

Supplementary Material

The Tomato Hybrid Proline-Rich Protein regulates the abscission zone competence to respond to ethylene signals

By

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Figure S1

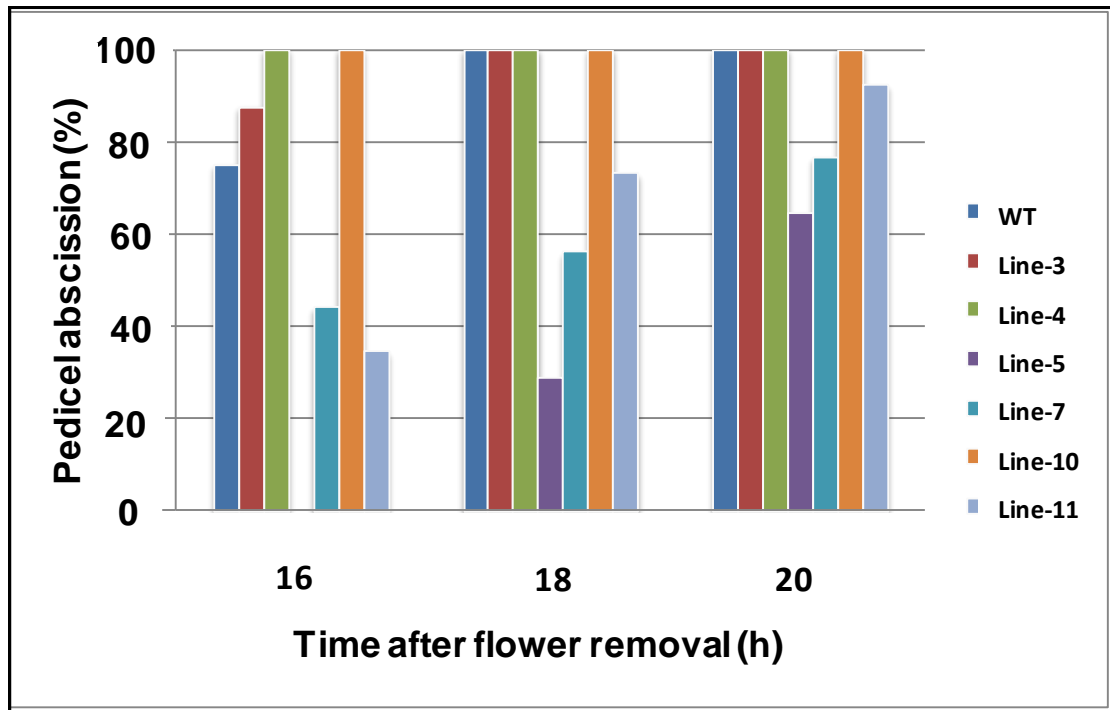


Figure S1. Screening of the kinetics of flower pedicel abscission following flower removal in several antisense silenced lines of the *THyPRP* gene. Flower explants were prepared and handled as previously described^{15, 34}. Wild type (WT) plants (cv. NY) and six silenced lines generation T1 were examined. Each transgenic line contained 4-6 plants, and 3-5 flowers in the right stage were harvested from each plant for abscission assay. The percentage of accumulated pedicel abscission was monitored at 16, 18, and 20 h after flower removal. The results are means of all explants per line (n = 16-24 explants).

