

**S1 Table. Strains and plasmids.**

Strain or plasmid	Genotype or description	Reference
<b><i>A. baumannii</i></b>		
ATCC 17978	cerebrospinal fluid isolate	[1]
EGA195	ATCC 17978 $\Delta bfmS::aacC1$ (Gm <sup>R</sup> )	[2]
EGA496	ATCC 17978 $\Delta bfmR::aacC1$	this study
EGA495	ATCC 17978 $\Delta bfmRS::aacC1$	this study
EGA497	ATCC 17978 $\Delta bfmRS::aacC1::pEGE152$	this study
EGA501	ATCC 17978 $\Delta bfmRS::aacC1::pEGE224$	this study
EGA507	ATCC 17978 $\Delta bfmRS::aacC1::pEGE234$	this study
EGA465	ATCC 17978 $\Delta adeIJK$	this study
EGA513	ATCC 17978 $\Delta adc$	this study
EGA514	ATCC 17978 $\Delta oxa51$	this study
EGA515	ATCC 17978 $\Delta adc \Delta oxa51$	this study
EGA516	ATCC 17978 $\Delta ampG$	this study
EGA517	ATCC 17978 $\Delta ampG \Delta bfmRS::aacC1$	this study
EGA518	ATCC 17978 $\Delta ampG \Delta bfmS::aacC1$	this study
EGA624	ATCC 17978 $\Delta adc \Delta bfmS::aacC1$	this study
EGA534	ATCC 17978 $\Delta bfmRS::aacC1 cspC(G17AfsX21)$	this study
EGA552	ATCC 17978 $\Delta bfmRS::aacC1 ACX60\_RS05385$ (NUDIX):: <i>ISAb1</i>	this study
EGA555	ATCC 17978 $\Delta bfmRS::aacC1 mnmA(G362S)$	this study
EGA692	ATCC 17978 $\Delta pbp2$	this study
EGA694	ATCC 17978 $\Delta pbp2 \Delta bfmS::aacC1$	this study
EGA695	ATCC 17978 $\Delta pbp2 \Delta bfmRS::aacC1$	this study
EGA68	ATCC 17978 $\Delta KL3::aacC1$	[2]
EGA127	ATCC 17978 $bfmS^{1-467}$ (G467DfsX19)	[2]
EGA187	ATCC 17978 $bfmS^{1-467}$ (G467DfsX19) $\Delta KL3::aacC1$	this study
ATCC 19606	urine isolate	[1]
EGA216	ATCC 19606 $\Delta bfmS::aacC1$	[2]
AB5075-UW	bone isolate/osteomyelitis	[3]
AB08299	AB5075-UW $bfmS::T26$	[3]
<b><i>E. coli</i></b>		
DH5 $\alpha$	<i>supE44</i> $\Delta lacU169$ ( $\phi 80lacZ\Delta M15$ ) <i>hsdR17 recA1 endA1 gyrA96 thi-1 relA1</i>	[4]
DH5 $\lambda$ pir	DH5 $\alpha$ ( $\lambda$ pir) <i>tet::Mu recA</i>	[5]
TO60	DH5 $\alpha$ $\lambda$ pir [F' <i>proAB lac<sup>q</sup>\Delta M15 Tn10</i> (Tc <sup>R</sup> )]	[6]
<b>plasmids</b>		
pUC18	<i>oriColE1</i> MCS Cb <sup>R</sup>	[7]
pSR47s	<i>oriTRP4 oriR6K</i> Km <sup>R</sup>	[8]
pEGE148	pSR47s:: <i>aacC1</i> Km <sup>R</sup> Gm <sup>R</sup>	[2]
pEGE152	pEGE148:: <i>bfmR bfmS</i>	[2]
pEGE224	pEGE148:: <i>bfmR bfmS</i> (H324Q, 3XFLAG)	this study
pEGE234	pEGE148:: <i>bfmR</i> (3XFLAG)	this study
pJB1801	<i>oriRSF1010</i> Cb <sup>R</sup>	[2]
pEGE144	pJB1801:: <i>bfmR bfmS</i>	this study

