## Supplementary Tables and Figures for "Genome-Wide Prediction of cis-Regulatory Regions Using Supervised Deep Learning Methods"

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Cell	A-E	I-E	A-P	I-P	A-X	I-X	UK	Total
A549	387	40,387	10,907	128,998	8,998	14,712	81,217	285,606
GM12878	$2,\!878$	28,156	10,816	73,891	8,226	19,078	80,004	223,049
HelaS3	1847	32,179	10,759	79,009	9,123	22,071	81,502	236,485
HepG2	1465	34,556	11,467	96,184	9,931	19,071	79,417	252,091
HUVEC	1226	$35,\!143$	$11,\!254$	101,861	9,739	18,249	80,333	257,805
K562	894	34,392	10,076	82,829	9,033	20,261	78,081	235,566
MCF7	249	36,873	10,733	81,510	10,829	$13,\!653$	82,663	236,510

Table S1: Numbers of labelled regions in our data.

Table S2: Number of features used for each cell type and number of common features between any two cell types in our labelled data.

Cell	A549	GM12878	HelaS3	HepG2	HMEC	HUVEC	K562	MCF7
A549	45	38	26	38	15	19	38	25
GM12878	-	101	50	54	16	21	69	26
HelaS3	-	-	74	43	16	22	56	23
HepG2	-	-	-	72	16	21	57	28
HMEC	-	-	-	-	16	16	16	9
HUVEC	-	-	-	-	-	24	24	13
K562	-	-	-	-	-	-	135	29
MCF7	-	-	-	-	-	-	-	38

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Original Class\DECRES Class	A-E	A-P	BG	Total
CRE-seq Positive Combined Predicted Enhancer in K562	254	0	132	386
CRE-seq Negative Combined Predicted Enhancer in K562	433	1	378	812
Combined Predicted Repressed Region in K562	0	0	298	298
Total	687	1	808	1496
MPRA Positive Enhancer in K562	120	0	2	122
MPRA Negative Enhancer in K562	179	0	15	194
Total	299	0	17	316
MPRA Positive Enhancer in HepG2	182	1	3	186
MPRA Negative Enhancer in HepG2	217	5	45	267
Total	399	6	48	453

Table S3: Confusion matrices of classifying CRE-seq and MPRA validated regions using DECRES.

Table S4: Numbers of predicted and cell-specific *cis*-regulatory regions in the whole human genome. Columns 2-3: results of two-class prediction. Columns 4-7: results of three class prediction.

Cell	A-E+A-P	Specific	A-E	Specific	A-P	Specific
GM12878	90,192	27,904	$70,\!905$	27,185	19,287	6,090
HelaS3	100,102	22,268	92,509	22,866	$7,\!593$	979
HepG2	$114,\!873$	$35,\!986$	$105,\!007$	41,109	9,866	$2,\!135$
HMEC	$104,\!621$	28,774	$88,\!803$	26,262	$15,\!818$	$3,\!043$
HUVEC	$110,\!347$	$16,\!415$	97,069	$35,\!073$	$13,\!278$	582
K562	$133,\!940$	$30,\!835$	$122,\!321$	$56,\!374$	$11,\!619$	371

Table S5: Numbers of predicted A-Es and A-Ps on the 102,021 BDT loci. AiA: Active in the FANTOM Enhancer Atlas. IiA: Inactive in the FANTOM Enhancer Atlas. NiA: Not included in the FANTOM Enhancer Atlas. Specific: Predicted cell-specific A-Es.

Cell	A-E	A-P	BG
GM12878	11,910	1,975	88,136
HelaS3	12,743	226	89,052
HepG2	10,761	488	90,772
HMEC	$13,\!356$	1,267	$87,\!398$
HUVEC	$17,\!192$	758	$84,\!071$
K562	$13,\!936$	288	87,797

Table S6: Overlap between our predicted A-Es and the FANTOM enhancer atlas.