Figure S2

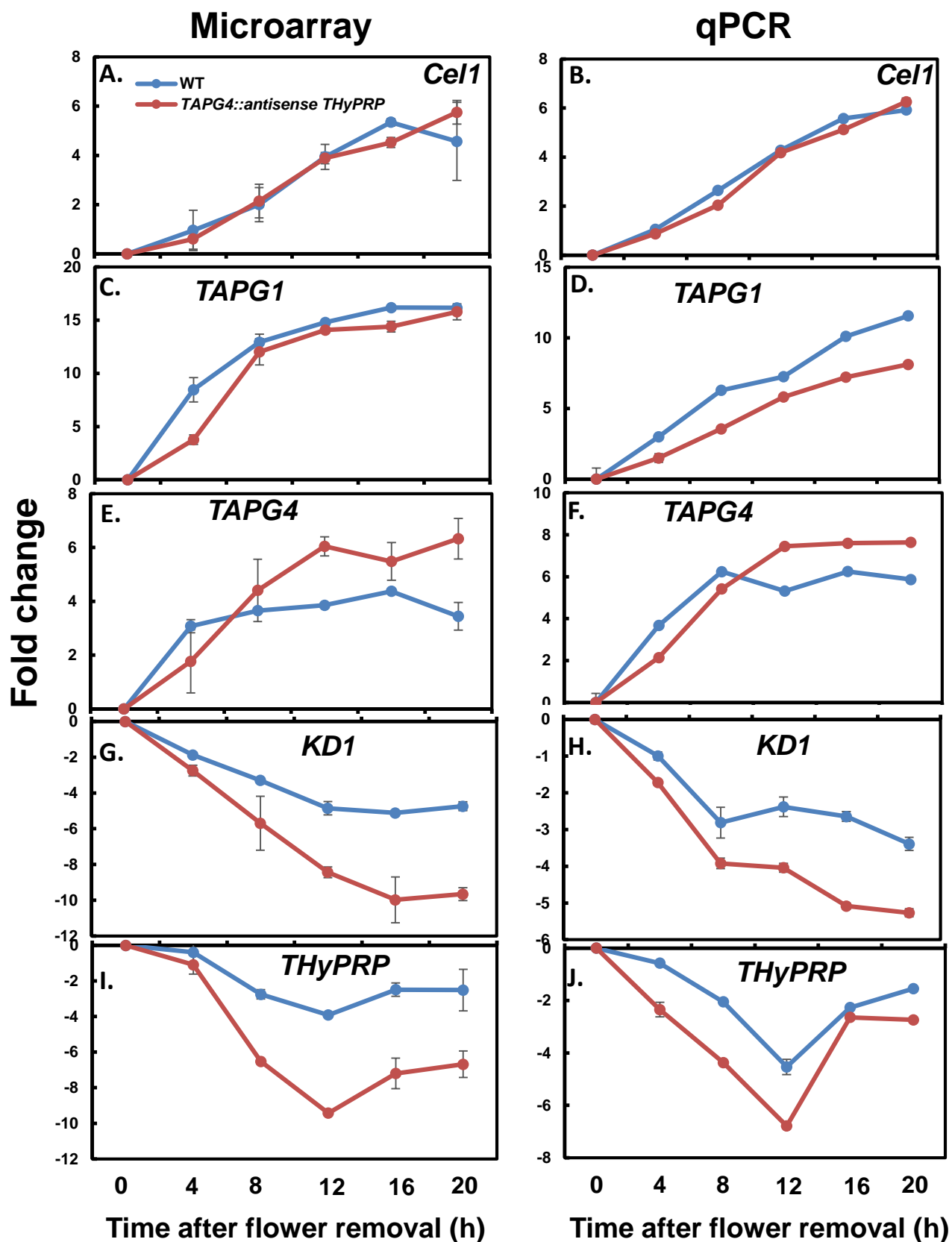


Figure S2. Validation by qPCR (B, D, F, H, J) of the microarray results (A, C, E, G, I) of the kinetics of changes in expression patterns of selected genes in the FAZ of WT and antisense silenced *THyPRP* plants following abscission induction by flower removal. *TAPG4::antisense THyPRP*-silenced line 11/generation T4 was used. Expression levels were measured for tomato *Cellulase1* (*Cell*) (ID - U13054/Solyc08g081620) (A, B); *Tomato Abscission Polygalacturonase1* (*TAPG1*) (ID - U23053/Solyc02g067630); (C, D); *TAPG4* (ID - U70481/Solyc12g096750) (E, F); *knotted1-like homeobox protein* (*KD1*) (ID - AF375969/Solyc06g072480) (G, H); and *THyPRP* (ID - X57076/Solyc07g043000) (I, J). The data indicate the fold change relative to time zero. The relative quantification of the gene expression level in the qPCR assay was determined by the comparative C_T method $2^{-\Delta\Delta CT}$ using *ACTIN* as a reference gene, and then calculating the fold change value. The ΔCT values for each gene were compared to the zero time ΔCT value for that gene to generate the $2^{-\Delta\Delta CT}$ values. The microarray and qPCR analyses were performed with different samples taken from independent biological replicates of two separate experiments. The results are means of two biological replicates \pm SD.