pEGE146	pJB1801:: <i>bfmR bfmS</i> (G494V)	this study
pWH1266	shuttle plasmid, Tc <sup>R</sup> Cb <sup>R</sup>	[9]
pFPV25	promoter trap vector containing promoterless <i>gfpmut3</i>	[10]
pEGE244	pWH1266 with <i>rrnB</i> , T7 tandem terminators replacing EcoRI-PstI fragment, Tc <sup>R</sup>	this study
pEGE245	pEGE244 with SacI-PstI fragment of pFPV25	this study
pEGE313	pEGE245 with <i>adcp</i> inserted upstream of <i>gfpmut3</i>	this study
pEGE314	pEGE245 with <i>oxa51p</i> inserted upstream of <i>gfpmut3</i>	this study
pEGE315	pEGE245 with <i>sItp</i> inserted upstream of <i>gfpmut3</i>	this study
pEGE253	pEGE245 with <i>ygeRp</i> inserted upstream of <i>gfpmut3</i>	this study
pEGE254	pEGE245 with <i>tolBp</i> inserted upstream of <i>gfpmut3</i>	this study
pEGE246	pEGE245 with <i>ACX60_RS18040p</i> inserted upstream of <i>gfpmut3</i>	this study
pEGE247	pEGE245 with <i>ompW(ACX60_RS05905)p</i> inserted upstream of <i>gfpmut3</i>	this study
pEGE248	pEGE245 with <i>ACX60_RS13710p</i> inserted upstream of <i>gfpmut3</i>	this study
pEGE251	pEGE245 with <i>ACX60_RS09685p</i> inserted upstream of <i>gfpmut3</i>	this study
pEGE304	pWH1266 with EcoRI-PstI fragment replaced by polylinker, Tc <sup>R</sup>	this study
pCA24N- <i>dinB</i>	Source of <i>lacI<sup>q</sup></i> , T5- <i>lacp</i> fragment	[11, 12]
pEGE305	pEGE304 with <i>lacI<sup>q</sup></i> , T5- <i>lacp</i> fragment, Tc <sup>R</sup>	this study
pEGE316	pEGE305 with <i>sItp</i> gene downstream of T5- <i>lacp</i> promoter	this study

### References

- Bouvet P, Grimont P. Taxonomy of the Genus *Acinetobacter* with the Recognition of *Acinetobacter baumannii* sp. nov. *Acinetobacter haemolyticus* sp. nov. *Acinetobacter johnsonii* sp. nov. and *Acinetobacter junii* sp. nov. and Emended Descriptions of *Acinetobacter calcoaceticus* and *Acinetobacter lwoffii*. *International Journal of Systematic Bacteriology*. 1986;36(2):228-40.
- Geisinger E, Isberg RR. Antibiotic modulation of capsular exopolysaccharide and virulence in *Acinetobacter baumannii*. *PLoS Pathog*. 2015;11(2):e1004691. Epub 2015/02/14. doi: 10.1371/journal.ppat.1004691 PPATHOGENS-D-14-02426 [pii]. PMID: 25679516.
- Gallagher LA, Ramage E, Weiss EJ, Radey M, Hayden HS, Held KG, et al. Resources for Genetic and Genomic Analysis of Emerging Pathogen *Acinetobacter baumannii*. *J Bacteriol*. 2015;197(12):2027-35. doi: 10.1128/JB.00131-15. PMID: 25845845.
- Hanahan D, Jessee J, Bloom FR. Plasmid transformation of *Escherichia coli* and other bacteria. *Methods Enzymol*. 1991;204:63-113. Epub 1991/01/01. doi: 0076-6879(91)04006-A [pii]. PMID: 1943786.
- Kolter R, Inuzuka M, Helinski DR. Trans-complementation-dependent replication of a low molecular weight origin fragment from plasmid R6K. *Cell*. 1978;15(4):1199-208. Epub 1978/12/01. doi: 0092-8674(78)90046-6 [pii]. PMID: 728998.
- O'Connor TJ, Adepoju Y, Boyd D, Isberg RR. Minimization of the *Legionella pneumophila* genome reveals chromosomal regions involved in host range expansion. *Proc Natl Acad Sci U S A*. 2011;108(36):14733-40. PMID: 21873199.
- Yanisch-Perron C, Vieira J, Messing J. Improved M13 phage cloning vectors and host strains: nucleotide sequences of the M13mp18 and pUC19 vectors. *Gene*. 1985;33(1):103-19. Epub 1985/01/01. PMID: 2985470.
- Andrews HL, Vogel JP, Isberg RR. Identification of linked *Legionella pneumophila* genes essential for intracellular growth and evasion of the endocytic pathway. *Infect Immun*. 1998;66(3):950-8. PMID: 9488381.
- Hunger M, Schmucker R, Kishan V, Hillen W. Analysis and nucleotide sequence of an origin of DNA replication in *Acinetobacter calcoaceticus* and its use for *Escherichia coli* shuttle plasmids. *Gene*. 1990;87(1):45-51. PMID: 2185139.
- Valdivia RH, Falkow S. Bacterial genetics by flow cytometry: rapid isolation of *Salmonella typhimurium* acid-inducible promoters by differential fluorescence induction. *Mol Microbiol*. 1996;22(2):367-78. PMID: 8930920.
- Kitagawa M, Ara T, Arifuzzaman M, Ioka-Nakamichi T, Inamoto E, Toyonaga H, et al. Complete set of ORF clones of *Escherichia coli* ASKA library (a complete set of *E. coli* K-12 ORF archive): unique resources for biological research. *DNA Res*. 2005;12(5):291-9. Epub 2006/06/14. doi: 10.1093/dnares/dsi012. PMID: 16769691.
- Macquire AE, Ching MC, Diamond BH, Kazakov A, Novichkov P, Godoy VG. Activation of phenotypic subpopulations in response to ciprofloxacin treatment in *Acinetobacter baumannii*. *Mol Microbiol*. 2014;92(1):138-52. Epub 2014/03/13. doi: 10.1111/mmi.12541. PMID: 24612352.