

Bacterial community changes in an industrial algae production system
Fulbright et al.

Supplemental Information

Figure S1. Observed OTUs increase with system scale. Rarefaction curve showing, on average, that there are increased OTUs at larger system scales than smaller ones. Small systems include volumes ranging from 5 mL – 4 L, plotted separately. Average observed OTUs for each system are listed in the sample legend. Error bars (plot) and ranges (\pm) represent ± 1 standard deviation.

Figure S2. Phylogenetic distance increases among communities in medium and large systems. Rarefaction curve showing phylogenetic distance (PD) increases on average at larger system scales. Small systems include volumes ranging from 5 mL – 4 L, plotted separately. Average PD for each system is listed in the sample legend. Error bars (plot) and ranges (\pm) represent ± 1 standard deviation.

Table S1. Abundance of bacterial orders across *N. salina* growth systems. Relative abundances of bacterial orders identified in *N. salina* growth systems are represented. Data are formatted as in Table 1. Ph, Phylum (for phylum abbreviations, see Table 1).

Table S2. Bacteria predominant in medium *N. salina* cultivation systems. The ten most abundant bacterial orders identified in medium growth systems are shown. Zero values represent relative abundances less than 0.05%. Ph, Phylum (for phylum abbreviations, see Table 1). Heat maps are formatted separately for each growth system as in Table 1. Σ , total abundance of these ten orders in each system.

Table S3. Comparison of prevalent bacteria in two studies of *Nannochloropsis* cultivations. Comparison of bacteria identified in the cultivations of *N. salina* reported here with those reported by Wang, et al. (2012) in cultivations of *N. oceanica* IMET1.

