

## Supporting Information

### Modular Synthesis of Biologically Active Phosphatidic Acid Probes Using Click Chemistry

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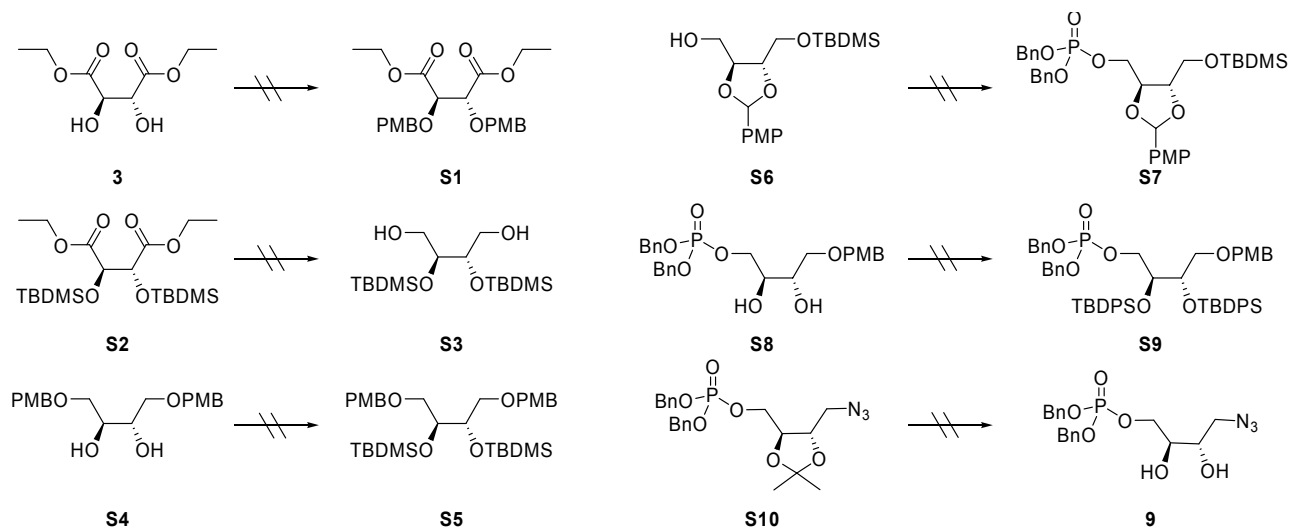
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46617 <sup>c</sup>Department of Chemistry and Biochemistry and the Walther Center for Cancer Research, University of Notre Dame, Notre Dame, IN 46656

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- V. Protecting Group Strategies Explored for the Interior Diol of Scaffold 2

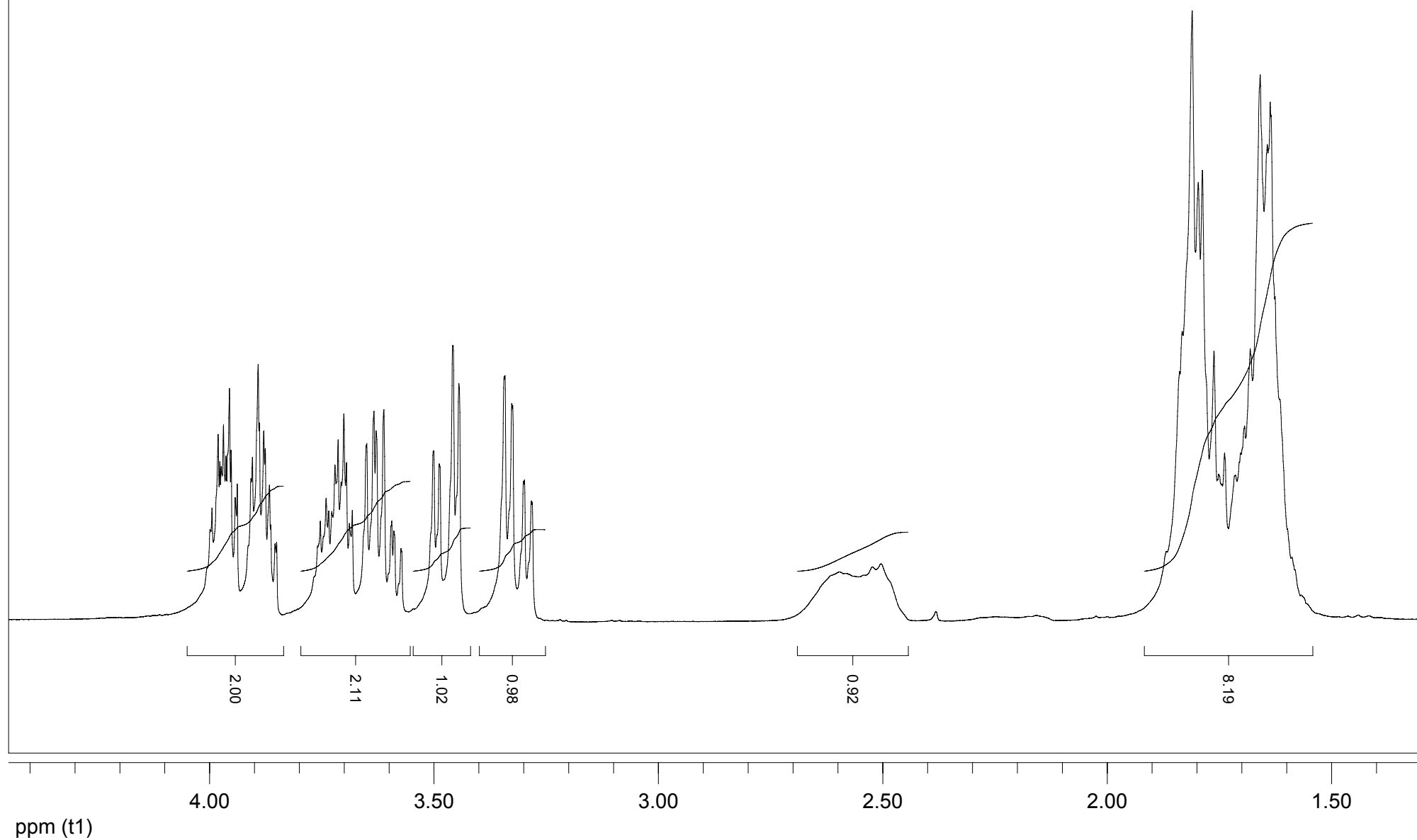
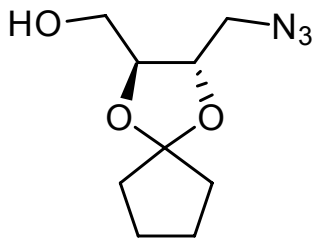
A key aspect of this synthesis was the determination of the optimal protecting group for the interior diol of **8** that could be removed without affecting the phosphotriester and azide groups. Several approaches were explored to determine that which is most effective. These routes are outlined in Scheme S1, in which the steps that were unsuccessful are indicated. First, the installation of two *p*-methoxybenzyl protecting groups onto the diol of **3** to **S1** was ineffective. While two silyl protecting groups could be introduced following literature procedures<sup>1,2</sup> to **S2**, these were removed during ester reduction in attempting to access **S3**. However, the introduction of two silyl groups onto the diol of **S4** to access **S5** was unsuccessful. While *p*-methoxybenzylidene-protected derivative **S6** was synthesized, all oxidation conditions attempted for the production of phosphotriester **S7** led to removal of this group. Bissilylation of **S8** to **S9** was once again unsuccessful. Finally, deprotection of the acetonide group of **S10** under acidic conditions yielded only ~10% of diol **9** due to decomposition.

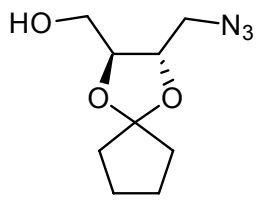


Scheme S1. Protecting group strategies explored in the synthesis of modular PA core 2.

#### References Cited

- 1 J. E. Baldwin; R. M. Adlington; A. T. Russell; M. L. Smith. *J. Chem. Soc., Chem. Commun.* 1994, 85- 86.
- 2 T. Hiyama; T. Minami; K. Takahashi. *Bull. Chem. Soc. Japan* 1995, **68**, 364-372.





119.828

78.543

76.066

62.035

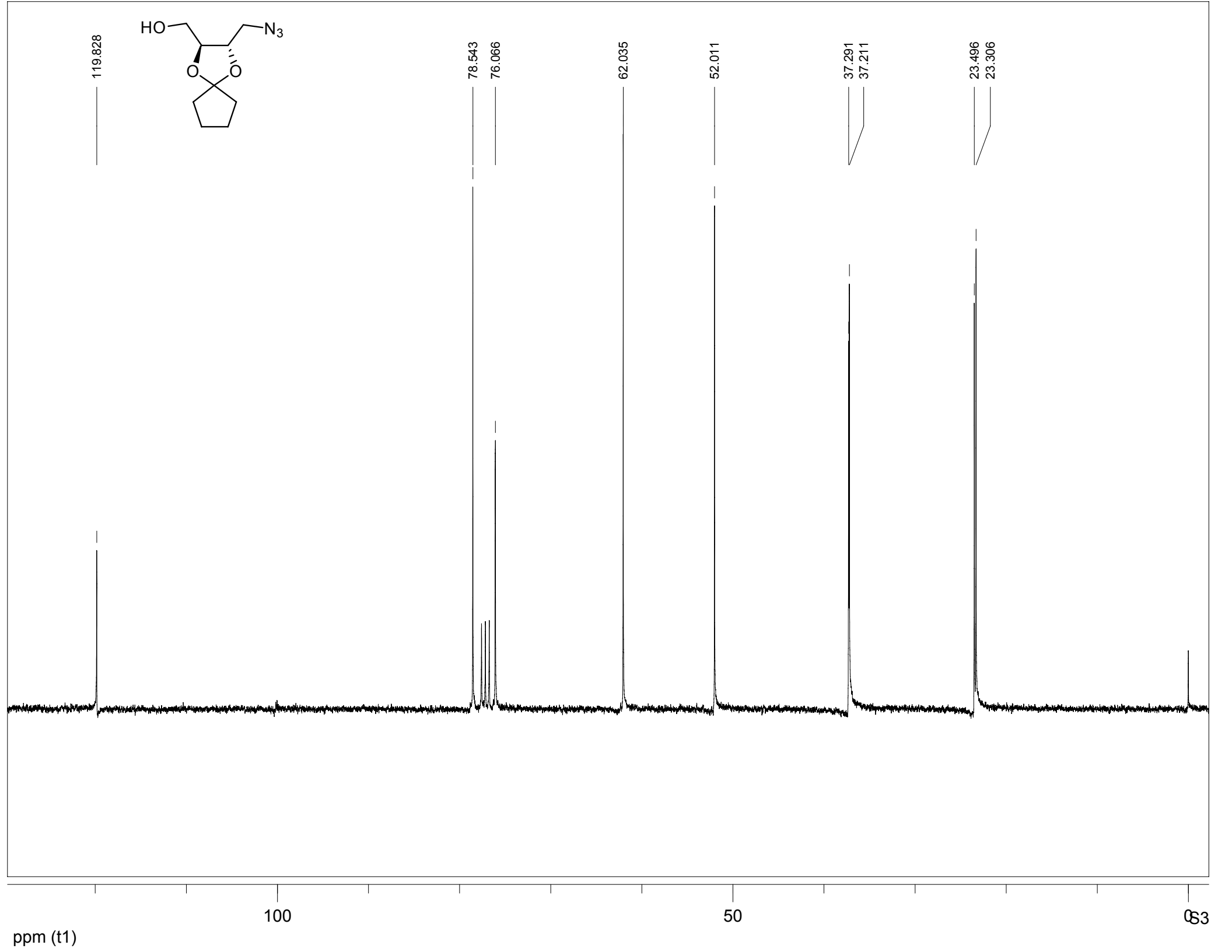
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37.211

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23.306

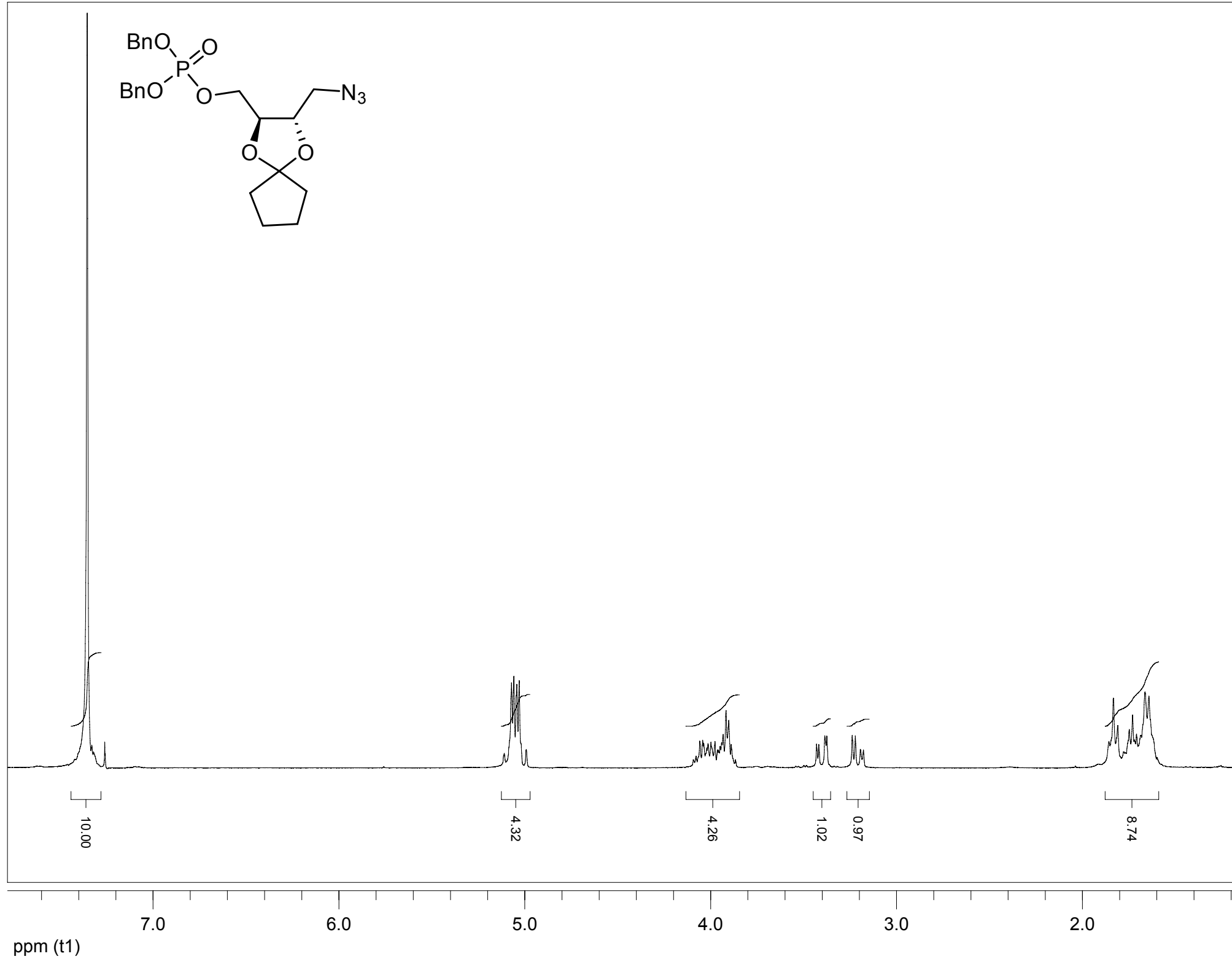
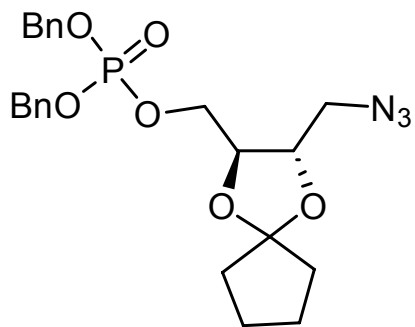


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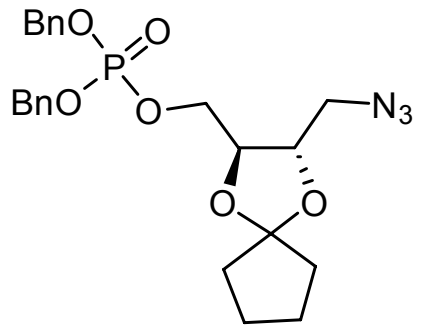
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0





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128.626  
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120.234



