc for $C_{40}H_{47}N_2O_4$ 619.35303, found 619.35260.

Compound 15

Starting from **14a** (43 mg, 76 μmol) and following **GP4**, the product was purified by flash column chromatography (DCM/MeOH, 99:1 \rightarrow 67:1) affording compound **15** as a colorless oil (32 mg, 75%). Using the same conditions, product **15** could be obtained from imidazoline **14b** (82 mg, 0.15 mmol) in 71% yield (58 mg). ¹H-NMR (500 MHz, CD₃CN) δ 7.51 (s, 1H), 7.42

– 7.18 (m, 20H), 5.05 (d, *J* = 11.5 Hz, 1H), 4.88 – 4.81 (m, 4H), 4.67 – 4.63 (m, 1H), 4.54 – 4.44 (m, 3H), 3.96 (dd, *J* = 9.0, 6.4 Hz, 1H), 3.86 (dd, *J* = 9.0, 3.9 Hz, 1H), 3.76 (t, *J* = 8.1 Hz, 1H), 3.59 (t, *J* = 7.5 Hz, 1H), 3.09 (m, 1H). ¹³C-NMR (125 MHz, CD₃CN) δ 140.3, 140.0, 139.8, 139.4, 136.8, 129.3, 129.2, 129.2, 129.0, 128.8, 128.8, 128.6, 128.5, 128.4, 128.3, 85.6, 79.5, 77.7, 75.5, 75.3, 73.7, 72.7, 70.0, 40.9. IR (neat, cm⁻¹): v 3028, 2862, 1496, 1454, 1359, 1087. HRMS (ESI) m/z: [M+H]⁺ calc for $C_{36}H_{37}N_2O_4$ 561.27478, found 561.27454.

Compound 17

Starting from **16a** (74 mg, 0.12 mmol) and following **GP5**, the product was purified by flash column chromatography (DCM/MeOH, 199:1 \rightarrow 99:1) affording compound **17** as a colorless oil (56 mg, 76%). Using the same conditions, product **17** could be obtained from imidazoline **16b** (40 mg, 64.7 μmol) in 70% yield (28 mg). ¹H-NMR (400 MHz, CD₃CN) δ 7.41 – 7.19 (m, 20H), 4.98 (d, *J* = 11.7 Hz, 1H), 4.87 – 4.76 (m, 4H), 4.63 (dd, *J* = 6.1, 1.5 Hz, 1H), 4.54 – 4.40 (m, 3H), 3.95 (dd, *J* = 8.8, 6.2 Hz, 1H), 3.84

(dd, *J* = 9.0, 4.1 Hz, 1H), 3.81 – 3.72 (m, 1H), 3.66 – 3.55 (m, 1H), 3.13 – 2.96 (m, 1H), 2.63 (d, *J* = 7.6 Hz, 2H), 1.68 – 1.59 (m, 2H), 1.35 (m, 2H), 0.92 (t, J = 7.4 Hz, 3H). ¹³C-NMR (101 MHz, CD₃CN) δ 150.4, 140.4, 140.0, 139.8, 139.5, 129.3, 129.2, 129.2, 129.1, 128.9, 128.8, 128.7, 128.5, 128.4, 128.4, 128.2, 85.4, 79.3, 77.7, 75.3, 75.2, 73.7, 72.5, 69.8, 41.3, 31.6, 29.0, 23.1, 14.1. IR (neat, cm-1): ν 2862, 1454, 1359, 1089. HRMS (ESI) m/z: [M+H]⁺ calc for C₄₀H₄₅N₂O₄ 617.33738, found 617.33710.

Compound 6 (gluco-1*H***-imidazole)**

Starting from **15** (26 mg, 46.4 μmol) following **GP6**, the pure product was afforded as a colorless oil (12 mg, quant.). ¹H-NMR (400 MHz, D₂O) δ 8.55 (s, 1H), 4.69 (d, *J* = 7.6 Hz, 1H), 4.14 (dd, *J* = 11.4, 2.8 Hz, 1H), 3.93 (dd, *J* = 11.4, 4.8 Hz, 1H), 3.79 (t, *J* = 9.4 Hz, 1H), 3.71 (dd, *J* = 9.8, 7.4 Hz, 1H), 3.01 (s, 1H). ¹³C-NMR (101 MHz, D₂O) δ 135.2, 128.2, 126.6, 76.9, 69.3, 66.6, 58.8, 40.9

HRMS (ESI-TOF) m/z: $[M+Na]^+$ calc for $C_8H_{12}N_2O_4$ 223.0689, found 223.0702.

