

Supporting Information

Hierarchical Assembly of an Interlocked M_8L_{16} Container

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Author Contributions

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Contents

1. Experimental section

1.2. Materials and measurements

Unless otherwise stated, all chemicals were obtained from commercial sources and used as received. 3,6-Dibromo-9,10-dihexyloxyphenanthrene,¹ 9,10-dipropoxyphenanthrene and 9,10-dimethoxy-3,6-bis(4-pyridyl)phenanthrene² were prepared according to literature procedures. GPC purifications were performed on a JAI 9210-II NEXT GPC System with a JAIGEL HH-2/HH-1 column combination running with CHCI3 (HPLC grade). Infrared spectra were collected on a Pelkin Elmer Spectrum Two spectrometer. Electrospray ionization (ESI) mass spectra were recorded on a Bruker Apex IV ESI-FTICR Mass Spectrometer with a dual electrospray ionization source. TEM measurements were performed on a Philips CM200 TEM high-resolution Transmission Electron Microscope equipped with an EDAX EDS system and Gatan 678 Image Filter and P/EELS, and Gatan 832 SC1000 CCD camera. DLS measurements were carried on a Malvern Zetasizer ZS equipped with a standard 633 nm laser. DLS samples were measured in acetonitrile in a 10 mm quartz cuvette. For Variable Temperature (VT) DLS measurements, the viscosities of CH3CN at different temperatures were extracted from published data.³ The peak size (d.nm) and polydispersity index (PDI) were determined from the autocorrelation functions using the Zetasizer software provided by the manufacturer.

1.3. Synthesis of ligands

Scheme 1

1.3.1. Synthesis of 3,6-dibromo-9,10-dipropoxyphenanthrene

The synthesis of 3,6-dibromo-9,10-dipropoxyphenanthrene was carried out according to a procedure described by Mazal and co-workers.⁴ 3,6-Dibromo-9,10-phenanthrenequinone (1.10 g, 3.00 mmol), Bu₄NBr (1.10 g, 3.41 mmol), and Na₂S₂O₄ (6.10 g, 35.0 mmol) were dissolved in aqueous THF (1:1, 44 mL) and stirred for 1 h. 1-Bromopropane (2.87 mL, 31.5 mmol) was added, followed by KOH (4.4 g, 78.4 mmol) in H₂O (22 mL). After stirring for 2 days at 25 °C, the reaction mixture was diluted with H₂O (75mL) and extracted with dichloromethane (3 \times 30 mL). The extracts were dried over MgSO4 and the solvent was removed under reduced pressure. The crude product was purified by column chromatography (CH₂Cl₂) and recrystallized from pentane to yield an off-white solid (0.92 g, 67%). Mp: 104.0 - 105.2 °C. ¹H NMR (500 MHz, CDCl₃) δ 8.65 (d, J = 1.8 Hz, 2H), 8.11 (d, J = 8.7 Hz, 2H), 7.71 (dd, J = 8.7, 1.8 Hz, 2H), 4.16 (t, J = 6.7 Hz, 4H), 1.91 (h, J = 7.3 Hz, 4H), 1.12 (t, J = 7.3 Hz, 6H). ¹³C NMR (150 MHz, CDCl₃) δ 143.3, 130.6, 129.0, 128.9, 125.5, 124.4, 120.5, 75.4, 23.8, 10.9. ESI-MS (C₂₀H₂₀O₂Br₂) calc: 450.9858 [M+H]⁺; found: 450.9796.

Figure S1. ¹H NMR spectrum (600 MHz/CDCl₃) of 3,6-dibromo-9,10-dipropoxyphenanthrene.

Figure S2. ¹³C NMR spectrum (150 MHz/ CDCl₃) of 3,6-dibromo-9,10-dipropoxyphenanthrene.

