

**Supplementary materials for**

**Placenta-Specific Drug Delivery by Trophoblast-Targeted Nanoparticles in Mice**

Baozhen Zhang<sup>1#</sup>, Lunbo Tan<sup>1,2#</sup>, Yan Yu<sup>3#</sup>, Baobei Wang<sup>1</sup>, Zhilong Chen<sup>1,2</sup>, Jinyu Han<sup>1</sup>, Mengxia Li<sup>1</sup>, Jie Chen<sup>1</sup>, Tianxia Xiao<sup>1</sup>, Balamurali K Ambati<sup>4</sup>, Lintao Cai<sup>5</sup>, Qing Yang<sup>2</sup>, Nihar R Nayak<sup>6\*</sup>, Jian Zhang<sup>1\*</sup>, and Xiujun Fan<sup>1\*</sup>

<sup>1</sup>Laboratory for Reproductive Health, Institute of Biomedicine and Biotechnology, Shenzhen Institutes of Advanced Technology, Chinese Academy of Sciences, Shenzhen, Guangdong, China, 518055.

<sup>2</sup>College of Veterinary Medicine, Hunan Agricultural University, Changsha, Hunan, China, 410128.

<sup>3</sup>Bao'an Maternal and Child Health Care Hospital, Shenzhen, China, 518133.

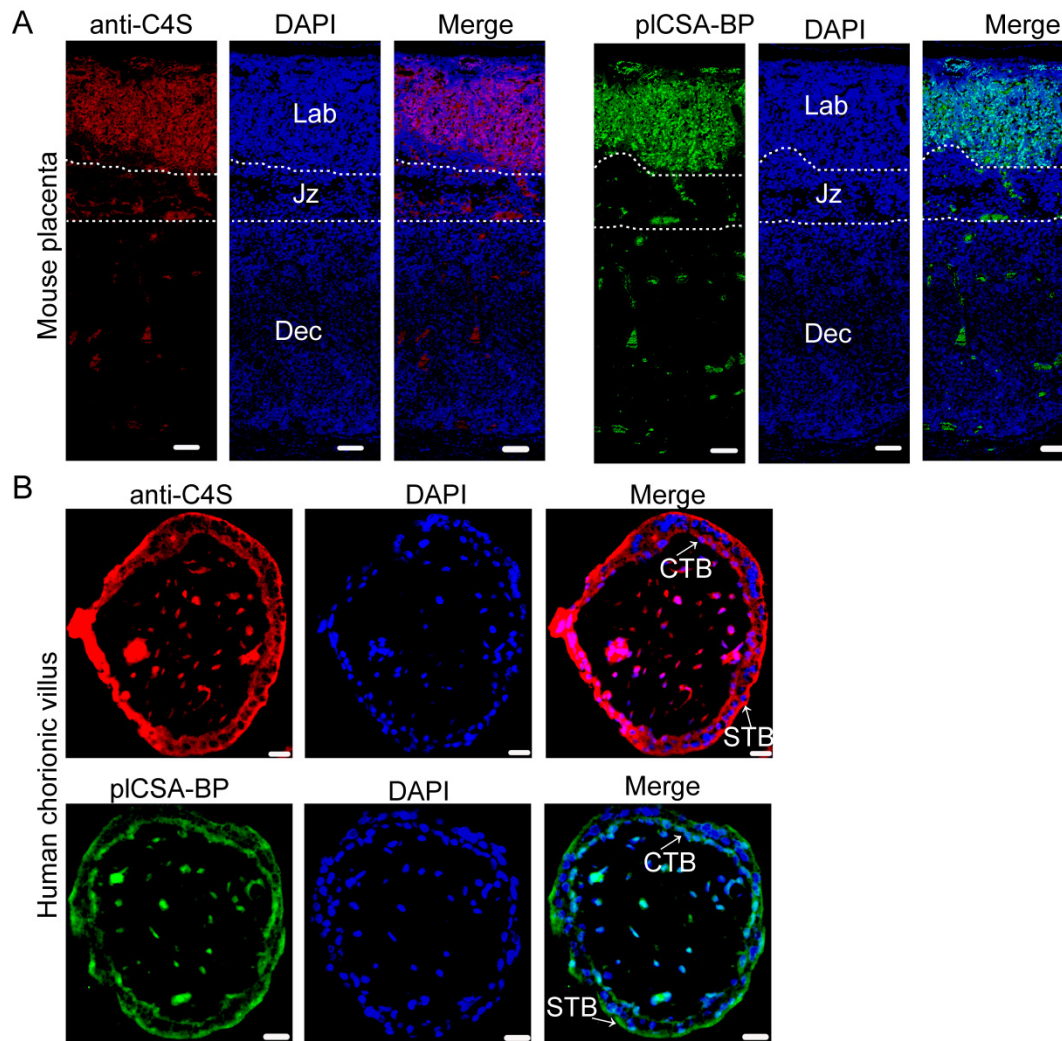
<sup>4</sup>John A. Moran Eye Center, University of Utah, Salt Lake City, Utah, USA, 84132.