Compound 7 (gluco-2-butyl-1*H***-imidazole)**

Starting from **17** (48 mg, 77.8 μmol) following **GP6**, the pure product was afforded as a colorless oil (24 mg, quant.). 1 H-NMR (400 MHz, MeOD) δ 4.53 (d, *J* = 7.4 Hz, 1H), 4.12 (dd, *J* = 10.8, 3.1 Hz, 1H), 3.89 (dd, *J* = 10.8, 5.0 Hz, 1H), 3.72 (t, *J* = 9.0 Hz, 1H), 3.60 (dd, *J* = 9.3, 7.4 Hz, 1H), 2.94 (t, *J* = 7.6 Hz, 2H), 2.87 (s, 1H), 1.69 – 1.58 (m, 2H), 1.29 (m, 2H), 0.86 (t, *J* = 7.3 Hz, 3H). ¹³C-NMR (101 MHz, MeOD) δ 150.5, 129.7, 127.6, 78.9, 71.1, 68.2, 60.5, 43.0, 31.0, 26.6, 23.1, 13.8. HRMS (ESI-TOF) m/z: [M+H]⁺ calc

for $C_{12}H_{20}N_2O_4$ 257.1496, found 257.1510.

Synthesis of conduritol B-1H-imidazoles

Compound S2

Starting from **S1**[7] (1.88 g, 3.48 mmol) and following **GP1**, the product was N_3 purified by flash column chromatography (pentane/EtOAc, 9:1) affording BnO. N_3 compound **S2** as a white solid (1.87 g, 91%). ¹H-NMR (400 MHz, CDCl₃) δ 7.45 – 7.27 (m, 20H), 4.98 – 4.91 (m, 3H), 4.89 – 4.82 (m, 3H), 4.82 – 4.72 (m, 2H), 4.01 BnO['] OBn (t, *J* = 3.1 Hz, 1H), 3.95 (t, *J* = 9.5 Hz, 1H), 3.85 (t, *J* = 9.7 Hz, 1H), 3.60 (dd, *J* = 9.6, ŌBn 3.2 Hz, 1H), 3.51 (t, *J* = 9.3 Hz, 1H), 3.41 (dd, *J* = 10.2, 3.1 Hz, 1H). ¹³C-NMR (101 MHz, CDCl₃) δ 138.3, 137.6, 137.4, 128.7, 128.5, 128.5, 128.3, 128.2, 128.1, 128.0, 127.9, 127.8, 127.7, 127.6, 83.9, 81.4, 80.2, 80.0, 76.2, 75.9, 73.4, 62.0, 61.9. HRMS (ESI) m/z: [M+H]⁺ calc for C₃₄H₃₅N₆O₄ 613.25337, found 613.25281. This analytical data is in accordance with literature. $^{[7]}$

Compound S3

Starting from **S2** (1.8 g, 3.0 mmol) and following **GP2**, the product was purified by flash column chromatography (DCM/MeOH, 99:1 \rightarrow 9:1) affording compound **S3** as a white solid (1.63 g, 99%). ¹H-NMR (400 MHz, CDCl₃) δ 7.43 – 7.13 (m, 20H), 5.03 – 4.87 (m, 3H), 4.82 (dd, *J* = 10.8, 4.4 Hz, 2H), 4.65 (m, 3H), 4.03 (t, *J* = 9.2 Hz, 1H), 3.76 (t, *J* = 9.6 Hz, 1H), 3.50 (m, 3H), 2.66 (dd, *J* = 10.0,

2.7 Hz, 1H), 1.53 (s, 4H, 2xNH₂). ¹³C-NMR (101 MHz, CDCl₃) δ 138.9, 138.8, 138.5, 128.6, 128.6, 128.5, 128.5, 128.1, 128.1, 127.9, 127.8, 127.8, 127.8, 127.7, 127.6, 85.3, 82.0, 81.9, 81.9, 76.0, 75.8, 75.6, 72.4, 54.0, 51.8. HRMS (ESI) m/z: $[M+H]^+$ calc for $C_{34}H_{39}N_2O_4$ 539.29043, found 539.29007. This analytical data is in accordance with literature.^[7]