1.3.2. Synthesis of 9,10-dipropoxy-3,6-bis(4-pyridyl)phenanthrene (\mathbf{L}^2)

3,6-Dibromo-9,10-dipropoxyphenanthrene (437 mg, 0.966 mmol), 4-pyridineboronic acid pinacol ester (594 mg, 2.90 mmol), K₃PO₄ (1.67, 7.9 mmol) and dioxane (32 mL) were combined in a schlenk vessel. Once the solvent was degassed (via freeze-thaw cycles), Pd(PPh₃)₄ (114 mg, 0.10 mmol) was added and the mixture was heated at 100 °C for 24 h. Once allowed to cool, water (40 mL) was added and the mixture was extracted with chloroform (4 \times 15 mL). The combined organic layers were dried over MgSO4 and the solvent was removed under reduced pressure. The residue was purified by column chromatography (CH₂Cl₂, 15:1 CH₂Cl₂/MeOH) and then further purified by GPC to yield the title compound as a white crystalline solid (185 mg, 42%). Mp: 160.6 - 162.7 °C; v_{max} (neat, cm⁻¹): 2965 (w), 1592 (s), 1540 (w), 1440 (s), 1331 (m); ¹H NMR (600 MHz, CD₃CN) δ 9.18 (d, J = 1.8 Hz, 2H, H_c), 8.73 – 8.70 (m, 4H, H_e), 8.39 (d, J = 8.5 Hz, 2H, H_a), 8.03 (dd, J = 8.5, 1.8 Hz, 2H, H_b), 7.90 – 7.87 (m, 4H, H_d), 4.23 (t, J = 6.6 Hz, 4H, N-CH₂), 1.99 – 1.93 (m, 4H, CH₂), 1.15 (t, J = 7.4 Hz, 6H, CH₃); ¹³C NMR (150 MHz, CD₃CN) δ 150.9, 148.3, 144.3, 136.0, 130.8, 129.4, 126.5, 123.9, 122.5, 122.4, 75.7, 24.0, 10.7. ESI-MS (C₃₆H₄₀N₂O₂) calc: 449.2224 [L²+H]⁺; found: 449.2284.

Figure S3. Partial ¹H NMR spectrum (600 MHz/CD₃CN) of L².

Figure S4. 13 C NMR spectrum (150 MHz/CD₃CN) of L^2 .

1.3.3. Synthesis of 9,10-dihexyloxy-3,6-bis(4-pyridyl)phenanthrene (\mathbf{L}^3)

3,6-Dibromo-9,10-dihexyloxyphenanthrene (300 mg, 0.557 mmol), 4-pyridineboronic acid pinacol ester (342 mg, 1.67 mmol), K_3PQ_4 (0.97 g, 4.55 mmol) and dioxane (19 mL) were combined in a schlenk vessel. Once the solvent was degassed (via freeze-thaw cycles), Pd(PPh₃)₄ (65.4 mg, 0.056 mmol) was added and the mixture was heated at 100 °C for 24 h. Once allowed to cool, water (50 mL) was added and the mixture was extracted with chloroform (4×20 mL). The combined organic layers were dried over MgSO₄ and the solvent was removed under reduced pressure. The residue was purified by column chromatography (CHCl₃, 20:1 CHCl₃/MeOH) and then further purified by GPC to yield the title compound as an off-white solid (150.8 mg, 53 %). Mp: 112.6 – 113.4 °C; v_{max} (neat, cm⁻¹): 2942 (m), 1594 (s), 1539 (w), 1439 (m), 1329 (m); ¹H NMR (500 MHz, CD₃CN) δ 9.18 (d, J = 1.8 Hz, 2H, H_c), 8.77 – 8.66 (m, 4H, H_e), 8.38 (d, J $= 8.5$ Hz, 2H, H_a), 8.03 (dd, J = 8.5, 1.8 Hz, 2H, H_b), 7.93 – 7.85 (m, 4H, H_d), 4.25 (t, J = 6.6 Hz, 4H, CH₂), 1.92 – 1.89 (m, 4H, CH2), 1.67 – 1.55 (m, 4H, CH2), 1.48 – 1.33 (m, 8H, 2 x CH2), 0.98 – 0.89 (m, 6H, CH3).¹³C NMR (150 MHz, CD3CN) δ 151.3, 148.6, 144.6,136.4, 131.2, 129.8, 126.9, 124.3, 122.9, 122.8, 74.6, 32.4, 31.1, 26.6, 23.3, 14.3. ESI-MS (C₃₆H₄₀N₂O₂) calc: 533.3163 [L³+H]⁺; found: 533.3152.

Figure S6. Partial ¹H NMR spectrum (500 MHz/CD₃CN) of L³.

1.4. Synthesis of metallosupramolecular assemblies

1.4.1. Synthesis of a mixture of $Pd_3L^1_{\omega}$ $Pd_4L^1_{\omega}$ (D_{4h}), and $Pd_4L^1_{\omega}$ (D_{2d}) (**1a, 1b** and **1c**)

The synthesis of a mixture of **1a, 1b,** and **1c** was carried out by heating a 2:1 mixture of **L'** and [Pd(CH₃CN)₄](BF₄)₂ in $CD₃CN$ for 2 h. Full experimental details including ¹H NMR and ESI-MS have been previously reported.⁵