Cell\Element	AiA	IiA	NiA	Specific
GM12878	2,088	5,144	$4,\!678$	$4,\!199$
HelaS3	$1,\!459$	$6,\!435$	$4,\!849$	1,073
HepG2	$1,\!178$	4,888	$4,\!695$	$1,\!495$
HMEC	$1,\!250$	$6,\!644$	$5,\!462$	1,916
HUVEC	879	8,716	$7,\!597$	$3,\!560$
K562	750	$7,\!383$	$5,\!803$	2,965

Cell	Transcription Factor	Functionality
GM12878	RUNX1	RUNX1 and other Runt-related factors play crucial roles in
		haematopoiesis. Translocation of RUNX1 leads to severe
		acute myeloid leukemia [11].
GM12878	$REL/NF-\kappa B$ Factors	Play a critical role in immune response to infection [4].
HelaS3	C/EBP-Related Factors	C/EBP-related factors, a subfamily of the basic leucine zip-
		per factors (bZIP), regulating genes involved in immune and
		inflammatory responses, are expressed in cervix [1].
HelaS3	CREB-Related Factors (Another bZIP	Possibly up-regulate Bcl-2 expression in apoptotic HeLa cells
	Subfamily)	induced by trichosanthin [15].
HelaS3	TEAD1	Plays a role of apoptotic resistance in Hela cells [9].
HelaS3	AP-2 Factors	Can act as tumor suppressor, and malfunction of AP-2 was
		found in cervical cancer cells $[2]$ .
HelaS3	HOX Factors	a subgroup of HOX genes involve in cervical carcinoma [8].
HepG2	HNF1A, HNF1B, FOXA1, FOXA2,	These factors are hepatocyte nuclear factors that regulate
	HNF4A, and HNF4G	liver-specific genes [3, 16].
HepG2	C/EBPs	Have a pivotal role in liver development and function [13].
HMEC	TEAD Family	Members of this family regulate epithelial-mesenchymal tran-
		sition [17].
HUVEC	GATA Factors	Involve in networks of key determinants of vascular endothelial
		cell identity [7].
HUVEC	SOX Factors	Regulate vascular cell development and growth (vasculogene-
		sis) [12].
HUVEC	CREB-Related Factors	Involve in several specific pathways in HUVEC cell [14].
K562	GATA-Type, NFYA, and NFYB USF1	The NFY factors cooperate with GATA1 to mediate erythroid-
		specific transcription, and coassociate with FOS binding to
		both promoters and enhancers in K562 [5]. The NFY fac-
		tors are also found cooperate with USF1 and USF2 to active
		HOXB4 for hematopoiesis [18].
K562	STAT5	Maintains the high-level cell proliferation of K562 [10].

Table S7: Transcription factors whose binding motifs are enriched in specific cells.



Figure S1: Mean performance and standard deviation of 10-fold cross-validations using the MLP model on our labelled data of eight cell types. A-E: Active Enhancer, A-P: Active Promoter, A-X: Active Exon, I-E: Inactive Enhancer, I-P: Inactive Promoter, I-X: Inactive Exon, UK: Unknown or Uncharacterized, BG: I-E+I-P+A-X+I-X+UK.



Figure S2: Cumulative DECRES membership probabilities (scores) of enhancers that were tested as positives and negatives by CRE-seq or MPRA. These enhancers were predicted as A-Es by DECRES (See Table S3).



Figure S3: Comparing the mean auPRCs over 100 resampling and retraining on our labelled regions using different feature sets. "Experimental" means our experimentally derived next generation sequencing feature set. "Sequence" means the set of 351 sequence properties used in [6]. "Experimental+Sequence" means the combination of these two sets. The *p*-values in each legend were obtained using two-tailed Student's t-test to compare "Experimental"-based results with "Experimental+Sequence"-based and "Sequence"-based results, respectively.



Figure S4: Feature importance and box plots of top features in the 3-class (A-E versus A-P versus BG) scenario. A: Feature importance discovered by randomized DFS (RDFS) and random forest (RF) on HelaS3, HepG2 and K562 cells. RF's feature importance scores were normalized to [0,1] for better comparison with RDFS. B: For the top 10 features of the 3-class models generated for four well-characterized cell lines, box plots depict the range of observed feature values (log2 scale) for 7 sequence classes.



Figure S5: Feature importance, classification performance, and top features in the 2-class (A-E+A-P versus BG) scenario. A: Feature importance discovered by randomized DFS (RDFS) and random forest (RF). The random forest's feature importance scores were normalized to [0,1] for better comparison with randomized DFS. B: auPRC versus the number of features incorporated into the RDFS and RF. The annotated points indicate where a line with slope 0.5 intersects a fitted curve). C: For the top 6 features of the 2-class models generated for four well-characterized cell lines, box plots depict the range of observed feature values (log2 scale) for 7 sequence classes.