Compound S4

Starting from **S3** (400 mg, 0.74 mmol) and following **GP3** using trimethyl orthoformate, the product was purified by flash column chromatography (DCM/MeOH, 99:1 \rightarrow 7:3) affording compound **S4** as a colorless oil (387 mg, 95%). ¹H-NMR (400 MHz, CD₃CN) δ 7.34 (m, 20H), 7.14 (s, 1H), 4.76 (s, 2H), 4.75 – 4.63 (m, 6H), 4.16 (dd, *J* = 10.7, 4.2 Hz, 1H), 3.97 – 3.87 (m, 2H), 3.82 – 3.73

(m, 2H), 3.57 (dd, J = 7.7, 5.6 Hz, 1H). ¹³C-NMR (101 MHz, CD₃CN) δ 155.2, 139.1, 138.9, 138.8, 138.8, 128.3, 128.3, 128.3, 128.2, 127.9, 127.9, 127.8, 127.8, 127.6, 127.6, 127.5, 127.4, 83.2, 81.3, 80.5, 77.0, 73.0, 72.9, 72.9, 72.4, 64.0, 59.5. IR (neat, cm-1): ν 3030, 2866, 1670, 1452, 1066. HRMS (ESI) m/z: $[M+H]^+$ calc for $C_{35}H_{37}N_2O_4$ 549.27478, found 549.27526.

Compound S5

Starting from **S4** (55 mg, 0.1 mmol) and using **GP4**, the product was purified by flash column chromatography (DCM/MeOH, 199:1 \rightarrow 67:1) affording compound **S5** as a colorless oil (34 mg, 62%). ¹H-NMR (400 MHz, CD₃CN) δ 7.57 (s, 1H), 7.44 – 7.14 (m, 20H), 4.92 – 4.83 (m, 4H), 4.80 (d, *J* = 11.2 Hz, 2H), 4.74 – 4.70 (m, 2H), 3.91 (dd, J = 4.4, 2.2 Hz, 2H). ¹³C-NMR (101 MHz, CD₃CN) δ 139.9, 137.9, 129.2, 129.2, 128.8, 128.4, 128.4, 84.5, 78.2 (broad, assigned with HSQC), 75.7, 73.2. IR (neat, cm⁻¹):

v 3030, 2866, 1585, 1496, 1452, 1344, 1053. HRMS (ESI) m/z: [M+H]⁺ calc for C₃₅H₃₅N₂O₄ 547.25913, found 547.25897.

Compound 8 (conduritol B-1*H***-imidazole)**

Starting from **S5** (18 mg, 32.9 μmol) and following **GP6**, the pure product was afforded as a colorless oil (8.0 mg, quant.). ¹H-NMR (400 MHz, D₂O) δ 8.70 (s, 1H), 4.76 (m, 2H), 3.74 – 3.58 (d, $J = 2.7$ Hz, 2H). ¹³C-NMR (101 MHz, D₂O) δ 135.8. 127.5, 75.9, 66.5. HRMS (ESI-TOF) m/z: $[M+Na]^+$ calc for $C_7H_{10}N_2O_4$ 209.0533, found 209.0542.

Compound S6

Starting from **S3** (53 mg, 0.1 mmol) and following **GP3** using trimethyl orthovalerate, the product was purified by flash column chromatography (DCM/MeOH, 99:1 \rightarrow 8:2) affording compound **S6** as a colorless oil (53 mg, 89%). ¹H-NMR (400 MHz, CD₃CN) δ 7.42 – 7.18 (m, 20H), 4.77 – 4.56 (m, 8H), 4.11 (dd, *J* = 10.2, 4.1 Hz, 1H), 3.89 – 3.77 (m, 2H), 3.75 – 3.66 (m, 2H), 3.51 (dd, *J* = 7.8, 5.7 Hz, 1H), 2.19 – 2.06 (t, *J* = 7.6 Hz, 2H), 1.55 – 1.42 (m, 2H), 1.29 (m, 2H), 0.84 (t, J = 7.3 Hz, 3H). ¹³C-NMR (101 MHz, CD₃CN) δ

168.9, 140.2, 139.9, 139.8, 129.2, 129.2, 128.9, 128.8, 128.6, 128.5, 128.4, 84.2, 82.3, 81.8, 78.5, 73.9, 73.8, 73.3, 65.6, 61.5, 29.5, 29.4, 23.1, 14.1. IR (neat, cm-1): ν 2870, 1606, 1454, 1357, 1064. HRMS (ESI) m/z: $[M+H]^+$ calc for $C_{39}H_{45}N_2O_4$ 605.33738, found 605.33722.

