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1170 1180 1190 1200
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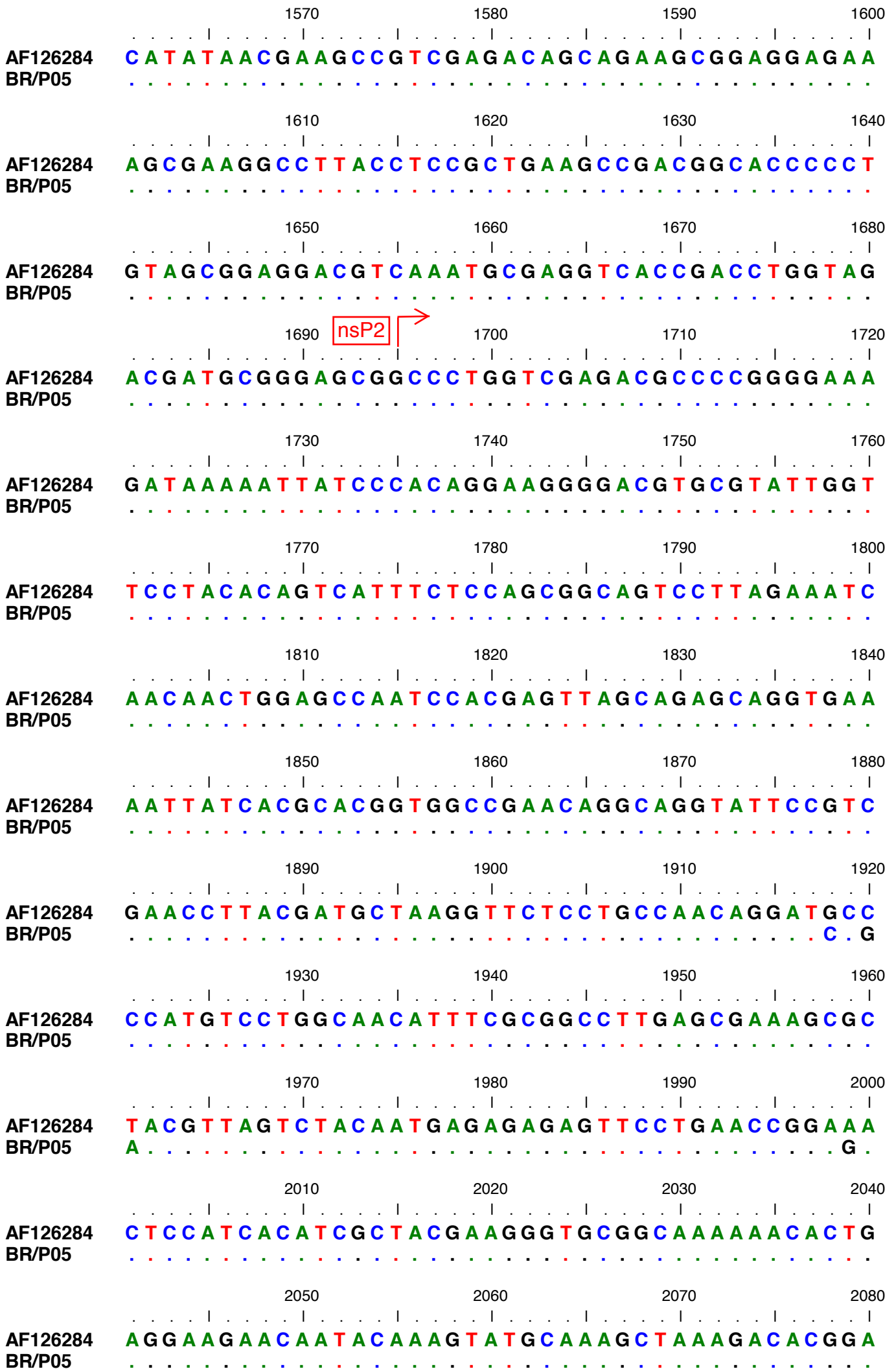
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 T C G G

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 BR/P05
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2690 2700 2710 2720
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2730 2740 2750 2760
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2770 2780 2790 2800
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2890 2900 2910 2920
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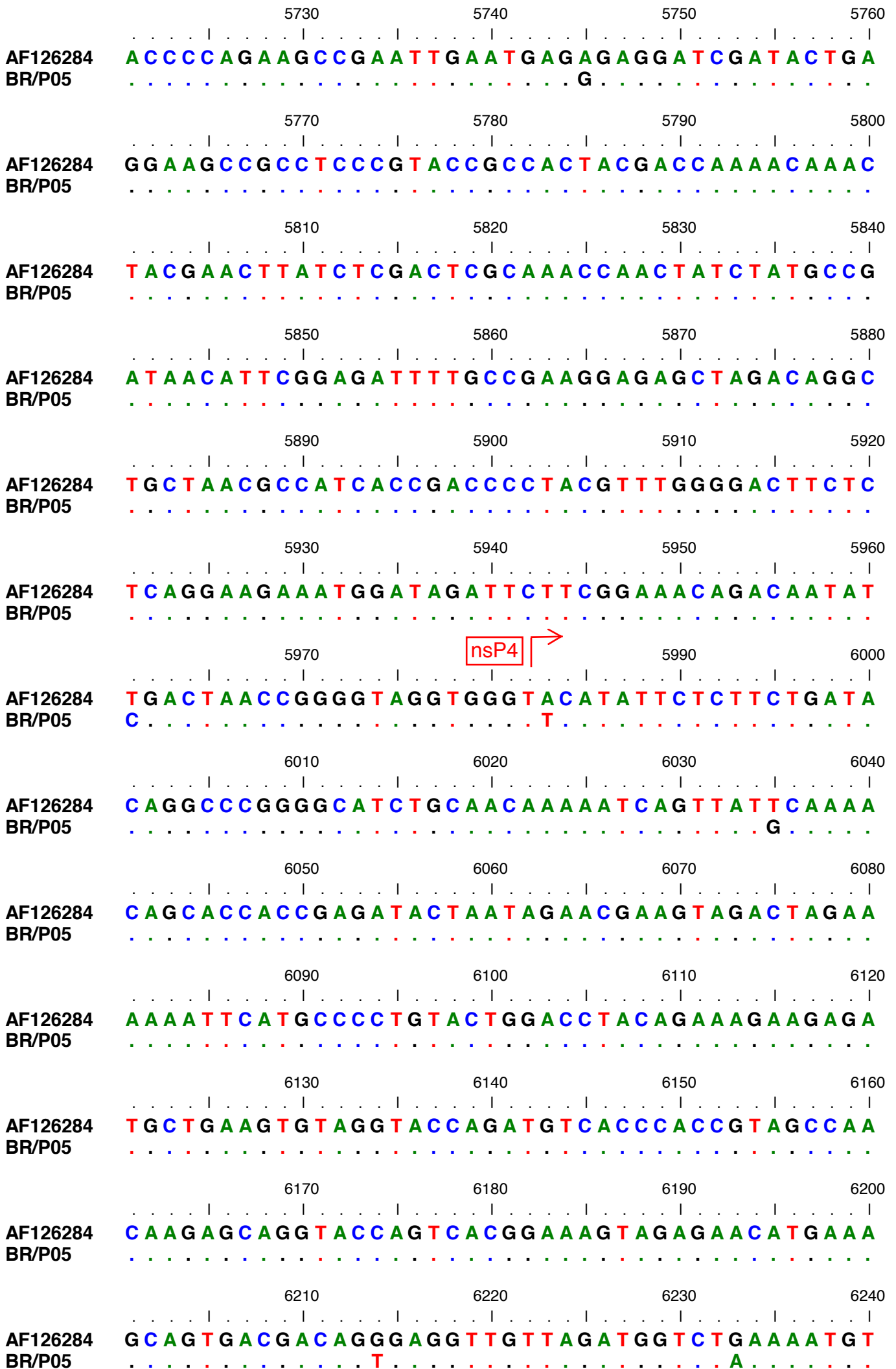
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AF126284 BR/P05
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BR/P05
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BR/P05
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7930 7940 7950 7960

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BR/P05
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C G

7970 7980 7990 8000

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BR/P05
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8010 8020 8030 8040

AF126284
BR/P05
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G

8050 8060 8070 8080

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G C G

8090 8100 8110 8120

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BR/P05
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8130 8140 8150 8160