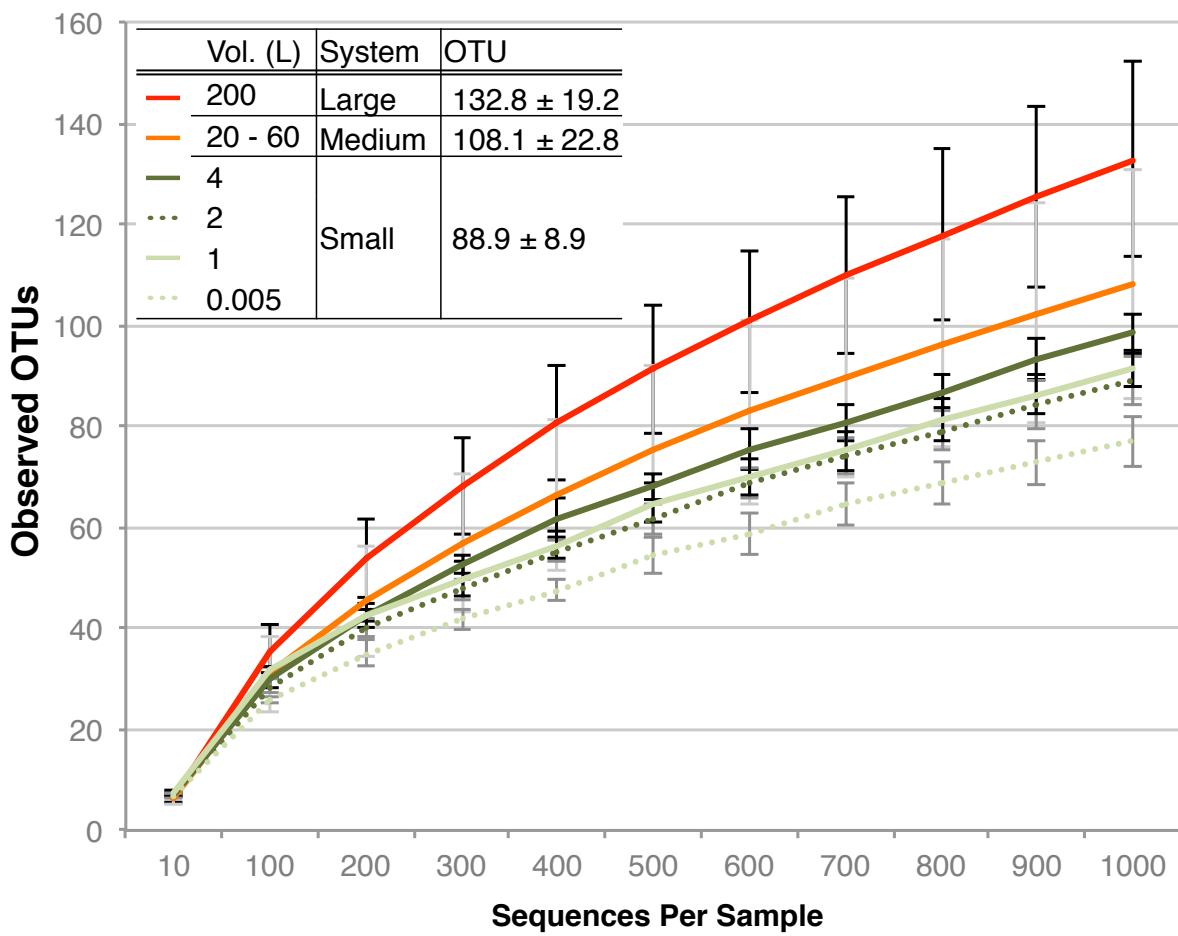


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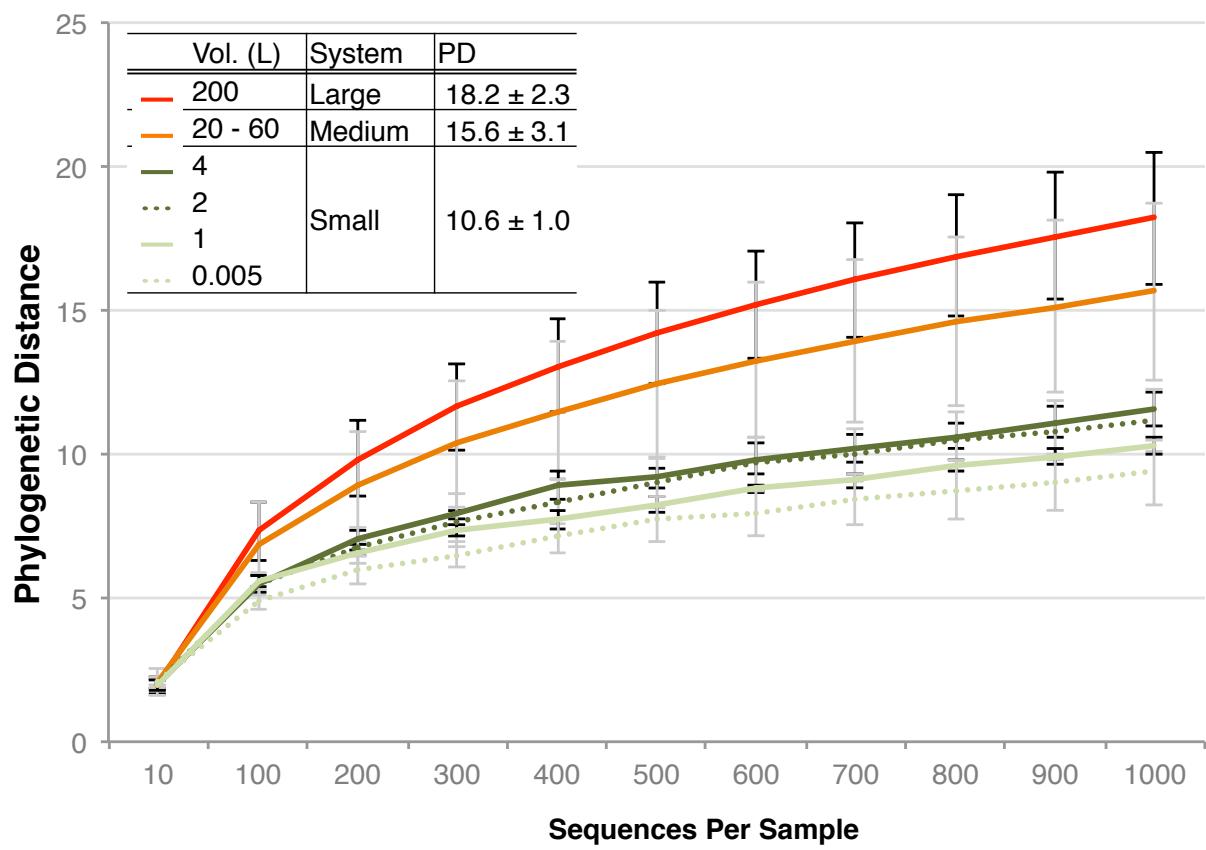


