

Online Resource 1. Biospecimen Reporting for Improved Study Quality (BRISQ): Items to Consider Reporting if Known and Applicable (Moore HM, Kelly AB, Jewell SD, et al. Biospecimen reporting for improved study quality (BRISQ). Cancer cytopathology. Apr 25 2011;119(2):92-101)

Apply to	Tier #	Item Description	Item #	Location
I. Pre-acquisition				
All	Tier 1	Biospecimen type. Solid tissue, whole blood, serum/plasma, isolated cells, urine, secretions, or another product derived from a human being.	Tissue, Abutment, Exudate	p. 5, 8, 9 _____
All	Tier 1	Anatomical or collection site. In standard terminology, organ(s) of origin or site of blood draw.	Retroauricular area	p. 7 _____
All	Tier 1	Biospecimen disease status. From controls or individuals with the disease of interest; in the case of solid tissue, whether it is from disease site or normal adjacent (not involved but from the same anatomical site as a disease specimen in the same patient).	Biospecimens from (i) titanium surface, (ii) exudate surrounding titanium surface, (iii) soft tissue biopsy 5 mm from titanium, at the treatment site	p. 7 _____
All	Tier 1	Clinical characteristics of patients. In standard terminology, available medical information known or believed to be pertinent to the condition of the biospecimens.	Hearing deficiency	p. 5, Table 1 _____
All	Tier 1	Vital state. Alive or deceased when biospecimens were obtained	Alive	p. 5 _____
All	Tier 3	<i>Disease state.</i> Patient condition relative to disease and treatment, if known (eg, during- or post-therapy; acute, chronic, or terminal stage).	Chronic	p. 5, Table 1 _____
All	Tier 3	<i>Cause of death.</i> For postmortem biospecimens, the cause of death and other diseases present at the time of death.	Not applicable	_____
All	Tier 3	<i>Agonal state.</i> The patients' physical condition immediately preceding death (eg, prolonged degeneration or relatively healthy)	Not applicable	_____
All	Tier 1	Diagnosis. Patient diagnoses pertinent to the study being conducted, using an accepted system of standards (eg, the Systemized Nomenclature of Medicine or the International Classification of Diseases). Please note that clinical and pathology diagnoses are not always the same.	Congenital conductive hearing loss, acquired conductive-mixed hearing loss, single sided deafness	Table 1 _____
All	Tier 1	Clinical. Patient clinical diagnoses (determined by medical history, physical examination, and analyses of a biospecimen) pertinent to the study being conducted.	Congenital conductive hearing loss, acquired conductive-mixed hearing loss, single sided deafness	Table 1 _____
All	Tier 1	Pathology. Patient pathology diagnoses (determined by macro and/or microscopic evaluation of a biospecimen at the time of diagnosis and/or prior to research use) pertinent to the study being conducted.	Not applicable	_____
All	Tier 2	Time between diagnosis and sampling. The time or range of time between disease diagnosis and sample acquisition.	Time between treatment and sampling: 0, 3, 12 months	p. 7 _____
All	Tier 3	Exposures. Neoadjuvant therapy, other current or past medical treatments or environmental factors that might influence the condition of the biospecimen (eg, chemo-and radiation therapy, blood thinner, smoking status).	Exclusion criteria, demographics	p. 7, Table 1 _____
All	Tier 3	Reproductive status. The hormonal or reproductive state of the patients (eg, pregnant, pre-pubescent, post-menopausal).	Not applicable	_____
All	Tier 2	Patient demographic information. Demographic information that might be relevant to the condition of the biospecimens (eg, age range, gender).	Demographics	Table 1 _____
All	Tier 2	Accrual scheme. Whether the biospecimens were obtained for the study being conducted or for a generalized collection such as a population-based biospecimen resource (i.e. retrospective or prospective procurement); whether any standard operating procedures (SOPs) were employed and whether these SOPs are available to others upon request. Reference any clinical trials relevant to the accrual scheme.	Obtained for the study; sampling SOPs available	p. 7 _____
All	Tier 2	Nature of the biobanking institution(s). The biobanking context in which the biospecimens were obtained (eg, as part of an internal collection or a biospecimen-acquisition network); include name, location, and primary contact details such as email address or Web site and reference to any pertinent SOPs.	Biobank 513	p. 7 _____
II. Acquisition				
All	Tier 1	Collection mechanism and parameters. How the biospecimens were obtained (eg, fine needle aspiration, pre-operative blood draw).	Tissue biopsy punch; retrieved abutment; paper-point absorption of exudate	p. 7 _____
Tissue	Tier 3	Time from cessation of blood flow in vivo to biospecimen excision/acquisition. The time or range of times that the biospecimens were ischemic in the body.	Not applicable	_____