Figure S9. ¹H NMR spectra (500 MHz/CD₃CN) of: a) a mixture of 1a, 1b and 1c (approximate ratio 1:2:0.2, respectively); b) a) heated for 24 h at 70 °C. The proton signals corresponding to 1b (green, broad) remain whilst the signals corresponding to 1a (orange) and 1c disappear. Furthermore, signals corresponding to L¹ (blue) appear suggesting that both 1**a** and 1**c** are not thermodynamically stable in CD₃CN; c) L¹. Note: the change from a) to b) is accompanied by a precipitate which does not dissolve in common organic solvents, indicative of a polymeric side-product. Whilst the full proton spectrum of 1c cannot be fully assigned in a) (due to peak overlap), the two methoxy proton signals at δ 4.10 and 3.97 are consistent with the D_{2d} symmetry of the tetrahedron 1c (see section 5). The ESI-MS spectrum of this sample has been previously reported.⁵

Figure S10. ¹H NMR spectra (500 MHz/DMSO) of: a) L¹; b) a 2:1 mixture of L¹ and Pd(NO₃)₂ (hydrate) in DMSO heated for 24 h at 70 °C. Proton signals corresponding to both 1b and L¹ (blue) are present; c) 1b, prepared from L¹ and Pd(CH₃CN)₄](BF₄)₂.² The small discrepancy in chemical shift of 1b in b) and c) is due to the presence of $NO₃$ counter-ions in b) and $BF₄$ counter-ions in c).

1.4.3. Synthesis of $[Pd_8L^1{}_{16}]^{16+}$ (2a)

A solution of palladium nitrate hydrate (368 µL, 15 mM/CD₃CN, 5.5 µmol) was combined with a solution of L¹ (4.21 mg, 10.7 μmol) in CD₃CN (3463 μL) and heated at 70 °C for 24 h to afford [Pd₈L¹16]¹⁶⁺. ¹H NMR (700 MHz, CD₃CN) δ 10.72 $(d, J = 6.1$ Hz, 16H), 10.16 $(d, J = 6.2$ Hz, 16H), 9.81 $(d, J = 5.9$ Hz, 16H), 9.59 $(s, 8H)$, 9.42 $(s, 8H)$, 9.19 $(d, J = 6.0$ Hz, 16H), 9.06 (s, 8H), 8.83 (d, J = 6.1 Hz, 16H), 8.68 (d, J = 6.0 Hz, 16H), 8.54 – 8.46 (m, 24H), 8.43 – 8.38 (m, 16H), 8.33 (d, J = 8.7 Hz, 8H), 8.09 (d, J = 8.8 Hz, 16H), 7.75 (d, J = 8.5 Hz, 8H), 7.26 (d, J = 8.6 Hz, 8H), 6.65 (d, J = 8.7 Hz, 8H), 4.09 (s, 24H), 4.07 (s, 24H), 3.87 (s, 24H), 3.44 (s, 24H). ¹³C NMR (150 MHz, CD3CN/DMSO 2:1): δ 152.4, 152.1, 152.0, 151.5, 151.3, 150.8, 145.3, 145.2, 144.8, 143.8, 135.0, 133.9, 132.6, 132.3, 131.5, 131.2, 130.2, 129.8, 129.3, 129.2, 128.8, 128.2, 127.5, 127.0, 126.8, 126.3, 126.0, 124.9, 124.8, 124.2, 124.2, 124.1, 123.9, 123.8, 123.50, 122.7, 122.4, 122.0, 61.39, 61.37, 61.1, 60.6. ESI HR-MS (C₄₁₆H₃₂₀N₃₂O₃₂Pd₈) calc: 840.5108 [2a+7NO₃]⁹⁺, 953.1982 [2a+8NO₃]⁸⁺, 1098.2248 [2a+9NO₃]⁷⁺, 1291.5939 $[2a+10NO₃]^{6+}$, 1562.3101 $[2a+11NO₃]^{5+}$, 1968.3849 $[2a+12NO₃]⁴⁺$; found: 840.5089 $[2a+7NO₃]⁹⁺$, 953.1991 $[2a+8NO₃]⁸⁺, 1098.2248 [2a+9NO₃]⁷⁺, 1291.5984 [2a+10NO₃]⁶⁺, 1562.3111 [2a+11NO₃]⁵⁺, 1968.3801 [2a+12NO₃]⁴⁺.$

Figure S12. Partial ¹³C NMR spectrum (150 MHz/ CD₃CN/DMSO 2:1) of 2a showing expansions of the regions containing overlapping signals. Of the 52 expected ¹³C signals, only 44 could be clearly identified.

Figure S13. 1 H – 1 H COSY spectrum (600 MHz/CD3CN) of 2a.

Figure S14. ¹H - ¹H NOESY spectrum (600 MHz/CD₃CN) of 2a with important NOESY contacts highlighted in red. A symmetry-by-color model of 2a based of the crystal structure of 2b is shown in the inset. The loss in two-fold ligand symmetry can be clearly visualized as the result of interpenetration, with the central and peripheral ligands possessing distinct chemical environments. The X-ray structure is therefore fully consistent with fourfold-split ¹H NMR spectroscopic signature of the molecule.