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51.788

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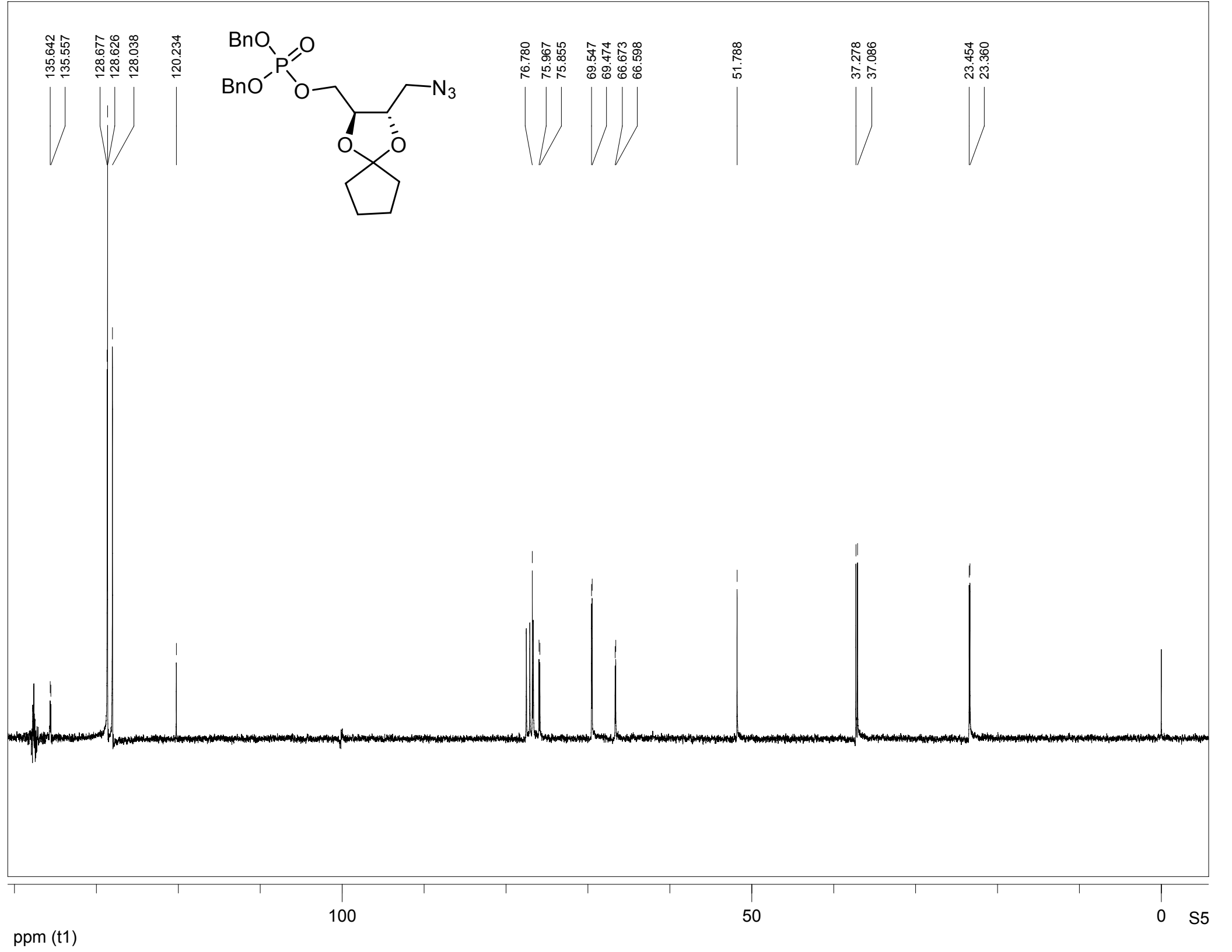
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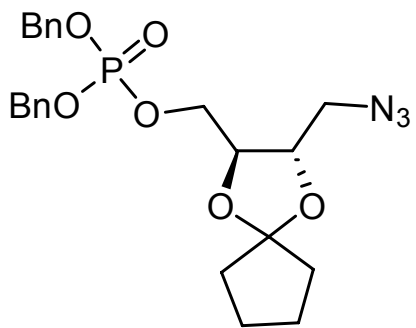
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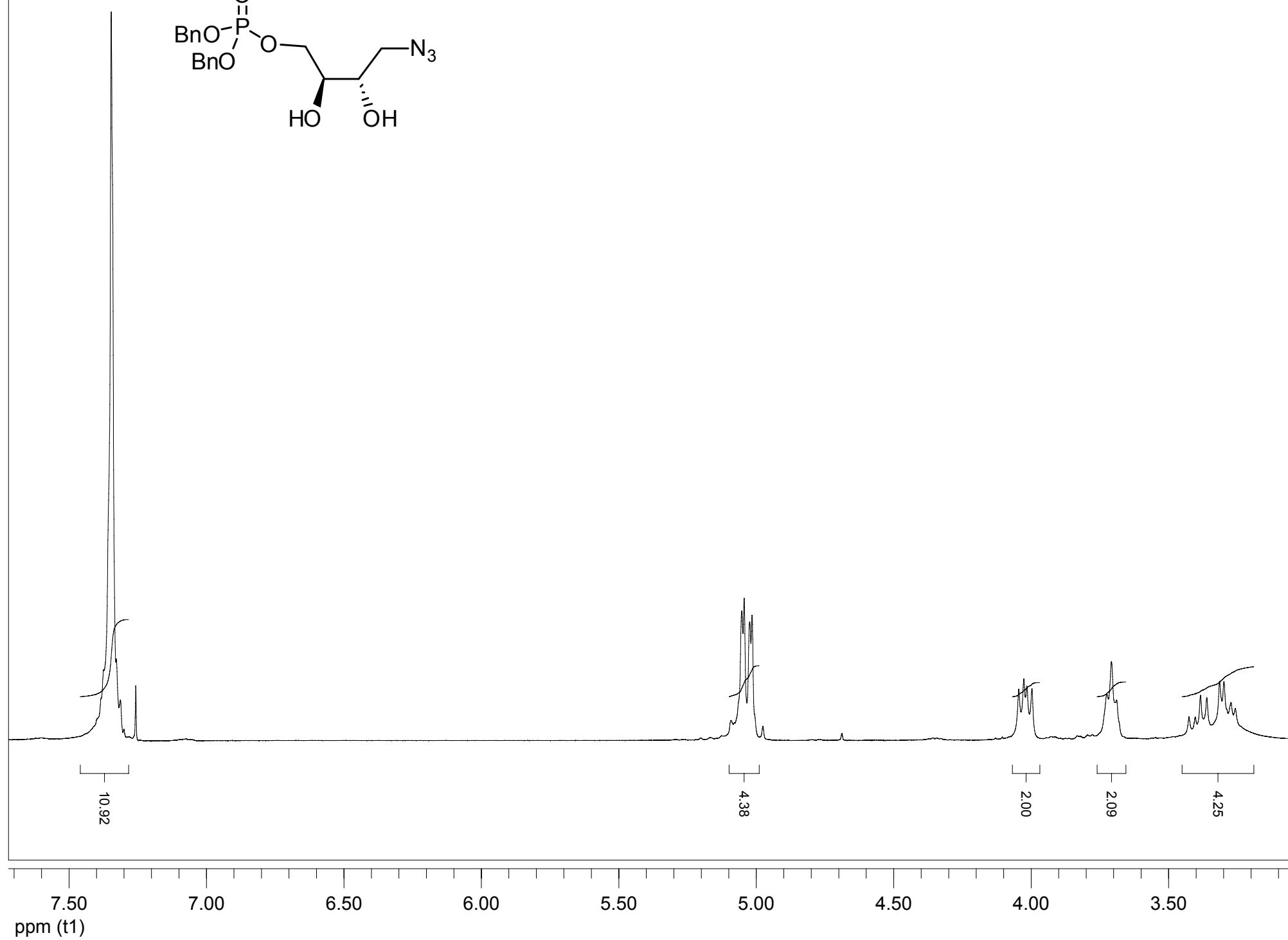
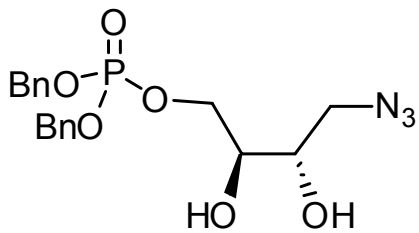
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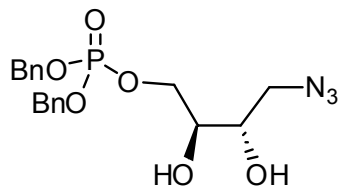
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S6

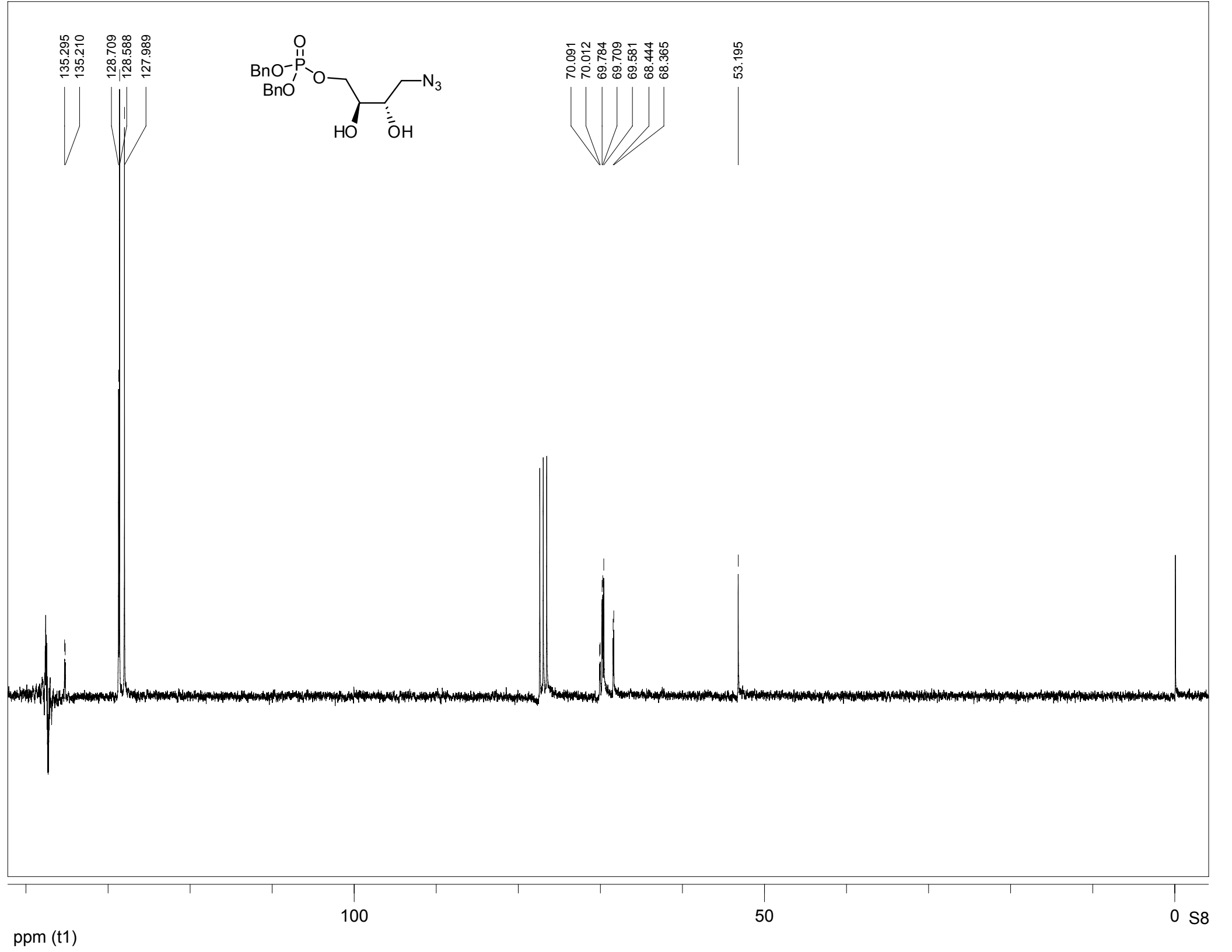


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128.588  
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70.091  
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53.195

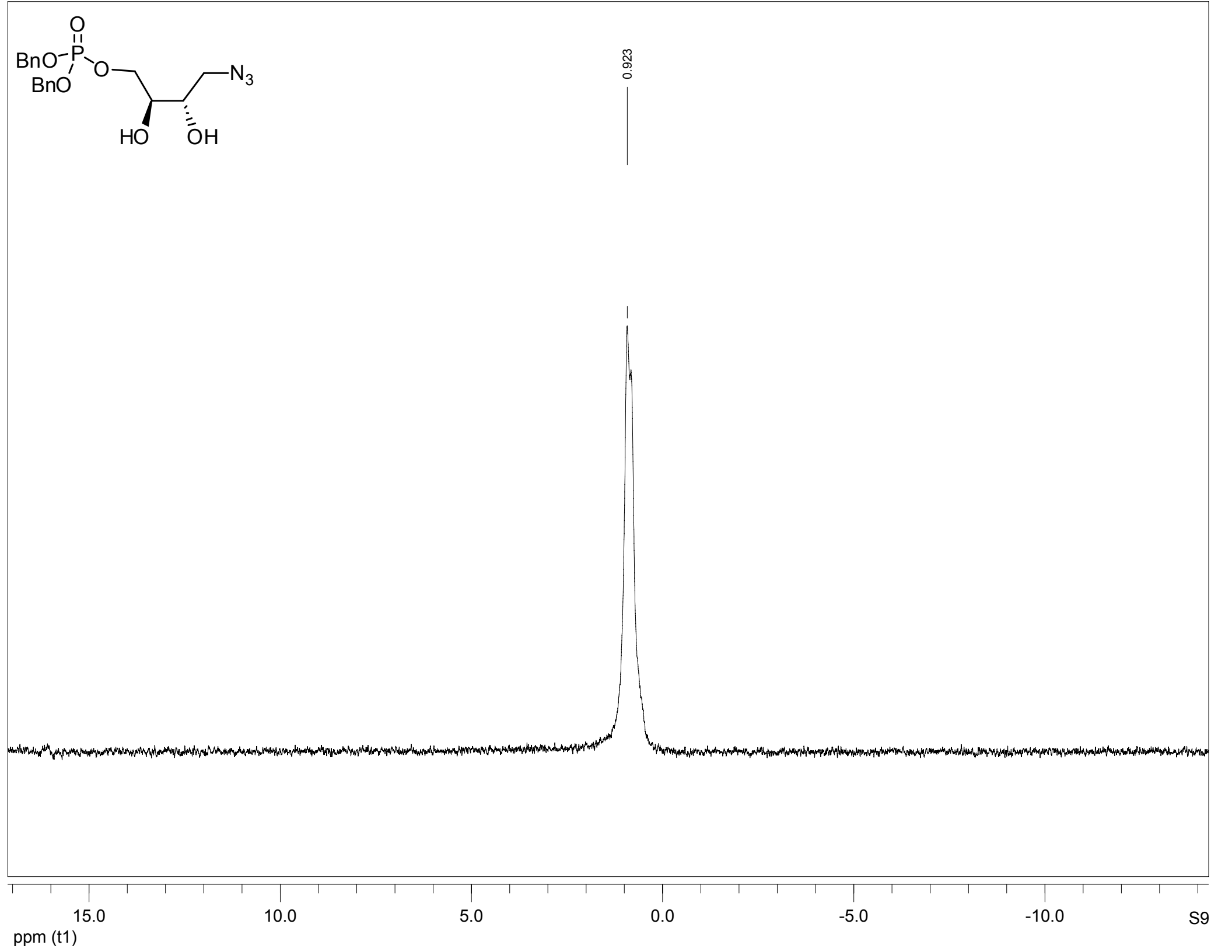
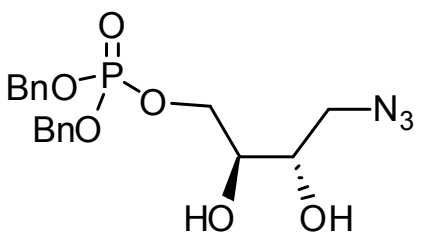


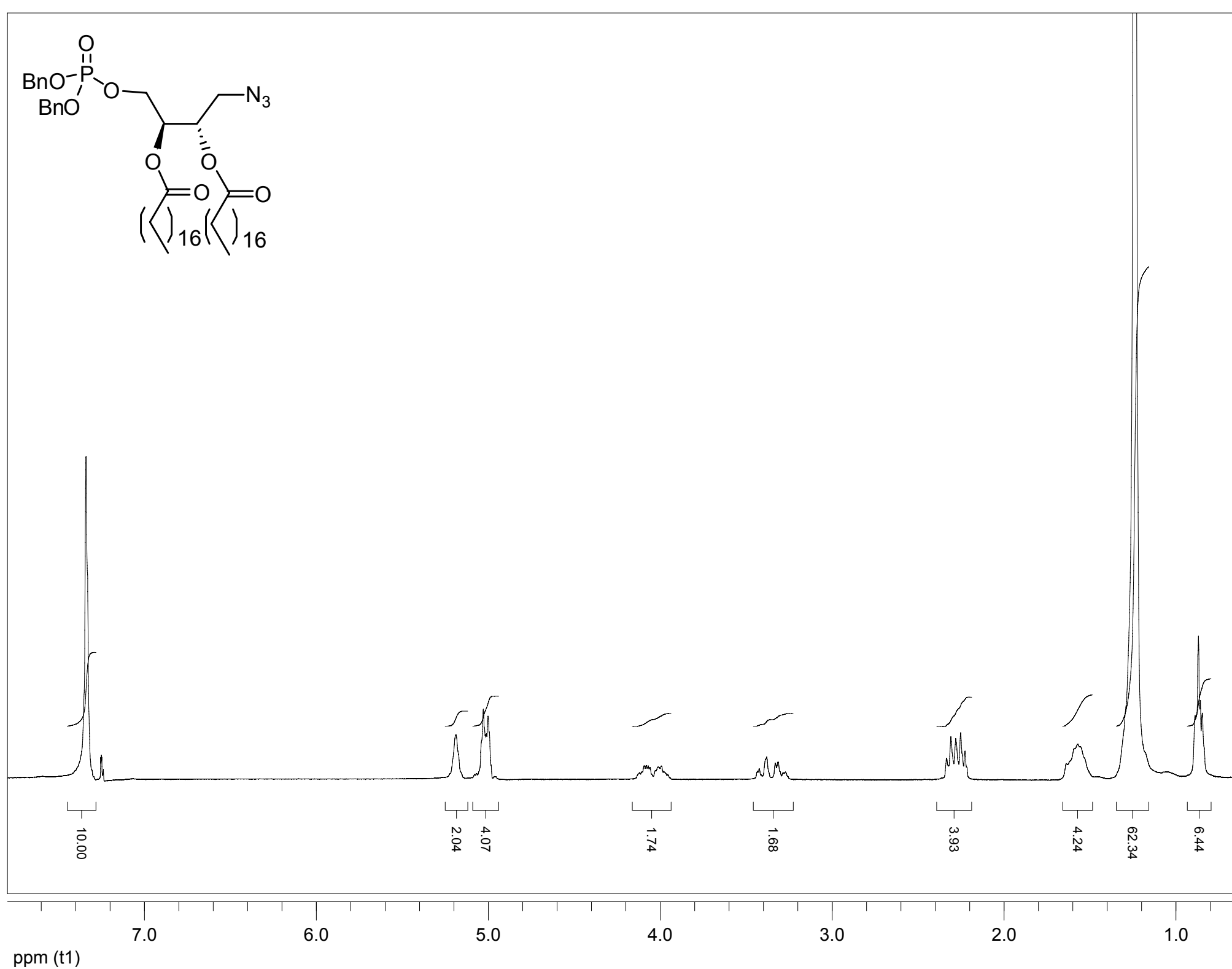
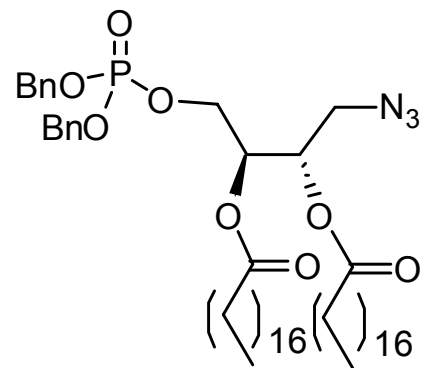
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0 S8

ppm (t1)