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Online Resource 1. (Continued)

Apply to	Tier #	Item Description	Item #	Location
All	Tier 2	<u>Time from biospecimen excision/acquisition to stabilization.</u> The time or time-range between when the biospecimens were obtained (eg, blood drawn or tumor surgically removed) and when they were stabilized. <i>For postmortem biospecimens</i> , list the postmortem interval range (i.e. the time from death to stabilization of the biospecimen).	Immediately	p. 7 _____
All	Tier 2	<u>Temperature between biospecimen excision/acquisition and stabilization.</u> The temperature or range thereof at which biospecimens were kept between when biospecimens were obtained (eg, blood drawn or tumor surgically removed) and when they were stabilized. <i>For postmortem biospecimens</i> , the temperature at which the cadaver was stored during the postmortem interval.	Room temperature	p. 7 _____
Fluid	Tier 2	<u>Collection container.</u> The kind of tube into which biospecimens were captured as they left the body.	ESwab™	p. 7 _____
III. Stabilization/Preservation				
All	Tier 1	<u>Mechanism of stabilization.</u> The initial process by which biospecimens were stabilized during collection [eg, snap or controlled-rate freezing, fixation, additive (heparin, citrate, or EDTA), none].	Total immersion in liquid Amies medium	p. 7 _____
All	Tier 1	<u>Type of long-term preservation.</u> The process by which the biospecimens were sustained after collection (eg, freezing and at which temperature; formalin fixation, paraffin embedding; additive; none). Please note, this might or might not differ from the mechanism of stabilization.	Total immersion in liquid Amies medium	p. 7 _____
All	Tier 1	<u>Constitution and concentration of fixative/preservation solution.</u> The make-up of any formulation employed to maintain the biospecimens in a non-reactive state (eg, 10 percent neutral-buffered formalin or 10 USP Heparin Units/mL).	Liquid Amies medium	p. 7 _____
Tissue	Tier 2	<u>Time in fixative/preservation solution.</u> The time or range thereof that biospecimens were exposed to the preservation medium.	1-2 days	p. 7, 8 _____
Tissue	Tier 2	<u>Temperature during time in preservation solution.</u> The temperature of the medium during the preservation process.	at 4°C and room temperature	p. 7 _____
Fluid	Tier 2	<u>Aliquot volume.</u> The amount in each liquid biospecimen sample.	Absorbed in 2 Roeko size 45 paper-points	p. 7 _____
Tissue	Tier 2	<u>Specimen size.</u> The approximate size or weight of solid biospecimen samples processed (eg, cubes approximately 0.5 cm on a side, 0.5 gram).	1 mm punch	p. 7 _____
IV. Storage/Transport				
<u>Storage parameters.</u> The conditions under which the biospecimens were maintained until analysis.				
All	Tier 1	<u>Storage temperature.</u> The temperature or range thereof at which the biospecimens were maintained until distribution or analysis.	4°C or room temperature	p. 7, 8 _____
All	Tier 1	<u>Storage duration.</u> The time or range thereof between biospecimen acquisition and distribution or analysis.	Maximum 2 days	p. 7, 8 _____
All	Tier 2	<u>Storage details.</u> Other conditions under which specimens were maintained during storage (eg, to minimize oxidation).	Samples completely submerged	p. 7 _____
All	Tier 3	<u>Type of storage container.</u> The vessel in which biospecimens were kept.	Plastic Tube	p. 7 _____
All	Tier 3	<u>Type of slide.</u> The microscope slides to which biospecimens were affixed.	Not applicable	_____
<u>Shipping parameters.</u> The conditions to which biospecimens were exposed during each shipment or inventory management.				
All	Tier 1	<u>Shipping temperature(s).</u> The temperature or range thereof at which biospecimens were maintained during each shipment or relocation.	Room temperature	p. 7, 8 _____
All	Tier 2	<u>Shipping duration.</u> The time, estimate, or range thereof that the biospecimens spent in shipment each time they were transported.	5-24 h	p. 7, 8 _____
All	Tier 3	<u>Type of transport container.</u> The type of vessel (eg, pre-manufactured shipping container, polystyrene box) and the packing material in which the biospecimens were transported.	Carton box	p. 7 _____
All	Tier 3	<u>Shipping parameters.</u> Other conditions under which the biospecimens were transported (eg, vacuum sealing, desiccant, packing material). Please note any deviations from standard operating procedures that might influence the condition of the biospecimens (eg, shipping anomalies that exposed paraffin blocks to high temperatures).	Carton box with an upright position of the tubes	p. 7 _____
<u>Freeze-thaw parameters.</u> The conditions to which biospecimens were subjected during any thaw events.				