Figure S15. a) Partial ¹H – ¹H COSY spectrum (600 MHz/CD₃CN) of 2a showing the weak ⁵J coupling between adjacent H_c protons in one of the L¹ ligands; b) Partial ¹H – ¹H NOESY spectrum (600 MHz/CD₃CN) of 2a showing the cross peaks between adjacent H_c protons in one of the L¹ ligands. This supports that, in 2a, the ligands have lost their two-fold symmetry.

Figure S16. DOSY spectrum (600 MHz, CD₃CN, 25 °C) of 2a. Diffusion coefficient: 4.79 x 10⁻¹⁰ m²/s, hydrodynamic radius = 1.24 nm.

Figure S17. ¹H NMR (500 MHz, CD₃CN) array experiment carried out at 70°C showing the gradual formation of 2a. The array experiment was carried out over 20.16 hrs in 15 min intervals. The inset shows the formation of 2a based on a final concentration of 0.175 mM, calculated by comparing the integration of the aromatic signals to a standard solvent signal.

1.4.4. Synthesis of $[Pd_8L^2_{16}]^{16+}$ (2b)

A solution of Pd(NO₃)₂ (387 µL, 15 mM/CD₃CN, 5.8 µmol) was combined with a solution of L² (5.05 mg, 11.3 µmol) in CD₃CN (3634 μL) and heated at 70 °C for 24 h to afford $[Pd_8L^2{}_{16}]^{16+}$. ¹H NMR (600 MHz, CD₃CN) δ 10.77 (d, J = 5.9 Hz, 16H), 10.15 (d, J = 6.0 Hz, 16H), 9.91 (s, 16H), 9.53 (s, 8H), 9.36 (s, 8H), 9.24 (d, J = 5.8 Hz, 16H), 9.11 (s, 8H), 8.88 (s, 16H), 8.67 (s, 16H), 8.54 – 8.44 (m, 24H), 8.41 (d, J = 8.8 Hz, 8H), 8.37 (s, 8H), 8.25 – 8.16 (m, 16H), 8.07 (d, J = 8.8 Hz, 8H), 7.74 (s, 8H), 7.16 (s, 8H), 7.08 (s, 16H), 6.89 (d, J = 8.6 Hz, 8H), 4.24 – 4.12 (m, 32H), 4.05 (s, 8H), 3.85 (s, 8H), 3.76 (s, 8H), 3.15 – 3.06 (m, 8H), 1.93 – 1.88 (m, 32H), 1.80 – 1.72 (m, 16H), 1.29 – 1.19 (m, 16H), 1.15 – 1.01 (m, 64H), 0.92 – 0.79 (m, 24H). ESI HR-MS (C₄₈₀H₄₄₈N₃₂O₃₂Pd₈) calc: 1065.5738 [**2b**+8NO₃]⁸⁺, 1226.5112 [**2b**+9NO₃]⁷⁺, 1441.2612 [**2b**+10NO₃]⁶⁺, 1741.7110 [**2b**+11NO₃]⁵⁺, 2193.1361 [**2b+**12NO₃]⁴⁺; found: 1065.5897 [**2b**+8NO₃]⁸⁺, 1226.5294 [**2b**+9NO₃]⁷⁺, 1441.2823 $[2b+10NO₃]⁶⁺$, 1741.9365 $[2b+11NO₃]⁵⁺$, 2193.1646 $[2b+12NO₃]⁴⁺$.

Figure S19. ESI-MS spectrum of $[Pd_8L^2_{16}+nNO_3]^{16-n+}$ with $n = 8-12$ (2b). The measured and calculated isotope patter of 2b+10·NO₃ shown in the inset. Each peak contains additional overlapping peaks corresponding closely to ions of 2b with partial cleavages of the alkyl chains.

Figure S21. 1 H – 1 H NOESY spectrum (600 MHz/CD₃CN) of 2b.

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1.4.5. Synthesis of $[Pd_8L^3{}_{16}]^{16+}$ (2c)