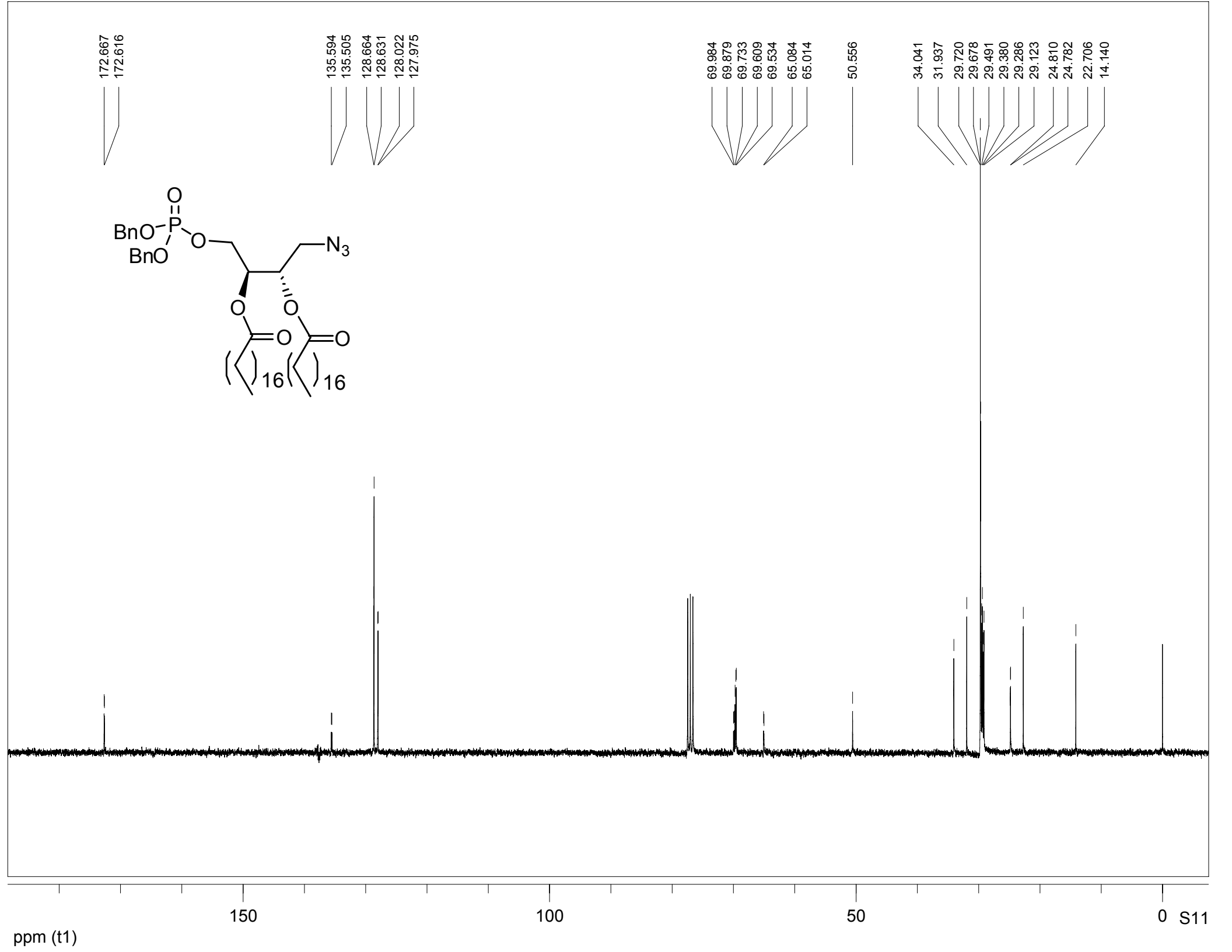
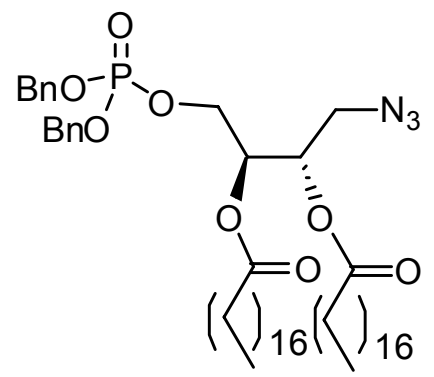
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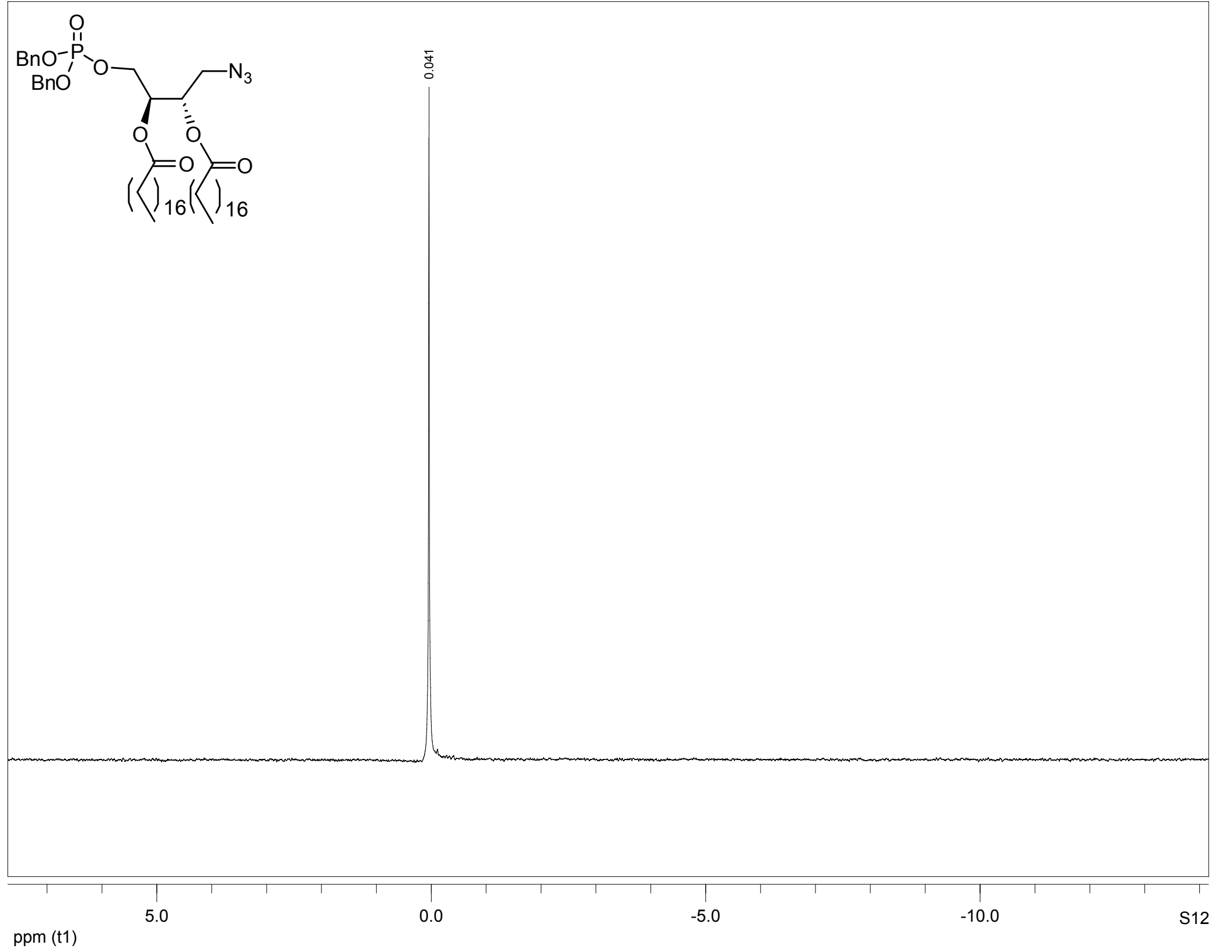
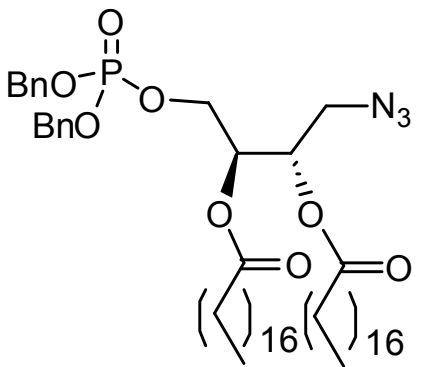
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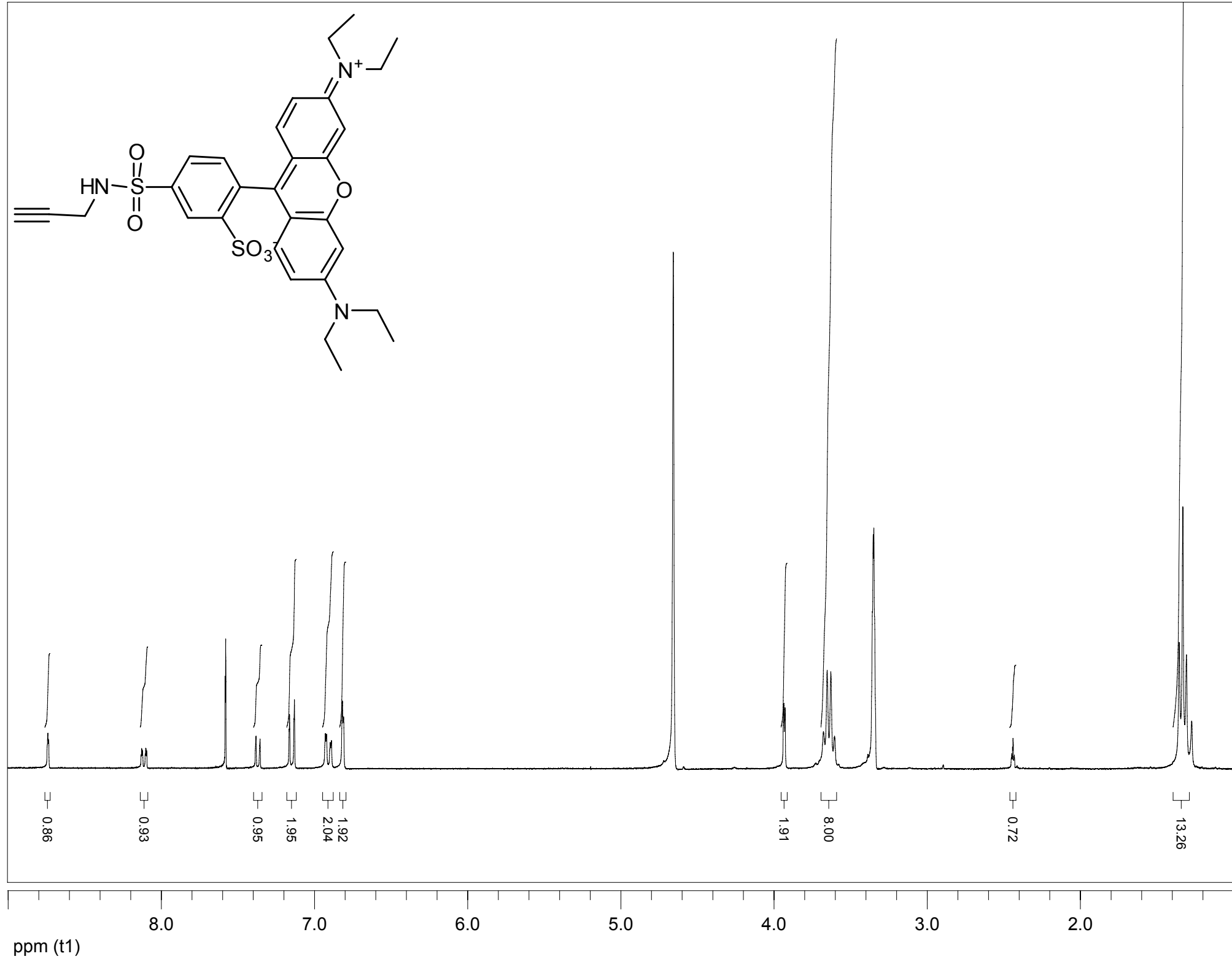
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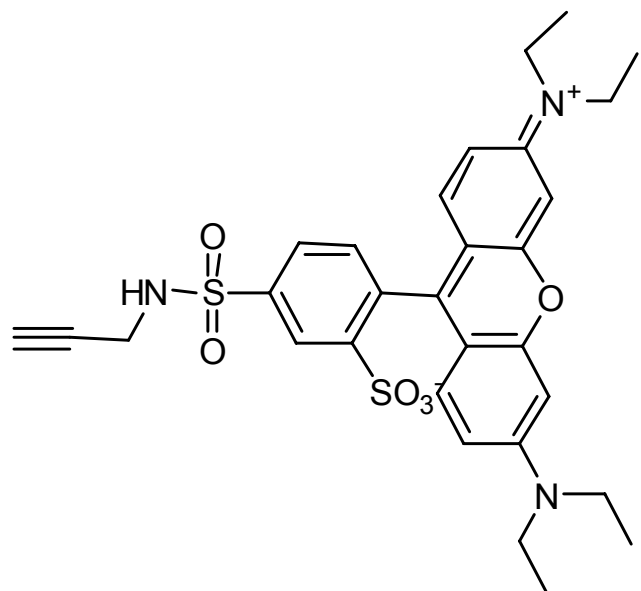




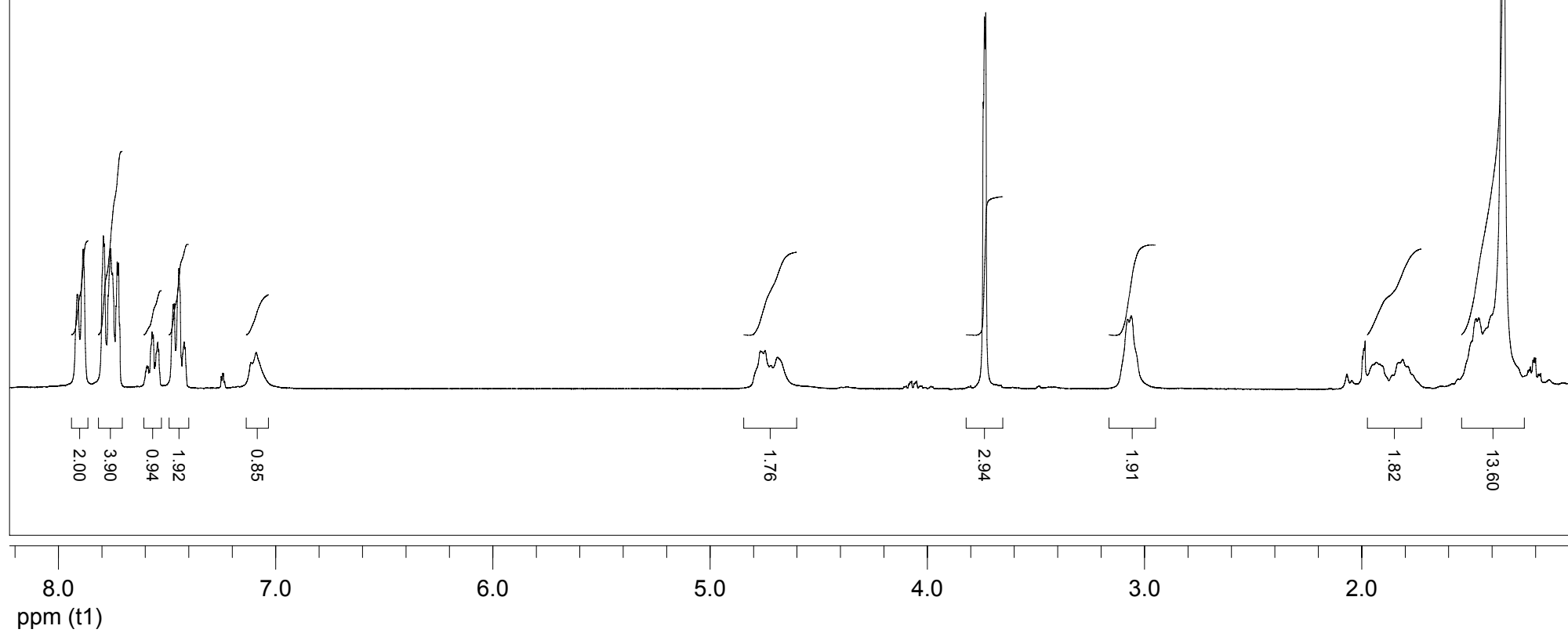
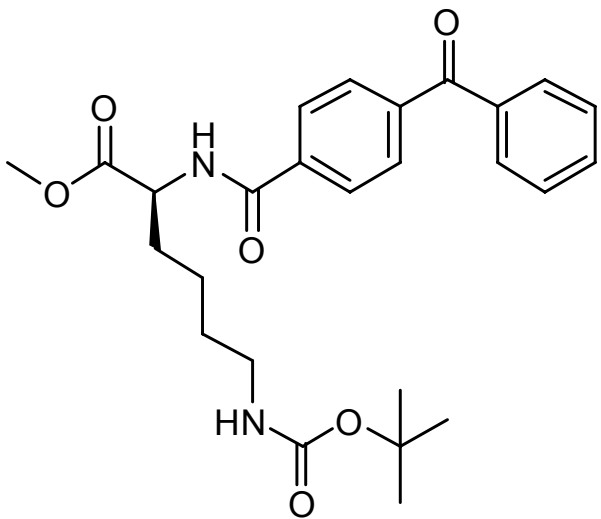