<sup>5</sup>Guangdong Key Laboratory of Nanomedicine, CAS Key Lab for Health Informatics, Institute of Biomedicine and Biotechnology, Shenzhen Institutes of Advanced Technology (SIAT), Chinese Academy of Sciences, Shenzhen, China, 518055.

<sup>6</sup>Department of Obstetrics and Gynecology, Wayne State University School of Medicine, C.S. Mott Center for Human Growth and Development, Detroit, Michigan, USA, 48201.

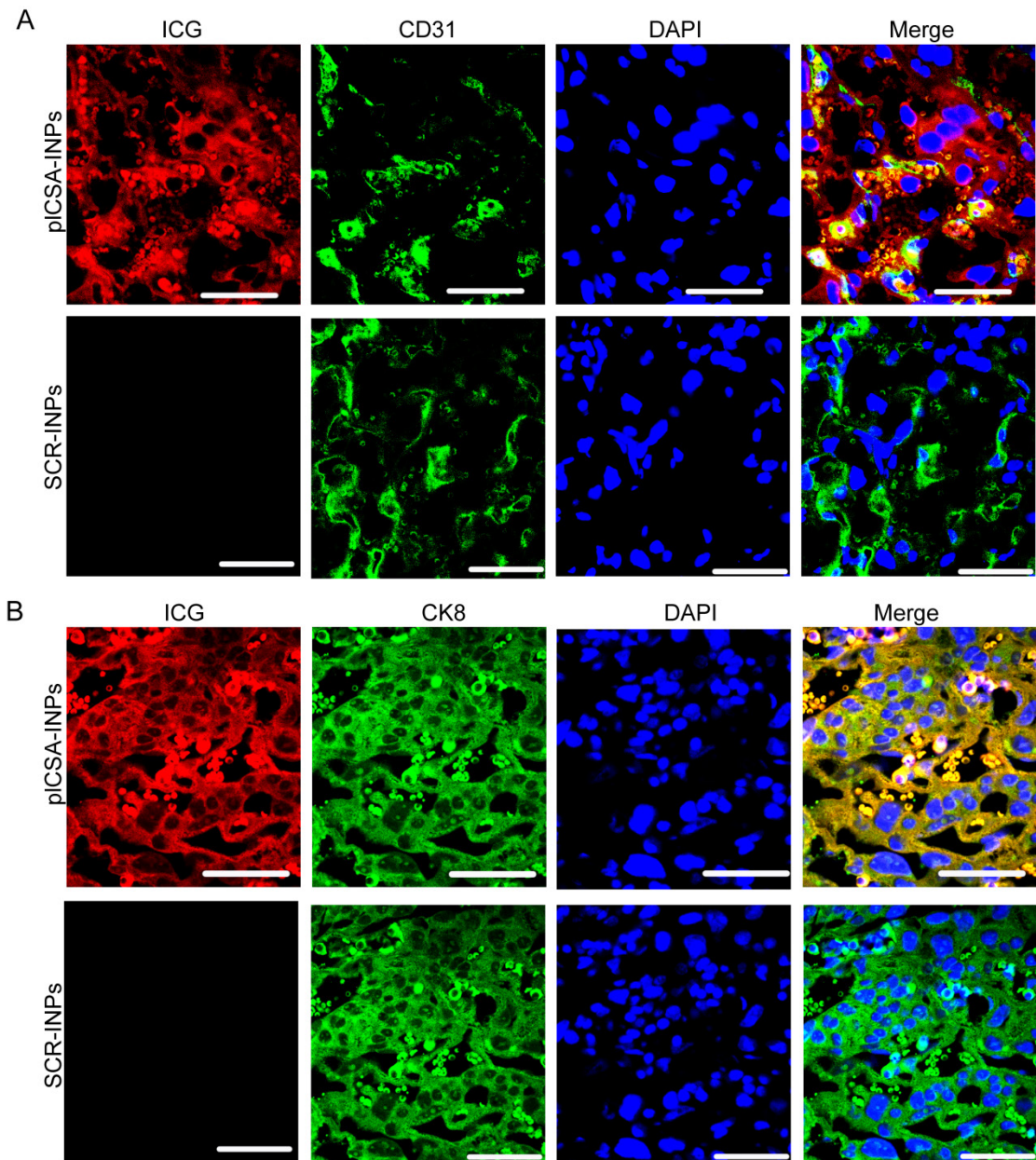
<sup>#</sup>Contributed equally to this work