Figure S6: Box plots of all features for A549.



Figure S7: Box plots of all features for GM12878.



Figure S8: Box plots of all features for HelaS3.



Figure S9: Box plots of all features for HepG2.



Figure S10: Box plots of all features for HMEC.



Figure S11: Box plots of all features for HUVEC.



Figure S12: Box plots of all features for MCF7.



Figure S13: Box plots of all features for K562.



Figure S14: An example of predicted regulatory regions in the UCSC Genome Browser.



Figure S15: Functional and motif analysis of the DECRES genome-wide predictions on cell line GM12878. A: Distance from the predicted A-Es to gene TSSs. B: Distance from the predicted A-Ps to gene TSSs. C,D,E: Top 20 enriched biological processes, pathways, and diseases, respectively, in the predicted cell-specific CRRs. F: Enriched *de novo* motifs in the predicted cell specific CRRs. Column 4: families of best-matched TFs. Column 5: best match scores.



Figure S16: Functional and motif analysis of the DECRES NiA enhancers on cell line GM12878. A,B,C: Top 20 enriched biological processes, pathways, and diseases, respectively, in the NiA enhancers. D: Enriched *de novo* motifs in the NiA enhancer regions. Column 4: the families of best-matched TFs. Column 5: best match scores.



Figure S17: Functional and motif analysis of the DECRES genome-wide predictions on cell line HelaS3. A,B: Top enriched biological processes and diseases (no pathways enriched for HelaS3), respectively, in the predicted cell-specific CRRs. C: Enriched *de novo* motifs in the predicted cell specific CRRs. Column 4: families of best-matched TFs. Column 5: best match scores.