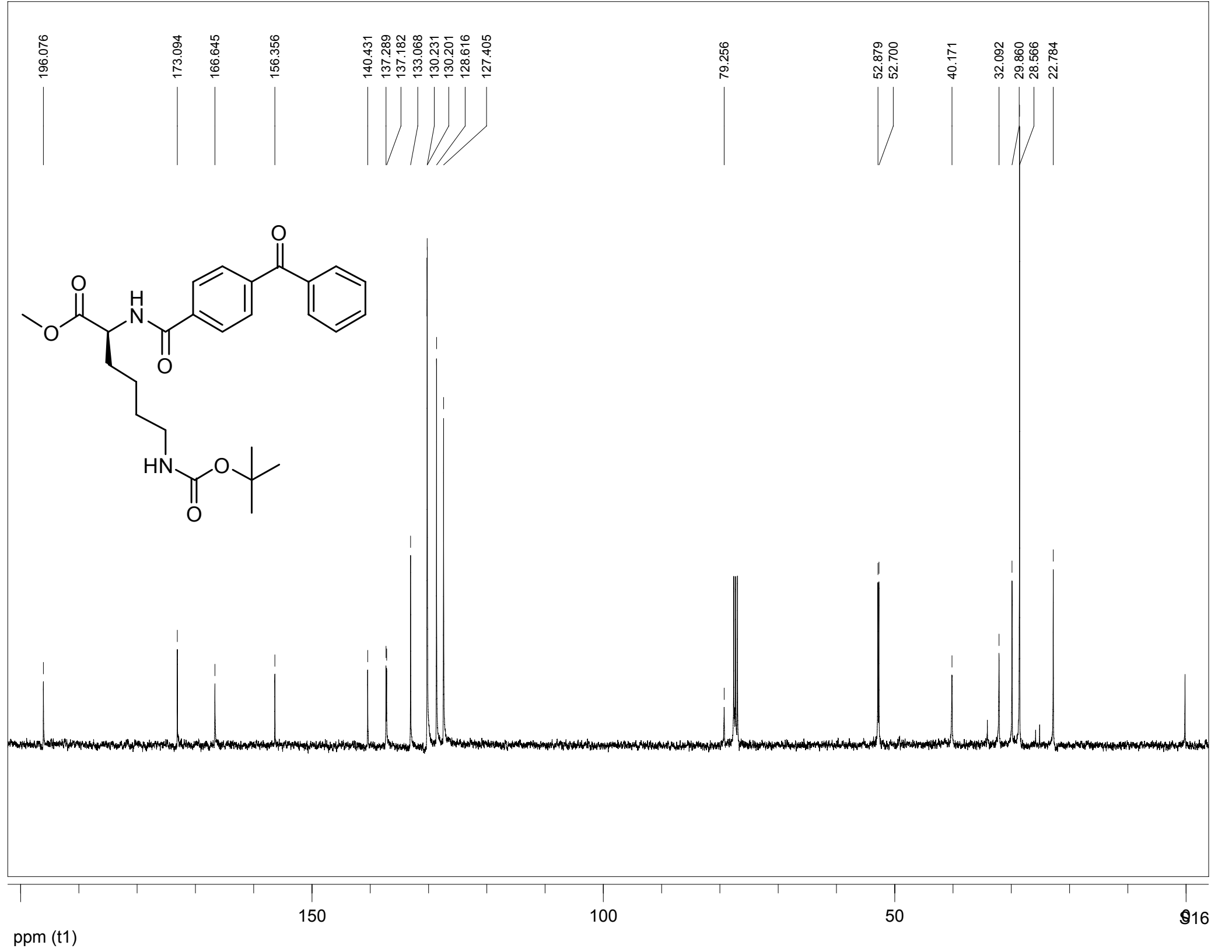
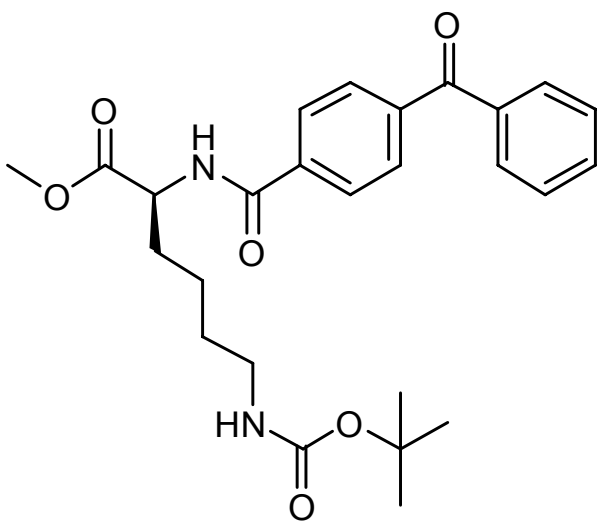


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134.402  
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127.649  
  
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114.094  
  
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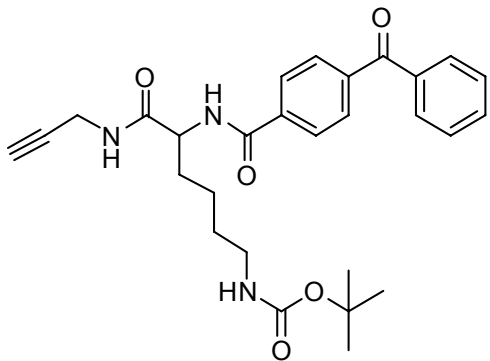
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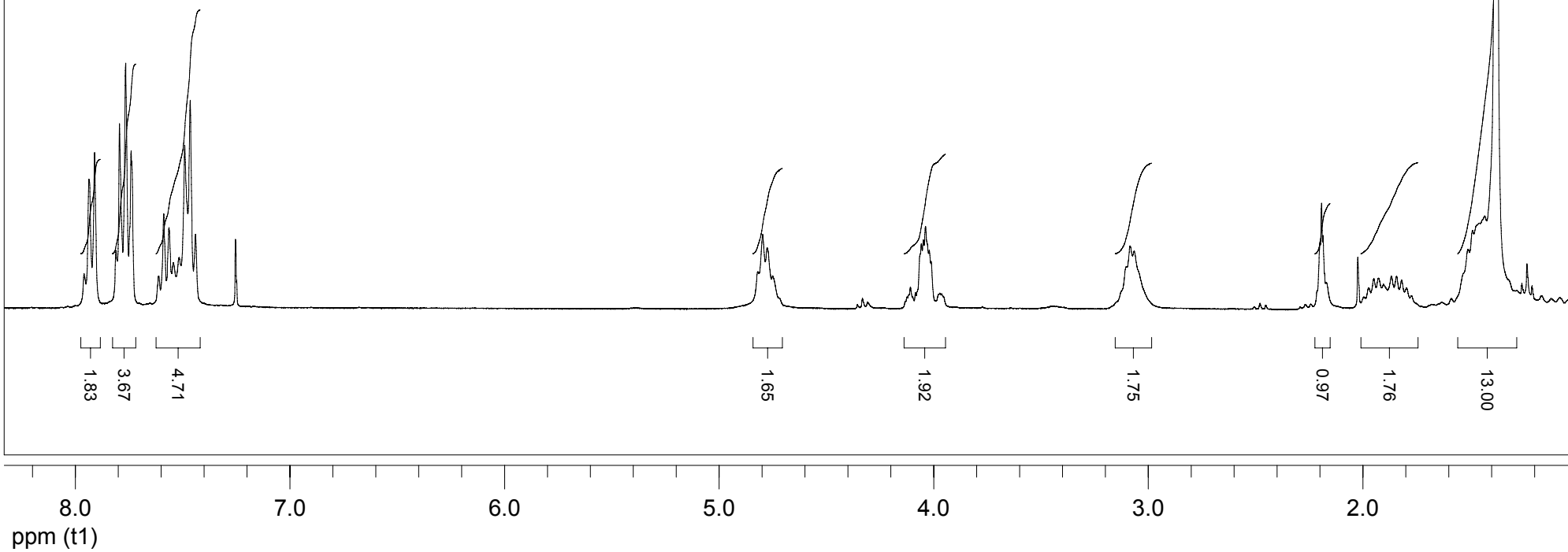


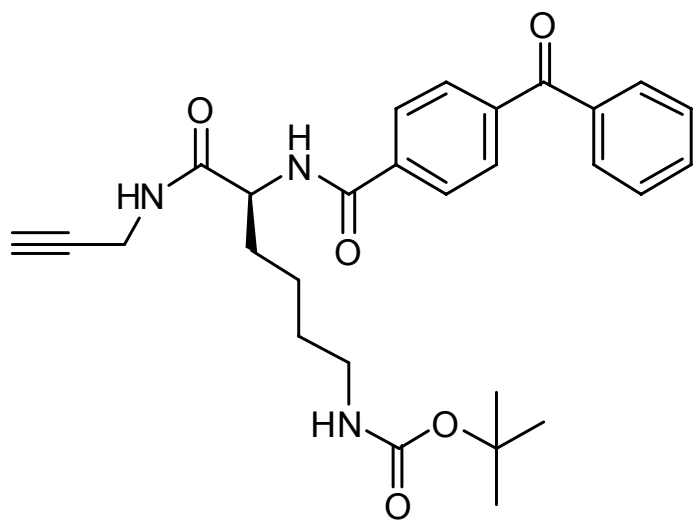
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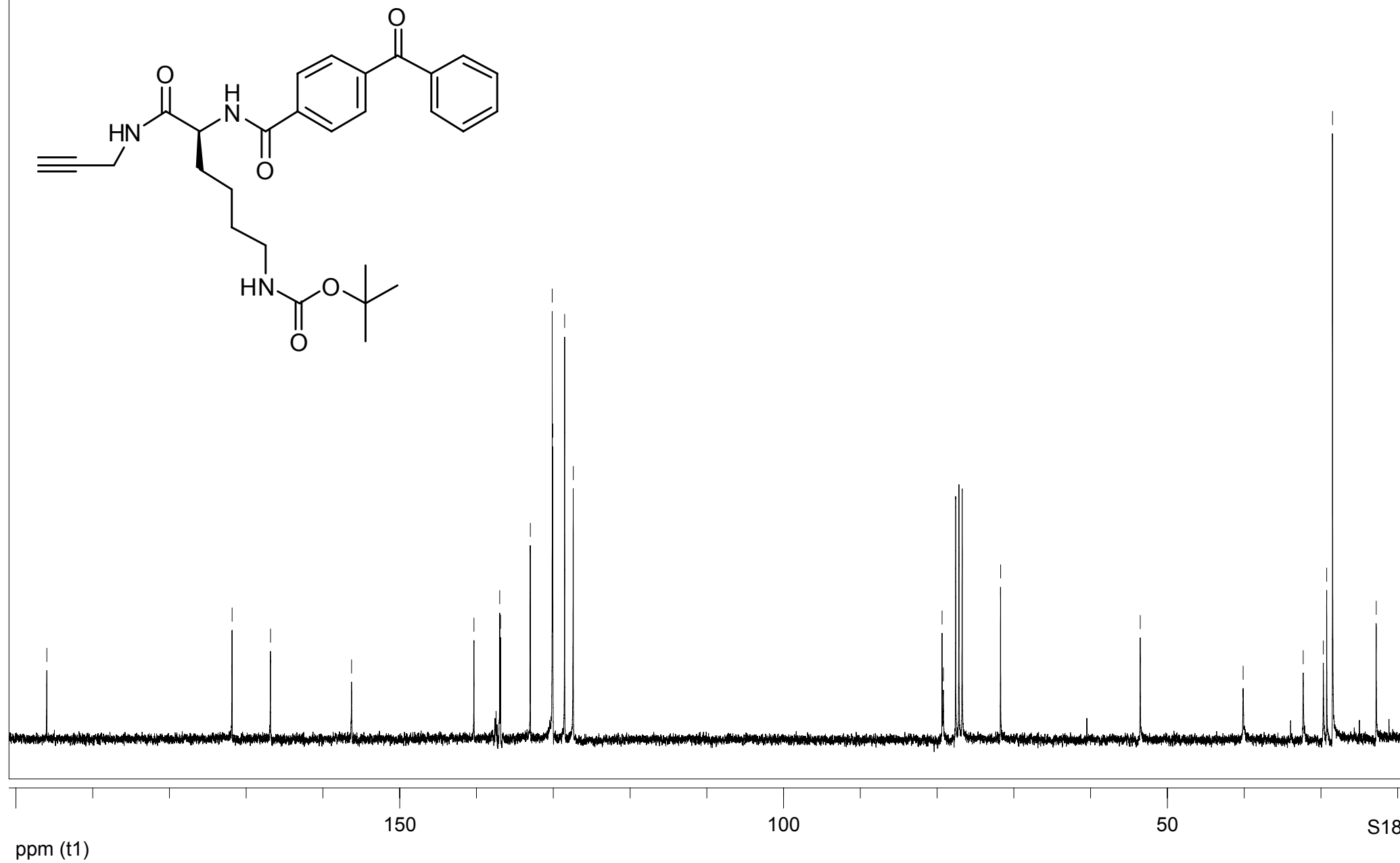
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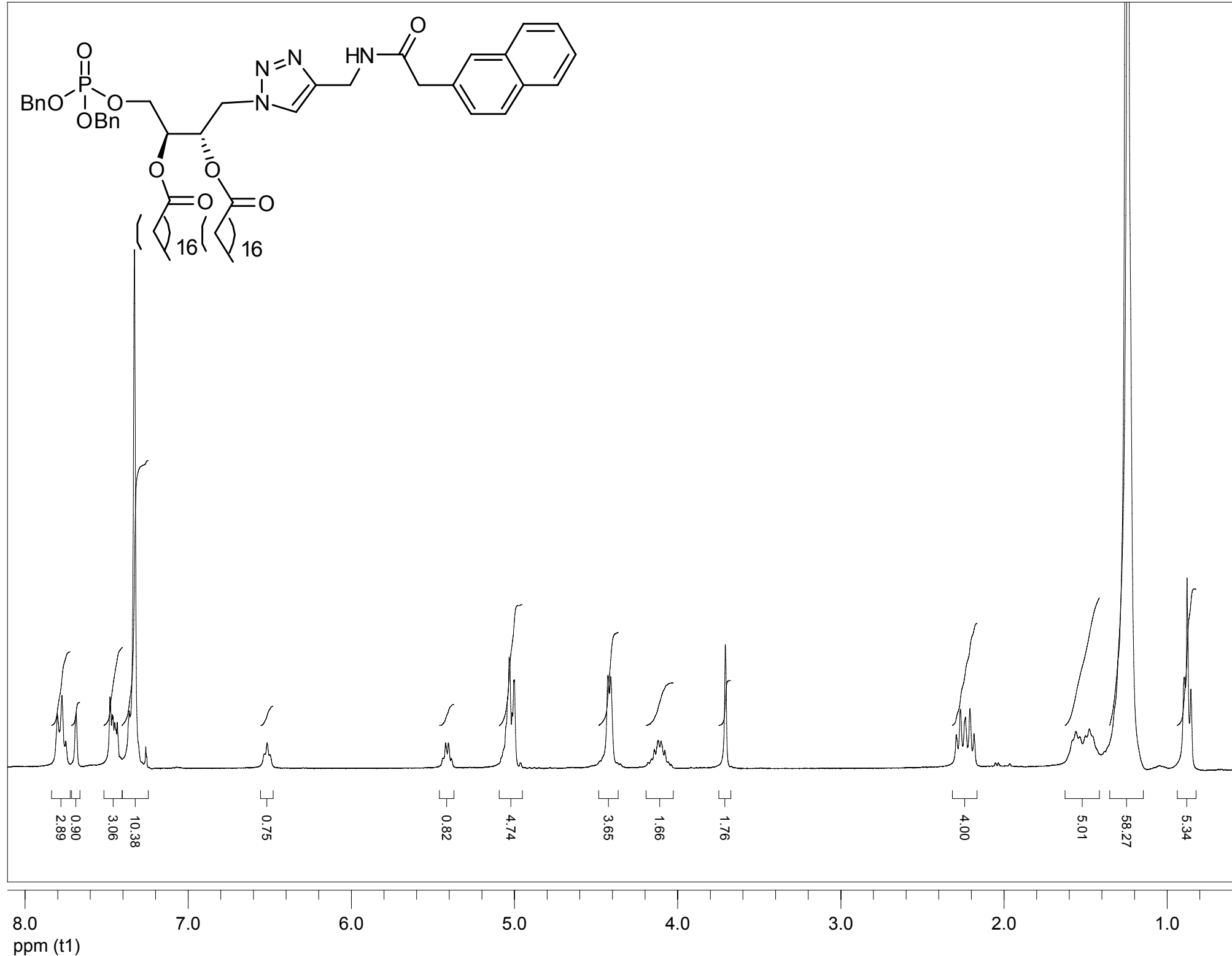
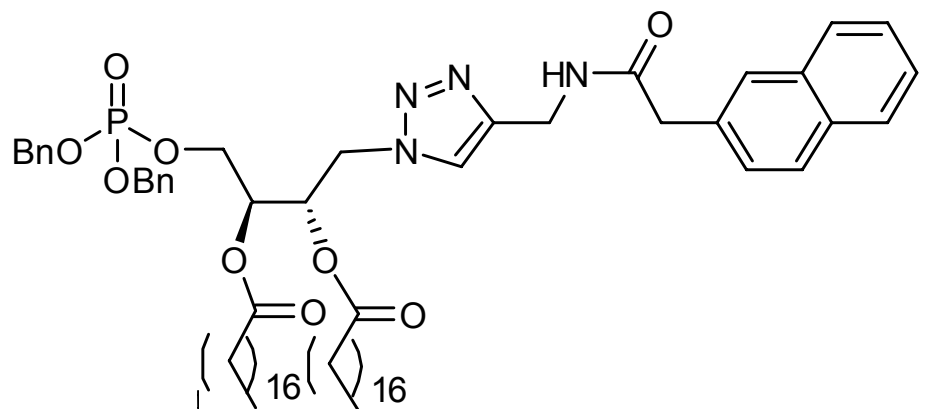




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128.507  
127.395

79.342  
79.180  
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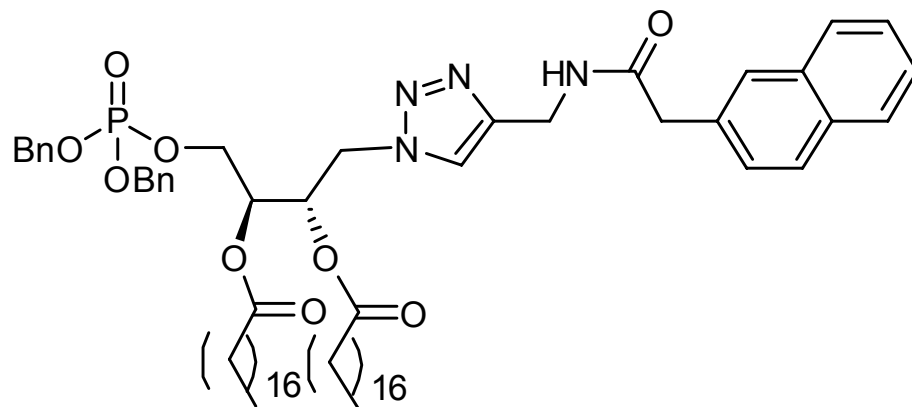
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49.858  
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31.921

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ppm (t1)