A solution of Pd(NO₃)₂ (312 µL, 15 mM/CD₃CN, 4.7 µmol) was combined with a solution of L³ (4.84 mg, 9.1 µmol) in CD₃CN (2933 µL) and heated at 70 °C for 24 h to afford [Pd₈L³₁₆]¹⁶⁺. Upon cooling to room temperature, the solution of 2c was observed to become cloudy (Figure S33). ¹H NMR (500 MHz, CD₃CN, 65 °C) δ 10.76 (d, J = 6.2 Hz, 16H), 10.14 (d, J = 6.1 Hz, 16H), 9.88 (d, J = 6.0 Hz, 16H), 9.54 (s, 8H), 9.38 (s, 8H), 9.24 (d, J = 6.0 Hz, 16H), 9.10 (s, 8H), 8.87 (d, J = 6.1 Hz, 16H), 8.64 (d, J = 6.0 Hz, 16H), 8.53 – 8.46 (m, 24H), 8.44 – 8.37 (m, 16H), 8.21 (dd, J = 13.7, 8.7 Hz, 16H), 8.08 (d, J = 8.9 Hz, 8H), 7.77 (d, J = 8.5 Hz, 8H), 7.18 (d, J = 8.7 Hz, 8H), 7.11 (d, J = 5.9 Hz, 16H), 6.94 (d, J = 8.6 Hz, 8H), 4.32 - 4.18 (m, 32H), 4.16 – 4.05 (m, 8H), 3.95 – 3.73 (m, 16H), 3.30 – 3.07 (m, 8H), 1.63 – 1.49 (m, 48H), 1.47 – 1.23 (m, 208H), 0.98 – 0.84 (m, 96H). ESI HR-MS $(C_{576}H_{640}N_{32}O_{32}Pd_8)$ calc: 1089.7898 [2c+7NO₃]⁹⁺, 1233.7620 [2c+8NO₃]⁸⁺, 1418.8692 $[2\mathsf{c}+9\mathsf{NO}_3]^{7+}$, 1665.6788 [$2\mathsf{c}+10\mathsf{NO}_3]^{6+}$; found: 1089.7793 [$2\mathsf{c}+7\mathsf{NO}_3]^{9+}$,1233.7542 [$2\mathsf{c}+8\mathsf{NO}_3]^{8+}$, 1418.8607 [$2\mathsf{c}+9\mathsf{NO}_3]^{7+}$, 1665.6679 [2c+10NO₃]⁶⁺.

Figure S23. Partial ¹H NMR spectrum (600 MHz/CD₃CN, 65 °C) of 2c.

Figure S24. ESI-MS spectrum of $[PdsL^{3}_{16}+nNO_3]^{16-n+}$ with $n = 7-11$ (2c) with the measured and calculated isotope patter of $[2c+9\cdot NO_3]^{7+}/$ [2c+Cl+8·NO₃]⁷⁺ shown in the inset. Each peak contains additional species of 2c with masses corresponding closely to the partial cleavages of the alkyl chain. 20% DMSO was added to disrupt the aggregation of 2c for the measurement.

Figure S25. 1 H – 1 H COSY spectrum (600 MHz/CD₃CN) of 2c.

Figure S26. 1 H – 1 H NOESY spectrum (600 MHz/CD₃CN) of 2c.

2. NO₃⁻ triggered transformation

Tetrabutylammonium nitrate (TBA NO₃) in CD₃CN (4, 8 or 12 equivalents, 17.5 mM solution) was added to the mixture of [Pd₃L¹₆](BF₄)₆, [Pd₄L¹₈](BF₄)₈ (D_{4h}), and [Pd₄L¹₈](BF₄)₈ (D_{2d}) (1a, 1b and 1c) in CD₃CN. The mixture was heated at 70 °C for 24 h and ¹H NMR spectra of the reaction mixture were recorded (below). For the ¹⁵N labelled experiment, TBA ¹⁵NO₃ was prepared from commercially available K¹⁵NO₃ (98% ¹⁵N labelled). A 17.5 mM stock solution (9:1 CD₃CN/D₂O) was made up by dissolving K¹⁵NO₃ in D₂O and subsequently adding TBA Cl in CD₃CN. A precipitate was observed (KCl) after 10 seconds and the mixture was heated at 70 °C for 3 h. After filtering off the precipitate the solution of TBA ¹⁵NO₃ was obtained.

Figure S27. ¹H NMR spectrum (500 MHz/CD₃CN) of: a) The mixture of **1a/1b/1c** obtained by heating L¹ with [Pd(CH₃CN)₄](BF₄)₂ for 2 h at 70 °C ; b) a) with 4 equiv. of NO₃⁻, after heating for 24 h; c) a) with 8 equiv. of NO₃⁻, after heating for 24 h; d) a) with 12 equiv. of NO₃⁻, after heating for 24 h; e) 2a obtained directly from Pd(NO₃)₂. Note: the best conversion of 1a/1b/1c to 2a occurs with 8 equiv. of NO₃[−] (spectrum shown in c)). Relative to c) the yield of 2a in b) and d) is 54 %, 33% respectively (calculated by comparing the average ratio of 4 aromatic protons of 2a to the TBA cation signal at 3.07 ppm). The lower yield of 2a in b) is presumably due to a minimum amount of NO3[−] anions required for the formation of the structure. In c) however, addition of more than 8 equivalents of NO₃⁻ results in the immediate formation of some precipitate. The small discrepancies in the shifts of the aromatic protons between b) – e) is presumably due to fast exchange of the BF₄⁻ anions at the periphery of the structure.

