

Supplementary Information for:

The Divergent Effects of Strong NHC Donation in Catalysis

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S1. NOESY spectra for *s*-**GIIm**, *u*-**GIIm**, *s*-**GII**, and *u*-**GII**

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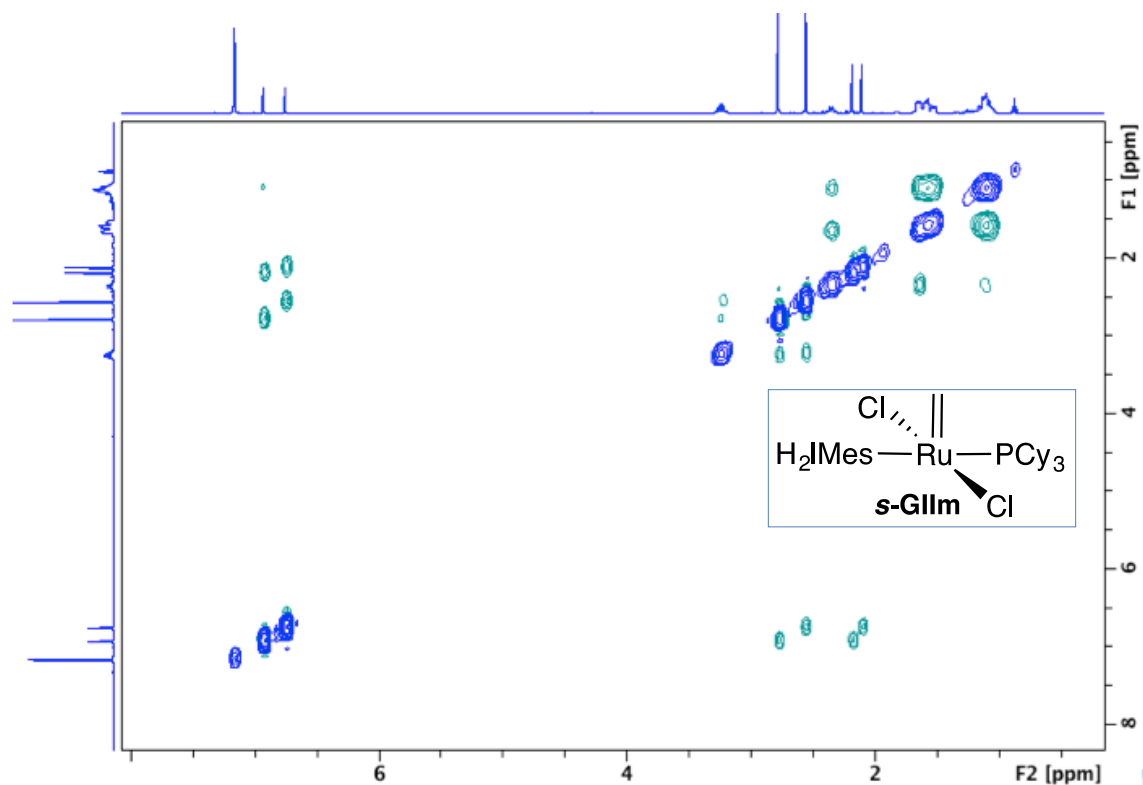


Figure S1. ^1H NMR NOESY spectrum (500.1 MHz, C_6D_6) for methyldene complex $\text{RuCl}_2(\text{H}_2\text{IMes})(\text{PCy}_3)(=\text{CH}_2)$, *s*-GIIIm.