Figure S2. Phylogenetic distance increases among communities in medium and large systems. Rarefaction curve showing phylogenetic distance (PD) increases on average at larger system scales. Small systems include volumes ranging from 5 mL – 4 L, plotted separately. Average PD for each system is listed in the sample legend. Error bars (plot) and ranges (\pm) represent ± 1 standard deviation.

Ph	Class	Order	Abundance		
			Sml	Med	Lrg
B	Bacteroidia	Bacteroidales	0.0	0.0	0.1
	Cytophagia	Cytophagales	5.0	18.9	22.3
	Flavobacteria	Flavobacteriales	20.1	4.6	3.4
	Sphingobacteria	Sphingobacteriales	0.0	5.6	3.0
	[Rhodothermi]	[Rhodothermales]	0.0	0.2	0.1
	[Saprosiriae]	[Saprosirales]	23.2	32.0	37.3
Pr	Alphaproteobacteria		0.1	0.4	0.2
		BD7-3	0.0	1.8	1.5
		Caulobacterales	0.0	0.1	0.1
		Kiloniellales	0.0	1.1	7.1
		Rhizobiales	19.7	8.0	6.2
		Rhodobacterales	8.7	3.6	2.9
		Rhodospirillales	1.1	1.3	2.0
		Rickettsiales	0.0	0.2	0.3
	Betaproteobacteria	Sphingomonadales	5.5	2.1	1.4
		Burkholderiales	0.0	0.0	0.1
		Rhodocyclales	0.0	1.3	0.3
	Deltaproteobacteria	Desulfovibrionales	0.0	0.0	0.4
		Myxococcales	0.0	0.0	0.2
		Spiroacillales	0.0	5.0	1.5
	Epsilonproteobacteria	Campylobacterales	0.0	0.0	0.1
G	Gammaproteobacteria	Alteromonadales	6.9	2.2	1.0
		Chromatiales	0.0	0.2	0.1
		Legionellales	0.0	0.1	0.0
		Oceanospirillales	1.6	0.8	0.1
		Vibrionales	0.0	0.0	0.1
		Pseudomonadales	0.0	0.1	0.0
		Thiotrichales	0.0	0.1	0.0
		Vibrionales	0.0	0.0	0.1
		Other	0.1	0.0	0.0
A	Solibacteres	Solibacterales	0.0	0.1	0.1
Ac	Acidimicrobidae	Acidimicrobiales	0.2	0.1	0.0
	Actinobacteria	Actinomycetales	0.2	0.1	0.0
Ar	[Fimbriimonadia]	[Fimbriimonadales]	0.0	0.1	0.0
Ch	Anaerolineae	SBR1031	0.0	0.1	0.1
	4C0d-2	SM1D11	0.0	0.2	0.0
Cy	ML635J-21		2.8	0.3	0.1
	Synechococcophycideae	Pseudanabaenales	0.0	2.2	0.0
F	Clostridia	Clostridiales	0.0	0.0	0.2
P	Planctomycetia	Pirellulales	1.4	1.5	0.7
		Planctomycetales	0.0	0.3	0.1
T	SC3		0.0	0.3	0.0
V	Opitutae	Opitutales	1.1	0.5	0.7
		Puniceicoccales	0.0	0.1	0.2
	Verrucomicrobiae	Verrucomicrobiales	0.0	0.3	1.2
cd	PRR-11 (BRC1)		0.0	0.0	0.1
	ZB2 (OD1)		0.0	0.2	0.2
	Unassigned		2.4	3.4	4.4
			$\Sigma = 100.1$	99.5	100.0
			$\Sigma_{(B+Pr)} =$	92.0	89.7
					91.9