Figure S28. a) ESI-MS spectrum of a mixture of 1a, 1b, and 1c (counter-ion BF₄-) after heating with 8 equiv. tetrabutylammonium nitrate at 70 ℃ for 24 h; [Pd₈L1₁₆+nX]^{16–n+} with n = 8–11 (X = Cl⁻, NO₃⁻, BF₄⁻). b) Calculated and measured isotope pattern of the 7+ peak; c) calculated and measured isotope pattern of the 6+ peak. Note: Cl[−] ions are present as a contaminant.

Figure S29. ¹⁵N NMR spectra (60.827 MHz/CD₃CN) measured on an AVANCE-III HD 600 equipped with a BBFO cryo-probe; a) TBA ¹⁵NO₃ (244.62 ppm). The signal at 379.71 ppm is presumed to be unreacted K¹⁵NO₃; b) 2a obtained by heating he mixture of 1a/1b/1c with 8 equiv. of TBA ¹⁵NO₃ for 24 h at 70°C.

Figure S30. ¹⁹F NMR spectra (377 MHz/CD₃CN) of 2a obtained by heating he mixture of 1a/1b/1c with 8 equiv. of TBA ¹⁵NO₃ for 24 h at 70°C.

Figure S31. ¹H NMR spectrum (500 MHz, CD₃CN) of; a) The mixture of 1a/1b/1c obtained by heating L¹ with [Pd(CH₃CN)₄](BF₄)₂ for 2 h at 70 °C; b) the same mixture after heating for 24 h at 70 °C in the presence of 8 equiv. of PF 6 .

3. Dynamic light scattering (DLS)

concentration with PDI shown (light blue) for each measurement. Note: each measurement is an average of three measurements. The vesicle-like particles of 2c show an inverse concentration dependent swelling.

Figure S33. a) DLS of 2c cycled between 25 and 70 °C. Upon heating (red), a smaller aggregate is observed (36.2 ± 20.4 nm) along with particles measuring 2.3 \pm 0.7 nm (corresponding to non-aggregated 2c). Upon cooling (grey), the original aggregate (157.6 \pm 44.9 nm) is recovered. Concentration of $2c = 5.25 \mu M$; b) a picture of $2c$ in MeCN at room temperature (left) and at 70 °C (right).

4. Transmission electron microscopy (TEM)

Samples of 2a and 2c were prepared in CH₃CN (0.0875 mM) and deposited on a copper electron microscopy grid. The solvent was allowed to evaporate and then the sample was put under vacuum for 30 minutes prior to immediate measurement.

Figure S34. TEM images of 2c at different magnifications.

Figure S35. An example of a raw EDX spectrum of 2c, confirming the palladium composition of the sample visualized by TEM.

Figure S36. TEM image of 2a. The dark circles measure 2 – 3 nm is diameter, which compares well to the diameter of 2a in solution ($D_H = 2.48 \text{ Å}$) and the solid-state (2.5 Å – longest dimensions of the cigar-shaped structure).

Figure S37. An example of a raw EDX spectrum of 2a, confirming the palladium composition of the sample visualized by TEM.

5. Formation of D_{2d} -[Pd₄L¹₈] (1c)

5.1. DFT calculations

All models were constructed using SPARTAN⁶ and were first optimized with the PM6 semi-empirical method. The resulting structures were then further refined by DFT calculations carried out with GAUSSIAN.⁷

As discussed here and elsewhere,⁵ when **L¹** is reacted with [Pd(CH₃CN)₄](BF₄)₂, D_{2d}-[Pd₄**L¹₈] (1c**) forms only as the minor product along with major products D_{4h} -[Pd₄L¹₈] (**1b**) and D_{3h} -[Pd₃L¹₆] (**1a**). We performed DFT calculations to provide insight into why 1c might be less stable than its structural isomer 1a. Indeed, DFT calculations (RB3LYP, def2/svp) revealed that 1c is 35.4 kcal/mol higher is energy than 1a, presumably due to the bond strain in one of the two bridging modes of L¹.

Level of theory: B3LYP/def2-SVP. D_{4h} box: E(hartree): -10604.3416736, D_{2d} tetrahedron: -10604.2852263

Figure S38. DFT calculated structures and relative energies for D_{4h} -[Pd₄L¹₈]⁸⁺ and D_{2d} -[Pd₄L¹₈]⁸⁺ (1**b** and 1c respectively).