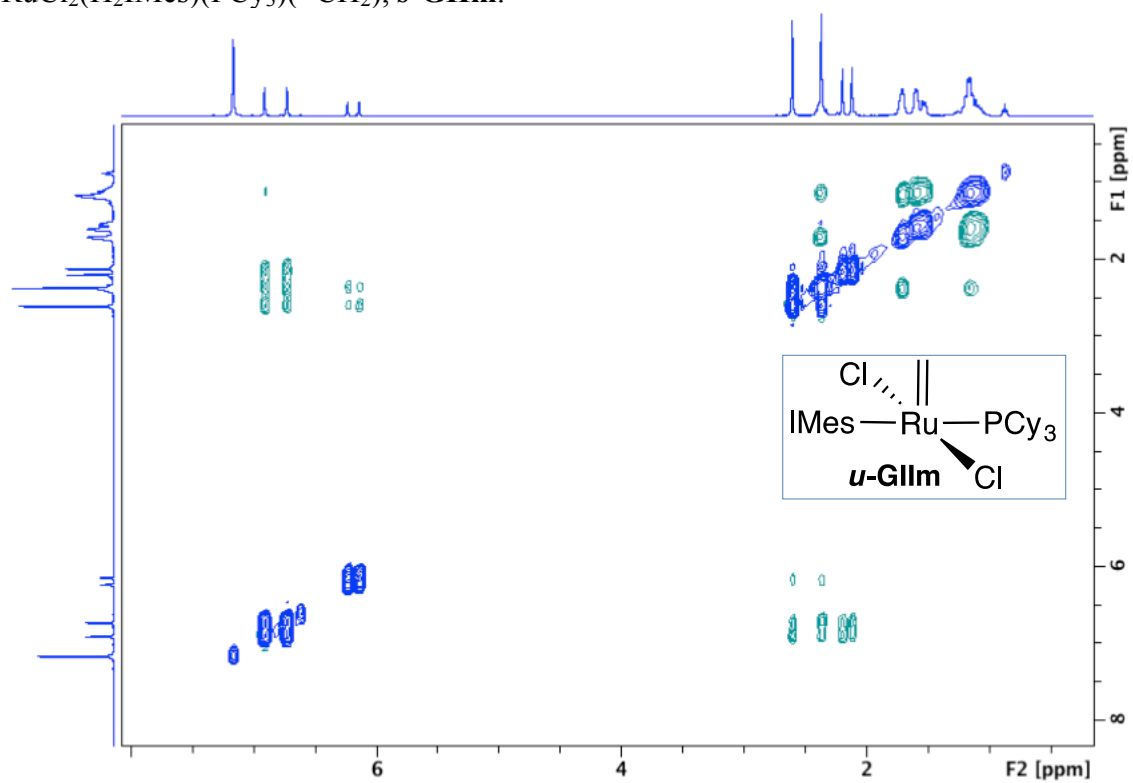


Figure S2. ^1H NMR NOESY spectrum (500.1 MHz, C_6D_6) for $\text{RuCl}_2(\text{IMes})(\text{PCy}_3)(=\text{CH}_2)$, *u*-GIIIm.