Figure S3

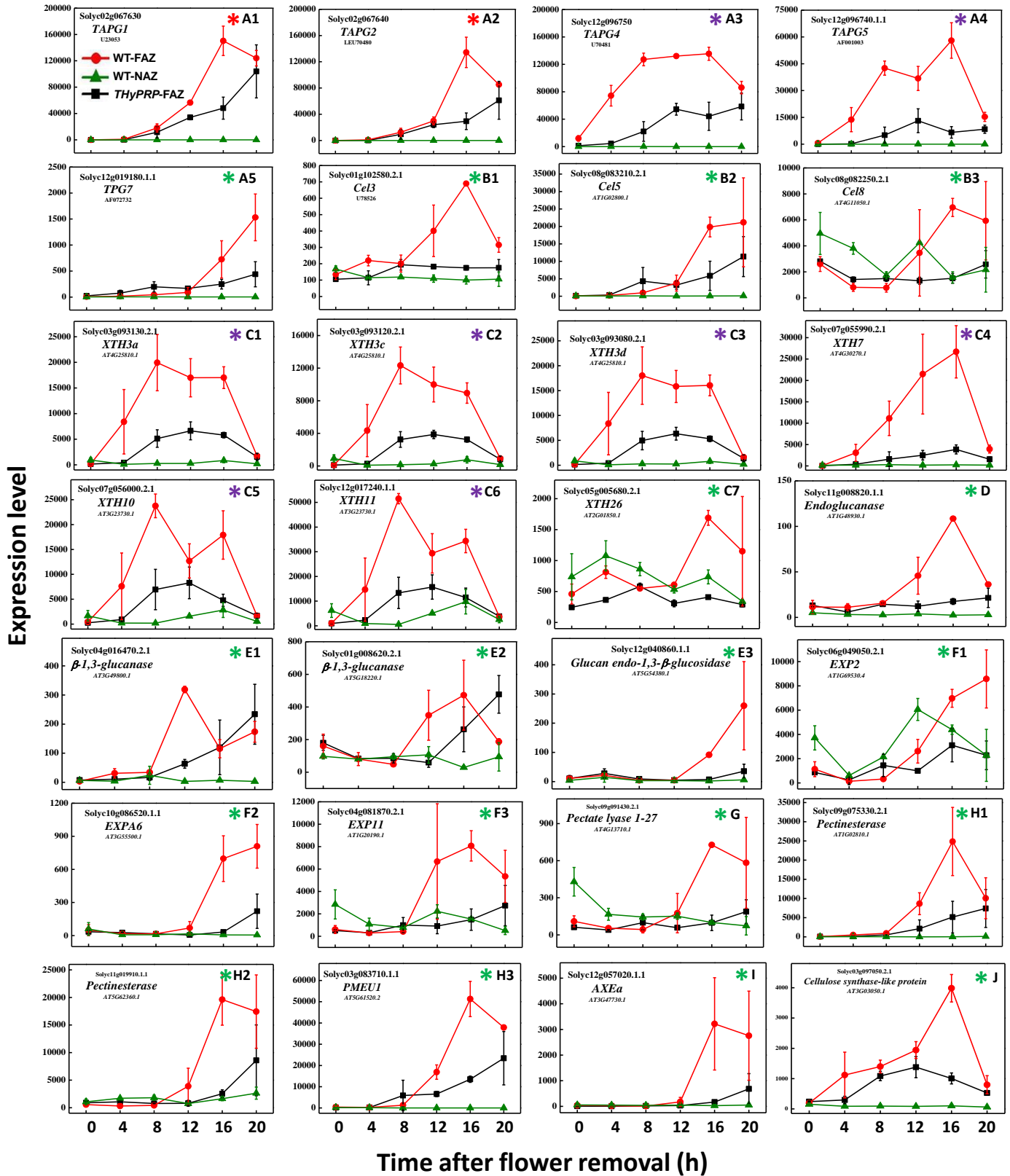


Figure S3. Effect of antisense silencing of *THyPRP* on the kinetics of changes in array-measured expression levels of genes coding for **cell wall modifying enzymes** that **were** specifically upregulated in the WT FAZ at various time points after flower removal: early - 4 (*) or 8 (*) or late 12-20 (*) h. The *TAPG4::antisense THyPRP*-silenced line 11/generation T4 was used. Expression levels were measured for *Tomato abscission polygalacturonase (TAPG)* genes (**A1-A5**); *Cellulase (Cel)* genes (**B1-B3**); *Xyloglucan endotransglucosylase/hydrolase (XTH)* genes (**C1-C7**); *Endoglucanase (D)*; β -1,3-glucanase genes (**E1-E3**); *Expansin (EXP)* genes (**F1-F3**); *Pectate lyase1-27 (G)*; *Pectinesterase* genes (**H1-H2**); *Pectin methyl esterase inhibitor U1 (PMEU1)* (**H3**); *Acetyl xylan esterase a (AXEa)* (**I**); and *Cellulose synthase-like protein (J)*. Transcript identities are indicated in the graphs by their gene ID and their Arabidopsis (At) gene number, and/or their nucleotide accession number. The results are means of two biological replicates \pm SD.