					GO Biological Process
					-log10(Binomial p value)
					0 5 10 15 20 25 29 29 29 29 29 29 29 29 29 29 29 29 29
Mark		Derrit	P	0	- response to oxygen levels 22.00
Moui	p-value	Best Match	Family	Score	response to hypoxia 27.58
<b>SET GASTGASSE</b>	1e-413	FOS, JUN	Fos-related factors, Jun-related factors	0.97	transforming growth factor beta receptor signaling pathway 22.46
TOCTOR	1e-79	KLF5	Three-zinc finger Kruppel-related factors	0.95	cellular regions to occessed oxygen levels 17.78 cellular regions to oxygen levels 17.88 regulation of carbohydrate metabolic process 18.84
<b>AST INCOM</b>	le-61	CEBPA	C/EBP-related factors	0.91	regulation of transforming growth factor beta production 16.81 cellular response to hypoxia 16.73 regulation of ducose metabolic process 16.21
SASCITEAC	1e-46	bZIP_CREB/G- box-	CREB-related factors	0.84	regulation of transforming growth factor beta2 production 15.46 extrinsic apoptotic signaling pathway 15.44 response to reactive exygen species 15.03
		like_subclass			muscle cell migration 14.44
<b>BUAGAARISE</b>	1e-45	FLI1	Ets-related factors	0.91	regulation of cellular carbohydrate metabolic process 14.07 A regulation of DNA-dependent transcription in response to stress 13.93
<b>CCATCTOG</b>	1e-32	CTCF	More than 3 adjacent zinc finger factors	0.87	positive regulation of cellular catabolic process 11.11
GAGIGITTAC	1e-30	FOXD1	Forkhead box factors	0.82	MSigDB Pathway
COSCIECC	1e-30	Sp4_2	BetaBetaAlpha-zinc finger	0.82	0 2 4 6 8 10 12 14 16 18 20 22 24 26
SCGAMATCCC	1e-26	REL	NFkB-related factors	0.92	PDGFR-beta signaling pathway 20.81 Keratinocyte Differentiation 16.50
AAGGGGGAGGGC	1e-24	SP1	Three-zinc finger Kruppel-related factors	0.73	Endocytosis 15.78 IL6-mediated signaling events 15.55 Diceo P2-a director
TAAATCSG	1e-24	Hoxc9	HOX-related factors	0.73	HIF-1-alpha transcription factor network 15.23 Pancreatic cancer 15.00
Z.SCA.SS	1e-22	NFIC	Nuclear factor 1	0.78	m TOR signaling pathway 14.29 PXR and PAR heterodimerization with other nuclear receptor 14.15 Pate of Coloradia MCR cleanaling to the machanite of the machani
GGTTCGAATC	1e-22	Zbtb12_2	BetaBetaAlpha-zinc finger	0.64	Genes involved in Signaling of VTG-beta Receptor Complex 13.28 Validated targets of C-MYC transcriptional repression 13.21
SCATICUE	1e-20	TEAD1	TEF-1-related factors	0.79	Bladder cancer 13.12 Cell cycle 21.96 Pegulation of nuclear SMAD2/3 cincation 12.96
TAMAGEOC	1e-19	Foxo1	Forkhead box factors	0.72	Cell Cycle: G1/S Check Point 12.68 C-MYB transcription factor network 12.56 B
CCAAGTCAGA	1e-19	Hand1::Tcfe2a	Helix-Loop-Helix	0.62	NFkB activation by Nontypeable Hemophilus influenzae
<b>CAGGAVACTUCT</b>	1e-18	ETS::E-box	Ets::bHLH	0.79	Disease Ontology
TGTTICACAAJC	1e-16	CEBPA	C/EBP-related factors	0.69	0 5 10 15 20 25 30 35 40 45 DNA virus infectious disease
<b>GECTITATATA</b>	1e-16	TBP	TBP-related factors	0.73	dsDNA virus infectious disease 38.74 neck neoplasm 38.30
GCTGGCAGCCA	1e-16	NFIC::TLX1	Nuclear factor 1::NK-related factors	0.65	neux canciora 37.96 neck carcinoma 32.11 thyroid neoplasm 28.79
GTTTTTCG	1e-16	Tcfap2e_2	Helix-Loop-Helix	0.76	head and neck squamous cell carcinoma 28.27 malignant neoplasm of thyroid 27.97
SEGATAGE	1e-15	Gata4	GATA-type zinc fingers	0.88	ter jaaniver virus intercitous disease 25.22 hepatitis B 25.22 Herpesviridae infectious disease 24.56
CACCEGETECCA	1e-14	NFIC	Nuclear factor 1	0.60	papillary epithelial neoplasm 23.46 upper respiratory tract disease 22.41
		D			transitional continuenti di scrimona di 21.04 large literitine admossrimona di 20.81 gastric admossrimona di 19.95 stornach carritorna 19.61 neopisan of bödy ol uterus 19.14 thyroid carritorna 15.99

Figure S18: Functional and motif analysis of the DECRES NiA enhancers on cell line HelaS3. A,B,C: Top 20 enriched biological processes, pathways, and diseases, respectively, in the NiA enhancers. D: Enriched *de novo* motifs in the NiA enhancer regions. Column 4: families of best-matched TFs. Column 5: best match scores.



Figure S19: Functional and motif analysis of the DECRES genome-wide predictions on cell line HepG2. A,B,C: Top enriched biological processes, pathways and diseases, respectively, in the predicted cell-specific CRRs. D: Enriched *de novo* motifs in the predicted cell specific CRRs. Column 4: families of best-matched TFs. Column 5: best match scores.



Figure S20: Functional and motif analysis of the DECRES NiA enhancers on cell line HepG2. A,B,C: Top 20 enriched biological processes, pathways, and diseases, respectively, in the NiA enhancers. D: Enriched *de novo* motifs in the NiA enhancer regions. Column 4: families of best-matched TFs. Column 5: best match scores.



Figure S21: Functional and motif analysis of the DECRES genome-wide predictions on cell line HMEC. A,B,C: Top enriched biological processes, pathways and diseases, respectively, in the predicted cell-specific CRRs. D: Enriched *de novo* motifs in the predicted cell specific CRRs. Column 4: families of best-matched TFs. Column 5: best match scores.



Figure S22: Functional and motif analysis of the DECRES NiA enhancers on cell line HMEC. A,B,C: Top 20 enriched biological processes, pathways, and diseases, respectively, in the NiA enhancers. D: Enriched *de novo* motifs in the NiA enhancer regions. Column 4: families of best-matched TFs. Column 5: best match scores.