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B	Sphingobacteria	Sphingobacteriales	0.0	5.6	3.0
Pr	Delta proteobacteria	Spirobacillales	0.0	5.0	1.5
B	Flavobacteria	Flavobacteriales	20.1	4.6	3.4
Pr	Alphaproteobacteria	Rhodobacteriales	8.7	3.6	2.9
Pr	Gammaproteobacteria	Alteromonadales	6.9	2.2	1.0
C	Synechococcophycideae	Pseudanabaenales	0.0	2.2	0.0
Pr	Alphaproteobacteria	Sphingomonadales	5.5	2.1	1.4
			$\Sigma = 89.1$	84.2	79.0

Table S2. Bacteria predominant in medium *N. salina* cultivation systems. The ten most abundant bacterial orders identified in medium growth systems are shown. Zero values represent relative abundances less than 0.05%. Ph, Phylum (for phylum abbreviations, see Table 1). Heat maps are formatted separately for each growth system as in Table 1. Σ , total abundance of these ten orders in each system.

Bacteria isolated from <i>Nannochloropsis oceanica</i> IMET1 cultures*		Organism Taxonomy** / Relationship to OTUs identified in Industrial <i>N. salina</i> Cultivations***										Comments	
Identity***	GenBank nt	GenBank Source	Organism	Temp: Day:	15 C 0	25 C 9	30 C 0	Phylum	Class	Order	Family	Genus	
98%	EU073067.1	Rhodococcus sp.	TS1					Actinobacteria	Actinobacteria	Corynebacteriales	Nocardiidae	Rhodococcus	
98%	EL1440982.1	Microbacterium	scheifelei					Actinobacteria	Actinobacteria	Micrococcales	Microbacteriaceae	Microbacterium	
95%	AB70576.2	Cytophaga	sp. MBIC04669					Bacteroides	Cytophagia	Cytophagales	Cytophagaceae	Cytophaga	
98%	DO080986.1	Roseivirga	spongicola					Bacteroides	Cytophagia	Cytophagales	Flammeovirgaceae	Roseivirga	
98%	EU443204.1	Jejuia	balduilutes					Bacteroides	Flavobacteriia	Flavobacteriales	Flavobacteriaceae	Jejuia	
95%	AB526333.1	Mariibacter	sp. JAM-BA06					Bacteroides	Flavobacteriia	Flavobacteriales	Flavobacteriaceae	Mariibacter	
95%	FJ29467.1	Mariibacter	sp. KLE-1063					Bacteroides	Flavobacteriia	Flavobacteriales	Flavobacteriaceae	Mariibacter	
100%	HM055805.1	Mureaudia	beigygionensis					Bacteroides	Flavobacteriia	Flavobacteriales	Flavobacteriaceae	Mureaudia	
98%	AU231180.1	Planctomyces	planctomycetia					Planctomyces	Planctomycetia	Planctomycetales	Planctomycetaceae	Planctomyces	
98%	AB257592.1	Hoilea	sp. KF7					Proteobacteria	Alphaproteobacteria	Rhizobiales	Phyllobacteriaceae	Hoilea	
98%	AY332116.1	Mesorhizobium	sp. GWS-BW-H238					Proteobacteria	Alphaproteobacteria	Rhizobiales	Phyllobacteriaceae	Mesorhizobium	
97%	FJ230835.1	Henicella	itonalis					Proteobacteria	Alphaproteobacteria	Rhizobiales	Hypomonadaceae	Henicella	
94%	AB372216.1	Hypomonadaceae	bacterium NAMA-F001					Proteobacteria	Alphaproteobacteria	Rhizobiales	Hypomonadaceae	Hypomonadaceae	
98%	AJ605746.2	Roseibaca	ekhonensis					Proteobacteria	Alphaproteobacteria	Rhizobiales	Rhizobacteriaceae	Roseibaca	