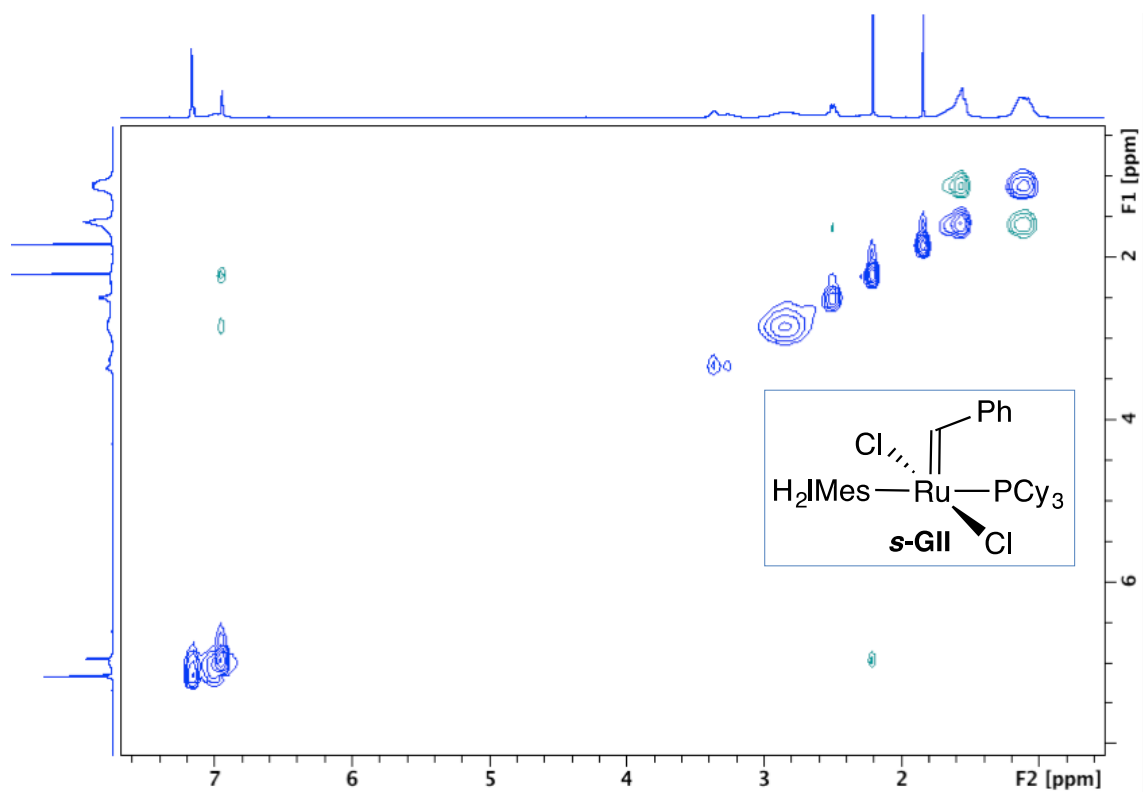


Figure S3. ^1H NMR NOESY spectrum (500.1 MHz, C_6D_6) for $\text{RuCl}_2(\text{H}_2\text{IMes})(\text{PCy}_3)(=\text{CHPh})$, *s*-GII.