					GO Biological Process
					-log10(Binomial p value)
					0 10 20 30 40 50 60 70 80 90 100 110
otif	p-value	Best Match	Family	Score	blood vessel morphogenesis 108
ATGASTCATS	1e-999	JUNB,	Jun-related factors, Fos-related factors	0.97	angiogenesis endocytosis 55.65
		JUND, FOS,			regulation of Rho protein signal transduction 51.42
ACACCAAGES		FO3L2			response to transforming growth factor beta stimulus 49.58
Sectores:	1e-689	FEV, FLII	Ets-related factors	0.93	cellular response to transforming growth factor beta stimulus transforming growth factor beta receptor signaling pathway
TCGGAAGGAA	1e-142	ELF5	Ets-related factors	0.69	phagocytosis 41.56
SCATIGITA	1c-64	SOX9	SOX-related factors	0.93	regulation of epithelial cell migration 35.67
CRACATAAG	1. 40	CUTU2	CATH Hard Street	0.07	regulation of cell shape 33.78
****GATAAQ	1e-48	GAIA3, Gata4 Gata1	GAIA-type zinc fingers	0.96	vascular endothelial growth factor receptor signaling pathway 32.73 regulation of endothelial cell migration 31.74
GIGITICTITG	1- 14	Enur?	Fashbard has feature	0.74	cell-substrate adhesion 31.72
elennechne	10-44	FOXA2, FOXA1	Porknead box factors	0.74	cytoskeleton-dependent intracellular transport [31,37]
GCTGGTTTCCTC	10.43	SPIR	Ete-related factors	0.65	Fc-gamma receptor signaling pathway involved in phagocytosis 28.83
	10-45	5116	EAS-TENANCE NACIONS	0.05	MSigDB Pathway
CAUGICE	1e-35	Mycn	bHLH-ZIP factors	0.96	-lop10(Binomial p value)
TGTTIANACAGE	1e-31	Foxa2	Forkhead box factors	0.75	0 10 20 30 40 50 60 70 80
AAAGTAAGTCTG	1e-30	EHE	Ets-related factors	0.55	Genes involved in Hemostasis 59.39
COCH CTENCE					Focal adhesion 52.40 Signaling events mediated by VEGEP1 and VEGEP2 48.81
GEGAGICAGE	1e-29	Matb	Mat-related factors	0.77	Genes involved in Signaling by TGF-beta Receptor Complex 46.72
TG.CLCCCCTCC	1e-24	Hic1_1	BetaBetaAlpha-zinc finger	0.67	Signaling events mediated by tocal adhesion kinase 46.48 Genes involved in Signaling by Rho GTPases 45.97
TGTTATCAGAAC	1e-23	Gata5_1	GATA-type zinc fingers	0.70 Genes in	TGF-beta receptor signaling 40.77
CACCANCANC	1. 00	cure.	E	0.70	Integrin-linked kinase signaling 34.76
STOLEN C	16-25	EHP	Ets-related factors	0.70	Genes involved in Response to elevated platelet cytosolic Ca2+ 34.51
SEFICICA	1e-21	NFIC	Nuclear factor 1	0.86	Integrin Signaling Pathway 34.04
CCACTTCACG	le-19	Nkx3-1_1	NK-related factors	0.71	PDGFR-beta signaling pathway 32.03
GAOSSOCIALOT	1 19	E3E6	E2E value of factors	0.60	VEGF and VEGFR signaling network 30.97
SHERICSHICK	10-10	E2F0	E2F-felated factors	0.00	Genes involved in Integrin cell surface interactions 30.55 Regulation of RhoA activity 30.51 B
aaaaaa	1e-15	Tcfap2b_2	Helix-Loop-Helix	0.72	Genes involved in Sema4D in semaphorin signaling
ADD P DO DO DO	1e-13	MZF1	More than 3 adjacent zinc finger factors	0.63	Disease Ontology
TGTGTCTTTCTC	1e-12	SMAD2	SMAD factors	0.62	-log10(Binomial p value)
		::SMAD3			0 5 10 15 20 25
		::SMAD4			nooplasm in vascular tissue 25.42
					Adenoviridae infectious disease 22.07 papillary adenocarcinoma 19.70
		П			hemangloma 17.72 Wirkett-Aldrich wordsome 15.73
		U			osteonecrosis 9.62
					angiosarcoma 8.96 proliferative diabetic retinopathy 8.57
					splenic disease 8.39
					lymphoid leukemia 5.21

Figure S23: Functional and motif analysis of the DECRES genome-wide predictions on cell line HUVEC. A,B,C: Top enriched biological processes,pathways and diseases, respectively, in the predicted cell-specific CRRs. D: Enriched *de novo* motifs in the predicted cell specific CRRs. Column 4: families of best-matched TFs. Column 5: best match scores.