Compound S7

Starting from **S6** (53 mg, 0.088 mmol) and following **GP5**, the product was purified by flash column chromatography (DCM/MeOH, 99:1) affording compound **S7** as a colorless oil (39 mg, 74%). ¹H-NMR (400 MHz, CD₃CN) δ 7.49 – 7.17 (m, 20H), 4.87 (m, 8H), 4.70 (dd, *J* = 4.3, 2.1 Hz, 2H), 3.90 (dd, *J* = 4.3, 2.1 Hz, 2H), 2.63 (dd, *J* = 8.2, 7.4 Hz, 2H), 1.64 (m, 2H), 1.35 (m, 2H), 0.91 (t, J = 7.4 Hz, 3H). ¹³C-NMR (101 MHz, CD₃CN) δ 151.6, 140.1, 139.9, 129.2, 128.8, 128.7, 128.4, 128.3, 84.3, 77.0, 75.6, 73.1, 31.5, 29.0, 23.1,

14.1. IR (neat, cm⁻¹): v 3030, 2870, 1454, 1355, 1058. HRMS (ESI) m/z: $[M+H]^+$ calc for $C_{39}H_{43}N_2O_4$ 603.32173, found 603.32178.

Compound 9 (conduritol B-2-butyl-1*H***-imidazole)**

Starting from **S7** (46 mg, 76.3 μmol) and following **GP6**, the pure product was afforded as a white solid (23 mg, quant.). ¹H-NMR (400 MHz, MeOD) δ 4.48 (s, 2H), 3.45 (s, 2H), 2.82 (t, *J* = 7.3 Hz, 2H), 1.70 – 1.54 (m, 2H), 1.27 (q, *J* = 7.1 Hz, 2H), 0.86 (t, J = 7.1 Hz, 3H). ¹³C-NMR (101 MHz, MeOD) δ 151.3, 129.1, 78.1, 68.4, 30.8, 26.6, 23.0, 13.8. HRMS (ESI-TOF) m/z: [M+Na]⁺ calc for $C_{11}H_{18}N_2O_4$ 265.1159, found 265.1172.

1H- and 13C-NMR spectra

p*K***AH determination**

The pK_{AH} of the imidazoles were determined with the method described by Gift *et al.*^[8] using a Metrohm 691 pH-meter and Hamilton spintrode. The compound was dissolved in D_2O (0.6 mL) and basified with NaOD (0.1 M in D_2O) to pH > 8. Then, the mixture was acidified by stepwise addition of DCl (0.1 M in D₂O) and a ¹H-NMR spectrum (Brüker DMX-300) was recorded after each addition. A correction for determination in D₂O instead of H₂O was applied according to Kręzel *et al*.^[9]

Figure S1 p K_{AH} determination of gluco-1H-imidazole by 1 H-NMR.

7.85 7.80 7.75 7.70 7.65 7.60 7.55 7.50 7.45 7.40 7.35 7.30 7.25 7.20 7.15 7.10 7.05 7.00 6.95 6.90 6.85 6.80 6.75 6.70 6.65 6.60

glucoimidazole

Figure S2 p K_{AH} determination of glucoimidazole by 1 H-NMR.

Determination of kinetic constant (*K***i) values**

Biochemical and Biological Methods

Enzyme preparations used for IC_{50} and kinetics measurements were as follows: Recombinant human β-glucosidase GBA1 (Cerezyme) and α-glucosidase recombinant human GAA (Myozyme) were obtained from Genzyme, USA. Bacterial β-glucosidase enzymes *Tm*GH1^[10] and *Tx*GH116^[11] were expressed as previously described. β-Glucosidase from almonds was purchased from Sigma Aldrich as lyophilized powder (7.9 U/mg solid). Cellular homogenates of a stable HEK293 over-expressing GBA2 cell line were obtained as previously described^[12] and were pre-incubated for 30 min with 1mM CBE. Proteins were stored in small aliquots at -80 °C until use. *p*-nitrophenyl-β-D-glucopyranoside was purchased from Sigma Aldrich, 4-MU-β-d-glucopyranoside was purchased from Glycosynth, and C6- NBD-ceramide (6-[N-methyl-N-(7-nitrobenz-2-oxa-1,3-diazol-4-yl)aminododecanoyl]sphingosine) from Molecular probes, GBA1 inhibitor Conduritol-β-Epoxide (CBE) was purchased from Enzo. 2,4 dinitrophenyl-β-D-glucopyranoside^[13] and 2,4-dinitrophenyl-α-D-glucopyranoside^[14] were synthesized following synthetic procedures previously described and their spectroscopic data are in agreement with those previously reported.