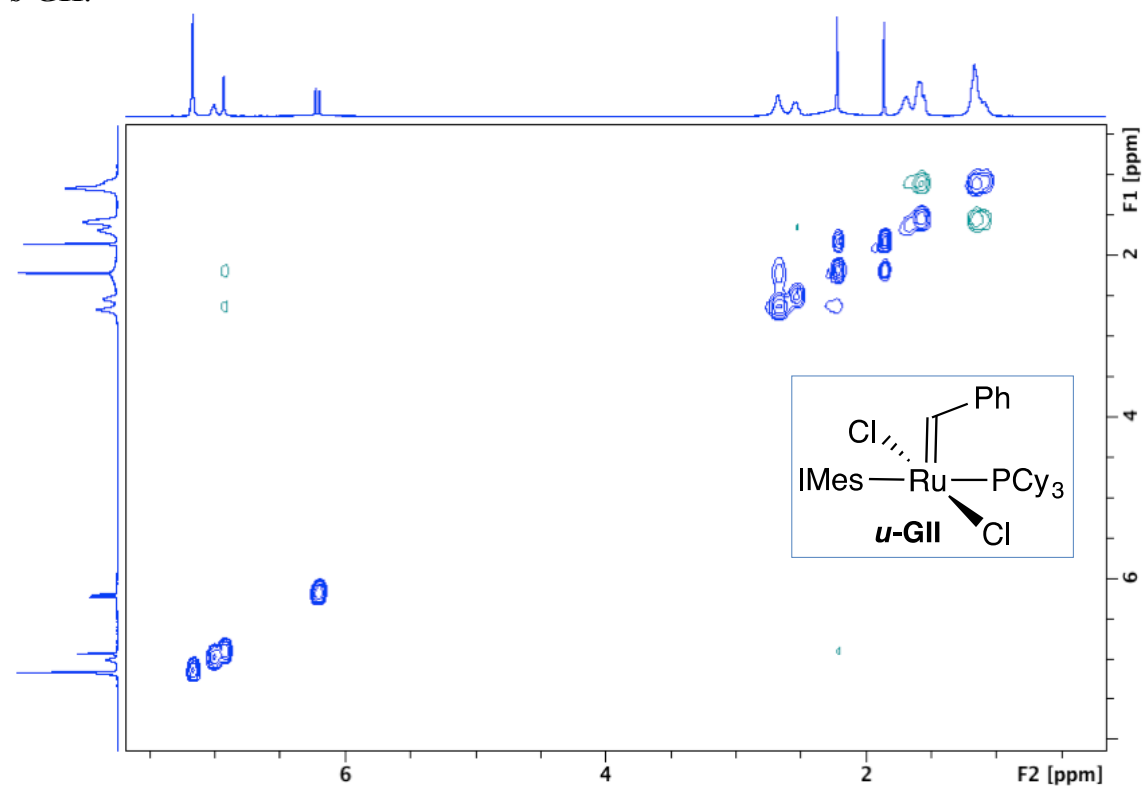


Figure S4. ^1H NMR NOESY spectrum (500.1 MHz, C_6D_6) for $\text{RuCl}_2(\text{IMes})(\text{PCy}_3)(=\text{CHPh})$, *u*-GII.

S2. Inducing rotation in *s*-GII by raising the temperature

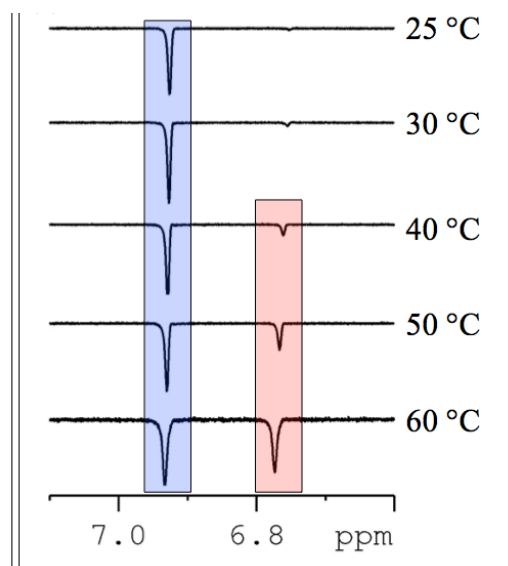
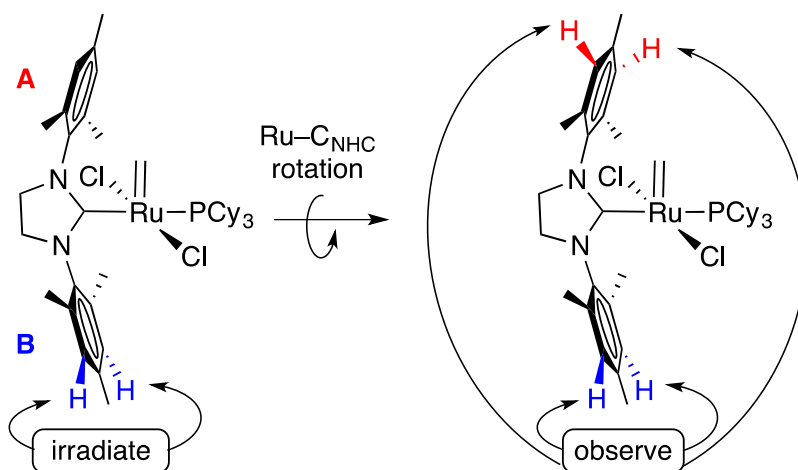
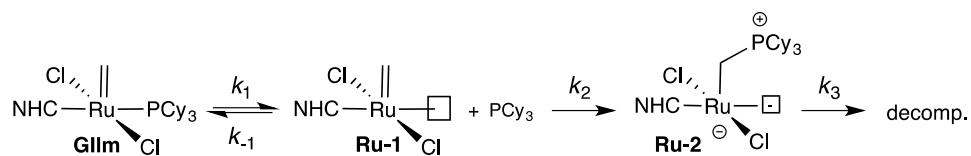


Figure S5. Inducing NHC rotation in *s-GIIIm*. (a) Distinguishing mesityl rings by selective irradiation of the Mes CH signal for Ring **B**. (b) Onset of EXSY enhancement of CH signals in Ring **A** at 40 °C. ^1H NMR, 500 MHz, C_6D_6 ; 2 s mixing.