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8170 8180 8190 8200

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C

8210 8220 8230 8240

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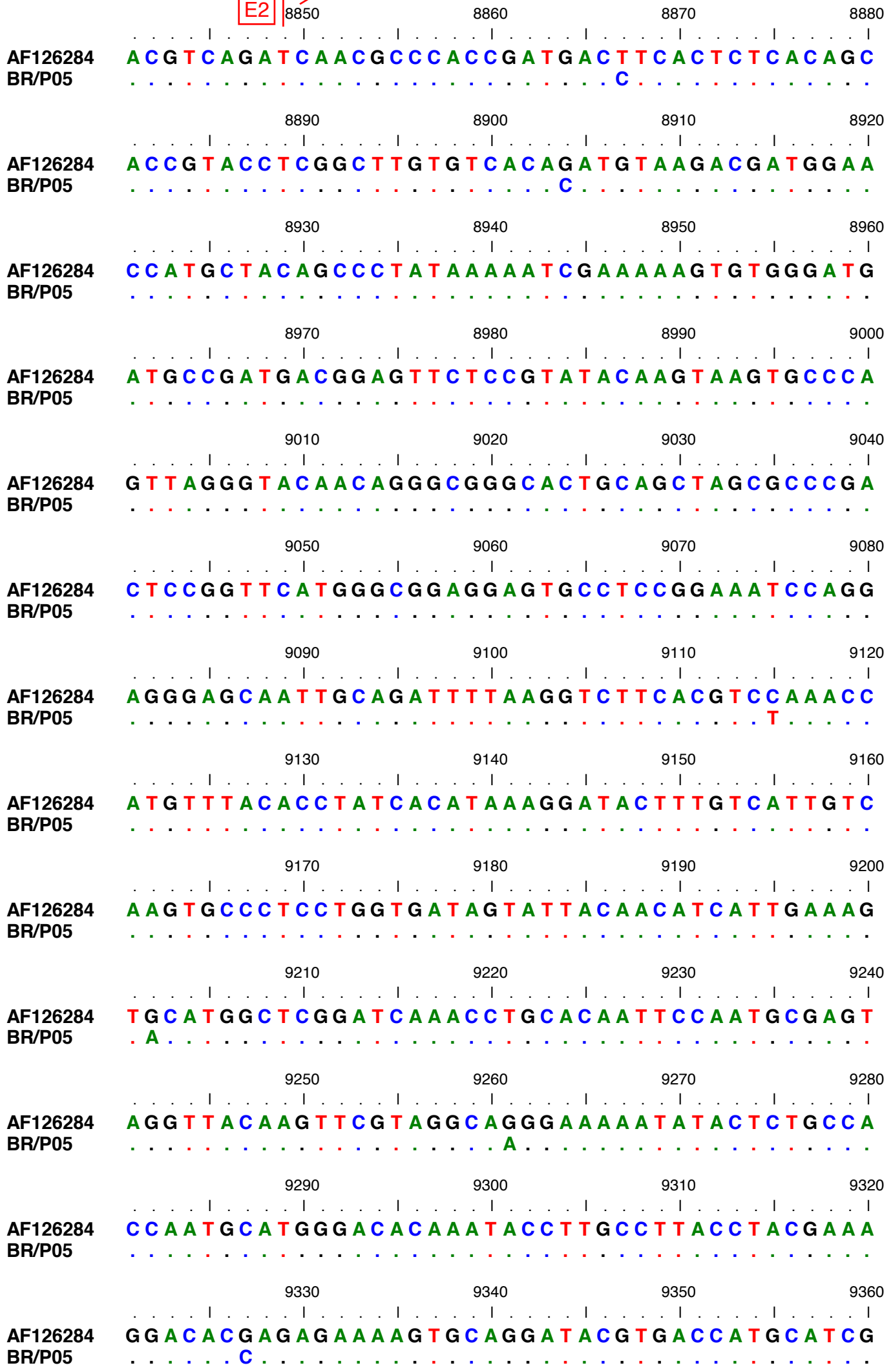
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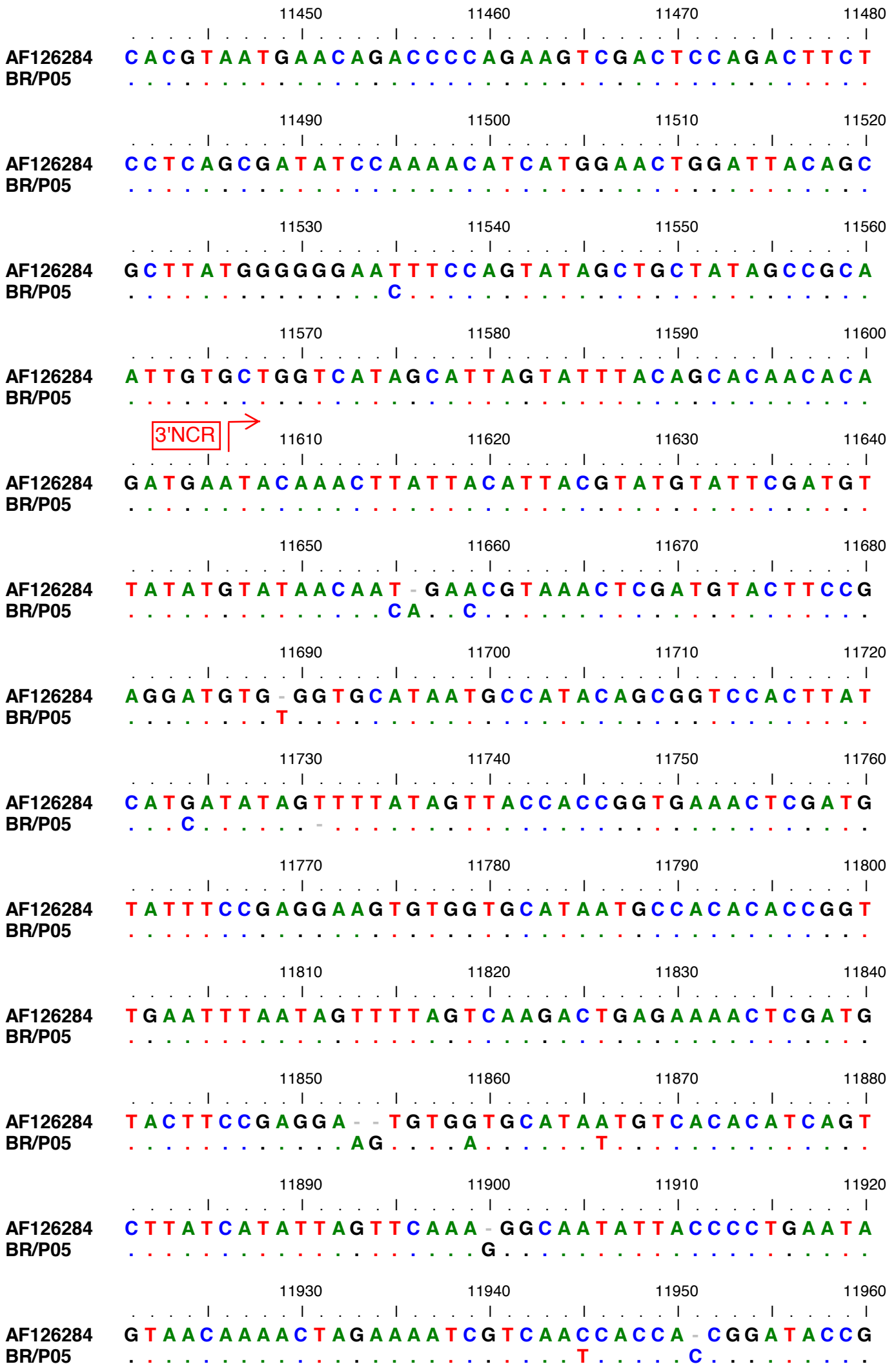
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ORF1