In vitro **apparent IC50 measurements**

To determine *in vitro* apparent IC₅₀ values, 25 µL of enzyme solution was pre-incubated with 25 µL of a range of 6 inhibitor dilutions for 30 min in a 96 well plate, using the following buffers: GBA1 in 150 mM McIlvaine buffer pH 5.2, 0.2% taurocholate (w/v), 0.1% Triton X-100 (v/v) and 0.1% bovine serum albumin (BSA) (w/v); GAA in 150 mM McIlvaine buffer pH 4.8 and 0.1% BSA (w/v); *Tm*GH1, *Tx*GH116 and β-glucosidase from sweet almonds in 50 mM NaHPO₄ pH 6.8 and 0.1% BSA (w/v).

After 30 min of pre-incubation, 50 µL of substrate solution in the same buffer was added to this E (25 μ L) + I (25 μ L) mixture (total reaction volume 100 μ L). GBA1 residual activity was measured using final 24 nM concentration of enzyme (Cerezyme) and 200 µM of 2,4-dinitrophenyl-β-Dglucopyranoside substrate, incubated for 30 min at 37 °C. GAA activity was measured using final concentrations of 156 nM and 200 µM of 2,4-dinitrophenyl-α-D-glucopyranoside substrate, for 30 min at 37 °C. *Tm*GH1, *Tx*GH116 and β-glucosidase from sweet almonds residual activity was measured using final concentrations of 37 nM, 82 nM and 0.125 U/mL respectively and 400 µM of *p*nitrophenyl-β-D-glucopyranoside, for 30 min at 37 °C. Finally, all enzyme reactions were monitored for 10 minutes and the release of 2,4-dinitrophenolate or *p*-nitrophenolate and UV-absorbance was measured at 420 nm in a Tecan GENios Microplate Reader. Values plotted for [I] are those in the final reaction mixture, containing $E + I + S$. Data was corrected for background absorbance, then normalized to the untreated control condition and finally curve-fitted via one phase exponential decay function (GraphPad Prism 5.0). Apparent in vitro IC₅₀ values were determined in technical triplicates.

For GBA2, 12.5 µL of lysate was pre-incubated with 12.5 µL of a range of 7 inhibitor dilutions for 30 min at 37°C. Afterwards, 100 µL of 3.7 mM 4-MU-β-d-glucopyranoside in 150 mM McIlvaine buffer pH 5.8 and 0.1% BSA (w/v) were added and incubated for 1h at 37°C. After stopping the substrate reaction with 200 µL 1M NaOH-Glycine (pH 10.3), liberated 4-MU fluorescence was measured with a fluorimeter LS55 (Perkin Elmer) using $\lambda_{\epsilon x}$ 366 nm and $\lambda_{\epsilon m}$ 445 nm. All IC₅₀ values were determined in duplicate.

In situ **apparent IC50 measurements**

IC50 values for GCS were determined with NBD-ceramide as substrate as previously described $^{[15]}$. RAW 264.7 (American Type culture collection) were cultured in RPMI medium (Gibco) supplemented with 10% FCS, 1 mM GlutaMAX™ and 100 units/mL penicillin/streptomycin (Gibco) at 37°C and 5% CO2. The RAW 264.7 cells were grown to confluence in 12-well plates and pre-incubated for 1h with 300 μ M CBE, followed by 1h incubation at 37°C in the presence of a range of 6 inhibitor concentrations and with 1 nmol C6-NBD-ceramide. The cells were washed 3x with PBS and harvested by scraping. After lipid extraction^[16], the C6-NBD lipids were separated and detected by HPLC (λ_{Ex} 470 nm and λ_{Em} 530 nm). IC₅₀ values were determined in duplicate from the titration curves of observed formed C6-NBD-glucosylceramide.

All reported values are the mean from two or three technical replicates.