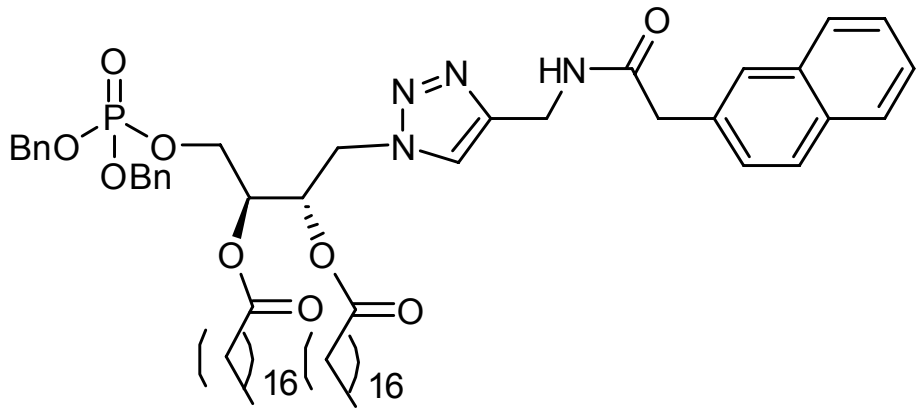
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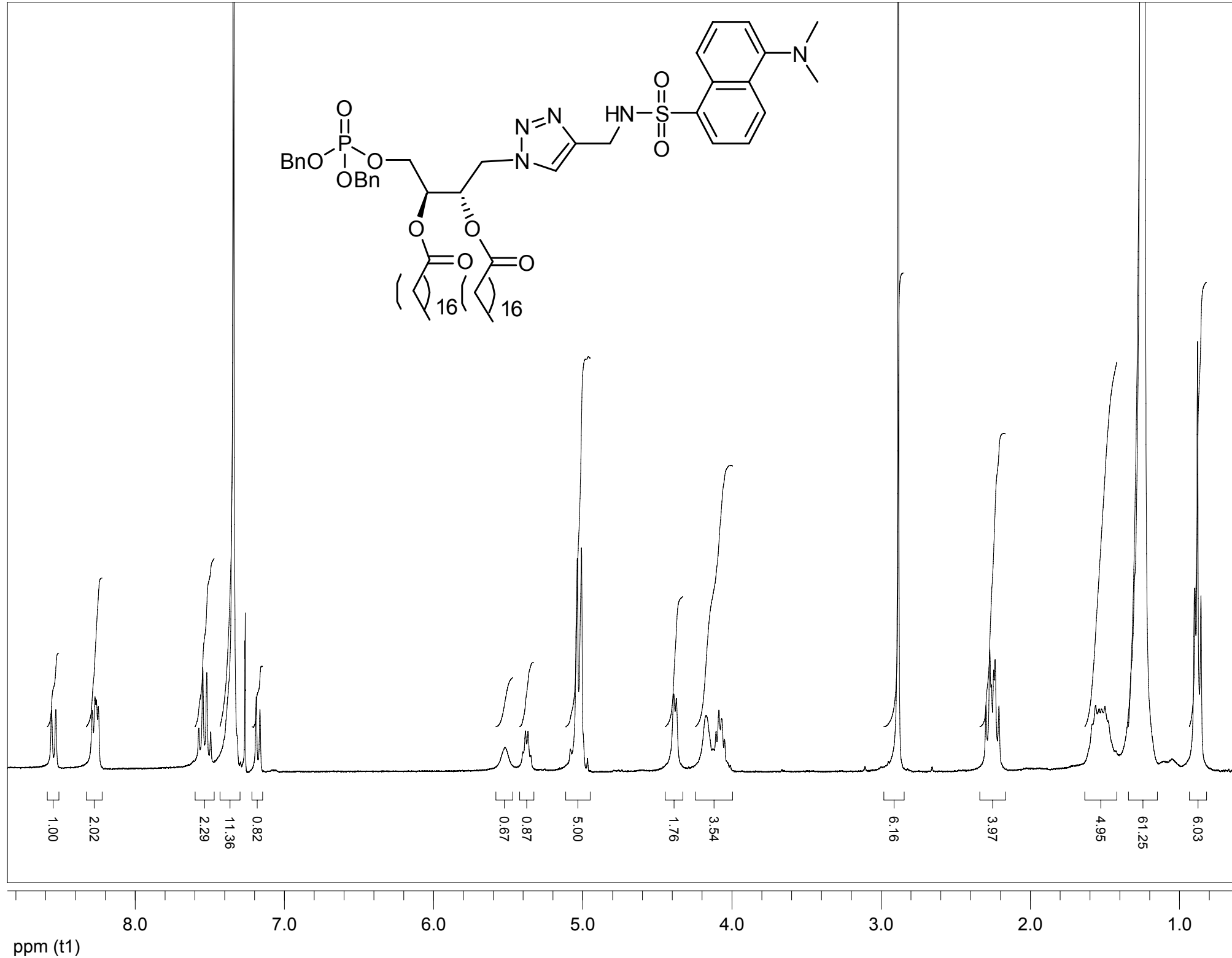
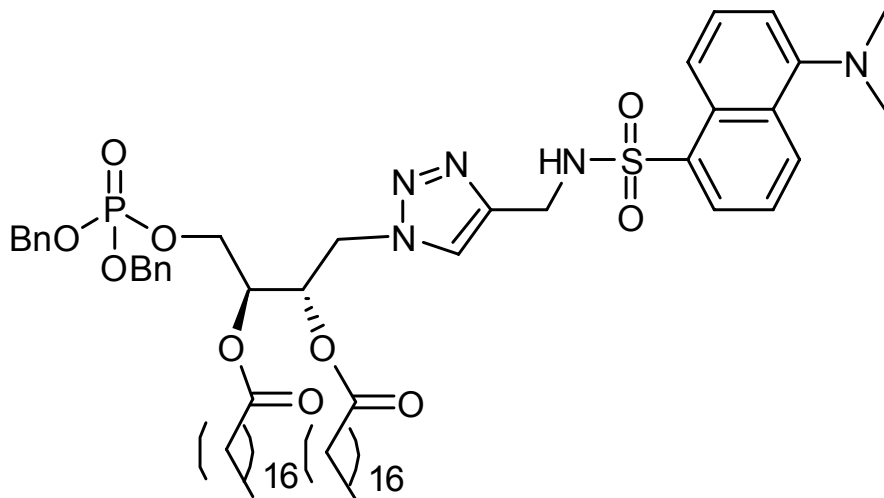
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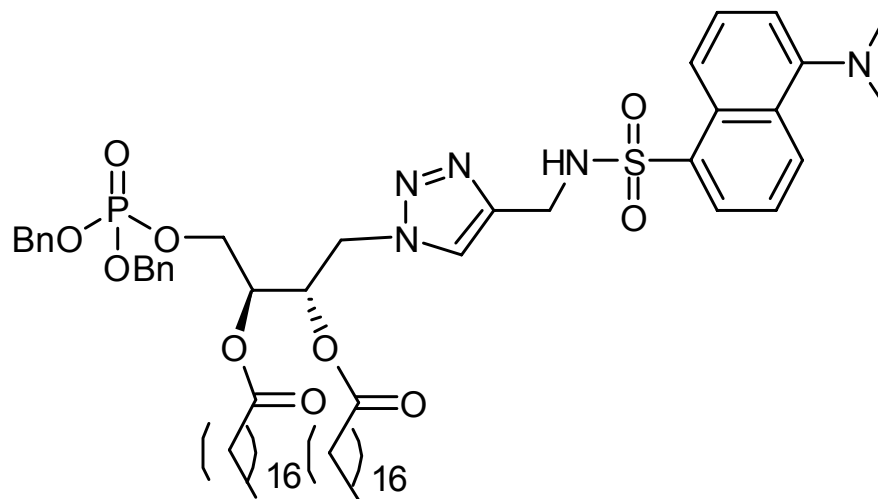
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69.758  
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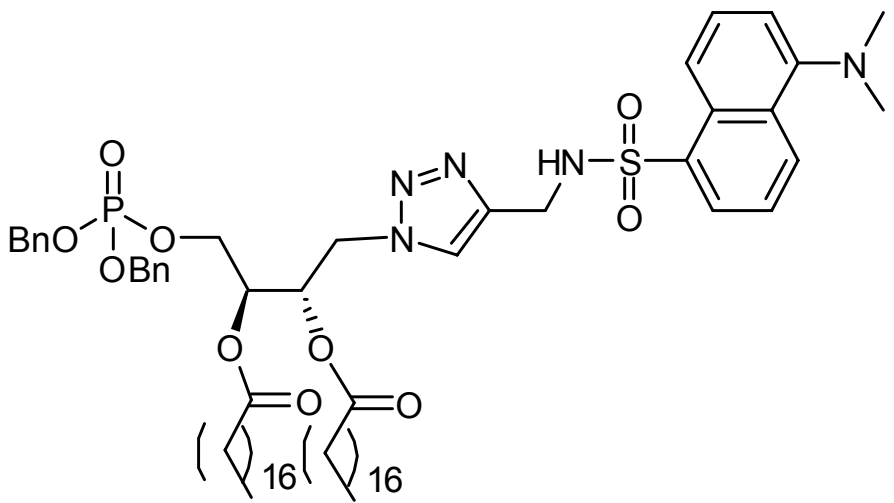
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S23



