

1 SUPPORTING INFORMATION (SI)

2 **Strain construction.** Fluorescent fusions to endogenous genes were constructed
3 as follows: The 500 bp regions flanking the C-terminus (stop codon) of each target
4 gene was amplified. These flanking regions were spliced together using a BamHI
5 site (GGTACC) replacing the stop codon of the target gene. (The BamHI sequence
6 acted as a two residue linker between the protein and fluorescent protein label.)
7 The spliced fragments were then cloned into the vector pEXG2 (1). Either mCherry
8 or superfolder-GFP (sfGFP) was cloned into the BamHI site to construct a C-terminal
9 fluorescent fusion to the target genes *clpV*, *tssB*, and *fha*. Finally, each fused gene was
10 inserted at the endogenous promoters by allelic exchange, as described in Ref. (1).

11 An in-frame deletion of *retS* was introduced to the ClpV-mCherry/TssB-sfGFP strain
12 using allelic exchange as described in (2).

13 Bacterial cultivation for strain construction was performed in Luria broth (LB)
14 medium supplemented with 25 µg/ml irgasan, 30 µg/ml gentamycin, and counter
15 selection for allelic exchange was performed on low-salt LB supplemented with 5%
16 wt/vol sucrose.

17 **Behavior of ClpV-TssB double fusion.** The ClpV-mCherry/TssB-GFP double fusion
18 measurably changed ClpV behavior compared to the ClpV-mCherry fusion alone. Although
19 the single-label ClpV-mCherry focus lifetime is identical in both wild type and $\Delta retS$, the
20 ClpV-mCherry/TssB-GFP foci lifetimes are 10% longer in $\Delta retS$ where T6SS is over-expressed.
21 Additionally, in wild type, the ClpV-mCherry/TssB-GFP strain has a diminished firing
22 rate compared to the ClpV-mCherry fusion alone, with at most one ClpV spike seen
23 over the 7–9 minute experiment with a spike lifetime five times as long as wild type
24 ClpV-mCherry. Although this observation of less active dynamics might suggest loss-of-function,
25 we performed a competition between *B. thailandensis* and the double-labeled wild
26 type and the double-labeled and wild type cells had statistically indistinguishable fitness.
27 We therefore believe that the qualitative localization dynamics observed in ClpV-mCherry/TssB-GFP
28 $\Delta retS$ is informative, despite the failure of TssB-GFP to completely complement TssB
29 with respect to the ClpV dynamics.

30 REFERENCES

1. LeRoux M, Kirkpatrick RL, Montauti EI, Tran BQ, Peterson SB, Harding BN, Whitney JC, Russell AB, Traxler B, Goo YA, Goodlett DR, Wiggins PA, Mougous JD. Kin cell lysis is a danger signal that activates antibacterial pathways of *Pseudomonas aeruginosa*. *eLife* 2015; p. e05701.
2. Mougous JD, Cuff ME, Raunser S, Shen A, Zhou M, Gifford CA, Goodman AL, Joachimiak G, Ordoñez CL, Lory S, Walz T, Joachimiak A, Mekalanos JJ. A Virulence Locus of *Pseudomonas aeruginosa* Encodes a Protein Secretion Apparatus. *Science* 2006; 312(5779):1526–1530.