Figure S24: Functional and motif analysis of the DECRES NiA enhancers on cell line HUVEC. A,B,C: Top 20 enriched biological processes, pathways, and diseases, respectively, in the NiA enhancers. D: Enriched *de novo* motifs in the NiA enhancer regions. Column 4: families of best-matched TFs. Column 5: best match scores.



Figure S25: Functional and motif analysis of the DECRES genome-wide predictions on cell line K562. A,B,C: Top enriched biological processes, pathways and diseases, respectively, in the predicted cell-specific CRRs. D: Enriched *de novo* motifs in the predicted cell specific CRRs. Column 4: families of best-matched TFs. Column 5: best match scores.



Figure S26: Functional and motif analysis of the DECRES NiA enhancers on cell line K562. A,B,C: Top 20 enriched biological processes, pathways, and diseases, respectively, in the NiA enhancers. D: Enriched de novo motifs in the NiA enhancer regions. Column 4: families of best-matched TFs. Column 5: best match scores.



Figure S27: Functional and motif analysis of the Combined genome-wide predictions on cell line GM12878. A: Distance from the predicted A-Es to gene TSSs. B: Distance from the predicted A-Ps to gene TSSs. C,D,E: Top 20 enriched biological processes, pathways, and diseases, respectively, in the predicted cell-specific CRRs. F: Enriched *de novo* motifs in the predicted cell specific CRRs. Column 4: families of best-matched TFs. Column 5: best match scores.



Figure S28: Functional and motif analysis of the Combined genome-wide predictions on cell line HelaS3. A: Distance from the predicted A-Es to gene TSSs. B: Distance from the predicted A-Ps to gene TSSs. C,D,E: Top 20 enriched biological processes, pathways, and diseases, respectively, in the predicted cell-specific CRRs. F: Enriched *de novo* motifs in the predicted cell specific CRRs. Column 4: families of best-matched TFs. Column 5: best match scores.



Figure S29: Functional and motif analysis of the Combined genome-wide predictions on cell line HepG2. A: Distance from the predicted A-Es to gene TSSs. B: Distance from the predicted A-Ps to gene TSSs. C,D,E: Top 20 enriched biological processes, pathways, and diseases, respectively, in the predicted cell-specific CRRs. F: Enriched *de novo* motifs in the predicted cell specific CRRs. Column 4: families of best-matched TFs. Column 5: best match scores.



Figure S30: Functional and motif analysis of the Combined genome-wide predictions on cell line K562. A: Distance from the predicted A-Es to gene TSSs. B: Distance from the predicted A-Ps to gene TSSs. C,D,E: Top 20 enriched biological processes, pathways, and diseases, respectively, in the predicted cell-specific CRRs. F: Enriched *de novo* motifs in the predicted cell specific CRRs. Column 4: families of best-matched TFs. Column 5: best match scores.