nsP1



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BR/P05

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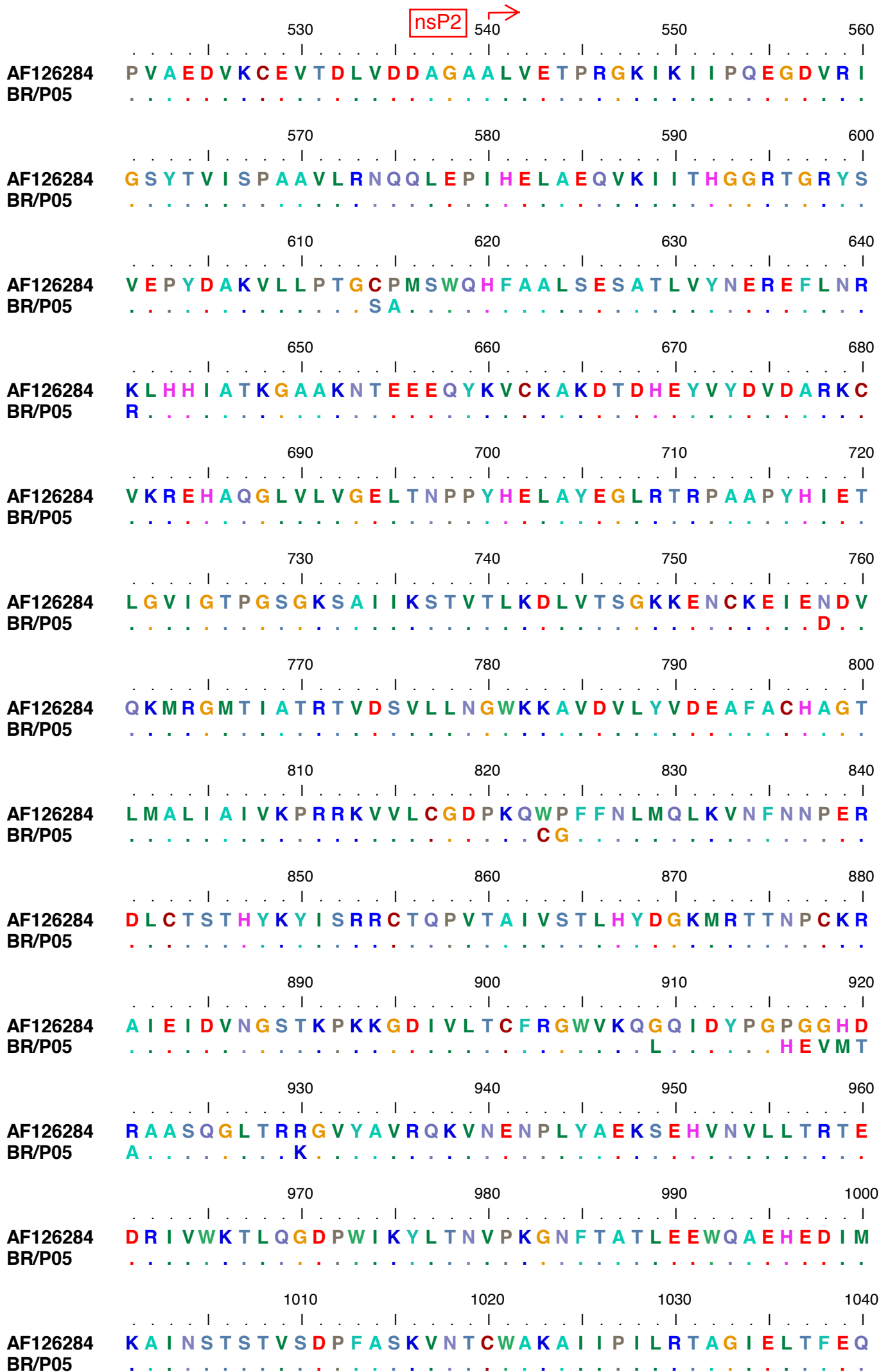
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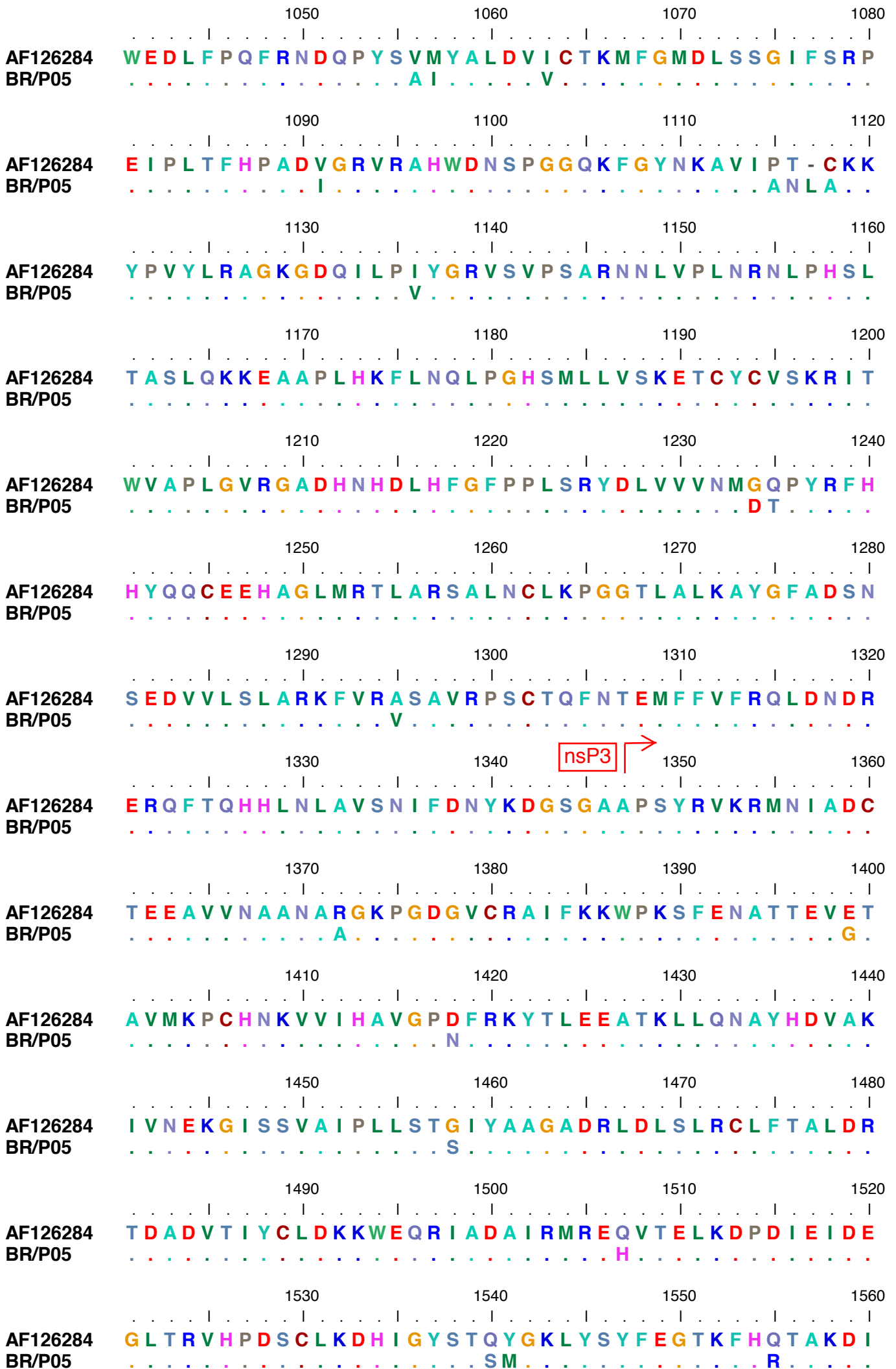
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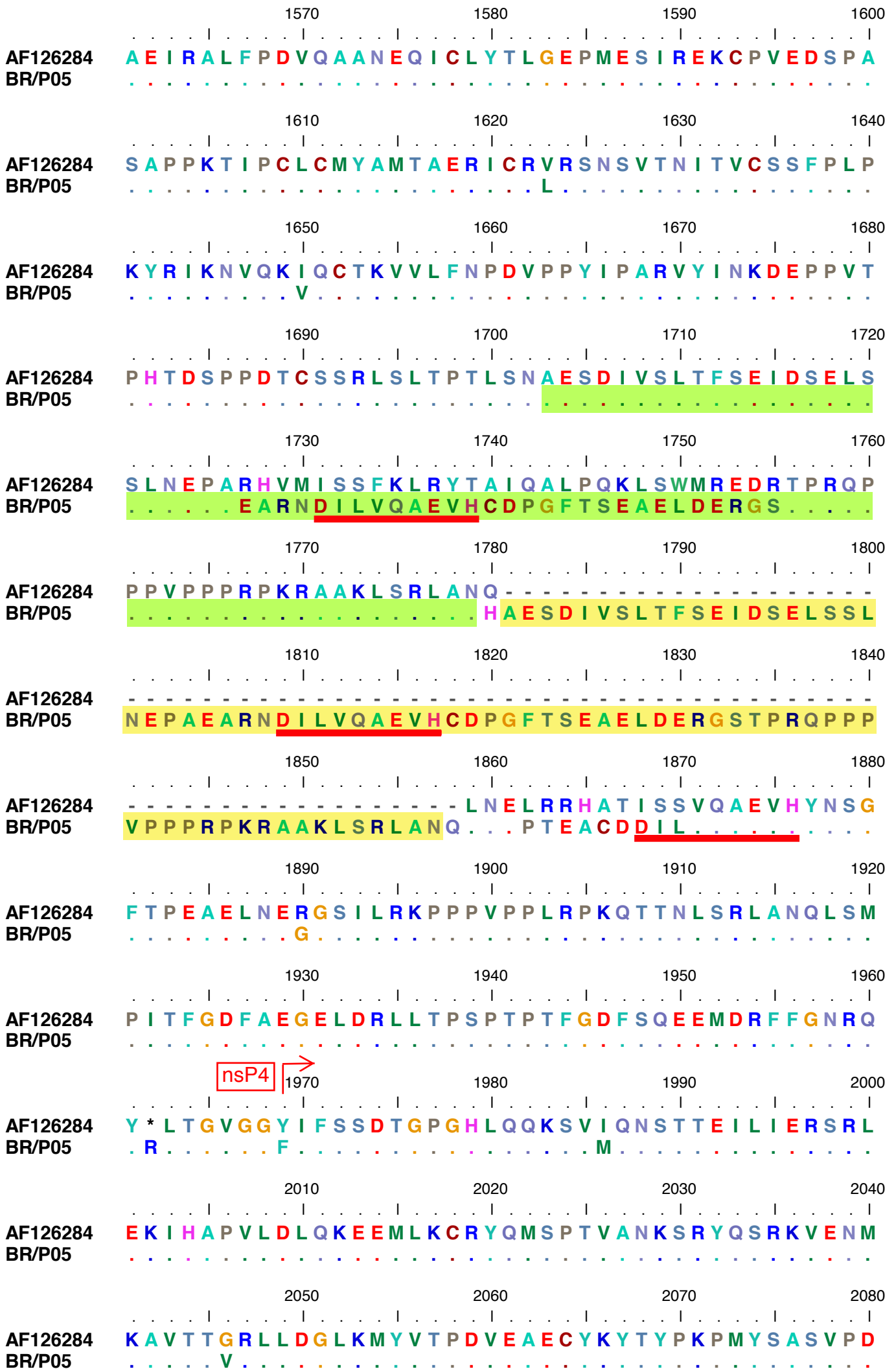
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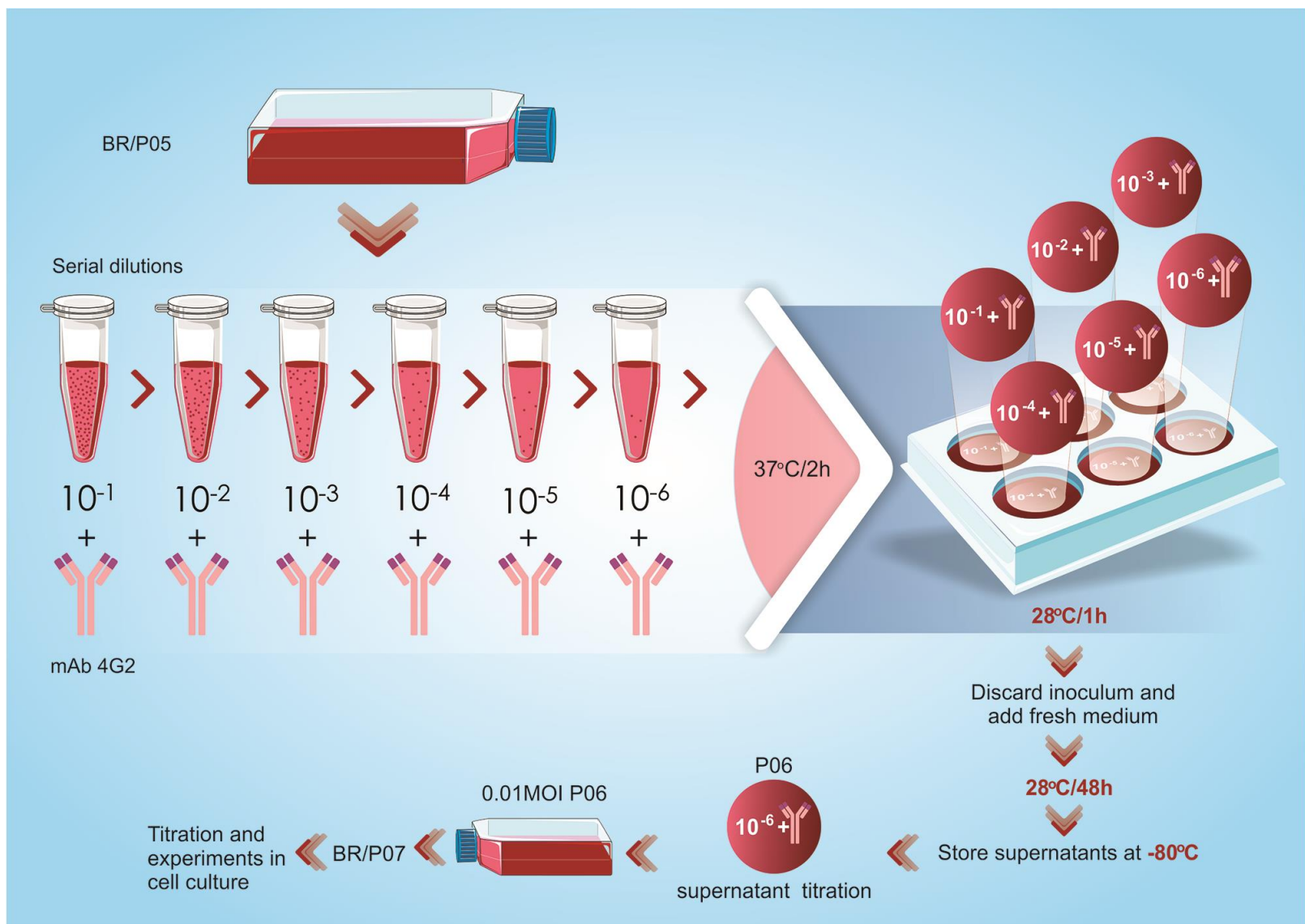


