The pyrroloquinoline-quinone dependent pyranose dehydrogenase from *Coprinopsis* cinerea (CcPDH) drives lytic polysaccharide monooxygenase (LPMO) action

Anikó Várnai^{a,*}, Kiwamu Umezawa^{b,‡,*}, Makoto Yoshida^{b,†,#}, Vincent G. H. Eijsink^{a,†,#}

^aFaculty of Chemistry, Biotechnology and Food Science, Norwegian University of Life Sciences (NMBU), 1432 Ås, Norway

^bDepartment of Environmental and Natural Resource Science, Tokyo University of Agriculture and Technology, Tokyo 183-8509, Japan

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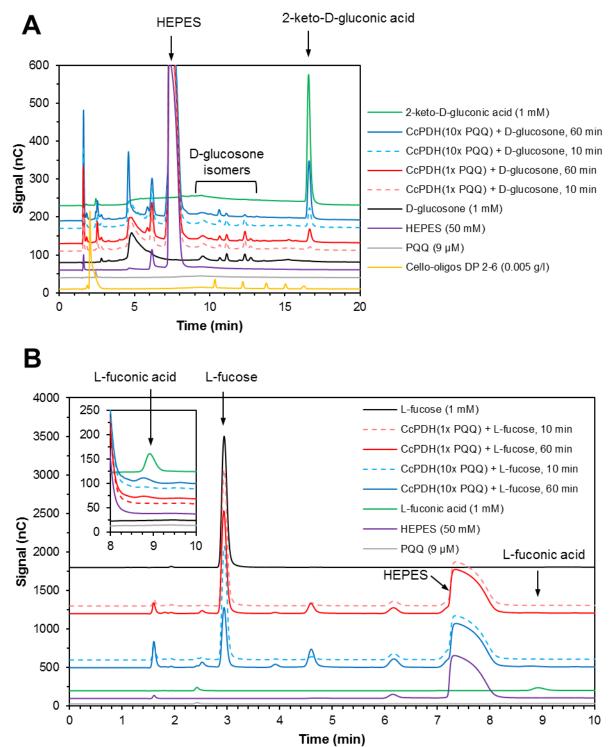
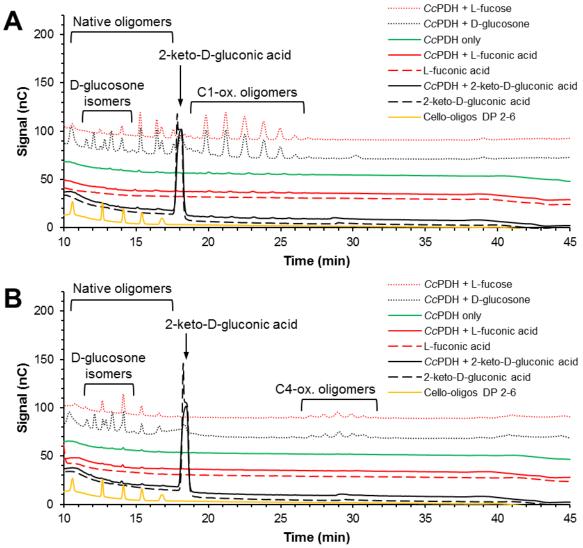


Figure S1. Identification of products generated by *Cc*PDH from (A) D-glucosone and (B) L-fucose with HPAEC-PAD. For reaction conditions, see Materials and methods. The retention times of the characteristic reaction products are 16.6 min (2-keto-D-gluconic acid generated from D-glucosone; Panel A) and 8.9 min (L-fuconic acid, generated from L-fucose; panel B). "1x PQQ" means PQQ-saturated *Cc*PDH (1 μ M), whereas "10x PQQ" means that an additional 9 μ M of PQQ was added to the reaction mixture. The insert in panel B shows the L-fuconic acid peak; note that the response factor for this compound is so low that it is hardly detectable in most experiments.



Time (min) Figure S2. Products generated by (A) *Nc*LPMO9F or (B) *Nc*LPMO9C from PASC using *Cc*PDH as electron donor. The figure shows that the LPMOs released oxidized oligosaccharides only when *Cc*PDH and its substrate, D-glucosone or L-fucose, were supplied together. Neither 2-keto-D-gluconic acid or L-fuconic acid, the corresponding products of *Cc*PDH, were able to activate the LPMOs, independently of the presence of *Cc*PDH. Reaction mixtures contained 0.2% (w/v) PASC in 20 mM HEPES buffer at pH 7.0 and were incubated at 30 °C for 10 min. The reactions contained 1 μ M *Cc*PDH with 1 mM D-glucosone (black dotted line) or L-fucose (red dotted line) as electron donating system. Control experiments

dotted line) or L-fucose (red dotted line) as electron donating system. Control experiments included 1 μ M *Cc*PDH with 1 mM 2-keto-D-gluconic acid (black solid line) or L-fuconic acid (red solid line), 1 μ M *Cc*PDH only (green line), 1 mM 2-keto-D-gluconic acid only (black dashed line) and 1 mM L-fuconic acid only (red dashed line).