Kinetic studies

The kinetic studies of reversible imidazole inhibitors in *Tm*GH1, *Tx*GH116 and β-glucosidase from sweet almonds were performed by monitoring the UV-absorbance of *p*-nitrophenolate released from *p*-nitrophenyl β-D-glucopyranoside. *Tm*GH1, *Tx*GH116 and β-glucosidase from sweet almonds (25 µL) at 37 nM, 82 nM and 0.125 U/mL respectively in 50 mM phosphate buffer (pH 6.8) and 0.1% BSA (w/v) were pre-incubated with a range of inhibitor dilutions (25 μ L) for 30 min at 37 °C in a 96 well plate. The reaction was then started by adding 50 µL of different *p*-nitrophenyl β-D-glucopyranoside substrate concentrations (0.05, 0.1, 0.25, 0.5, 0.75, 1.0, 2.5 and 5 mM) in 50 mM phosphate buffer (pH 6.8) to the 50 µL enzyme-inhibitor mixture. For kinetic studies in human recombinant βglucosidase, 25 µL of 24 nM Cerezyme in 150 mM McIlvaine buffer pH 5.2 supplemented with 0.2% taurocholate (w/v), 0.1% Triton X-100 (v/v) and 0.1% bovine serum albumin (BSA) (w/v), was incubated with a range of inhibitor dilutions (25 μ L) for 30 min at 37 °C in a 96 well plate. The reaction was then started by adding 50 µL of different 2,4-dinitrophenyl-α-D-glucopyranoside

substrate concentrations (0.05, 0.1, 0.2, 0.3, 0.4, 0.5, 0.6 and 0.7 mM) in the previously described 150 mM McIlvaine buffer (pH 5.2) to the 50 µL enzyme-inhibitor mixture.

The release of *p*-nitrophenolate or 2,4-dinitrophenolate was monitored by absorbance at 420 nm for 10 min (at 25 °C for *Tm*GH1, *Tx*GH116 and β-Glucosidase from almonds or 37 °C for human Cerezyme and Myozyme) in a Tecan GENios Microplate Reader to determine the hydrolysis rate. The K_i values of reversible competitive or linear mixed inhibition were determined by Michaelis-Menten model using standard nonlinear regression (GraphPad Prism 5.0). K_i values were determined in technical triplicates.

Protein expression and crystallography

*Tm***GH1**

*Tm*GH1 was produced by expression of the construct pET-28a-*Tm*GH1-His₆ and purified as described by Zechel et al.^[17] TmGH1 was crystallized by sitting drop vapour diffusion, with the protein at 10 mg/ml in 50 mM imidazole pH 7.0 and the well solution comprised of 11 % polyethylene glycol (PEG) 4000, 0.1 M imidazole pH 7.0, 50 mM calcium acetate, 100 mM trimethylamine *N*-oxide. The protein drop was seeded with a seed stock grown under similar conditions. To generate the ligand complex, a crystal of *Tm*GH1 was soaked with 10 mM gluco-1*H*-imidazole **6** for 4 days, and fished into liquid nitrogen via a cryoprotectant solution comprised of the well solution supplemented with 25 % (v/v) ethylene glycol. Data were collected at Diamond beamline I03, processed using *DIALS*[18] and scaled using *AIMLESS*^[19] to a resolution of 1.7 Å. The structure was solved using 1OD0 without the water molecules as the starting model for *REFMAC*^[20], and refined by manual rebuilding in *Coot*^[21] combined with further cycles of refinement using *REFMAC*. Crystal structure figures were generated using Pymol.

There are two molecules in the asymmetric unit of the *Tm*GH1 crystal structure. **6** is modelled in the active site of chain B only at an occupancy of 0.8, whilst the equivalent site in chain A has been modelled with ethylene glycol in two alternative conformations and two water molecules. The authors have observed that crystal structures of ligand complexes obtained with *Tm*GH1 crystals sometimes yield ligand in only one out of two molecules in the asymmetric unit. It may be that some of the active sites are blocked by N-terminal residues on adjacent chains, as observed for 1OD0.pdb (where 5 residues at the start of chain B extend into the active site of mol A), but for this complex it has not been possible to definitively model N-terminal residues before Val3.

*Tx***GH116**

*Tx*GH116 was produced by expression of construct pET30a-*Tx*GH116 Δ1-18 with a C-terminal His₆ tag and purified as described by Charoenwattanasatien *et al*.^[22] *Tx*GH116 was crystallized by the sitting drop vapour diffusion method, with a well solution of 0.2M ammonium sulfate, 20 % (v/v) PEG 3350,

0.1 M Bis-Tris pH 6.8. To generate ligand complexes, crystals of *Tx*GH116 were soaked with 10 mM gluco-1*H*-imidazole **6** for 20 hours, before fishing via a cryoprotectant solution with 25 % (v/v) ethylene glycol. Data were collected at Diamond beamline I03, processed using *DIALS* and scaled using *AIMLESS* to a resolution of 2.1 Å. The structure was solved using *MOLREP*[23], with 5BVU as the model, and the solved structure refined by cycles of manual rebuilding in *Coot* and refinement using *REFMAC*. Crystal structure figures were generated using Pymol.