Figure S1. Workflow depicting the protocol used for DENV neutralization and production of the AURAV viral stock (BR/P07) from BR/P05. Different dilutions (10^{-1} - 10^{-6}) of the supernatant of BR/P05 were incubated at 37 °C with an anti-flavivirus monoclonal antibody (4G2) for 2 h. This mixture was incubated with C6/36 cells (3.5×10^5 cells/well, seeded the day before in a 6-well plate) for 1 h at 28 °C. The inoculum was discarded, and the cell monolayer washed once with sterile PBS, after which 3 ml/well of medium added. The supernatants were collected at two days post-infection, aliquoted and stored at -80 °C. The supernatant of the infection (P06) that was carried out with the least amount of virus (10^{-6}) was titrated and used to infect C6/36 cells at a multiplicity of infection (MOI) of 0.01 to produce the viral stock (BR/P07) for biological characterization experiments. Figure by Wagner Nagib de Souza Birbeire.

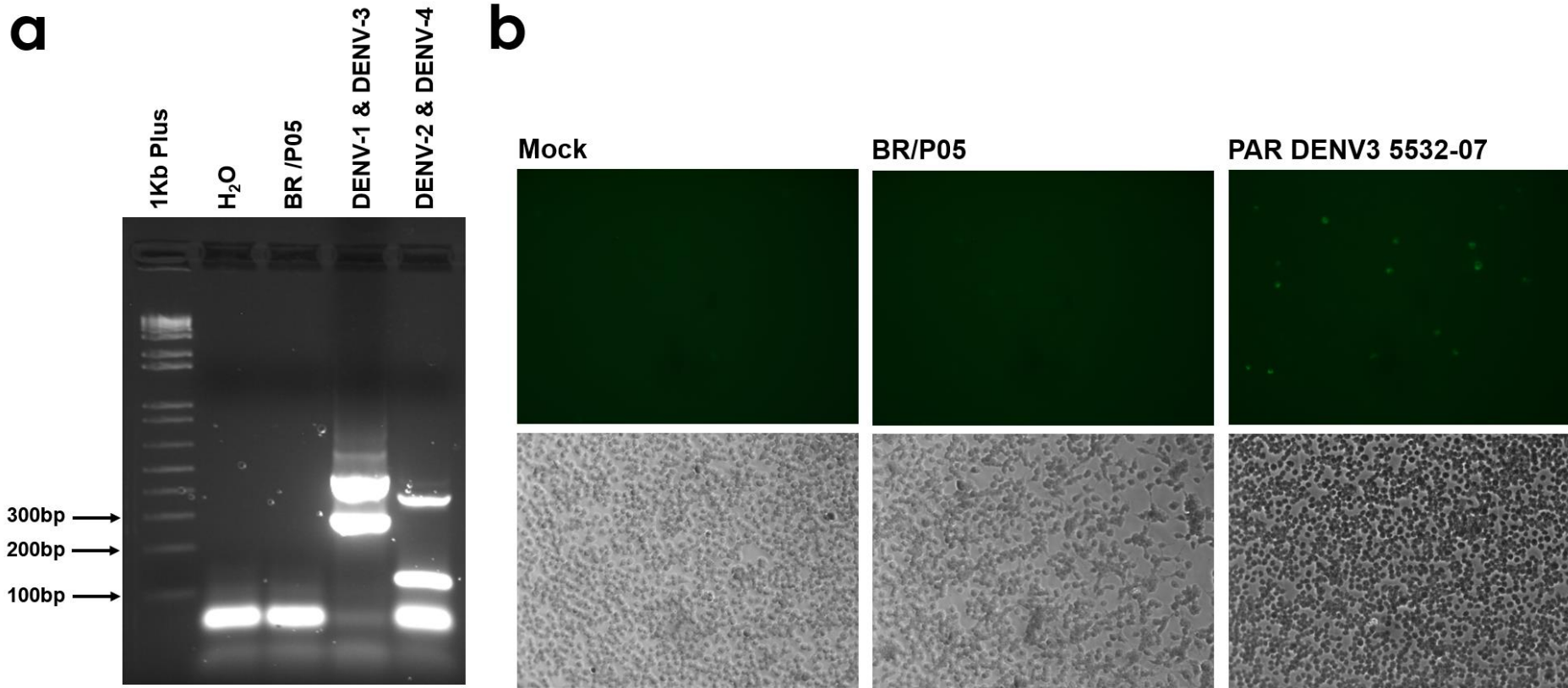


Figure S2. DENV detection in BR/P05. **a** One-step RT-PCR for DENV serotyping. RNA extracted from the supernatant of BR/P05 was used as the template. Water was used as negative control, and RNAs extracted from DENV-1, -2, -3 and 4 were used as positive controls. **b** IFA of C6/36 cells infected with the supernatant of BR/P05 at 2 days post-infection. Cells were fixed with 3% paraformaldehyde, permeabilized with 0.5% triton and incubated with an anti-flavivirus monoclonal antibody (4G2) followed by a goat anti-mouse IgG (H+L), Alexa Fluor 488 conjugate. Images were obtained at 200x magnification using a Nikon Eclipse TE300 inverted microscope attached to a CoolSNAP-ProCf camera and visualized and processed using Image-Pro Plus version 4.5.1.22 software.