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Online Resource 1. (Continued)

Apply to	Tier #	Item Description	Item #	Location
Fluid	Tier 2	<u>Number of freeze-thaw cycles.</u> The number, estimate, or range thereof of thaw-refreeze events to which biospecimens were subjected prior to analysis.	Not applicable	_____
Fluid	Tier 3	<u>Duration of thaw events.</u> The amount of time or range thereof the biospecimens spent thawed prior to the final thaw before processing.	Not applicable	_____
Fluid	Tier 3	<u>Time from last thaw to processing.</u> The time or range of times between unfreezing and analysis.	Not applicable	_____
All	Tier 3	<u>Temperature between last thaw and processing.</u> The temperature at which biospecimens were kept between unfreezing and analysis.	Not applicable	_____

V. Quality Assurance Measures Relevant to the Extracted Product and Processing Prior to Analyte Extraction and Evaluation

All	Tier 1	Composition assessment and selection. Any parameters that were used to evaluate and/or choose biospecimens for inclusion in the study.	Visual inspection	p. 8 _____
All	Tier 2	<u>Gross and microscopic review.</u> The anatomical characteristics of the biospecimens in the study and the relevant qualifications of the individual performing the review (eg, anatomist, pathologist, hematologist, microbiologist, or researcher).	Visual inspection by a microbiologist (MT)	_____
Tissue	Tier 2	<u>Proximity to primary pathology of interest.</u> Whether the biospecimen was taken from a region adjacent to or distal from another region of interest, such as a tumor or area of necrosis. Give approximate distances if known.	All specimens taken from the immediate site of interest. Soft tissue biopsy (3 m, 12 m) obtained 5 mm from abutment	p. 7 _____
All	Tier 2	<u>Method of enrichment for relevant component(s).</u> The method by which pertinent portions of the biospecimen were separated from the rest of the biospecimen (eg, laser-capture microdissection of tissue, block selection for region of lesion, centrifugation of blood).	Cultures in thioglycolate broth	p. 9 _____
All	Tier 2	<u>Details of enrichment for relevant component(s).</u> The parameters used to separate pertinent portions of the biospecimen from the rest of the biospecimen, if applicable (eg, centrifugation speed and temperature).	5 days additional enrichment	p. 9 _____
Tissue	Tier 3	<u>Embedding reagent/medium.</u> Any formulation used to enclose the biospecimens (eg, paraffin).	Not applicable	_____
All	Tier 2	<u>Quality assurance measures.</u> Any methods used to assess the quality of the biospecimens relevant to the biomolecular analyte, when these methods were employed (eg, prior to long-term storage or immediately before experimental analysis), and the results (eg, RNA integrity number, hemolysis assessment).	Testing of selective media with relevant control strains	p. 9 _____

Bold: Tier 1–Recommended to report.

Plain: Tier 2–Beneficial to report.

Italics: Tier 3–Additional items to report.

The clinical outcome and microbiological profile of bone anchored hearing systems (BAHS) with different abutment topographies – A prospective pilot study

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