Table S2 - Data collection and refinement statistics

Electron density and B-factor comparisons for complexes of 5 and 6

Figure S3 Electron density and B-factor comparisons for complexes of **5** and **6**. **a** Superposed electron densities for **5** and **6** in complex with *Tm*GH1 (pink ligand for **5**, cyan for **6**; chain B of each structure) and *Tx*GH116 (salmon for **5**, blue for **6**). The small 'upwards' shift at the apical carbon of the imidazole in complexes of **6** compared to **5** is well supported by the diverging electron densities at this region. In contrast, electron densities overlay well in the 'glucose' portion of the ligands. Densities shown are REFMAC maximum-likelihood/ σ_A weighted 2Fo-Fc contoured between 1.5–2.0 r.m.s.d (0.38–0.48 e⁻/Å³ for *Tm*GH1-**5**, *Tm*GH1–**6**, *Tx*GH116-**6**; 0.89 e- /Å³ for *Tx*GH116-**5**). **b** Superposition of *Tm*GH1-**6** (cyan), against **5** from chains A (orange) and B (pink) of the *Tm*GH1-**5** complex. **6** shows a clear 'upwards' shift compared to both molecules of **5**, which overlay well with each other. **c** Ligands in complex with *Tm*GH1 and *Tx*GH116 colored by B-factor, with B-factors of peripheral atoms annotated. B-factors increase substantially towards the imidazole portion of **6** in both *Tm*GH1 and *Tx*GH116 complexes, indicating greater crystallographic disorder at this region of the ligand. B-factors are more consistent in complexes with **5**. The *Tm*GH1-**5** ligand shown is from chain B; the ligand from chain A shows a similar B-factor trend.

ITC

ITC experiments were carried out using a MicroCal AutoITC200 (Malvern Instruments, formerly GE Healthcare). All titrations were run at 25 °C in 50 mM Sodium Phosphate, pH 5.8 or 6.8. Proteins were buffer exchanged into ITC buffer via at least 3 rounds of dilution/concentration using an Amicon Ultra spin concentrator (Millipore), and further degassed under vacuum prior to use. Cell concentrations of 100 μ M (protein) and syringe concentrations of 2 mM (ligand) were used for titrations using gluco-1*H*-Imidazoles **6** and **7**. Cell concentrations of 50 µM and syringe concentrations of 500 µM were used for titrations using **5**. Analyses were carried out using the MicroCal PEAQ-ITC analysis software (Malvern Instruments).

All reported values are the mean ± standard deviation from three (ligands **6**, **7**) or four (ligand **5**) technical replicates.

Representative ITC traces pH 6.8

Representative ITC traces pH 5.8

DFT calculations

Geometry optimization

All calculations were performed with DFT as level of theory in combination with the B3LYP hybrid functional. A conformer distribution search option included in the Spartan 04 program^[24], in gasphase with the use of 6-31G(d) as basis set, was used as starting point for the geometry optimization. All generated structures were further optimized with Gaussian $03^{[25]}$ at 6-311G(d,p). Optimization was done in gas-phase and subsequently corrections for solvent effects were done by the use of a polarizable continuum model using water as solvent parameter. The free Gibbs energy of the computed conformations was calculated using Equation (1) in which ΔE_{gas} is the gas-phase energy (electronic energy), ΔG_{RRHO}^T (T= 298.15 K and pressure= 1 atm.) is the sum of corrections from the electronic energy to free Gibbs energy in the rigid-rotor-harmonic-oscillator approximation (RRHO) also including zero-point-vibrational energy, and ΔG_{Solv}^{T} is their corresponding free solvation Gibbs energy.

$$
\Delta G_{aq}^T = \Delta E_{gas} + \Delta G_{gas,RRHO}^T + \Delta G_{solv}
$$
\n
$$
= \Delta G_{gas}^T + \Delta G_{solv}
$$
\n(1)

The denoted free energies include unscaled zero-point vibrational energies. Visualisation of the conformations of interest was done with CYLview.[25] The three lowest energy geometries for **6** were all calculated to adopt a ${}^{4}H_3$ conformation, differing only in the rotation angle around the C5-C6 axis: *gt*, *gg* and *tg* respectively.