S3. Derivation of the [PCy₃] dependence of GIIIm decomposition

(a) Dissociative pathway



$$\begin{aligned} \text{decomp. rate} &= -d[\text{GIIIm}] / dt \\ &= k_3 [\text{Ru-2}] \end{aligned}$$

Applying the steady state approximation to spectroscopically unobservable **Ru-2**,

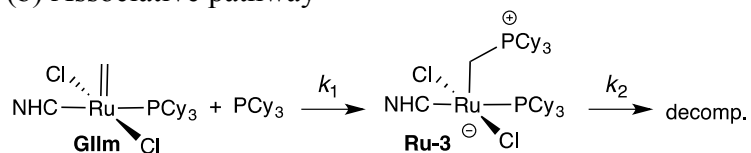
$$\begin{aligned} k_3 [\text{Ru-2}] &= k_2 [\text{Ru-1}] [\text{PCy}_3] \\ [\text{Ru-2}] &= k_2 / k_3 [\text{Ru-1}] [\text{PCy}_3] \\ \text{then rate} &= k_2 [\text{Ru-1}] [\text{PCy}_3] \end{aligned}$$

Applying the steady state approximation to spectroscopically unobservable **Ru-1**,

$$\begin{aligned} k_1 [\text{GIIIm}] &= k_{-1} [\text{Ru-1}] [\text{PCy}_3] + k_2 [\text{Ru-1}] [\text{PCy}_3] \\ [\text{Ru-1}] &= \frac{k_1 [\text{GIIIm}]}{(k_{-1} + k_2) [\text{PCy}_3]} \\ \text{then rate} &= \frac{k_1 k_2 [\text{GIIIm}] [\text{PCy}_3]}{(k_{-1} + k_2) [\text{PCy}_3]} \\ &= \frac{k_1 k_2}{(k_{-1} + k_2)} [\text{GIIIm}] \end{aligned}$$

Thus the rate of decomposition is independent of [PCy₃]

(b) Associative pathway



$$\begin{aligned} \text{decomp. rate} &= -d[\text{GIIIm}] / dt \\ &= k_2 [\text{Ru-3}] \end{aligned}$$

Applying the steady state approximation to spectroscopically unobservable **Ru-3**,

$$\begin{aligned} k_2 [\text{Ru-3}] &= k_1 [\text{GIIIm}] [\text{PCy}_3] \\ [\text{Ru-3}] &= k_1 / k_2 [\text{GIIIm}] [\text{PCy}_3] \\ \text{then rate} &= k_1 [\text{GIIIm}] [\text{PCy}_3] \end{aligned}$$

Thus the rate of decomposition has a first-order dependence on [PCy₃]