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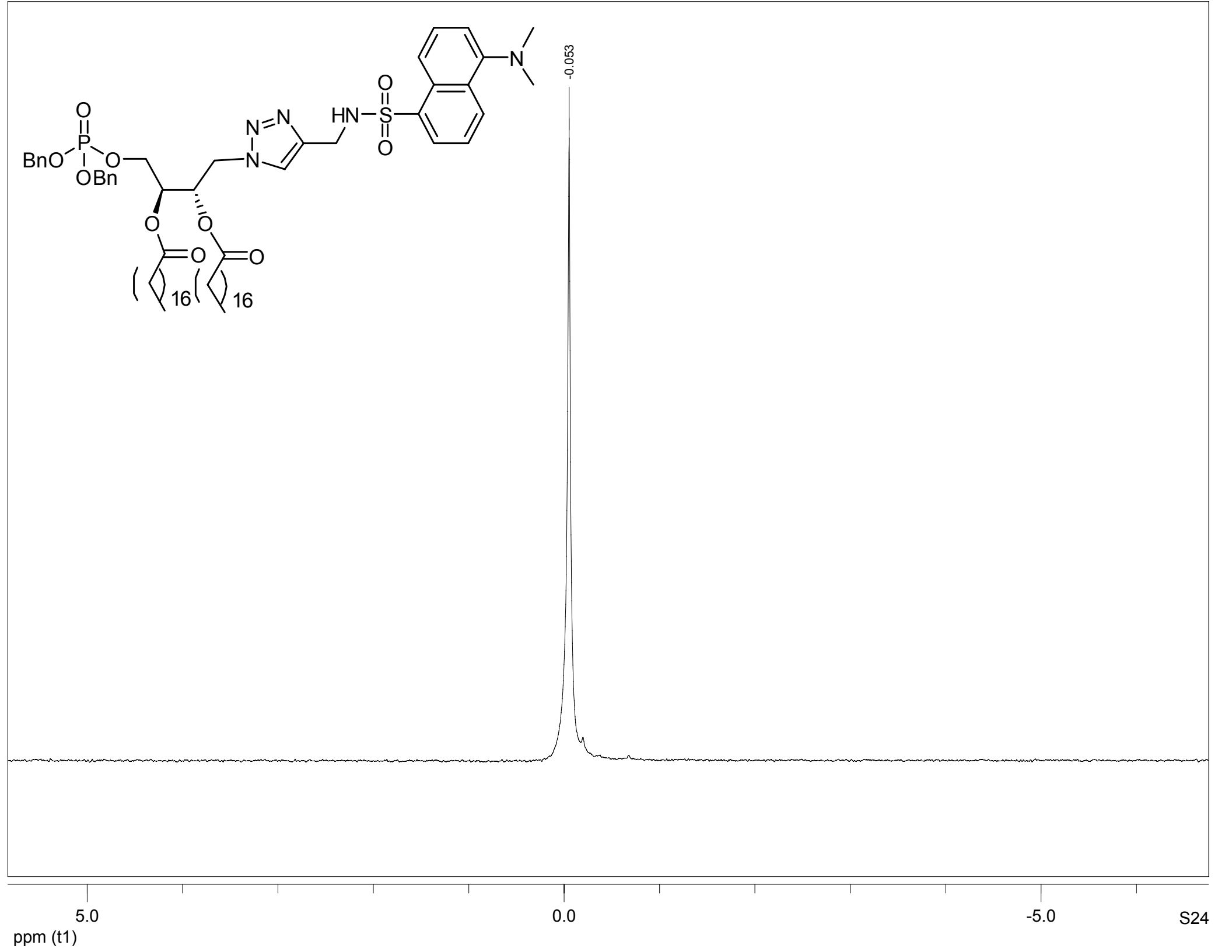
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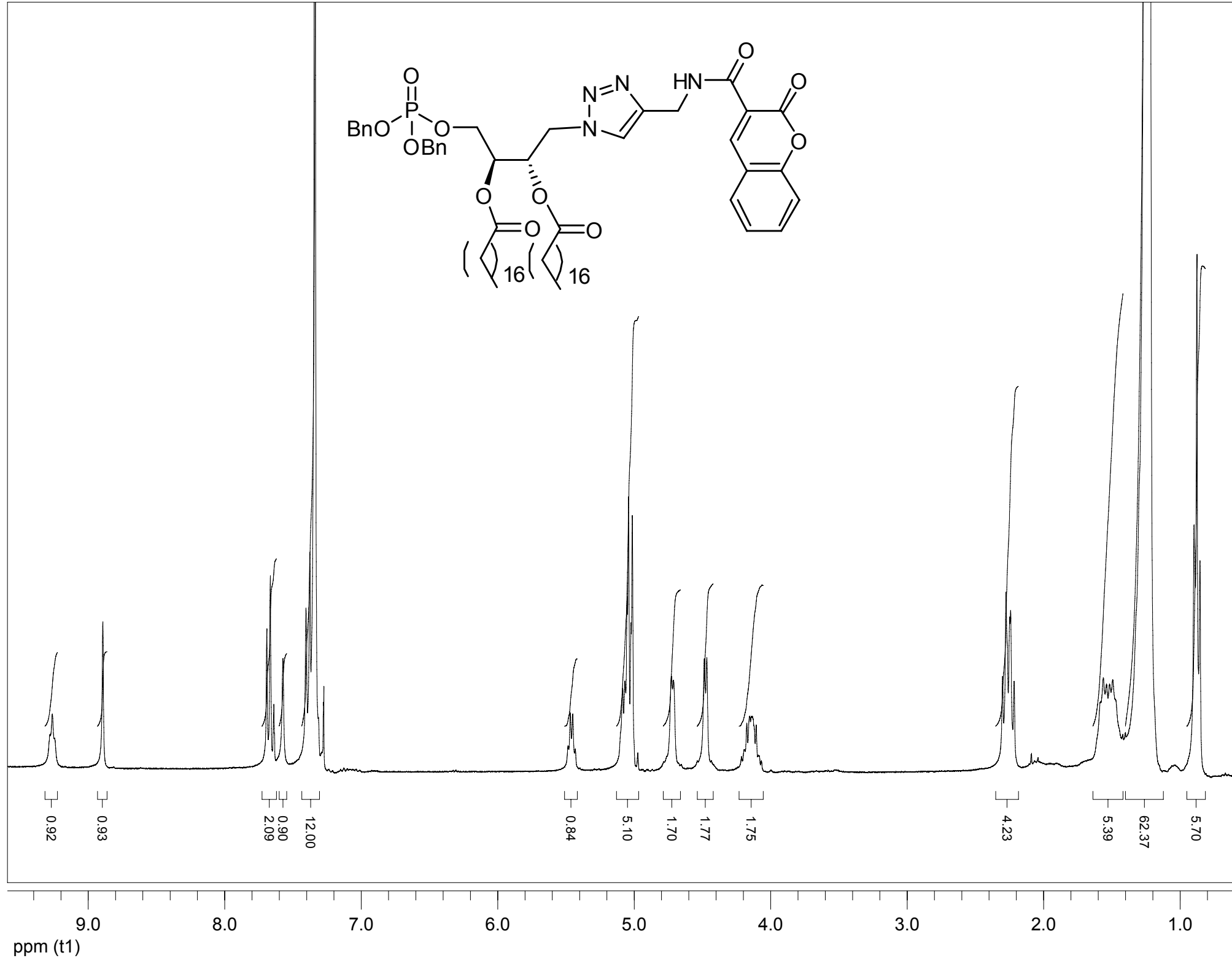
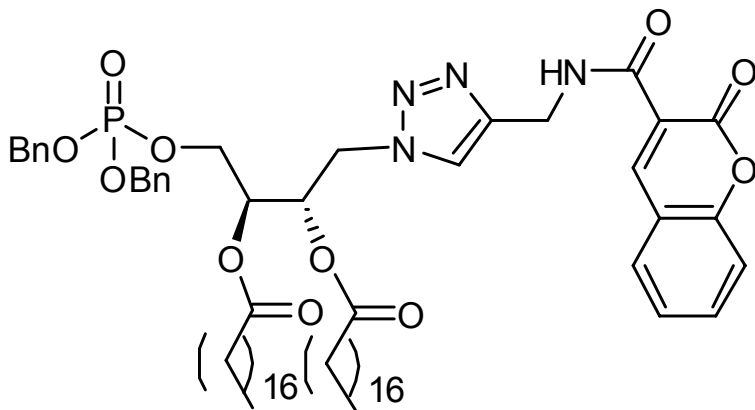
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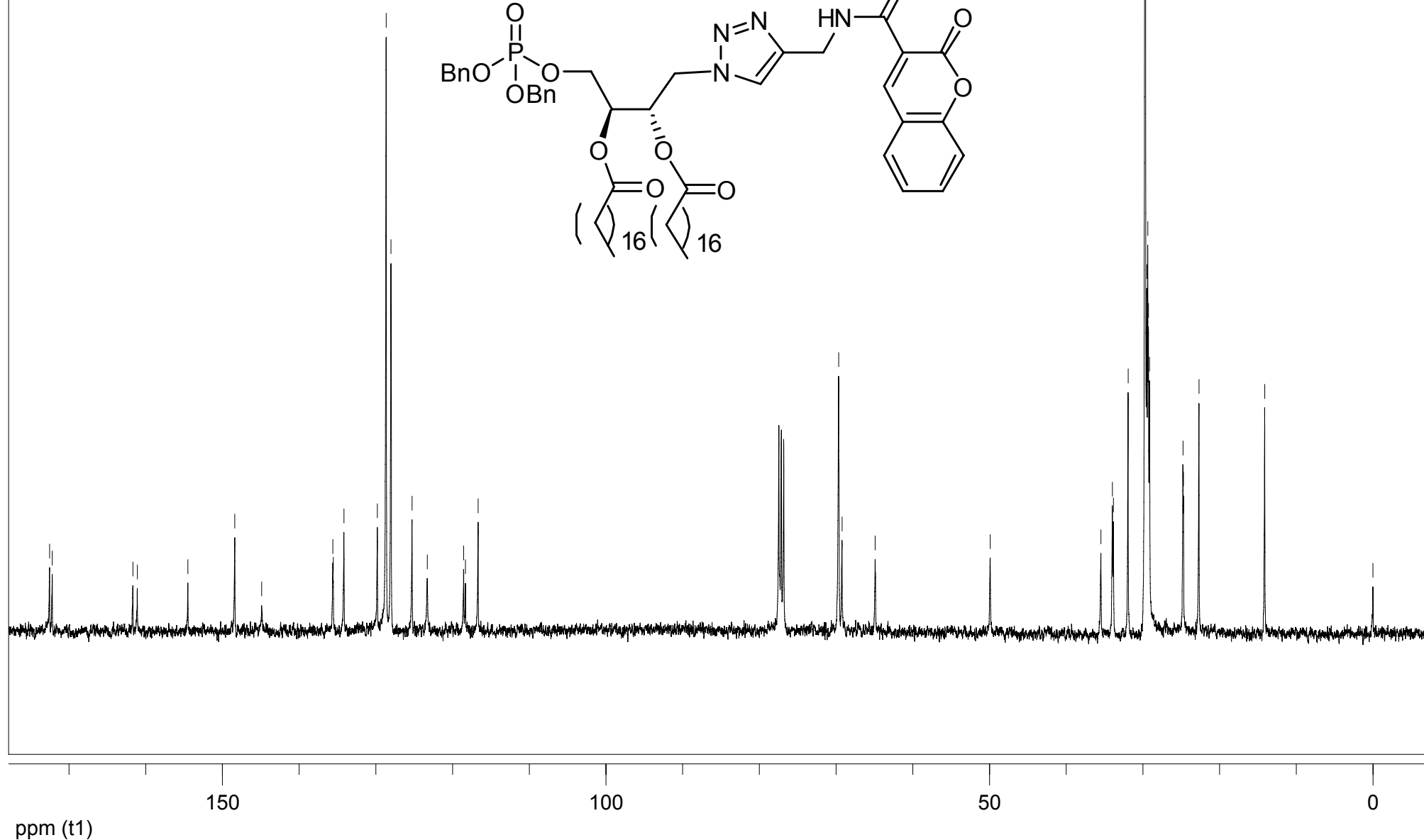
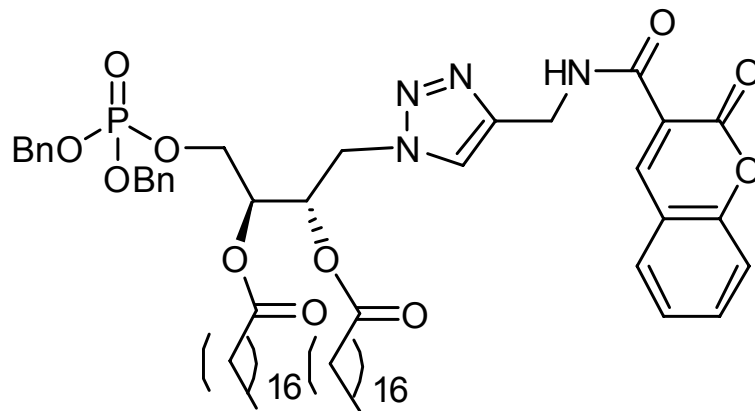
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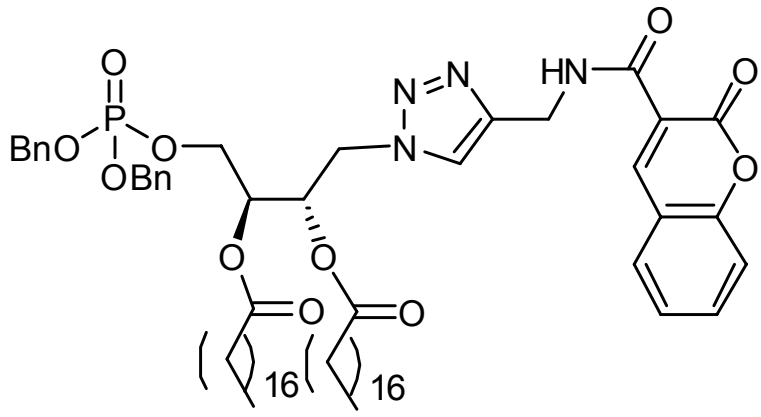
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118.310  
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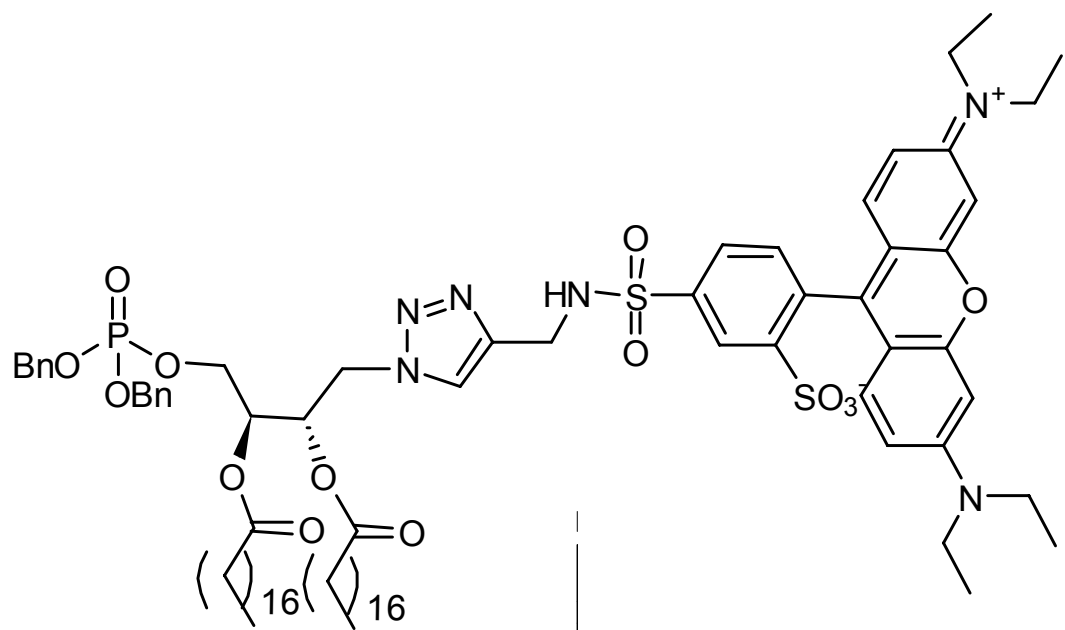
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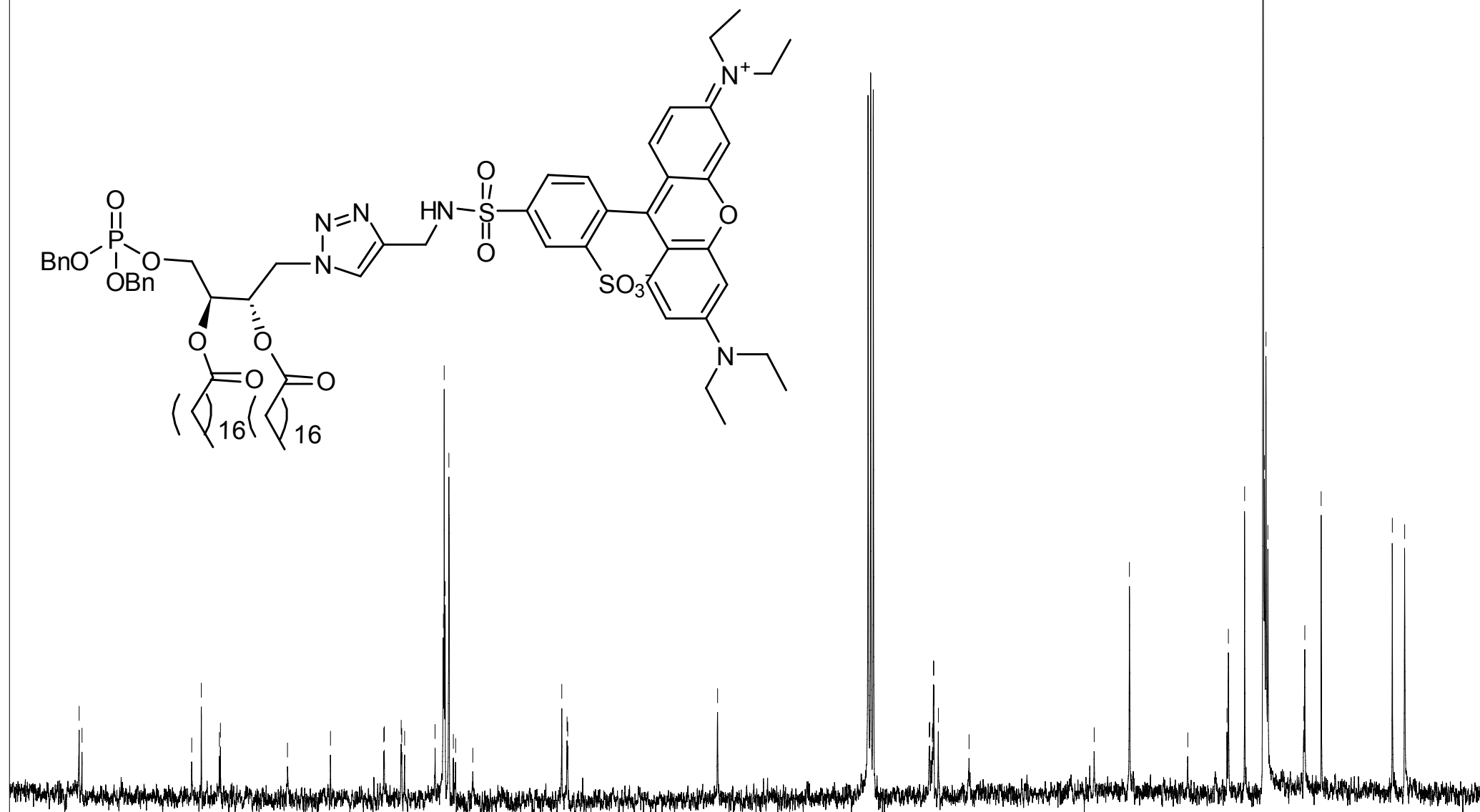
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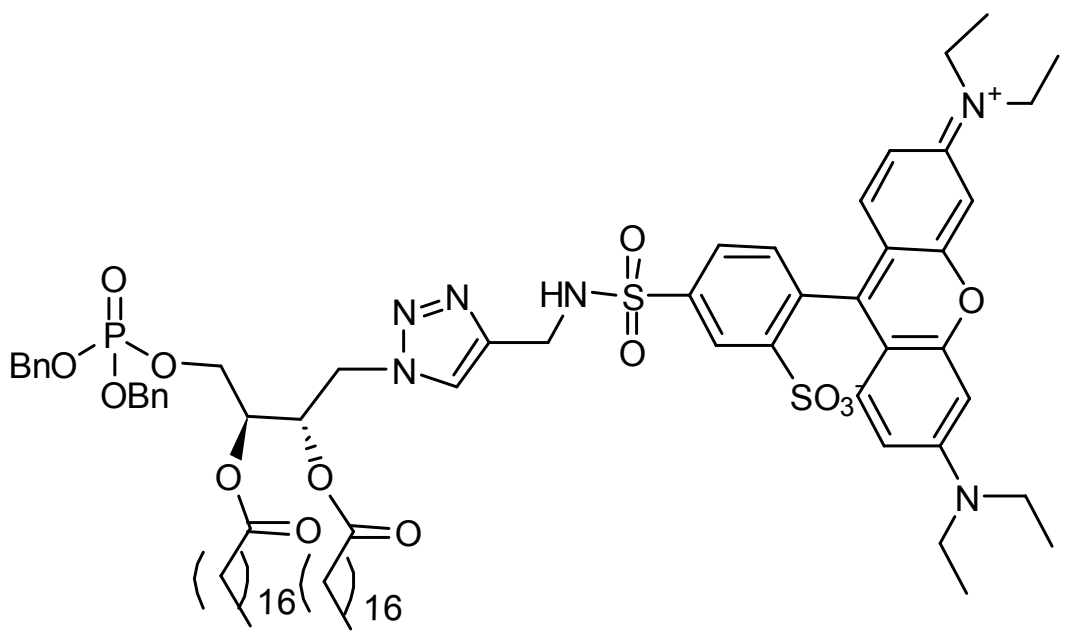




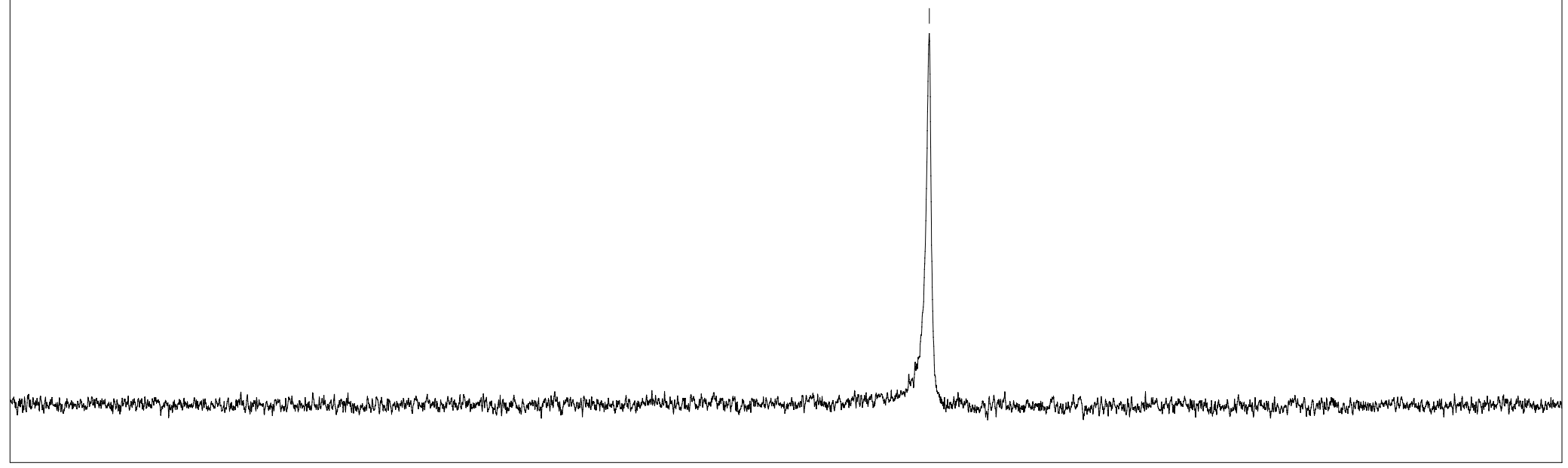


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125.084  
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95.551  
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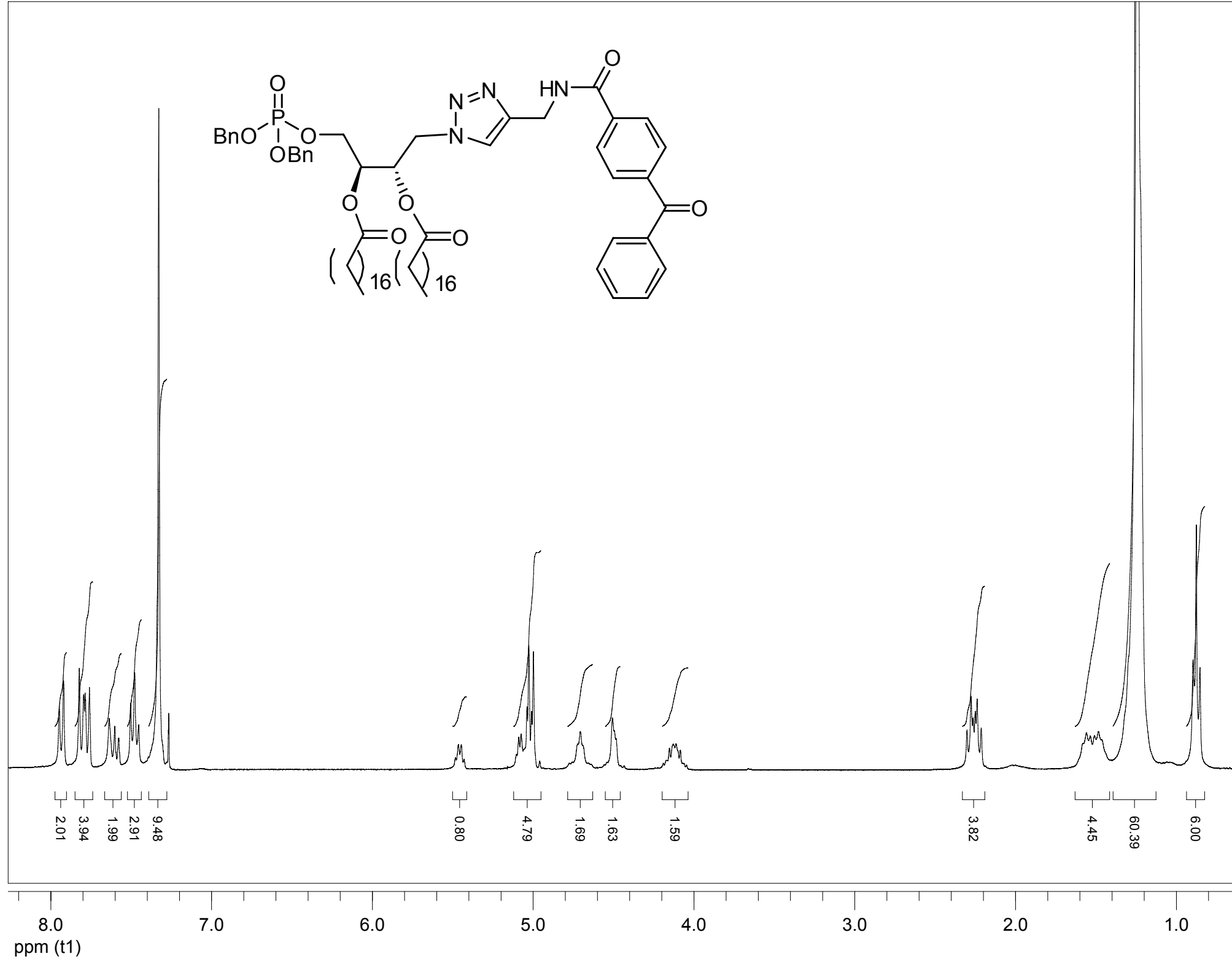
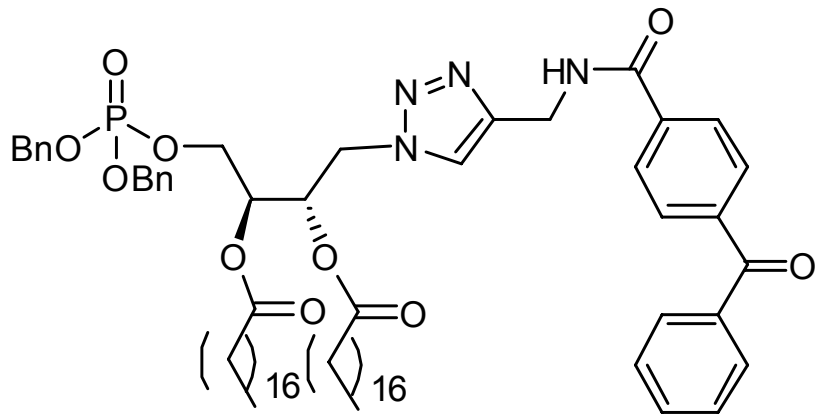