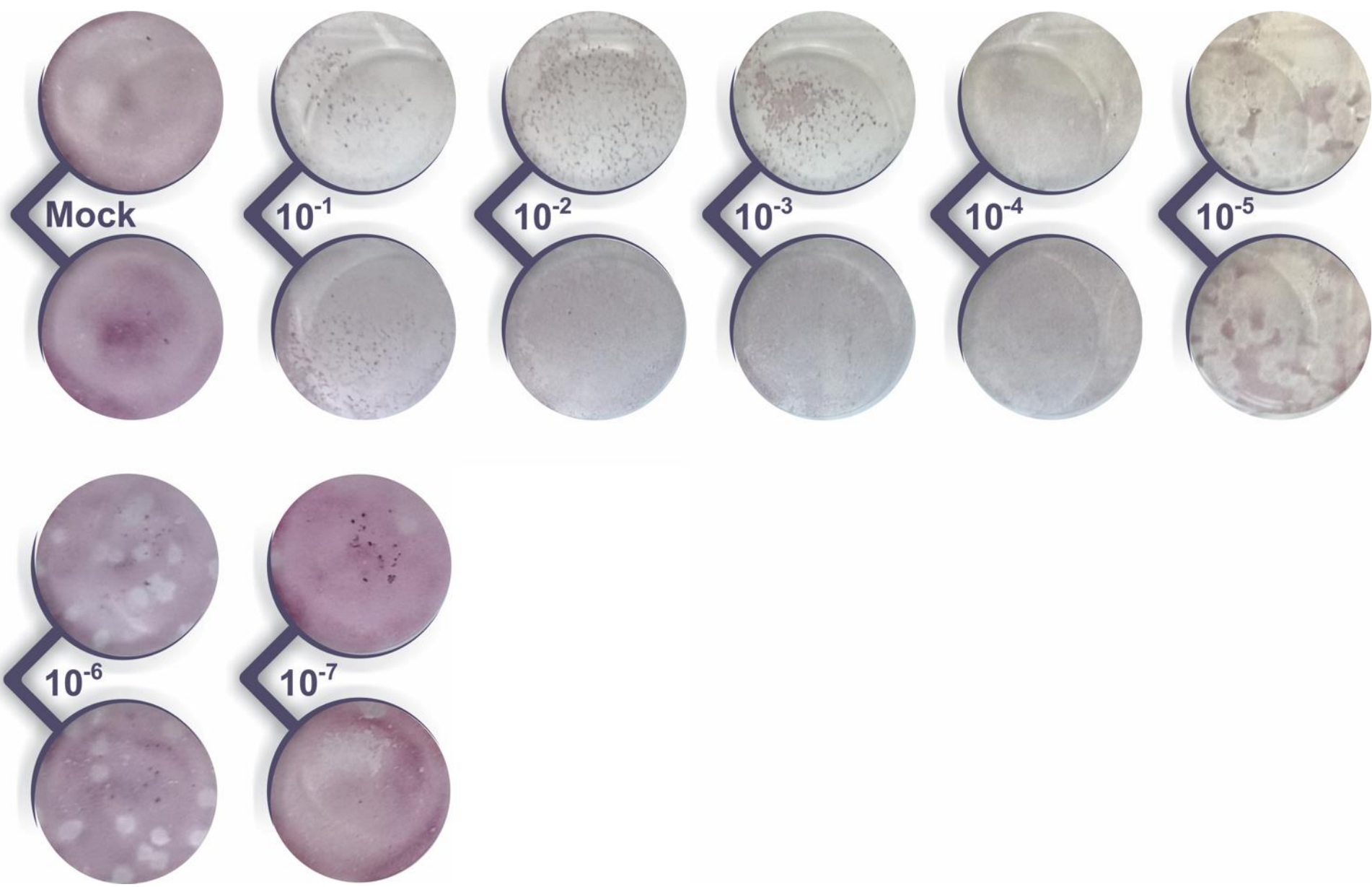
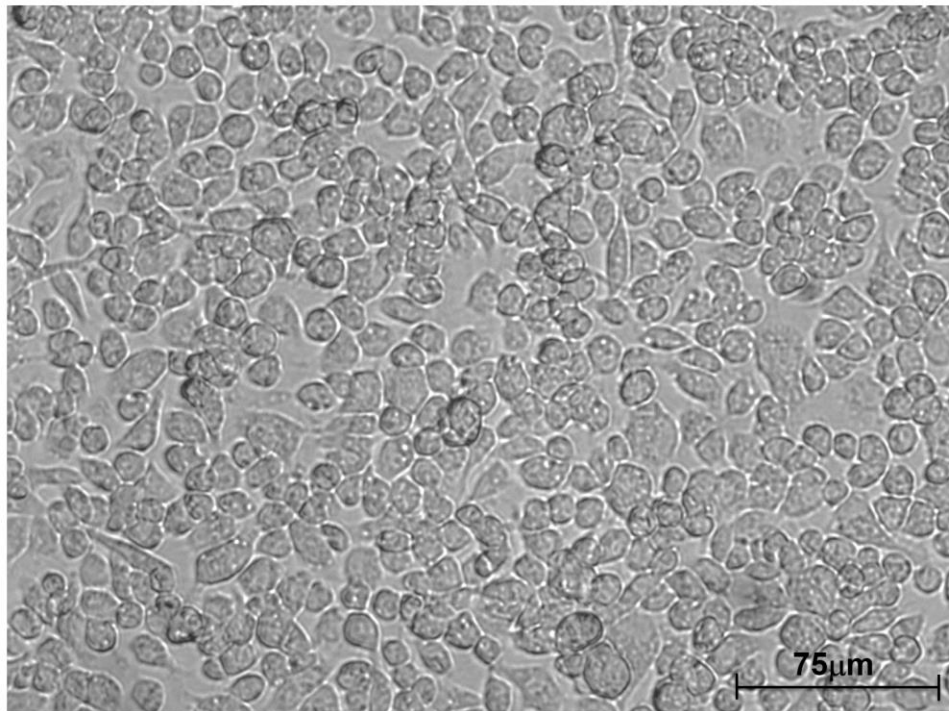


Figure S3. Result of BR/P07 titration by plaque assay in C6/36 cells.

MOCK



BR/P07

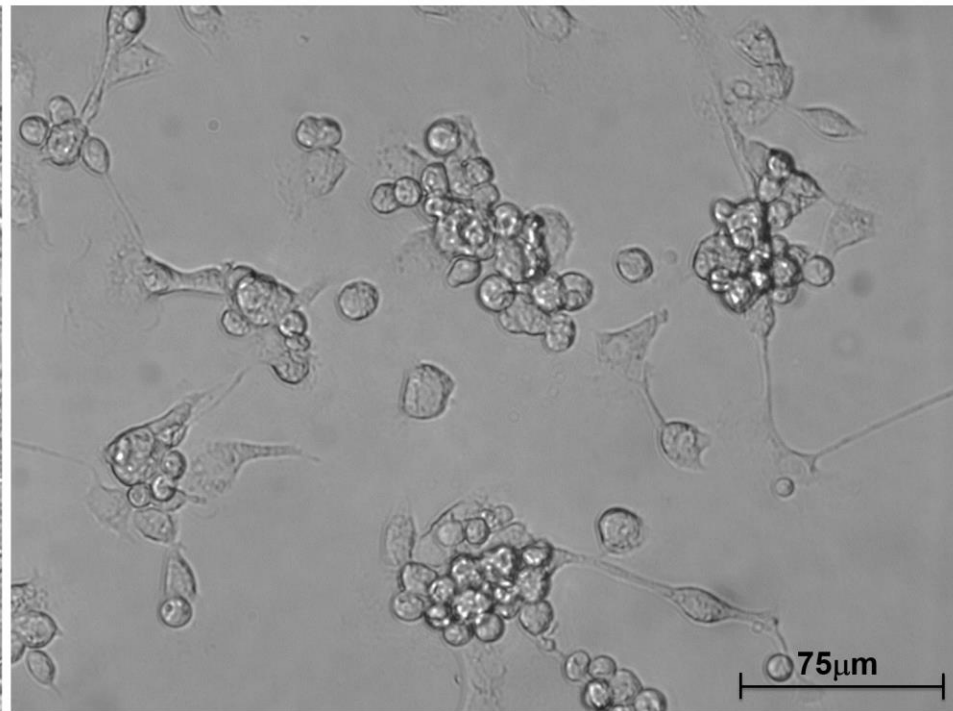


Figure S4. Brightfield images of C6/36 cells infected or not (mock) with 0.1 MOI of BR/P07 at 3 days post-infection. Images were obtained at 400x magnification using a Nikon Eclipse TE300 inverted microscope attached to a CoolSNAP-ProCf camera and visualized and processed using Image-Pro Plus version 4.5.1.22 software.