S4. Rate profiles for PCy₃ loss from *s*-GIIIm and *u*-GIIIm at 40 °C and 80 °C

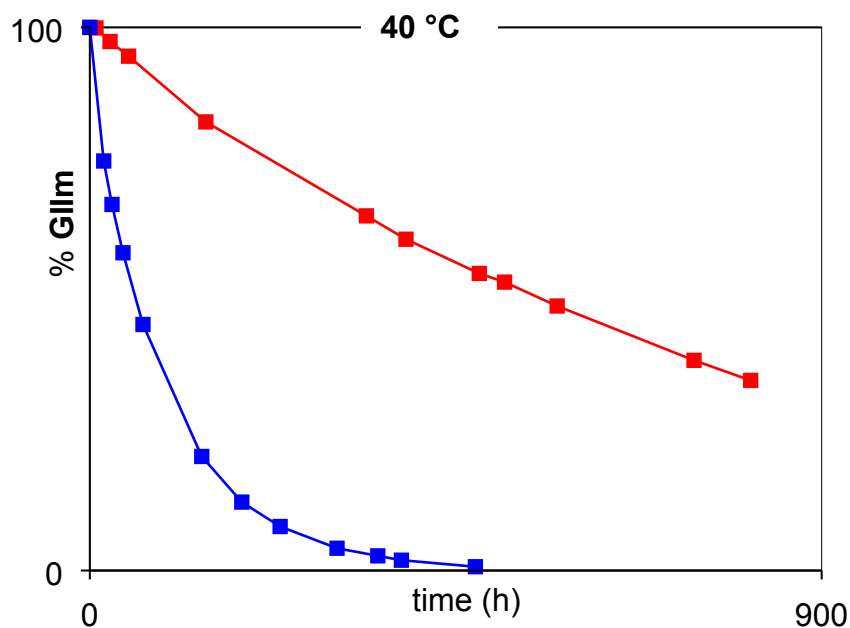


Figure S6. Assessing the stability of *s*-GIIIm (blue) and *u*-GIIIm (red) at 40 °C in C₆D₆ (¹H NMR integration vs. TMB as internal standard; measuring decreases in integration following t₀).

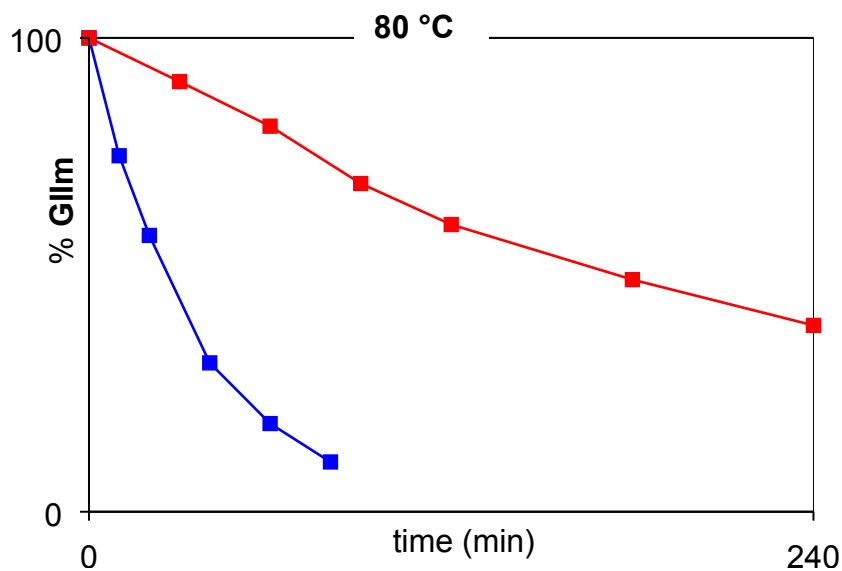


Figure S7. Assessing the stability of *s*-GIIIm (blue) and *u*-GIIIm (red) at 80 °C in C₆D₆ (¹H NMR integration vs. TMB as internal standard; measuring decreases in integration following t₀).

S5. Half-lives for *s*-GII_m and *u*-GII_m at 40 °C, 60 °C, and 80 °C in C₆D₆

Table S1. Half-lives data corresponding to the thermolysis experiments of Fig. 4.^a

T (°C)	<i>u</i> -GII _m	<i>s</i> -GII _m
40	545	63
60	34	4.1
80	2.9	0.4

^aAssessed by ¹H NMR: 300 MHz, TMB used as internal standard

S6. Key chemical shifts for GII_m and GII in common solvents

Table 2. Diagnostic ¹H and ³¹P chemical shifts for GII_m and GII complexes.^a

Solvent	δ_{H} (Ru=CHR)		δ_{P} (Ru-PCy ₃)	
	<i>u</i> -GII _m	<i>s</i> -GII _m	<i>u</i> -GII _m	<i>s</i> -GII _m
R = H				
C ₆ D ₆	18.77	18.42	40.8	38.2
C ₇ D ₈	18.65	18.29	40.7	38.0
CD ₂ Cl ₂	18.09	17.73	39.9	37.1
CDCl ₃	18.15	17.78	39.2	36.2
THF-d ₈	18.21	17.86	40.3	37.6
R = Ph				
C ₆ D ₆	19.94	19.65	32.0	30.1
C ₇ D ₈	19.84	19.55	32.0	30.0
CD ₂ Cl ₂	19.38	19.08	32.1	29.9
CDCl ₃	19.43	19.13	31.8	29.6
THF-d ₈	19.55	19.25	31.9	29.9

^a 300.1 MHz (¹H NMR) or 121.6 MHz (³¹P{¹H} NMR); 22 °C.