5.2. Discussion

We were surprised when a single crystal of 1**c** was isolated after slow vapour diffusion of THF into a solution of [Pd $_8$ L 1 16] (2a) in MeCN. It is important to note that the crystallization occurred over a period of 3 months, and only three small $(0.1 \, \text{mm}^3)$ crystals were observed on the walls of the vial. Therefore, only a very small amount of the material crystallized with the remaining material observed in the form of a precipitate. An explanation for this could be that, after the initial precipitation, significant dilution of the 2a by slow diffusion of THF shifts the equilibrium toward noninterpenetrated products. One would therefore expect 1b to be the major product (as it is the non-interpenetrated monomeric form of 2a). An explanation for this could be that the presence of NO₃⁻ anions stabilizes 1c, or that 1c packs more favourably in the solid-state than 1a or 1b. To test the former hypothesis, we titrated the mixture of 1a/1b/1c with TBA NO₃[−] and allowed the mixture to equilibrate over 3 days before measuring the ¹H NMR spectrum. As can be seen from the figure below, addition of NO_3^- anions does indeed shift the equilibrium towards 1c as the major product.

Figure S39. ¹H NMR spectra (500 MHz/CD₃CN) of: a) a mixture of 1a, 1b and 1c (approximate ratio: 1:2:0.2, respectively); b) the same sample with 1 equivalent of TBA NO₃⁻ after 3 days at 25 °C. Ratio of 1a to 1c: 3:4. 1b (D_{4h} -[Pd₄L¹₈]) could not be detected.

6. X-ray crystallography

6.1. Crystal structure of 1c

Single crystals of 1c were grown over a period of 3 months by slow vapour diffusion of THF into a 0.175 mM CD₃CN solution of 2a. A single crystal of 1c in mother liquor was pipetted onto a glass slide containing NVH oil. To avoid collapse of the crystal lattice, the crystal was quickly mounted onto a 0.1 mm nylon loop and immediately flash cooled in liquid nitrogen. Crystals were stored at cryogenic temperature in dry shippers, in which they were safely transported to macromolecular beamline PXII at the Swiss Light Source (SLS), Paul Scherrer Institute, Villigen, Switzerland. A wavelength of $\lambda = 0.9994$ Å was chosen using a liquid N₂ cooled double crystal monochromator. Single crystal X-ray diffraction data was collected at 100(2) K on a single axis goniometer, equipped with an Oxford Cryostream 800 a Pilatus 6M. 3600 diffraction images were collected in a 360° φ sweep at a detector distance of 165 mm, 100% filter transmission, 0.1° step width and 1 second exposure time per image. The data were integrated with $XDS.⁸$

The structure was solved by intrinsic phasing/direct methods using SHELXT⁹ and refined with SHELXL¹⁰ using 22 cpu cores for full-matrix least-squares routines on F^2 and ShelXle¹¹ as a graphical user interface and the DSR program plugin was employed for modelling.¹²

6.1.1. Specific refinement details of 1c.

Stereochemical restraints for the PMP ligands, THF solvent molecule and NO3 $[NO₃]$ counter ions of the structure were generated by the GRADE program using the GRADE Web Server (http://grade.globalphasing.org) and applied in the refinement. A GRADE dictionary for SHELXL contains target values and standard deviations for 1,2-distances (DFIX) and 1,3-distances (DANG), as well as restraints for planar groups (FLAT). Due to the location of the THF on a special position (2-fold axis) the GRADE restraint dictionary was modified manually to cover symmetry generated atoms. All displacements for non-hydrogen atoms were refined anisotropically. The refinement of ADP's for carbon, nitrogen and oxygen atoms was enabled by a combination of similarity restraints (SIMU) and rigid bond restraints (RIGU).13 The contribution of the electron density from disordered counterions and solvent molecules, which could

not be modelled with discrete atomic positions were handled using the SOUEEZE¹⁴ routine in PLATON¹⁵. The solvent mask file (.fab) computed by PLATON were included in the SHELXL refinement via the ABIN instruction leaving the measured intensities untouched.

6.1.2. Description of the structure of $1c$

1c crystallizes in the tetragonal space group P-4n2, with two molecules of L^1 , one Pd(II) centre, one NO₃⁻ counter-ion and half of a THF molecule in the asymmetric unit (the other nitrate counter-ion could not be adequately modelled from the electron density in the difference map. In the structure of $1c$, four Pd atoms form an ideal tetrahedron, with the doubly-bridged Pd – $(L)_2$ – Pd distances (12.02 Å) being almost equal to singly bridged P – L – Pd connections (12.07 Å). One bridging mode of L¹ results in an out-of-plane bending of the pyridyl donors, consistent with the bond strain predicated in the calculated model.

Figure S40. X-ray structure of 1c; a) Space-filling overlay; b) perspective view with hydrogen atoms omitted for clarity; c) the two types of ligand geometries in 1c. The pyridyl donors of ligand 'A' are significantly twisted out of plane (with respect to the phenanthrene core) which is in accordance with the DFT calculations.