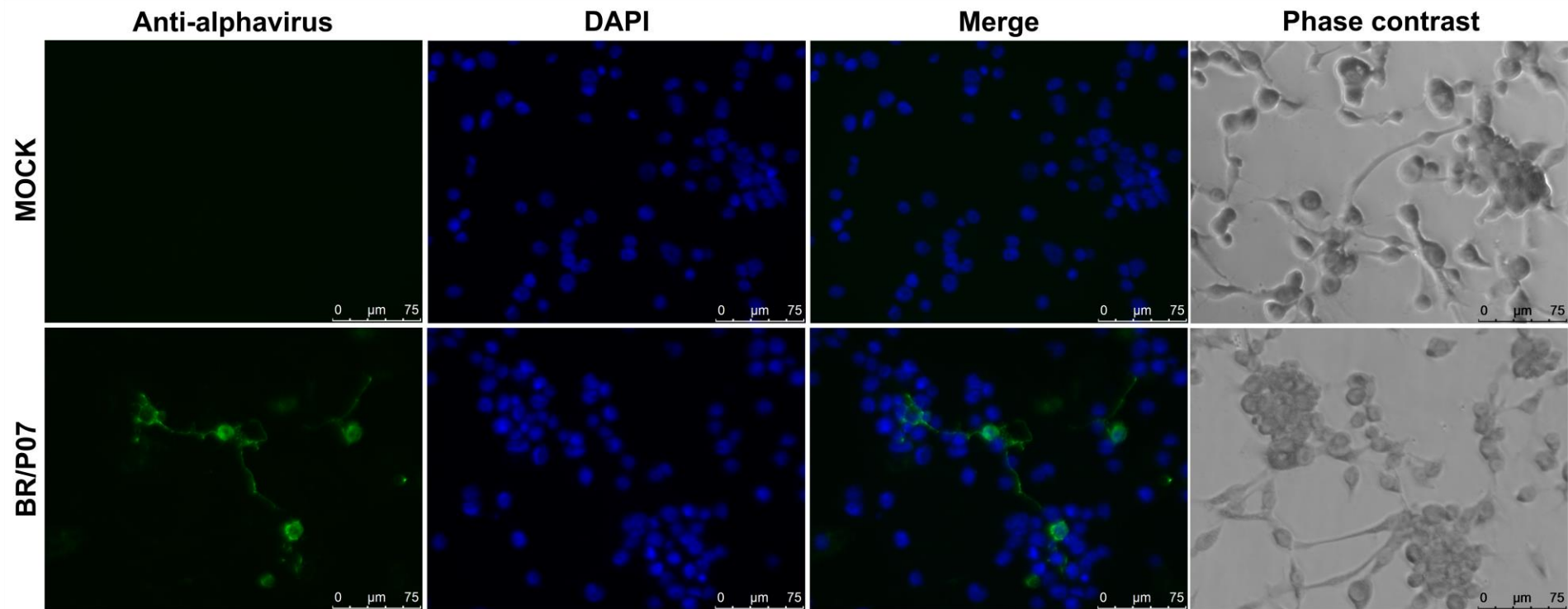


Figure S5. IFA of AP-61 cells infected or not (mock) with 1 MOI of BR/P07. At 3 days post-infection, the cell monolayer was fixed and permeabilized with methanol:acetone (1:1) at -20 °C and then incubated with an anti-alfavirus monoclonal antibody (mAb) clone 1A4B-6 followed by a goat anti-mouse IgG (H+L), Alexa Fluor 488 conjugate. Images were obtained using a Leica DMI 6000B inverted microscope attached to a Leica DFC365 FX camera and visualized and processed using Leica Application Suite Advanced Fluorescence 3.1.0 software.

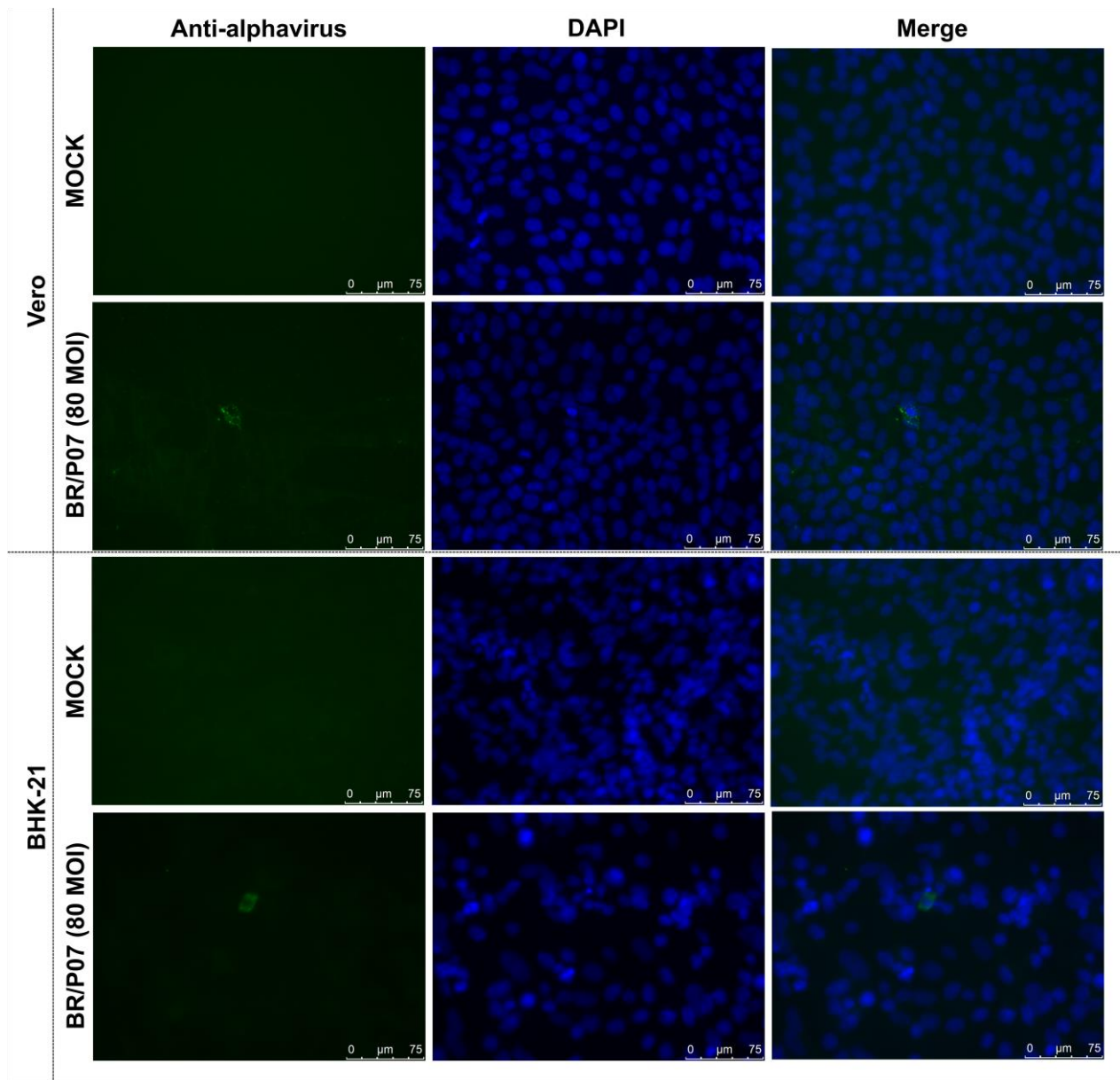


Figure S6. IFA of Vero and BHK-21 cells infected or not (mock) with 80 MOI of BR/P07. At 3 days post-infection, cell monolayers were fixed and permeabilized with methanol:acetone (1:1) at -20 °C and then incubated with an anti-alphavirus monoclonal antibody (mAb) clone 1A4B-6 followed by a goat anti-mouse IgG (H+L), Alexa Fluor 488 conjugate. Images were obtained using a Leica DMI 6000B inverted microscope attached to a Leica DFC365 FX camera and visualized and processed using Leica Application Suite Advanced Fluorescence 3.1.0 software.