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ppm (t1) 10.0 5.0 0.0 -5.0 S30



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172.208

166.524

140.144

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137.078

135.535

135.472

132.787

130.037

128.648

128.405

128.013

127.971

127.148

123.621

69.691

69.644

69.238

64.873

64.832

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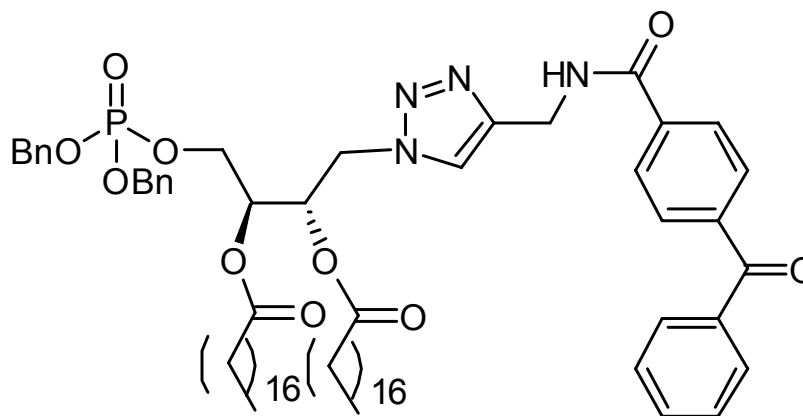
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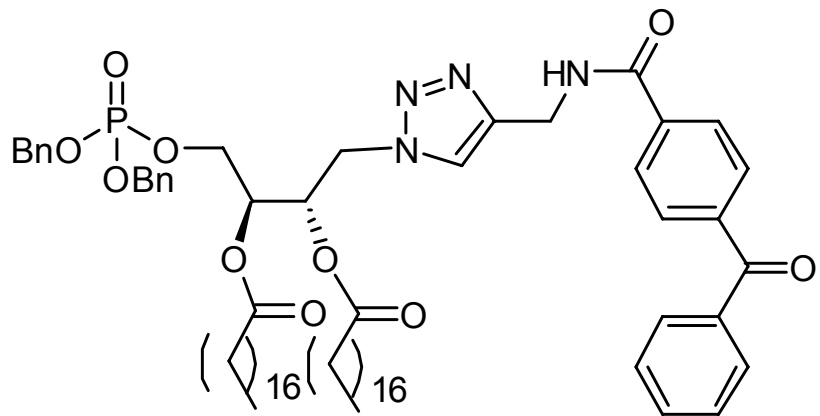
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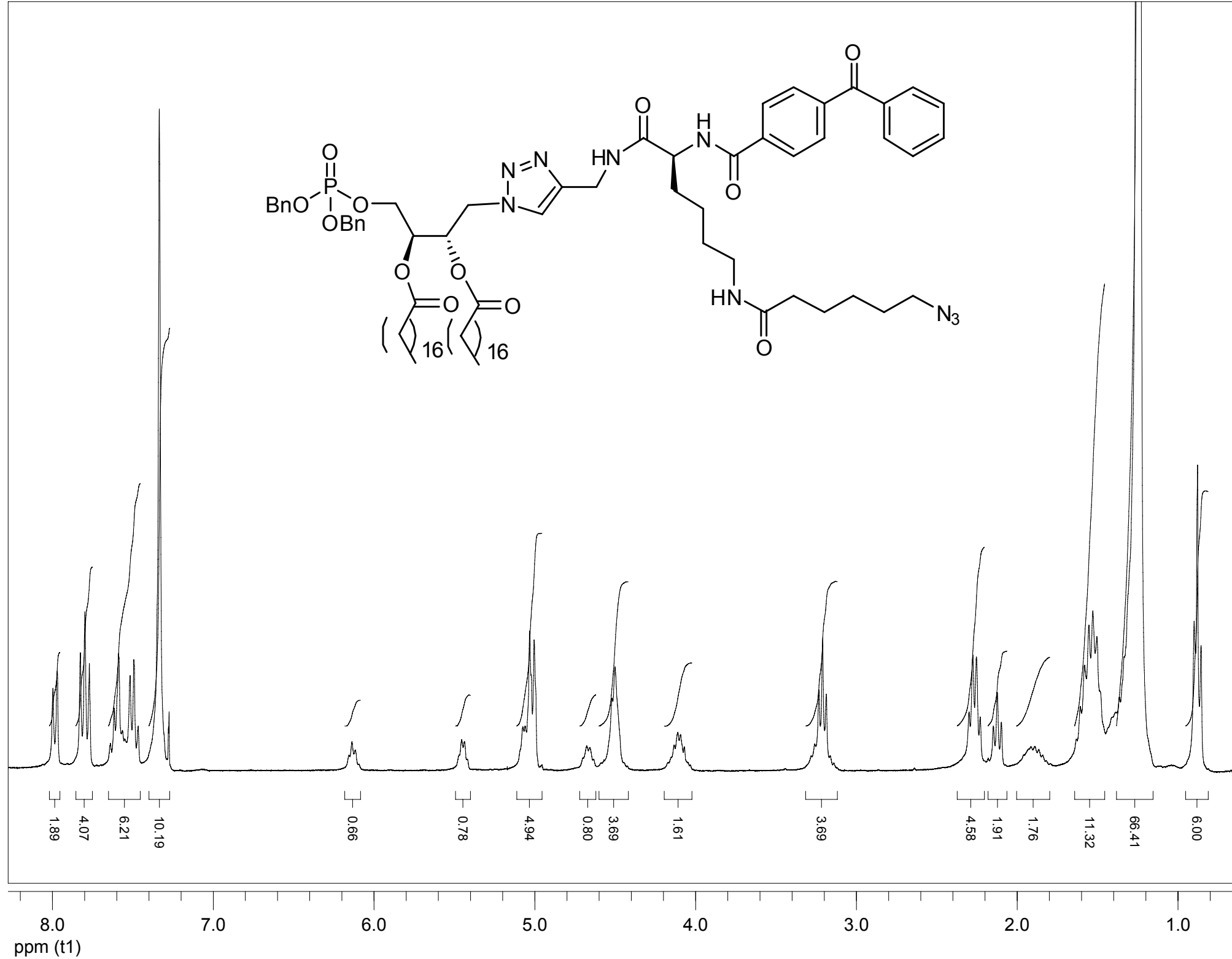
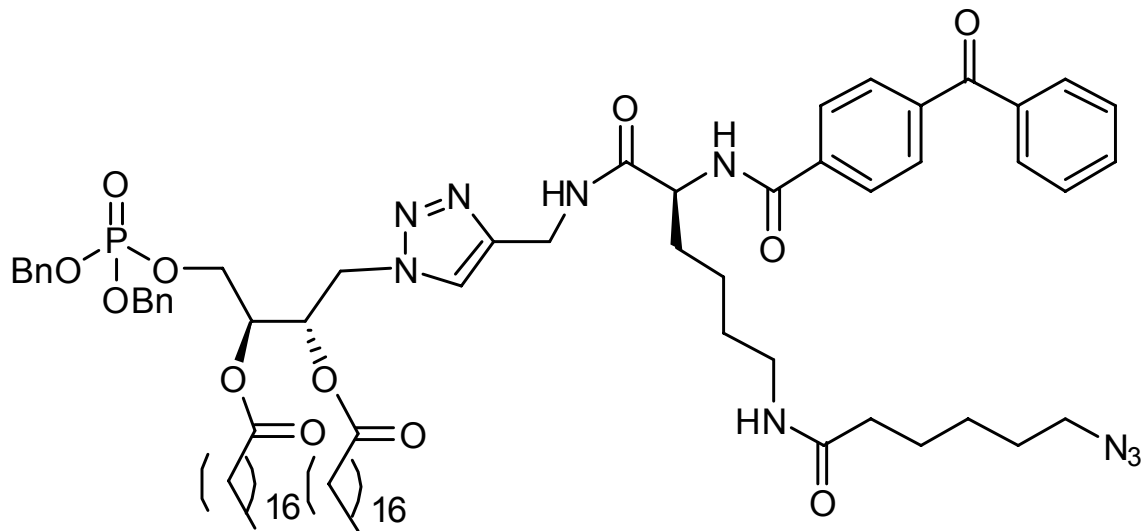
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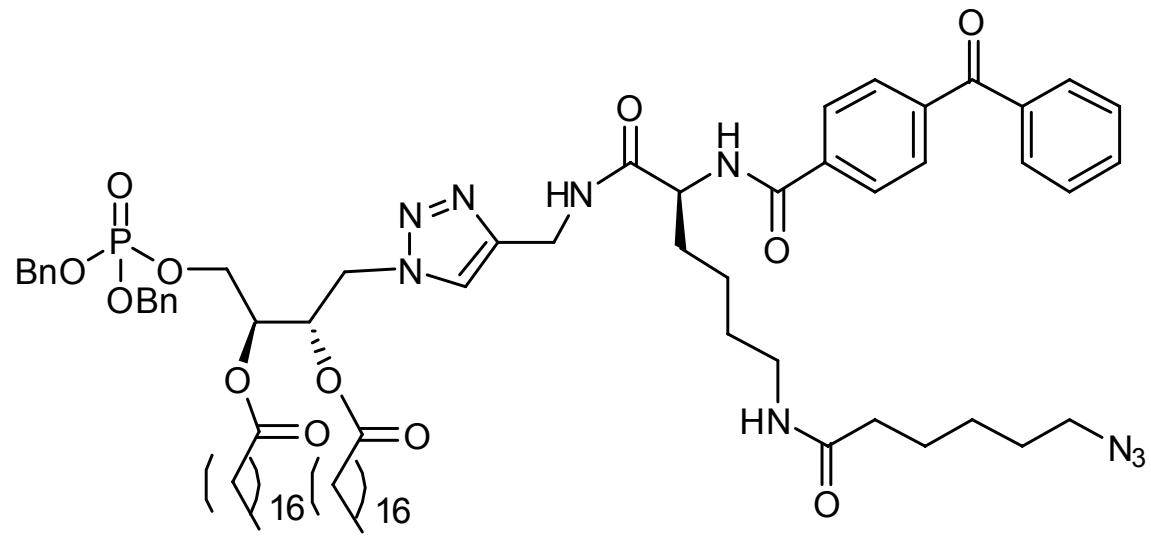
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S33

ppm (t1)



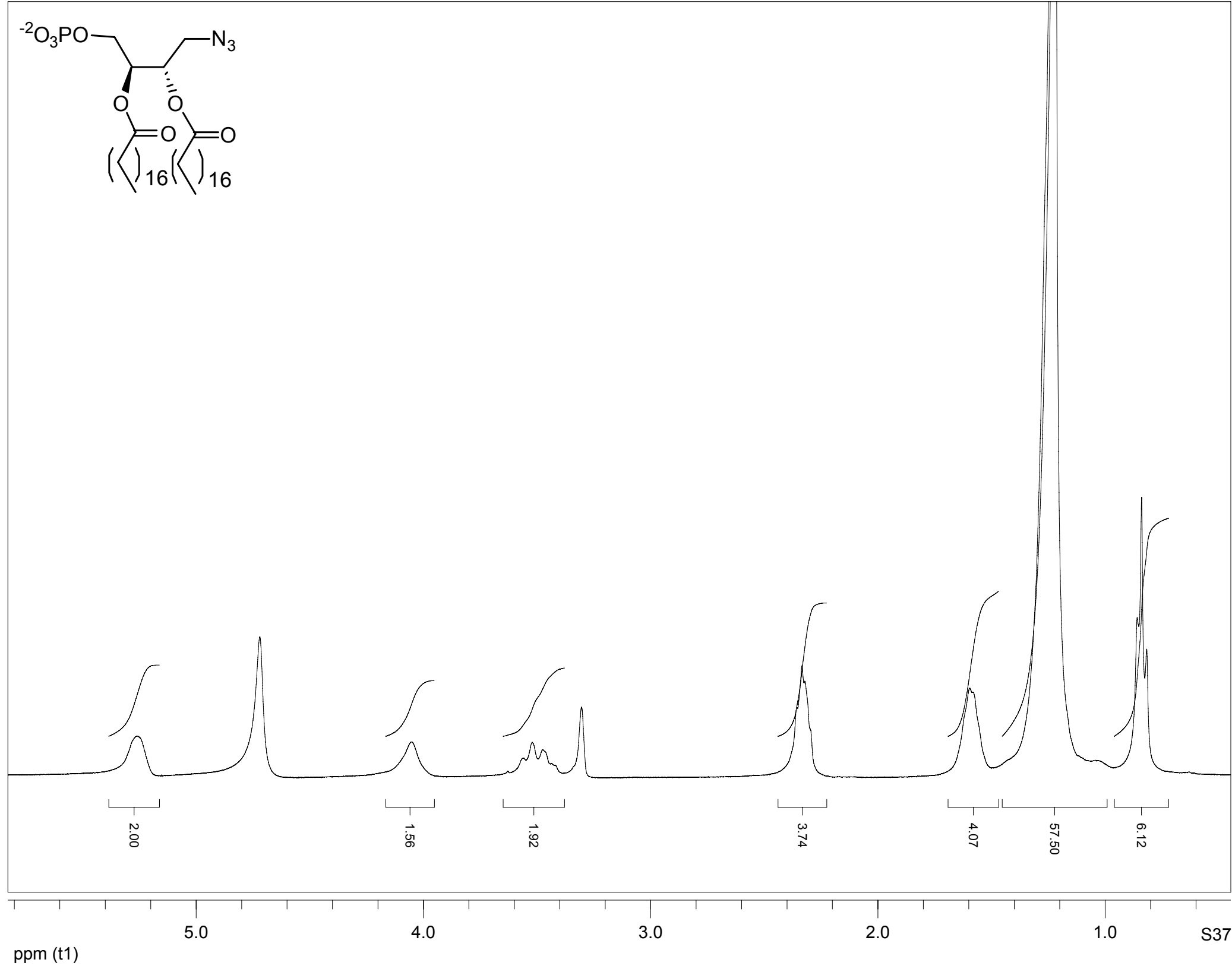
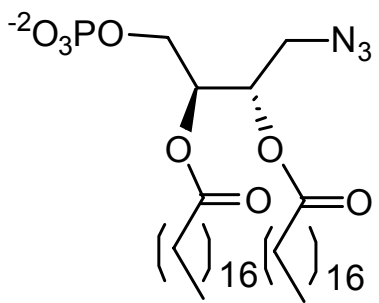




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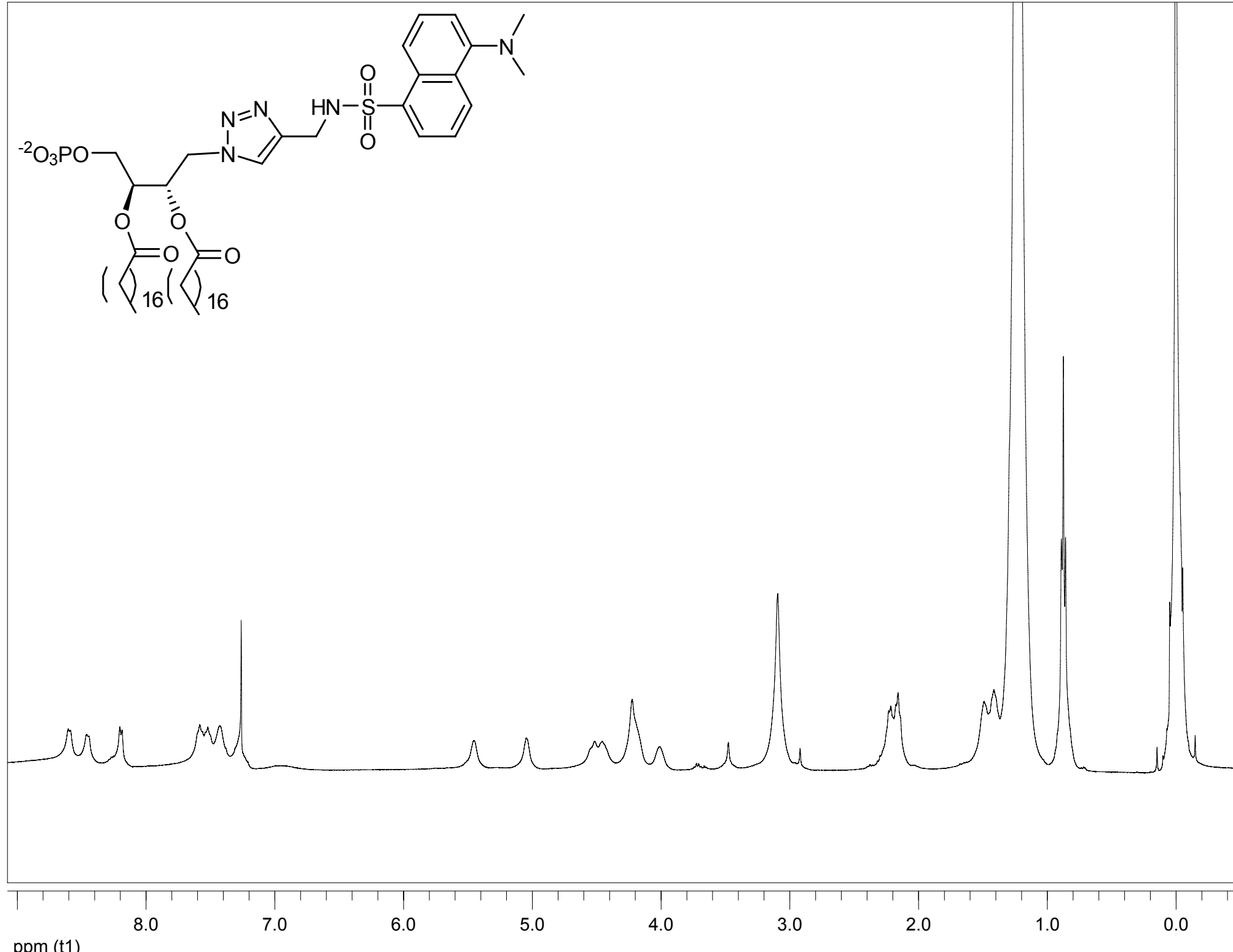
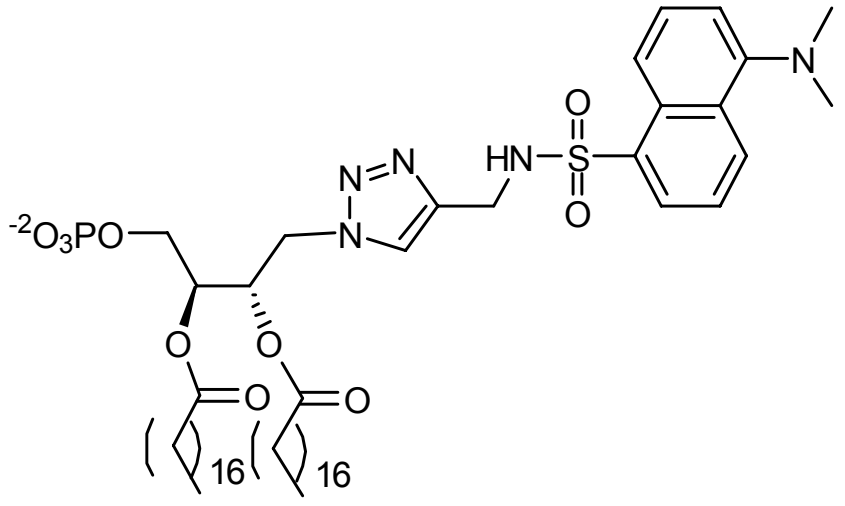