98%	FJ088585.1	Roseobacter	sp. RAS3					Proteobacteria	Alphaproteobacteria	Rhizobiales	Rhizobacteriaceae	Roseobacter	
98%	AY326633.1	Roseobacter	sp. TM1042					Proteobacteria	Alphaproteobacteria	Rhizobiales	Rhizobacteriaceae	Roseobacter	
98%	AY682197.1	Sulfobacter	sp. KMM-3457					Proteobacteria	Alphaproteobacteria	Rhizobiales	Rhizobacteriaceae	Sulfobacter	
96%	GU036120.1	Thalassobacter	sp. W-2-2					Proteobacteria	Alphaproteobacteria	Rhizobiales	Rhizobacteriaceae	Thalassobacter	
97%	AY320370.1	Topicalbacter	naphthalenivorans					Proteobacteria	Alphaproteobacteria	Rhizobiales	Rhizobacteriaceae	Topicalbacter	
98%	EF512131.1	Rhodobacteraeace	bacterium CSQ-8					Proteobacteria	Alphaproteobacteria	Rhizobiales	Rhodobacteraceae	Rhodobacter	
99%	AB062106.1	Porphyrobacter	sanguineus					Proteobacteria	Alphaproteobacteria	Rhizobiales	Sphingomonadales	Erythrobacteraceae	Porphyrobacter
99%	DQ112064.1	Mariobacter	sp. LOB-4					Proteobacteria	Gamma-proteobacteria	Alteromonadales	Alteromonadaceae	Mariobacter	
98%	DQ85041.1	Halomonas	sp. JL-1044					Proteobacteria	Gamma-proteobacteria	Oceanospirillales	Halomonadaceae	Halomonas	
89%	GQ867230.1	Pseudomonas	sp. BRAZ6-1					Proteobacteria	Gamma-proteobacteria	Pseudomonadales	Pseudomonadaceae	Pseudomonas	

* Data adapted from Wang, et al. (2012) Supplemental Table S2: "16S rRNA gene sequence identities of isolates from *N. oceanica* IMET1 culture", bacteria from *N. oceanica* cultures grown at 15 C, 25 C, or 30 C were isolated on Marine Agar at Day 0 and Day 9; 16S rRNA genes were amplified, sequenced, and used to identify the closest cultured organism in the GenBank nucleotide database; heat map shows bacteria isolated from cultures maintained at the indicated temperature at Day 0 (light green) or Day 9 (dark green)

** Taxonomy based on indicated GenBank Nucleotide Records

*** Highlighting indicates taxonomic classifications shared with OTUs identified in *N. salina* cultivations in the present study. For additional information, see "Color coding of bacterial types" key (below) and "Comments" (far right), plus Tables 1, 2, 3, S1 and S2.

**** Percent identity of 16S rRNA gene sequences of bacteria isolated in Wang, et al. (2012) compared with respective Genbank Source Organism. Identities $\geq 97\%$ are shown in red.

Color coding of bacterial types:
Isolated from *N. oceanica* cultures at Day 0*

Associated with 90% of *N. salina* cultivations in this study

Associated with 95% of *N. salina* cultivations in this study
Other notable bacteria;
see Comments

Table S3. Comparison of prevalent bacteria in two studies of *Nannochloropsis* cultivations. Comparison of bacteria identified in the cultivations of *N. salina* reported here with those reported by Wang, et al. (2012) in cultivations of *N. oceanica* IMET1.