FIGURE S2

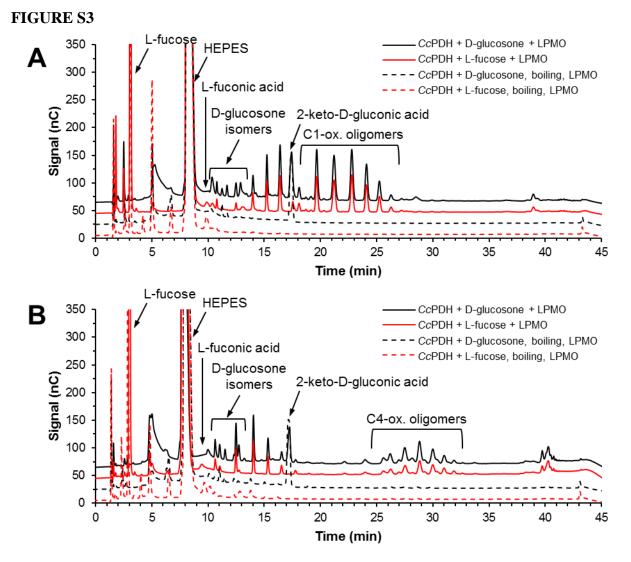


Figure S3. HPAEC-PAD chromatograms showing lack of (A) *Nc*LPMO9F or (B) *Nc*LPMO9C activity by boiled *Cc*PDH products. Products generated by (A) *Nc*LPMO9F or (B) *Nc*LPMO9C from PASC, using *Cc*PDH as electron donor or boiled *Cc*PDH products. The LPMOs were co-incubated with *Cc*PDH and D-glucosone (black solid line) or *Cc*PDH and L-fucose (red solid line), or with a boiled reaction mixture of *Cc*PDH pre-incubated with D-glucosone (black dashed line) or L-fucose (red dashed line). Reaction mixtures (yielding the solid lines) contained 0.2% (w/v) PASC, 1 μ M LPMO and 1 μ M *Cc*PDH in 20 mM HEPES buffer at pH 7.0 and were supplemented with 10 mM CaCl₂, 1 mM PQQ and 1 mM D-glucosone (black) or L-fucose (red) and were incubated for 10 min at 30 °C. Reactions done to assess the effect of possibly unknown additional reaction products of *Cc*PDH were incubated at 30 °C for 10 min and boiled for 5 min prior to addition of 1 μ M LPMO, followed by another incubation at 30 °C for 10 min.

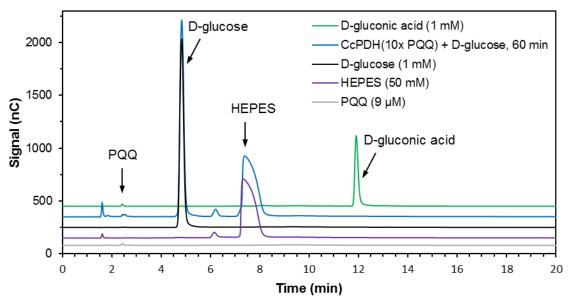


FIGURE S4

Figure S4. HPAEC-PAD chromatograms showing the lack of activity of *Cc*PDH on D-glucose. Comparison of the four standards with the reaction mixture obtained after incubating *Cc*PDH with 1 mM D-glucose and an additional 9 μ M of PQQ in 50 mM HEPES pH 7.0, at 30 °C, for 60 minutes, shows that glucose had not been converted by *Cc*PDH.

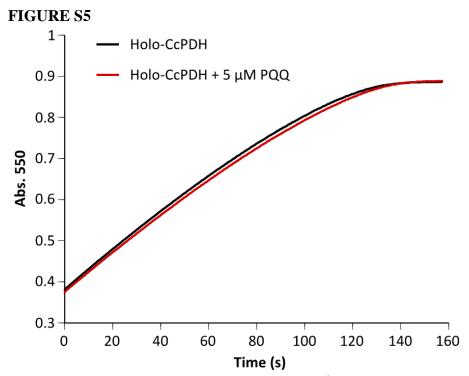


Figure S5. The effect of adding extra PQQ and Ca²⁺ on the activity of holo-*Cc*PDH. The curves show reduction of cytochrome *c* over time in reactions containing 200 nM presumably saturated *Cc*PDH and 50 μ M cytochrome *c* in 50 mM PIPES-NaOH (pH 7.0), 1 mM L-fucose. The curve labeled "+ 5 μ M PQQ" shows results for a similar reaction mixture which was supplemented with additional PQQ and CaCl₂ (50 μ M). The fact that the two curves are identical indicates that the presumably saturated *Cc*PDH indeed was saturated.