## References

- B. Arnett, P. Soisson, B.S. Ducatman, and P. Zhang. Expression of CAAT enhancer binding protein beta (C/EBP beta) in cervix and endometrium. *Molecular Cancer*, 2:21, 2003.
- [2] M. Beger, K. Butz, C. Denk, T. Williams, H.C. Hurst, and F. Hoppe-Seyler. Expression pattern of AP-2 transcription factors in cervical cancer cells and analysis of their influence on human papillomavirus oncogene transcription. *Journal of Molecular Medicine*, 79(5-6):314–320, 2001.
- [3] R.H. Costa, V.V. Kalinichenko, A.X. Holterman, and X. Wang. Transcription factors in liver development, differentiation, and regeneration. *Hepatology*, 38(6):1331–1347, 2003.
- [4] A. Dev, S. Iyer, B. Razani, and G. Cheng. NF-κB and innate immunity. Current Topics in Microbiology and Immunology, 349:115–143, 2011.
- [5] J.D. Fleming, G. Pavesi, P. Benatti, C. Imbriano, R. Mantovani, and K. Struhl. NF-Y coassociates with FOS at promoters, enhancers, repetitive elements, and inactive chromatin regions, and is stereopositioned with growth-controlling transcription factors. *Genome Research*, 23(8):1195–1209, 2013.
- [6] D. Kleftogiannis, P. Kalnis, and V.B. Bajic. DEEP: A general computional framework for predicting enhancers. *Nucleic Acids Research*, 43(1):e6, 2015.
- [7] A.K. Linnemann, H. O'Geen, S. Keles, P.J. Farnham, and E.H. Bresnick. Genetic framework for GATA factor function in vascular biology. *Proceedings of the National Academy of Sciences*, 108(33):13641– 13646, 2011.
- [8] R. Lopez, E. Garrido, G. Vazquez, P. Pina, C. Perez, I. Alvarado, and M. Salcedo. A subgroup of HOX Abd-B gene is differentially expressed in cervical cancer. *International Journal Gynecological Cancer*, 16(3):1289–1296, 2006.
- [9] A.L. Malt, J. Cagliero, K. Legent, J. Silber, A. Zider, and D. Flagiello. Alteration of TEAD1 expression levels confers apoptotic resistance through the transcriptional up-regulation of Livin. *PLoS One*, 7(9):e45498, 2012.
- [10] E.V. Mityushova, N.D. Aksenov, and I.I. Marakhova. STAT5 in regulation of chronic leukemia K562 cell proliferation: Inhibitory effect of WHI-P131. *Cell and Tissue Biology*, 4(1):63–69, 2010.
- [11] T. Okuda, M. Nishimura, M. Nakao, and Y. Fujita. RUNX1/AML1: A central player in hematopoiesis. International Journal of Hematology, 74(3):252–257, 2001.
- [12] G.V. Samant, M.O. Schupp, M. Francois, S. Moleri, R.K. Kothinti, C.Z. Chun, I. Sinha, S. Sellars, N. Leigh, K. Pramanik, M.A. Horswill, I. Remadevi, K. Li, G.A. Wilkinson, N.M. Tabatabai, M. Beltrame, P. Koopman, and R. Ramchandran. Sox factors transcriptionally regulate ROBO4 gene expression in developing vasculature in zebrafish. *The Journal of Biological Chemistry*, 286:30740–30747, 2011.
- [13] H. Schrem, J. Klempnauer, and J. Borlak. Liver-enriched transcription factors in liver function and development. Part II: The C/EBPs and D site-binding protein in cell cycle control, carcinogenesis, circadian gene regulation, liver regeneration, apoptosis, and liver-specific gene regulation. *Pharmacological Reviews*, 56(2):291–330, 2004.
- [14] C.C. Thornton, F. Al-Rashed, D. Calay, G.M. Birdsey, A. Bauer, H. Mylroie, B.J. Morley, A.M. Randi, D.O. Haskard, J.J. Boyle, and J.C. Mason. Methotrexate-mediated activation of an AMPK-CREBdependent pathway: A novel mechanism for vascular protection in chronic systemic inflammation. *Annals* of the Rheumatics Diseases, 75(2):439–448, 2015.
- [15] P. Wang, J. Xu, and C. Zhang. CREB, a possible upstream regulator of Bcl-2 in trichosanthin-induced HeLa cell apoptosis. *Molecular Biology Reports*, 37(4):1891–1896, 2010.
- [16] Z. Wang, E.P. Bishop, and P.A. Burke. Expression profile analysis of the inflammatory response regulated by hepatocyte nuclear factor 4α. BMC Genomics, 12:128, 2011.

- [17] H. Zhang, C. Liu, Z. Zha, B. Zhao, J. Yao, S. Zhao, Y. Xiong, Q. Lei, and K. Guan. TEAD transcription factors mediate the function of TAZ in cell growth and epithelial-mesenchymal transition. *The Journal* of *Biological Chemistry*, 284(20):13355–13362, 2009.
- [18] J. Zhu, D.M. Giannola, Y. Zhang, A.J. Rivera, and S.G. Emerson. NF-Y cooperates with USF1/2 to induce the hematopoietic expression of HOXB4. *Blood*, 102(7):2420–2427, 2003.