\*Corresponding authors: Xiujun Fan, E-mail: xj.fan@siat.ac.cn, Phone: 86-755-86392360. Fax: 86-755-86392282. Jian Zhang, E-mail: jian.zhang@siat.ac.cn, Nihar R. Nayak, E-mail: nnayak@med.wayne.edu.

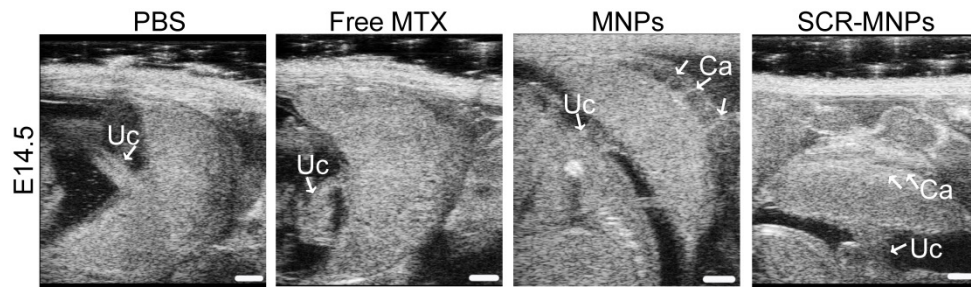


**Figure S1. pICSA-BP and anti-C4S are colocalized, shown by adjacent section immune staining.**

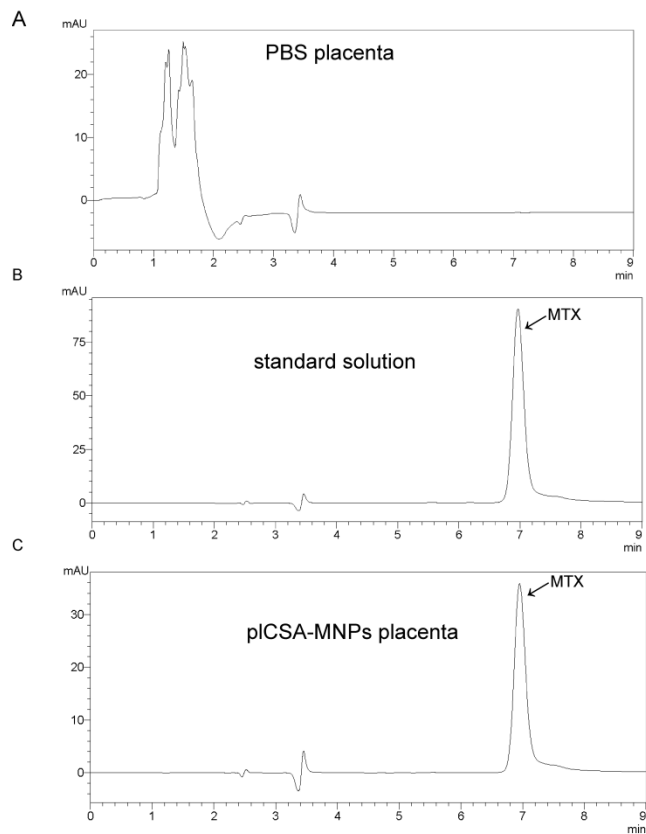
Mouse (A) and human (B) placental tissue sections ( $5\mu\text{m}$ ) stained using biotin-pICSA-BP (green) and anti-C4S (red) and counter-stained with DAPI (blue). Images representative of  $n=5$ . The scale bar represents  $100\ \mu\text{m}$  (A) and  $20\ \mu\text{m}$  (B). Lab, labyrinth; Jz, junctional zone; Dec, decidua. CTB, cytotrophoblasts; STB, syncytiotrophoblasts. **Related to Figure 1.**