6.2. Crystal structure of 2b

Single crystals of 2b were grown over a period of 11 days by slow vapour diffusion of diisopropyl ether into a concentrated (0.35 mM) CD₃CN solution of 2b. A single crystal of 2b in mother liquor was pipetted onto a glass slide containing NVH oil. To avoid collapse of the crystal lattice, the crystal was quickly mounted onto a 0.2 mm nylon loop and immediately flash cooled in liquid nitrogen.

Crystals were only poorly diffracting up to 2 Å resolution on our in-house diffractometer Bruker D8 venture equippend with an Incoatec microfocus source (Iμs 2.0) using Cukα radiation and 240 seconds exposure time per 1° rotation. In order to achieve a higher resolution sufficient for structure solution using direct methods (1.4 Å), an extremely bright 3rd generation source of synchrotron radiation was required. For this purpose, 20 crystals mounted on loops were placed in UNI Pucks and stored at cryogenic temperature in dry shippers, in which they were safely transported to macromolecular beamline P11¹⁶ at Petra III, DESY, Germany. UNI Pucks were transferred to the sample dewar and all samples were mounted using the StäubliTX60L robotic arm. A wavelength of 0.6889 Å was chosen using a liquid N₂ cooled double crystal monochromator. Single crystal X-ray diffraction data was collected at 80(2) K on a single axis goniometer, equipped with an Oxford Cryostream 800 a Pilatus 6M fast. As initial screening showed significant dispersive scattering contributions and a distinct diffraction limit of approximately 1.4 - 1.3 Å, 1800 diffraction images were collected in a 360° φ sweep at a detector distance of 300 mm, 30% filter transmission, 0.2° step width and 0.2 seconds exposure time per image. Data integration and reduction were undertaken using XDS.⁸ The data was cut at 1.28 Å, as the signal to noise ratio has dropped below I/σ(I) < 2.0. Due to high mosaicity, disorder in the solvent region and significant dispersive scattering contributions a higher resolution could not be achieved. The structure was solved by intrinsic phasing/direct methods using SHELXT⁹ and refined with SHELXL¹⁰ using 22 cpu cores and ShelXle¹¹ as a graphical user interface and the DSR program plugin was employed for modelling.¹² All modelling cycles were refined against F^2 until convergence using the conjugate-gradient algorithm (CGLS). Only for computing the crystallographic information file (CIF) the full-matrix least-squares routine was employed. Hydrogen atoms were included as invariants at geometrically estimated positions

6.2.1. Specific refinement details of 2b

Due to the limited resolution the data/parameter ratio is poor for a small molecule structure, but not at all critical in macromolecular structure. In order to generate a molecular model and increase robustness of the refinement we have adapted and exploited techniques commonly applied in macromolecular structure refinement. Stereochemical

restraints for the PPP ligands and NO3 $[NO₃]$ counter ions of the structure were generated by the GRADE program using the GRADE Web Server (http://grade.globalphasing.org) and applied in the refinement. This macromolecular refinement technique has been adapted to be used in the program SHELXL¹⁰ and successfully applied to other supramolecular structures behaving like macromolecules.^{17−20} Also for this structure it was found to drastically increase the robustness of the refinement, especially when combined with the rigid bond restraint for atomic displacements.¹³ A GRADE dictionary for SHELXL contains target values and standard deviations for 1,2-distances (DFIX) and 1,3-distances (DANG), as well as restraints for planar groups (FLAT). Non-crystallographic symmetry restraints (NCSY) were employed for 1,4-distances to enhance local structural similarity in between the 16 independent PPP ligands and thereby additionally stabilize their highly flexible propoxy side chains. All displacements for non-hydrogen atoms were refined anisotropically. The refinement of ADP's for carbon, nitrogen and oxygen atoms was enabled by a combination of similarity restraints (SIMU) and rigid bond restraints (RIGU).¹³ For the NO₃ counter ions, isotropic restraints (ISOR) were employed additionally. As the observed electron density was compelling, NO₃ anion (residue 24) located in a pocket between two Palladium cations was modelled although the PLATON routine of CHECKCIF reported a short contacts B alert. It is possible, that there are multiple positions for this counterion, which we cannot resolve at the resolution achieved. The contribution of the electron density from disordered counterions and solvent molecules, which could not be modelled with discrete atomic positions were handled using the SQUEEZE¹⁴ routine in PLATON.¹⁵ The solvent mask file (.fab) computed by PLATON were included in the SHELXL refinement via the ABIN instruction leaving the measured intensities untouched.

6.3. Thermal ellipsoid plots

Figure S41. The asymmetric unit of the X-ray structure of 1c, with all non-hydrogen atoms shown as ellipsoids at the 50% probability level.

Figure S42. The asymmetric unit of the X-ray structure of 2b, with all non-hydrogen atoms shown as ellipsoids at the 50% probability level.

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