Gluco-1*H***-imidazole (both tautomers)**

Gluco-1*H***-imidazole (protonated)**

Mulliken charges

Mulliken atomic charges of lowest energy geometry of **6** (Gluco-1*H*-imidazole)

 1 C 0.061369 2 C 0.104400 3 C 0.042436 **4 C 0.106302** 5 C -0.195326 6 C 0.055654 7 H 0.143586 8 H 0.136044 9 H 0.121522 10 H 0.125417 11 N -0.429194 12 N -0.386821 **13 C 0.145851** 14 H 0.152469 15 H 0.306533 16 C 0.043782 17 H 0.092401 18 H 0.107852 19 O -0.463283 20 H 0.244617 21 O -0.463231 22 H 0.283708 23 O -0.472542 24 H 0.297766 25 O -0.452488 26 H 0.291176

Mulliken atomic charges of lowest energy geometry of **6** (Gluco-1*H*-imidazole (protonated))

 1 C 0.052501 2 C 0.125295 3 C 0.177257 **4 C 0.112704** 5 C -0.186080 6 C 0.049958 7 H 0.162319 8 H 0.172471 9 H 0.135404 10 H 0.143009 11 C 0.037413

- 12 H 0.117263
- 13 H 0.119184
- 14 O -0.475253
- 15 H 0.314409
- 16 O -0.463192
- 17 H 0.302591
- 18 O -0.470192
- 19 H 0.306331
- 20 O -0.462279
- 21 H 0.307617
- 22 N -0.356638
- 23 H 0.319731
- 24 N -0.361151
- 25 H 0.343516
- **26 C 0.263310**
- 27 H 0.212501

Mulliken atomic charges of lowest energy geometry of **5** (Glucoimidazole)

- 1 C 0.003047
- 2 C 0.078107
- 3 C 0.007354
- 4 C 0.061292
- 5 H 0.143840
- 6 H 0.168935
- 7 H 0.127926
-
- 8 H 0.141160
- 9 C 0.017720
- 10 H 0.126707
- 11 H 0.124005
- 12 O -0.453463
- 13 H 0.300757
- 14 O -0.452985
- 15 H 0.293432
-
- 16 O -0.471394
- 17 H 0.303466
- 18 O -0.463576
- 19 H 0.295509
- 20 N -0.395317
- **21 C 0.306442**
- 22 C -0.000203

- 23 H 0.132330
- **24 C -0.117443**
- 25 H 0.125825
- 26 N -0.403472

Mulliken atomic charges of lowest energy geometry of **5** (Glucoimidazole (protonated))

Restricted conformational energy surface calculations

The geometry with the lowest free Gibbs energy was selected as the starting point for the partial conformational energy surface calculation. A survey of the possible neighbouring conformational space was made by scanning two dihedral angles, including the C1-C2-C3-C4 (D1), C3-C4-C5-O (D3) ranging from −60° to −20°. The C5-O-C1-C2 (D5) was fixed at 0° since this is highly favoured. The resolution of this survey is determined by the step size which was set to 5° per puckering parameter. These structures were calculated with Gaussian 03 with a $6-311G(d,p)$ as basis set. Furthermore, solvation effects of H_2O were taken into account with a polarizable continuum model function.

Gluco-1*H***-imidazole-protonated (6)**

Glucoimidazole (5)

Glucoimidazole-protonated (5)

NMR calculations

Based on the optimized structures the spin-spin coupling constants were calculated according to the work of Rablen and Bally^[26] with the use of 6-311g(d,p) u+1s as basis set and PCM(H₂O) as solvent model. The calculated total nuclear spin-spin coupling terms were used as calculated spin-spin coupling constants. The calculated ${}^{3}J_{(H,H)}$ coupling constants for these low energy ${}^{4}H_3$ rotamers of 6 matched well with experimental ${}^{3}J_{(H,H)}$ coupling constants, suggesting that that 6 most likely adopts a $^{4}H_{3}$ conformation in solution.

Gluco-1*H***-imidazole**

Gluco-1*H***-imidazole (protonated)**

gt (0.0 kcal/mol) *gg* (+0.0 kcal/mol) *tg* (+2.3 kcal/mol)

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