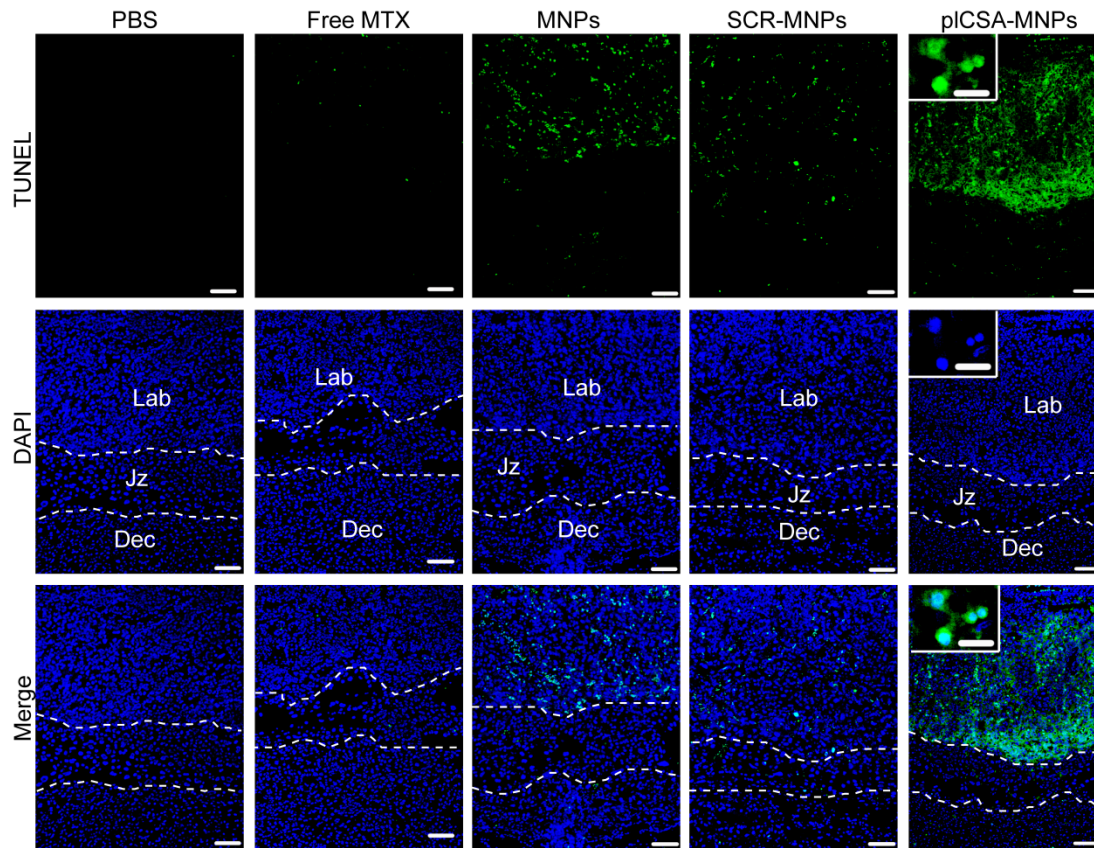
**Figure S2. pICSA-INPs accumulate in mouse placental trophoblast cells.** Placenta tissues immunostained for CK8 (green) and CD31 (green) and counter-stained with DAPI (blue) as in Figure 4(D-E). These sections are 100  $\mu\text{m}$  away from the placental center sections. Images representative of  $n=6$ . The scale bar represents 50  $\mu\text{m}$ . **Related to Figure 4.**



**Figure S3. Representative B-mode images of embryo placenta at E14.5.** The placenta is visualized as a homogenous mass in the PBS and free MTX groups (n=6). The placentas display hyperechogenic calcification deposits (Ca, arrows) in MNP and SCR-MNP groups. **Related to Figure 6.**



**Figure S4. Representative HPLC chromatograms of a pICSA-MNP group placental sample. (A)** PBS placental sample. **(B)** Standard solution of 25  $\mu\text{g/ml}$  MTX, with UV detection at 313 nm; retention time for MTX was 7 min. **(C)** Placental sample 24 h post-pICSA-MNP injection in tail vein. **Related to Figure 7.**



**Figure S5. Apoptosis is induced by pICSA-MNPs in the placenta, as confirmed by TUNEL assay.**

Section selected 100  $\mu\text{m}$  away from the middle sections stained using the TUNEL (green) method; blue indicates DAPI-stained nuclei. Lab, labyrinth; Jz, junctional zone; Dec, decidua. Scale bar=100  $\mu\text{m}$ , and white boxes represent magnifications of the indicated areas (scale bar 20  $\mu\text{m}$ ). Images representative of n=6. **Related to Figure 8.**