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## **Appendix Figure legends**

### **Appendix Figure S1.**

(A and C) A schematic diagram showing the route for the generation of single- and multi-BCR component KOs from WT Ramos B cells by the CRISPR/Cas9 method. (B and D) Expression of the indicated BCR component in WT and the different Ramos KO cells as measured by flow cytometry after staining with the corresponding antibodies. (E) Western blot analysis of the expression of the indicated BCR component in WT and the different KO Ramos cells. One clone is used for each genotype.

### **Appendix Figure S2.**

Cell proliferation assay of WT and BCR component KO Ramos cells using CytoTell™ Ultragreen. The data are representative of three independent experiments. One clone is used for each genotype.

### **Appendix Figure S3.**

Growth competition between BCR component KO cells and gWT Ramos cells. The GFP<sup>+</sup> viable cells were single-cells sorted into 96-well plates 48h post transfection of the CRISPR/Cas9 KO plasmid. The sorted cells were cultured for 10 days and then transferred into larger wells for expansion for another 4 days. After the deletion of the target gene has been confirmed by staining with the corresponding antibodies, the freshly generated single colonies were immediately used for growth competition assay. The competition between GFP-WT and gWT cells serves as a control. The data represent the mean and standard error of a minimum of three independent experiments. Differences between the WT and KO cells were evaluated by first performing the multiple t-test for each day without assuming the same SD and the multiple t-test results were then further analyzed using the False Discovery Rate (FDR, Q) approach with Two-stage linear step-up procedure. Data showed significant difference ( $Q < 0.01$ ) were marked by stars. One clone (Different than those used in Fig 1) is used for each genotype.

### **Appendix Figure S4.**

**(A-D)** Calcium responses of WT and the indicated BCR component KO Ramos cells upon the stimulation with anti-Ig $\alpha$  antibodies. HL KO = HC/LC double KO; HL $\alpha$  KO = HC/LC, Ig $\alpha$  triple KO; HL $\beta$  KO = HC/LC, Ig $\beta$  triple KO. One clone is used for each genotype.

**Appendix Figure S5.**

**(A)** A schematic diagram showing the generation of SLP65 KO from HL KO Ramos B cells by the CRISPR/Cas9 method. **(B)** Western blot analysis of SLP65 expression in HL double KO and HLSLP65 triple KO Ramos cells. The staining for GAPDH served as loading control. One clone is used for each genotype.

**Appendix Figure S6.**

**(A)** A schematic diagram showing the generation of CD19 or CD81 deficient cells from HL KO Ramos B cells by the CRISPR/Cas9 method. **(B)** FACScan analysis of CD19 or CD81 expression on HL KO, HLCD19 KO or HLCD81 KO Ramos B cells as measured by flow cytometry. One clone is used for HLKO. HLCD19KO and HLCD81 KO are batch sorted.