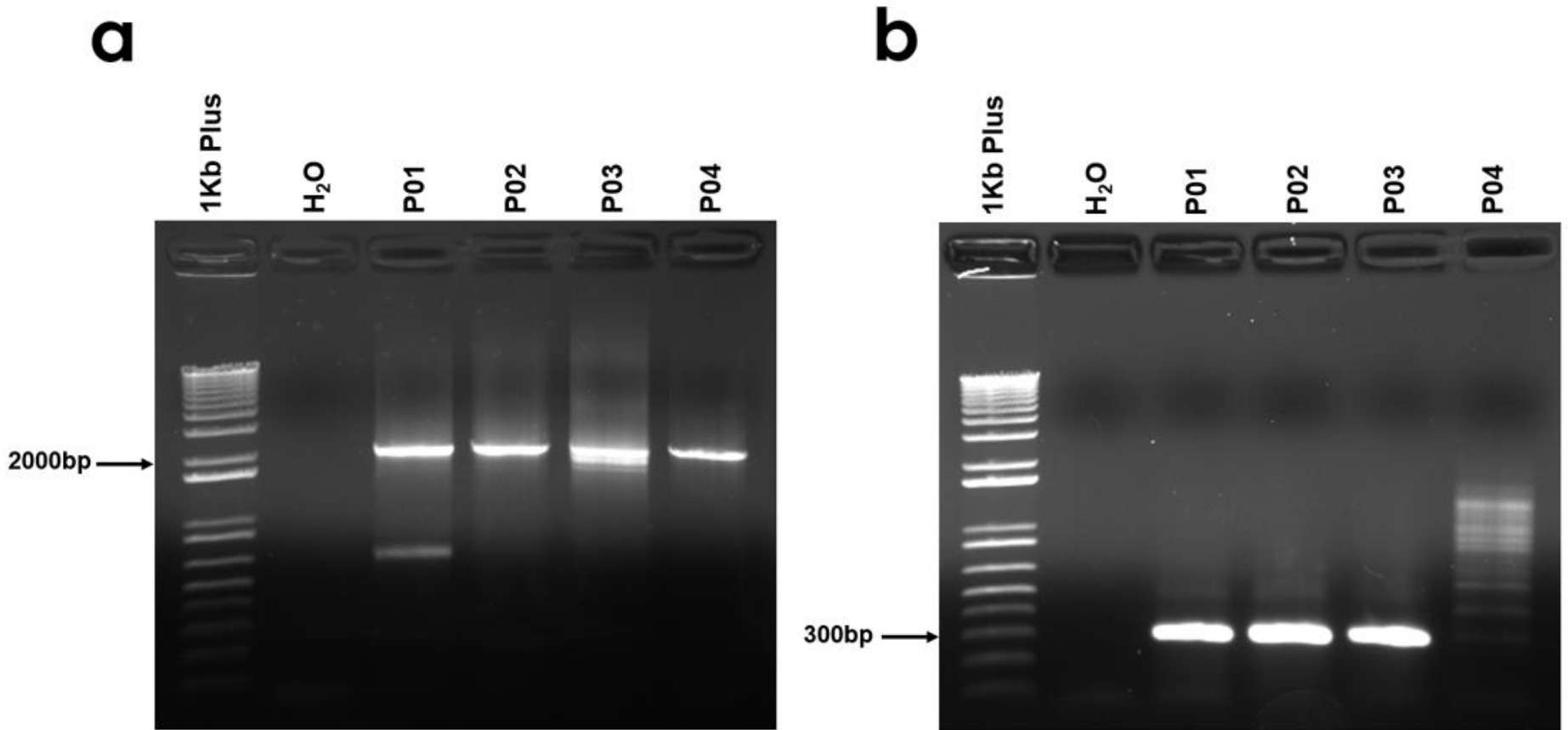
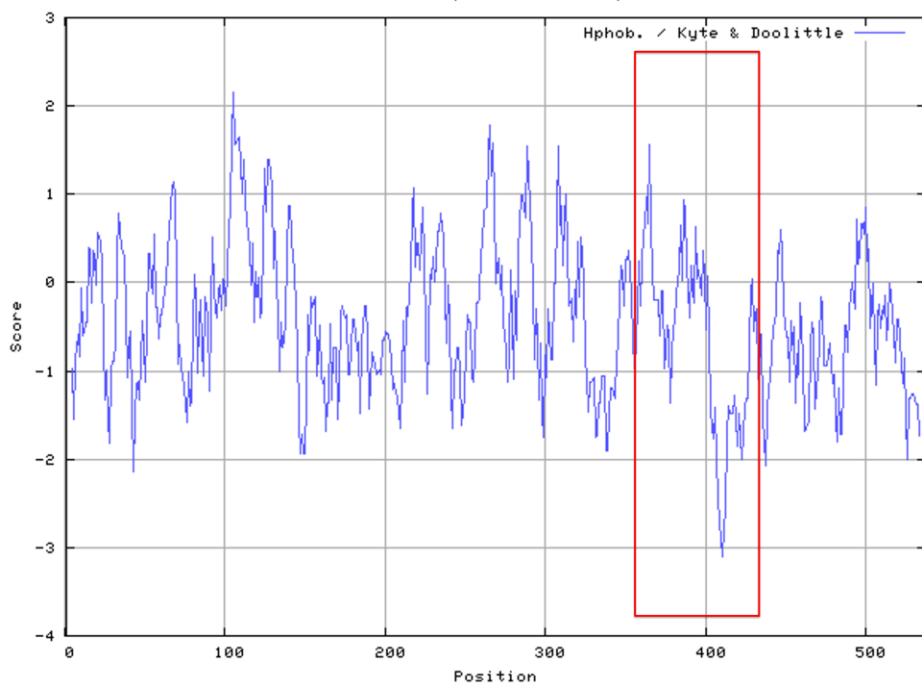


Figure S7. RT-PCR of P01-P04 for **a** AURAV and **b** DENV. RT-PCR was carried out following standard protocols. The primers used for AURAV testing were NsP3F and NsP3R (see Supplementary Table 2 for primer sequence). The primers used for DENV testing were D1 (5'-TCAATATGCTGAAACGCGCGAGAAACCG-3') and TS3 (5'-TAACATCATCATGAGACAGAGC-3'). The expected amplicon size for AURAV and DENV is 1995 bp and 290 bp, respectively.

AF126284

ProtScale output for user sequence



BR/P05

ProtScale output for user sequence

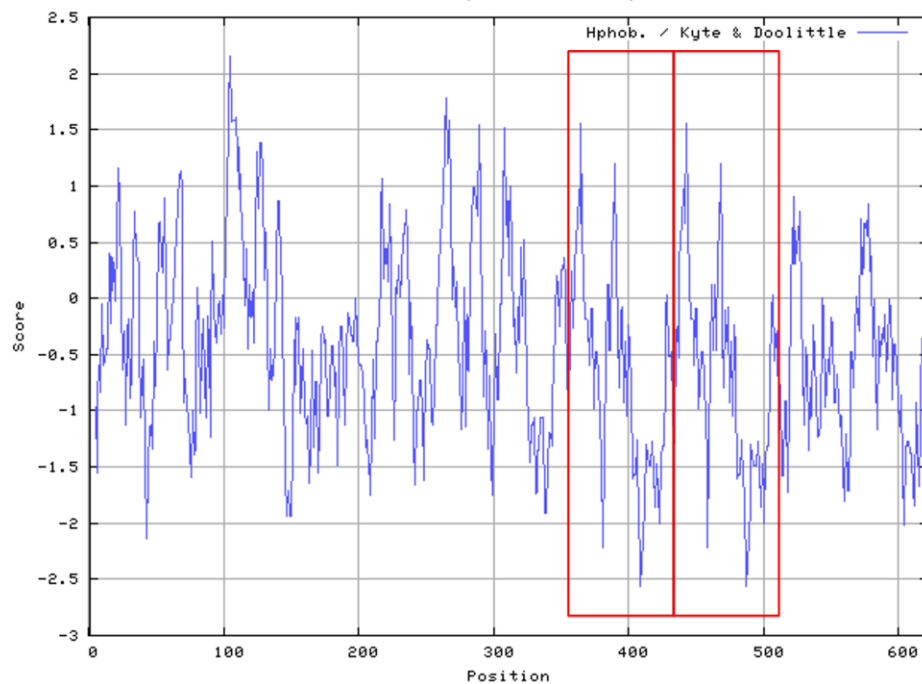


Figure S8. nsP3 hydrophobicity plot of AF126284 and BR/P05. The region corresponding to the duplication is highlighted by the red box. The Kyte & Doolittle hydrophobicity plot was drawn using the default settings, as available at the ProtScale website (<http://web.expasy.org/protscale/>).

a

Score	Expect	Identities	Gaps	Strand
176 bits(194)	1e-45	149/181(82%)	2/181(1%)	Plus/Plus
AF126284	5415	CTAAACGAGCTACGGAGGCATGCGACGATATCCTCCGTTCAAGCTGAGGTACACTACAAT	5474	
AF126284	5238	CTAAACGAGCCTGCGAGGCACGTAATGATATCCTC-GTTCAAGCTGAGGTACACTGCGAT	5296	
AF126284	5475	TCAGGTTTTACCCAGAAAGCCGAATTGAATGAGAGAGGATCG-ATACTGAGGAAGCCGCC	5533	
AF126284	5297	CCAGGCTTTACCTCAGAAGCTGAGTTGGATGAGAGAGGATCGCACACCGAGACAGCCGCC	5356	
AF126284	5534	TCCCGTACCGCCACTACGACCAAACAAACTACGAACTTATCTCGACTCGCAAACCAACT	5593	
AF126284	5357	TCCTGTACCGCCACCACGACCAAACGAGCCGCGAAATTATCCCGACTAGCAAACCAGCT	5416	
AF126284	5594	A	5594	
AF126284	5417	A	5417	