Figure S4

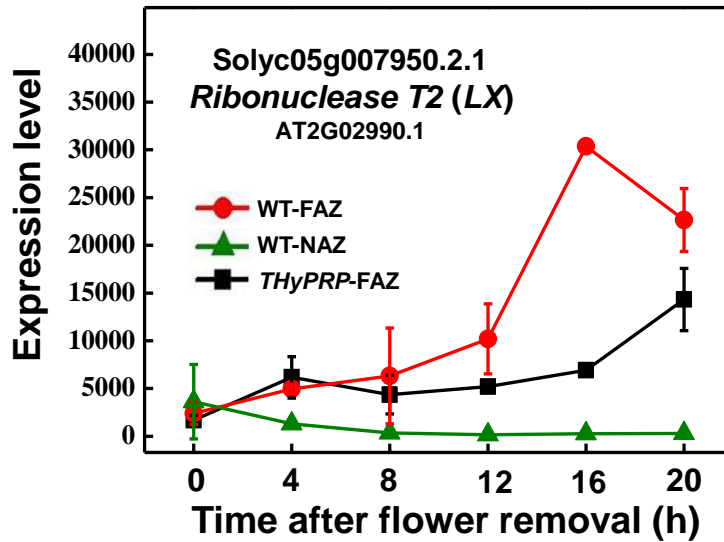


Figure S4. Effect of antisense silencing of *THyPRP* on the kinetics of changes in array-measured expression levels of *Ribonuclease T2 (LX)* gene that was specifically upregulated in the WT FAZ at the late stages (12-20 h after flower removal) of abscission. The *TAPG4::antisense THyPRP*-silenced line 11/generation T4 was used. The results are means of two biological replicates \pm SD.

Table S1: List of primers and their sequences used in the qPCR assay.

No	Primer name	Gene bank ID	Solyc ID	Primer sequence (5' to 3')	Tm (°C)
1	Actin_F	AB199316	Solyc03g078400	GTGTTGGACTCTGGTGATGG	60
2	Actin_R			GTAGTCAAGAGCCACATAAGC	60
3	TAPG1_F	U23053	Solyc02g067630	GCAGTGAAACTTGATTGTAGC	56
4	TAPG1_R			CCATTCTTGTGATAGTATACAC	56
5	TAPG4_F	U70481	Solyc12g096750	GAACATCAGCTACAGAAATCG	60
6	TAPG4_R			ACCAGAAGCTCTTCCTCCAG	60
7	Cel1_F	U13054	Solyc08g081620	GGATTAATGCCAGAAAGTAGC	60
8	Cel1_R			GTAACCTCCTCCTATTCCAGC	60
9	TPRP-F1_F	X57076	Solyc07g043000	GCACAAC TATTAGACTCAAGC	60
10	TPRP-F1_R			TGCCTTCAACTATGACAATGC	60
11	KD1_F	AF375969	Solyc06g072480	CTCACTCACAATGGATCAACC	60
12	KD1_R			GGAGTGGAAGTAGAAGTAGG	60