Fig. S2

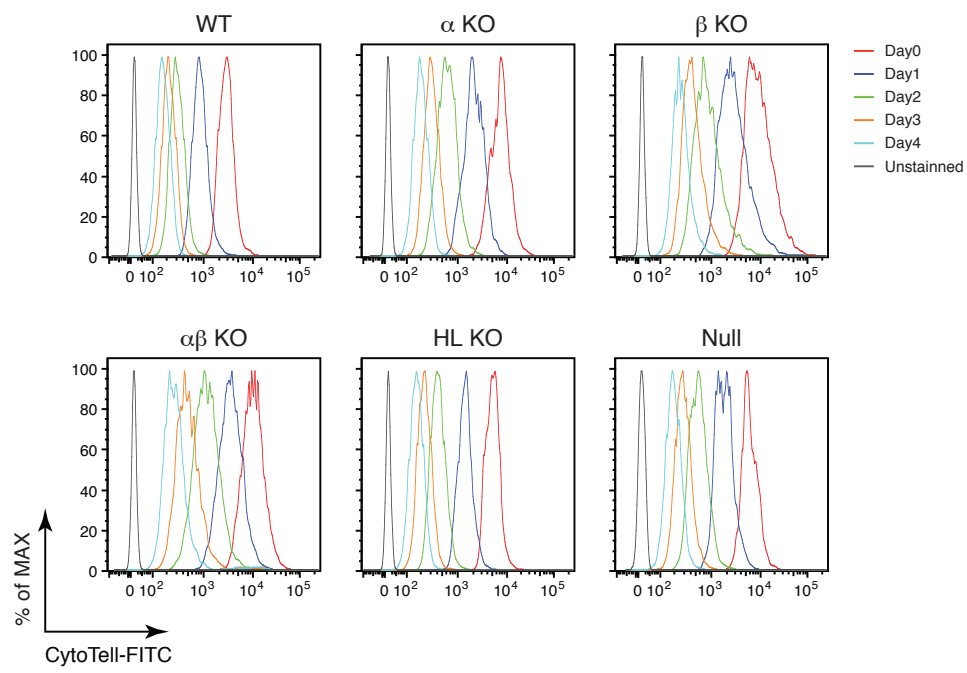


Fig. S3

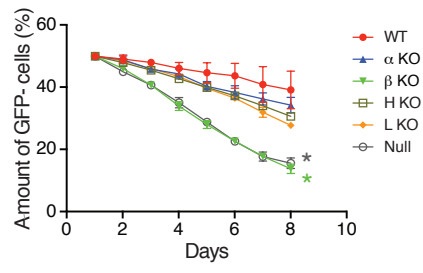


Fig. S4

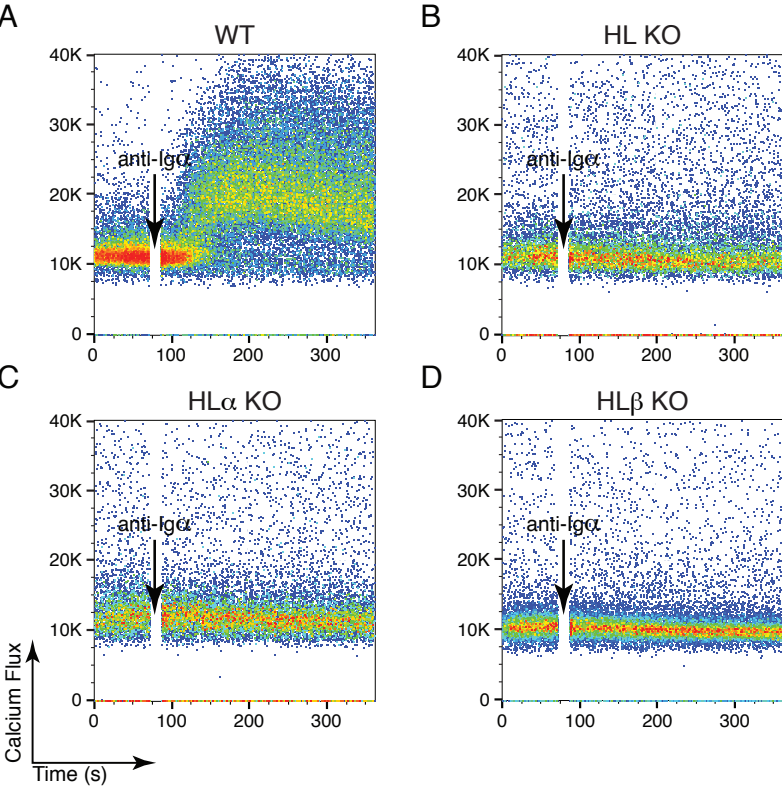


Fig. S5

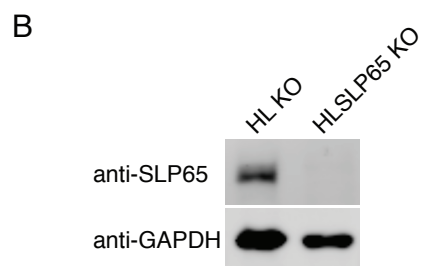
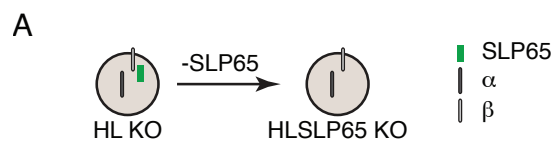
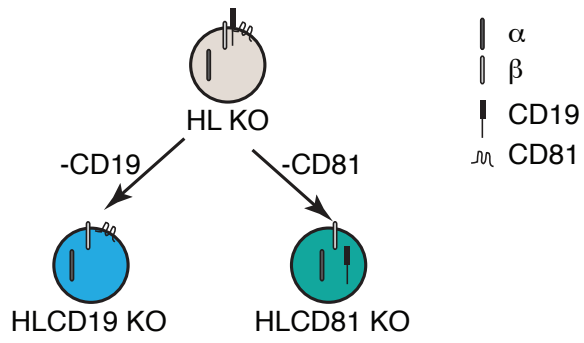