7. Crystallographic details for *u*-GIIIm.

Table S2. Crystallographic Details for *u*-GIIIm.

<i>A. Crystal data</i>	
formula	C ₄₀ H ₅₉ Cl ₂ N ₂ PRu
formula weight	770.83
crystal dimensions (mm)	0.16 × 0.14 × 0.02
crystal system	monoclinic
space group	<i>P</i> 2 ₁ / <i>n</i> (an alternate setting of <i>P</i> 2 ₁ / <i>c</i> [No. 14])
unit cell parameters ^a	
<i>a</i> (Å)	12.2117 (3)
<i>b</i> (Å)	18.0562 (6)
<i>c</i> (Å)	18.2926 (6)
β (deg)	103.741 (2)
<i>V</i> (Å ³)	3918.0 (2)
<i>Z</i>	4
ρ _{calcd} (g cm ⁻³)	1.307
μ (mm ⁻¹)	5.088
<i>B. Data collection and refinement</i>	
Diffractionmeter	Bruker D8/APEX II CCD ^b
radiation (λ [Å])	Cu Kα (1.54178) (microfocus source)
temperature (°C)	-100
scan type	ω and φ scans (1.0°) (5 s exposures)
data collection 2θ limit (deg)	135.98
total data collected	25622 (-14 ≤ <i>h</i> ≤ 14, -21 ≤ <i>k</i> ≤ 21, -21 ≤ <i>l</i> ≤ 21)
independent reflections	6912 (<i>R</i> _{int} = 0.1444)
number of observed reflections (<i>NO</i>)	4015 [<i>F</i> _o ² ≥ 2σ(<i>F</i> _o ²)]
structure solution method	Patterson/structure expansion (<i>DIRDIF-2008</i>) ¹
refinement method	full-matrix least-squares on <i>F</i> ² (<i>SHELXL-97</i>) ²
absorption correction method	Gaussian integration (face-indexed)
range of transmission factors	0.8876–0.4910
data/restraints/parameters	6912 / 1 ^c / 429
extinction coefficient (<i>x</i>) ^d	0.00016(3)
goodness-of-fit (<i>S</i>) ^e [all data]	0.954
final <i>R</i> indices ^f	
<i>R</i> ₁ [<i>F</i> _o ² ≥ 2σ(<i>F</i> _o ²)]	0.0517
<i>wR</i> ₂ [all data]	0.0517
largest difference peak and hole	0.499 and -0.699 e Å ⁻³

^aObtained from least-squares refinement of 3223 reflections with 7.92° < 2θ < 105.86°.

^bPrograms for diffractometer operation, data collection, data reduction and absorption correction were those supplied by Bruker.

^cThe C1–H1A and C1–H1B distances were constrained to be equal (within 0.06 Å) during refinement.

^d*F*_c^{*} = *kF*_c [1 + *x*{0.001*F*_c²λ³/sin(2θ)}]^{-1/4} where *k* is the overall scale factor.

^e*S* = [Σ*w*(*F*_o² - *F*_c²)²/(*n* - *p*)]^{1/2} (*n* = number of data; *p* = number of parameters varied; *w* = [σ²(*F*_o²) + (0.0490*P*)²]⁻¹ where *P* = [Max(*F*_o², 0) + 2*F*_c²]/3).

^f*R*₁ = Σ||*F*_o|| - |*F*_c||/Σ||*F*_o||; *wR*₂ = [Σ*w*(*F*_o² - *F*_c²)²/Σ*w*(*F*_o⁴)]^{1/2}.

8. References

- (1) Beurskens, P. T.; Beurskens, G.; de Gelder, R.; Smits, J. M. M.; Garcia-Granda, S.; Gould, R. O. *The DIRDIF-2008 Program System*, Crystallography Laboratory, Radboud University: Nijmegen, The Netherlands, 2008.
- (2) Sheldrick, G. M. *Acta Crystallogr.* **2008**, *A64*, 112–122.