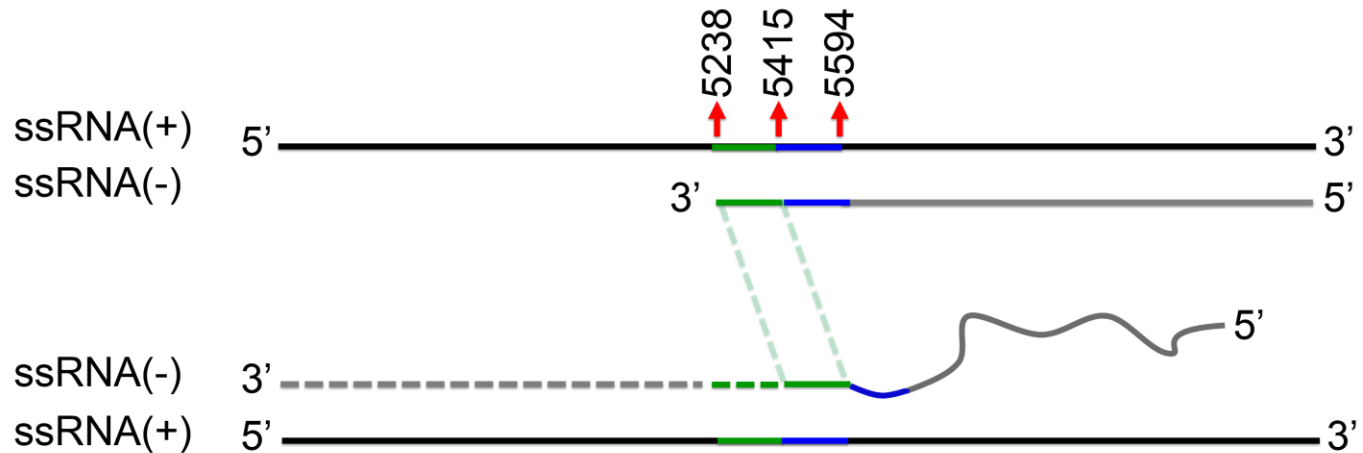
b

Figure S9. Mechanism of duplication hypothesis. **a** Partial results of BLAST 2 sequences search of AF126284 against itself. **b** Possible mechanism involved in the duplication. During the synthesis of the negative strand, one of these segments in the negative strand (represented in green) might hybridize with the neighboring segment in the positive strand (represented in blue), resulting in duplication of the segment represented in green (green dashed line).

Table S1
BLAST analyses of DOP-PCR sequencing results

Clone	Region of homology with AF126284	Identity	E-value
MP1	6413-6772	99%	8.10 ⁻¹⁷⁶
MP3	6413-6796	99%	0.0
	7826-7884	100%	2.10 ⁻¹⁹
MP5	8715-9366	99%	0.0
MP7	7463-7884	98%	0.0
MP9	7398-7885	98%	0.0
MP11 ¹	-	-	-
MP13	7463-7884	98%	0.0
MP15	6413-6599	98%	9.10 ⁻⁸⁵
	10326-10394	94%	1.10 ⁻¹⁹
MP17	6413-7884	99%	0.0
MP19	7463-7884	98%	0.0
MP21	8715-9366	99%	0.0
MP23	7463-7884	98%	0.0
MP25	8715-9363	99%	0.0
MP27	7463-7885	97%	0.0
MP31	7463-7884	98%	0.0
MP33	4385-5118	98%	0.0
MP35	7409-7886	99%	0.0
MP37	7463-7884	98%	0.0

¹Blast hits presented low coverage (<17%) and high E-values (>0.11)

Table S2
Primers used in RT-PCR and sequencing

Primer	Sequence (5' → 3')	Position (AF126284)
AURAV1F	ATAGCGGACGGACTAGTACTTGTACTAC	1-28
AURAV2R	TAGGGCCTGATGGTAAATGGTC	581-560
AURAV3F	GGTCAAAGGGGAGCGTGTATCGTTC	1040-1064
AURAV4R	ATAGTTCTGCATCGTGTGGTGGTTCT	1226-1200
AURAV5F	TAGATGCCAGAAAATGCGTAAAAAGAG	2101-2127
AURAV6R	ATAGTCATTCCCCGCATTTTCTGG	2380-2357
AURAV7F	AATGACCAACCTTACTCCGTGATGT	3225-3249
AURAV8R	GTATTTGGTCCCCTTTTCTGCTCTTA	3474-3448
AURAV9F	GTTGGAGGAAGCGACGAAGCTA	4343-4364
AURAV10R	ATCTGCTATGCGTTGCTCCCCTT	4574-4551
AURAV11F	AACGAGCTACGGAGGCATGCGACGATA	5418-5444
AURAV12R	CCAAACGTAGGGGTTCGGTGATG	5674-5653
AURAV13F	GCAGCACTATTCGCAAAGACACATCGA	6537-6563
AURAV14R	GGTCTGCTGCTTGAATAACTTGT	6691-6668
AURAV15F	CACTACGCGGGAGCCCAAGACACCT	7534-7558
AURAV16R	TAGTGAGCTGTTGGATCTGGGTAG	7760-7737
AURAV17F	GTTCTGCTGGACGCTGCTCTGA	8564-8585
AURAV18R	CGTCATCGGCATCATCCCACAC	8735-8714
AURAV19F	CACCTCTATCCCTTCTACACCGTTACA	9695-9721
AURAV20R	CATTGTTTGACTGTGTTGCCATAGGTA	9934-9908
AURAV21F	CCGGATCAGATCTATTAGCAAACACAGC	10671-10698
AURAV22R	GGCTCGGATACGCGTGTAAATGCAG	10923-10899
AURAV23F	GCTGCTATAGCCGCAATTGTGCTGG	11303-11327
AURAV24R	GAAATATTA AAAACAAAATTTTGAAATAAT	11824-11795
Nsp3F	GATCGTGAGCGCCAATTCAC	4029-4048
Nsp3R	TTTTTGTTCAGATGCCCG	5789-5770
DOP	CCGACTCGAGINNNNNNTGTGG	not applicable

Table S3
Dataset used in the phylogenetic analysis

GenBank Accession number	Abbreviation	Name
AJ316244	SPDV	Salmon pancreas disease virus complete genome, isolate F93125, genomic RNA
AJ316246/NC003433	SDV	Sleeping disease virus complete genome
HM147990	SESV	Southern elephant seal virus, complete genome
U73745	BFV	Barmah Forest virus strain BH2193, complete genome
EF536323	MIDV	Middelburg virus strain MIDV857 non-structural polyprotein and structural polyprotein genes, complete cds
HM147989	NDUV	Ndumu virus, complete genome
AB032553	SAGV	Sagiyama virus genomic RNA, complete genome
AY702913	GETV	Getah virus from South Korea, complete genome
GQ433354	RRV	Ross River virus strain QML 1, complete genome
HM147985	BEBV	Bebaru virus, complete genome
NC_003417	MAYV	Mayaro virus, complete genome
HM147992	UNAV	Una virus non structural polyprotein gene, partial cds; and structural polyprotein gene, complete cds
HM045788	CHIKV	Chikungunya virus strain PO731460, complete genome
AF079456	ONNV	O'nyong-nyong virus strain SG650, complete genome
L01442	VEEV IAB	Venezuelan equine encephalitis virus, complete genome
AF375051	VEEV IC	Venezuelan equine encephalitis virus nonstructural polyprotein and structural polyprotein genes, complete cds
L00930	VEEV ID	Venezuelan equine encephalitis virus strain 3880, complete genome
U34999	VEEV IE	Venezuelan equine encephalitis virus nonstructural and structural polyprotein genes, complete cds
AF075257	VEEV IF	Venezuelan equine encephalitis virus strain 78V-3531, complete genome
AF075251	EVEV	Venezuelan equine encephalitis virus strain Everglades Fe3-7c, complete genome
AF075254	TONV	Venezuelan equine encephalitis virus strain Tonate CaAn 410d, complete genome
AF075253	MUCV	Venezuelan equine encephalitis virus strain Mucambo BeAn 8, complete genome
AF075255	VEE 71D1252	Venezuelan equine encephalitis virus strain 71D-1252, complete genome
AF075256	PIXV	Venezuelan equine encephalitis virus strain Pixuna BeAr 35645, complete genome
AF075259	CABV	Venezuelan equine encephalitis virus strain Cabassou CaAr 508, complete genome
AF075258	RNV	Venezuelan equine encephalitis virus strain AG80-663, complete genome
EF151502	EEEV LinI	Eastern equine encephalitis virus strain FL93-939, complete genome
DQ241303	EEEV LinII	Eastern equine encephalitis virus strain PE-3.0815, complete genome
DQ241304	EEEV_LinIII	Eastern equine encephalitis virus strain PE-0.0155, complete genome
EF151503	EEEV_LinIV	Eastern equine encephalitis virus strain BeAr436087, complete genome

HM147991	TROV	Trocar virus non structural polyprotein gene, partial cds; and structural polyprotein gene, complete CDs
NC_018615	EILV	Eilat virus isolate EO329, complete genome
AF126284	AURAV	Aura virus polyprotein 1 and polyprotein 2 genes, complete CDs
NC001547	SINV	Sindbis virus, complete genome
M69205	SINV Ockelbo	Ockelbo virus strain Edsbyn, complete genome
HM147984	SINV Babanki	Babanki virus, complete genome
AF103728	SINV XJ160	Sindbis virus strain XJ-160, complete genome
AF429428	SINV SW6562	Sindbis virus isolate SW6562, complete genome
HM147993	WHATV	Whataroa virus, complete genome

Supplementary sequence data

In red: pGEM-T-easy sequence

>MP1

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