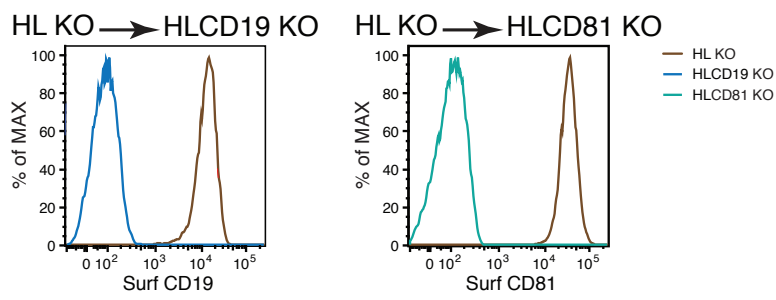


Fig. S6

A



B



**Appendix Table S1. Information of CRISPR/Cas9 KO plasmids**

<b>Target gene</b>	<b>Description</b>
Ig $\alpha$	Purchased from Santa Cruz
Ig $\beta$	Purchased from Santa Cruz
mIg Heavy chain	sgRNA 5'-CAGCTACAGCAGTGGGGCGC-3'
mIg Light chain	sgRNA 5'-GGGGTTTCTAATCGCTTCTC-3'
SLP65	Purchased from Santa Cruz
CD19	sgRNA 5'-AAGCGGGGACTCCCGAGACC-3'
CD81	Purchased from Santa Cruz

**Appendix Table S2. Information of primers for cloning**

Name	Sequences 5'-3'	Template	Final plasmid description
Igβ65F	GTTTGGGGTCGGGGAC AG	Ramos cDNA	pJET-Igβ
Igβ704 R	CCCGTCCATCCCCAGA GA		
pMIGIgβ_F	ATTAGATCTCTCGAGCC ACATGGCCAGGCTGGC GTTG	pJET-Igβ	PCR fragments were fused into the Xho I linearized pMIG to obtain pMIG-Igβ
pMIGIgβ_R	GCGGAATTCGTAACTC ACTCCTGGCCTGGGTG		
pMIGHAIgβ_F1	TATCCATATGATGTTCC AGATTATGCTGCCAGA TCGGAGGACCGG	pMIG-Igβ	HA tagged Igβ plasmid: pMIG-HA-Igβ
pMIG_R1	CCGTAGAAAAGATCAA AG		
pMIG_F2	TGATCTTTTCTACGGGG T	pMIG-Igβ	
pMIGHAIgβ_R2	AACATCATATGGATAT GCTGGTACTGGCTCAG C		
Igα74F	GCCACTGGAGCCCATC TC	Ramos cDNA	pJET-Igα
Igα771R	CCTGGCAGGAGTAGGG GT		
pMIGIgα_F	ATTAGATCTCTCGAGCC ACATGCCTGGGGGTCC AGGA	pJET-Igα	PCR fragments were fused into the Xho I linearized pMIG to obtain pMIG-Igα
pMIGIgα_R	GCGGAATTCGTAACTC ACGGCTTCTCCAGCTG		
pMIGIgβAt_F1	CCTATCTTCTGCTGAG GAAACGATGGCAGAAC	pMIG-Igα	Two PCR fragments were fused together to

pMIG_R1	See above		obtain pMIG-HA-Ig $\beta$ - Ig $\alpha$ tail
pMIG_F2	See above	pMIG-HA-Ig $\beta$	
pMIGIg $\beta$ At_R2	CTGCCATCGTTTCCTCA GCAGGAAGATAGGCAC		
Ig $\beta$ Y196F_F	GGAAGATCACACCTTC GAGGGCCTGGACA	pMIG-HA-Ig $\beta$	Ig $\beta$ ITAM Y196 was mutated into F
Ig $\beta$ Y196F_R	TGTCCAGGCCCTCGAA GGTGTGATCTTCC		
Ig $\beta$ Y207F_F	CCAGACAGCCACCTTT GAGGACATAGTGA	pMIG-HA-Ig $\beta$	Ig $\beta$ ITAM Y207 was mutated into F
Ig $\beta$ Y207F_R	TCACTATGTCCTCAAAG GTGGCTGTCTGG		
Ig $\beta$ TailL_F1	AAGGATGACAGCAAGT GAGTTAACGAATTCCG C	pMIG-HA-Ig $\beta$	Two PCR fragments were fused together to obtain pMIG-HA-Ig $\beta$ tail less
pMIG_R1	See above		
pMIG_F2	See above	pMIG-HA-Ig $\beta$	
Ig $\beta$ TailL_R2	GAATTCGTTAACTCACT TGCTGTCATCCTTGTC		
CD19_F1	CCCCGGAGAGTCTGAC CA	Ramos cDNA	pJET- CD19
CD19_R1	AGGAGATCCAGGCTGG CC		
pMIGCD19_F	ATTAGATCTCTCGAGCC ACATGCCACCTCCTCGC CTC	pJET- CD19	PCR fragments were fused into the Xho I linearized pMIG to obtain pMIG-CD19
pMIGCD19_R	GCGGAATTCGTTAACTC ACCTGGTGCTCCAGGT		