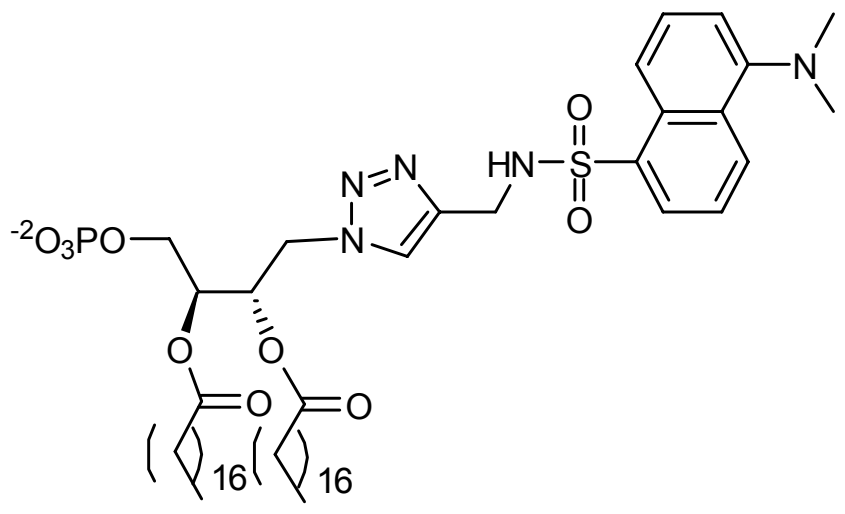




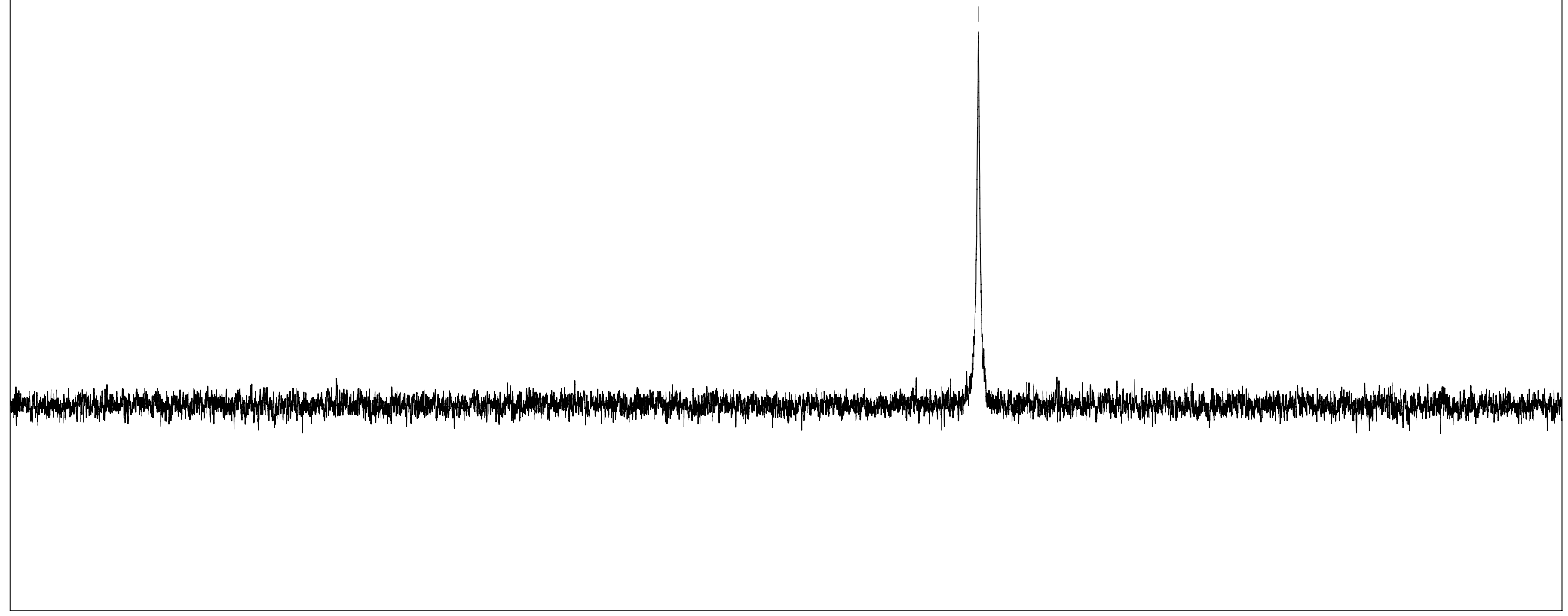




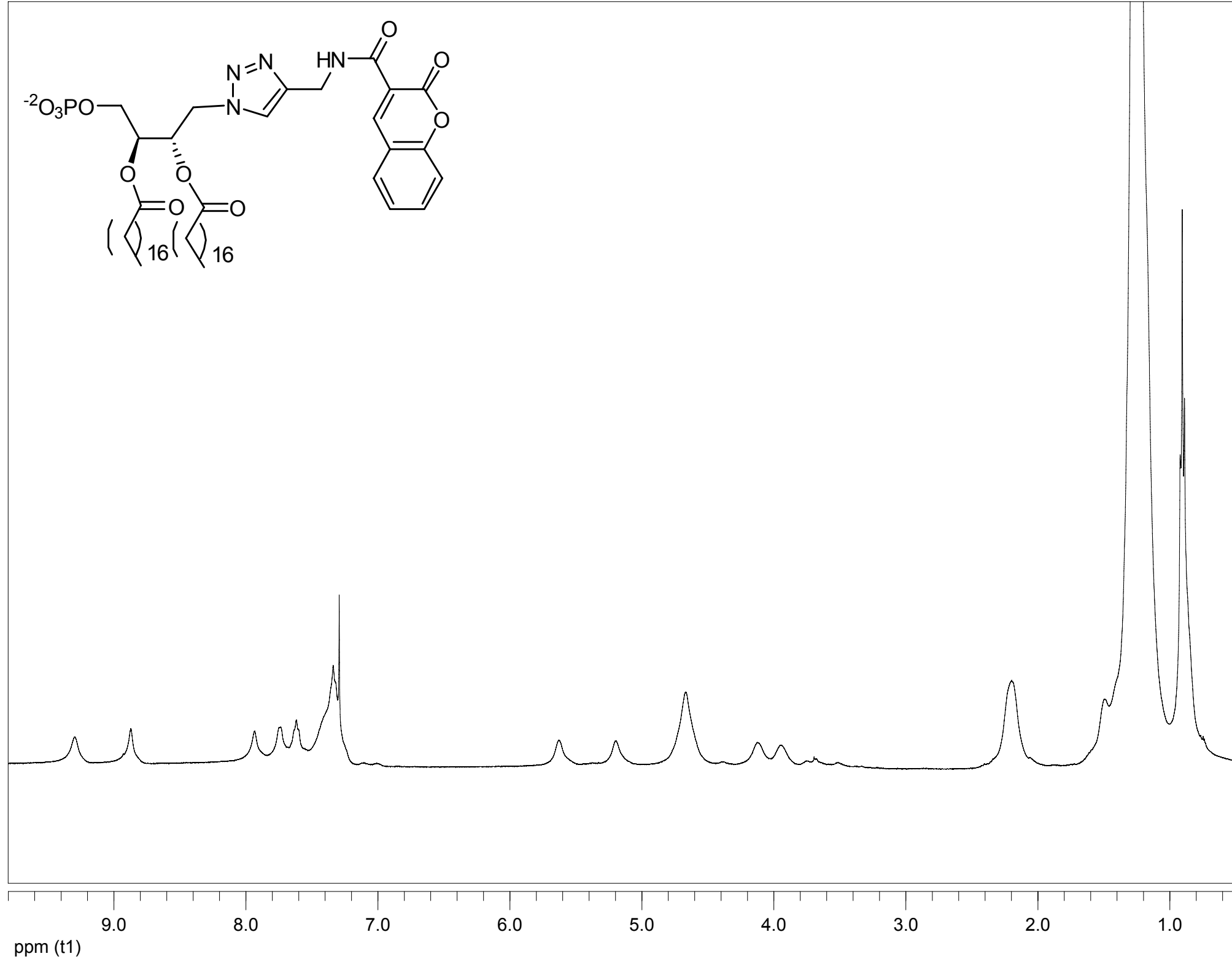
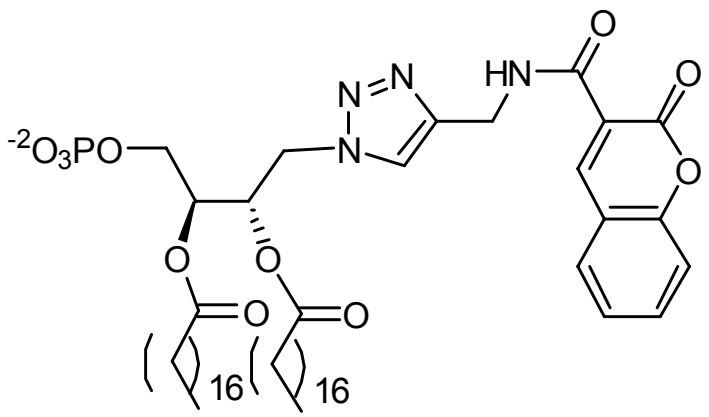


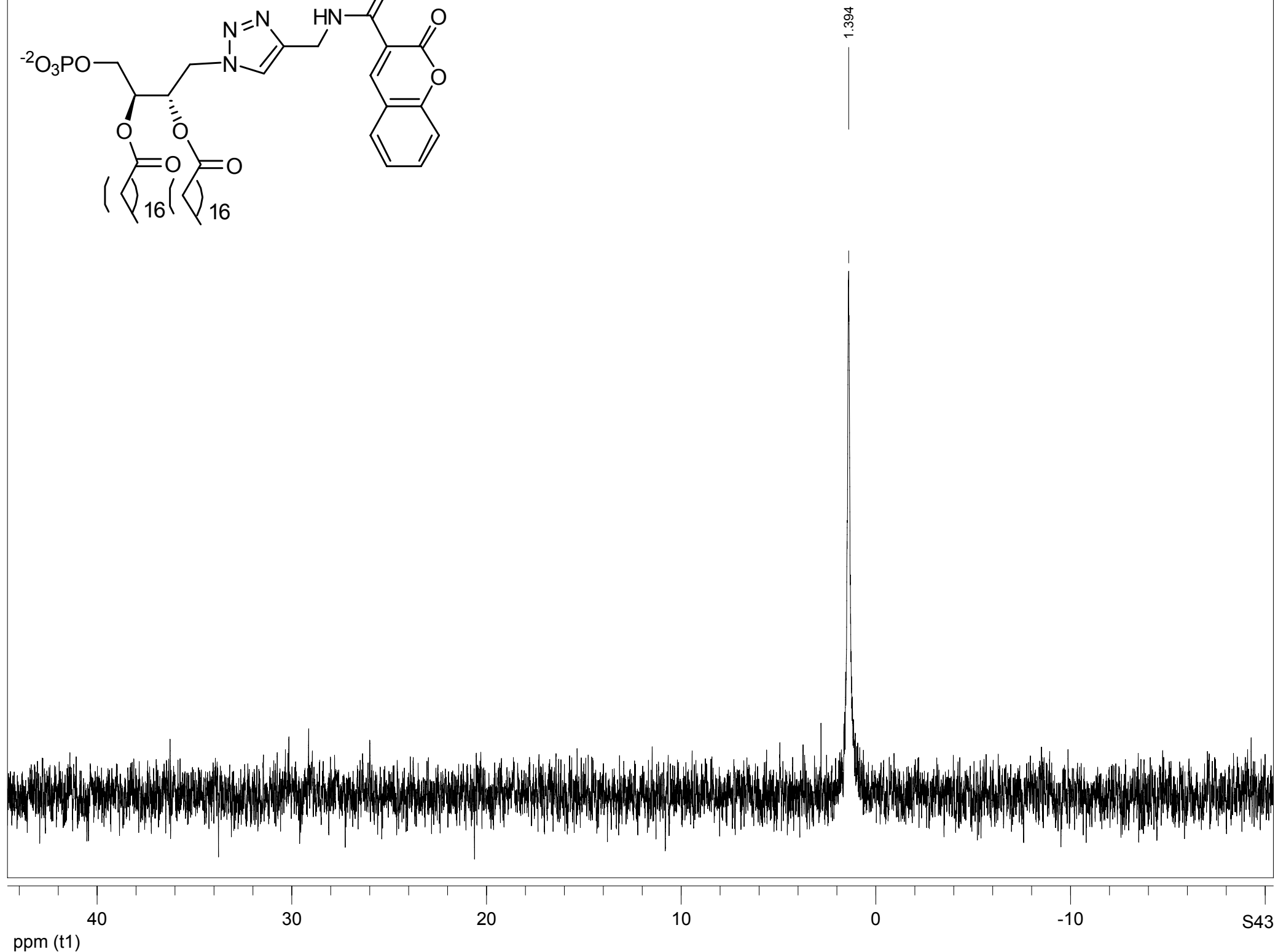
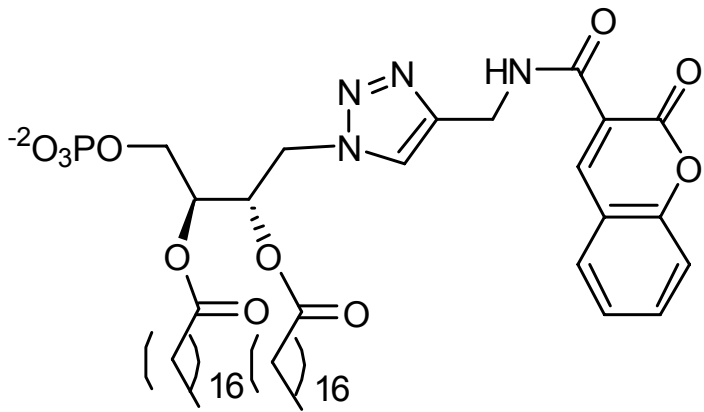


1.094



ppm (t1) 40 30 20 10 0 -10 -20 S41

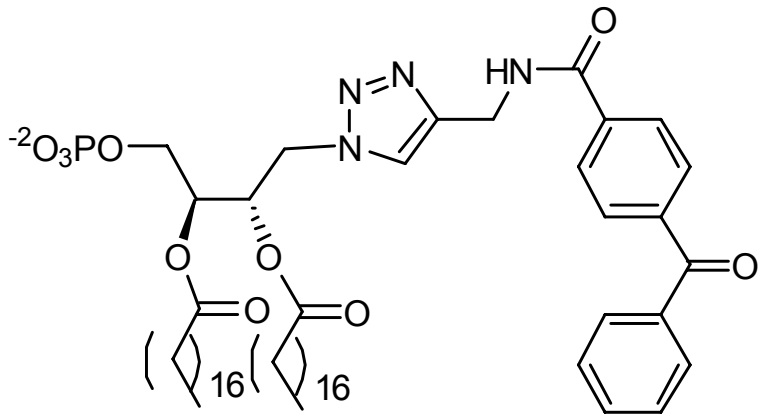




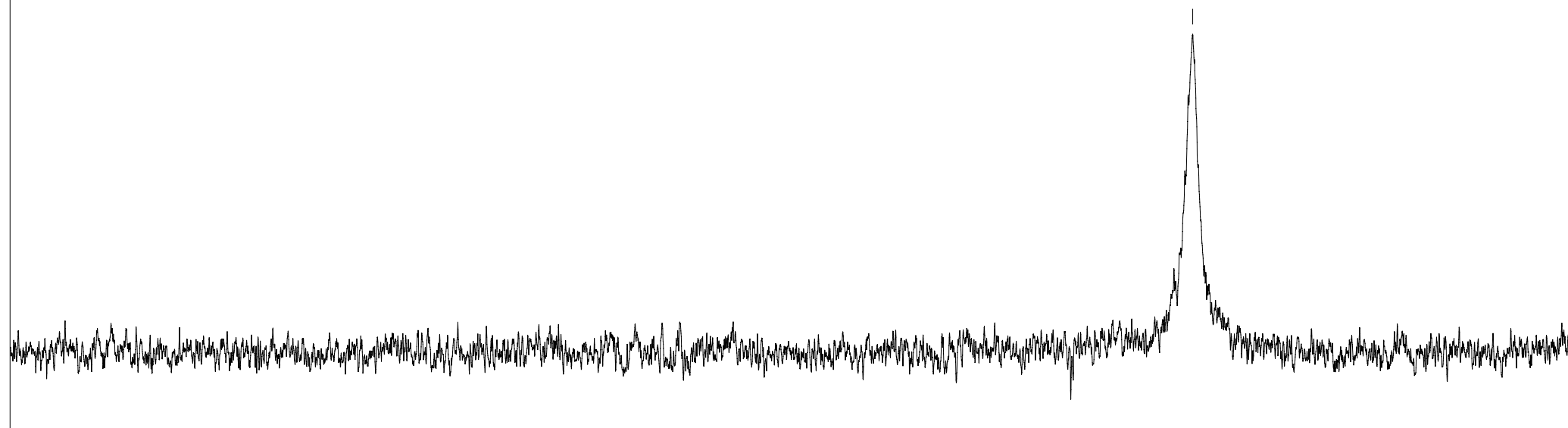
S43



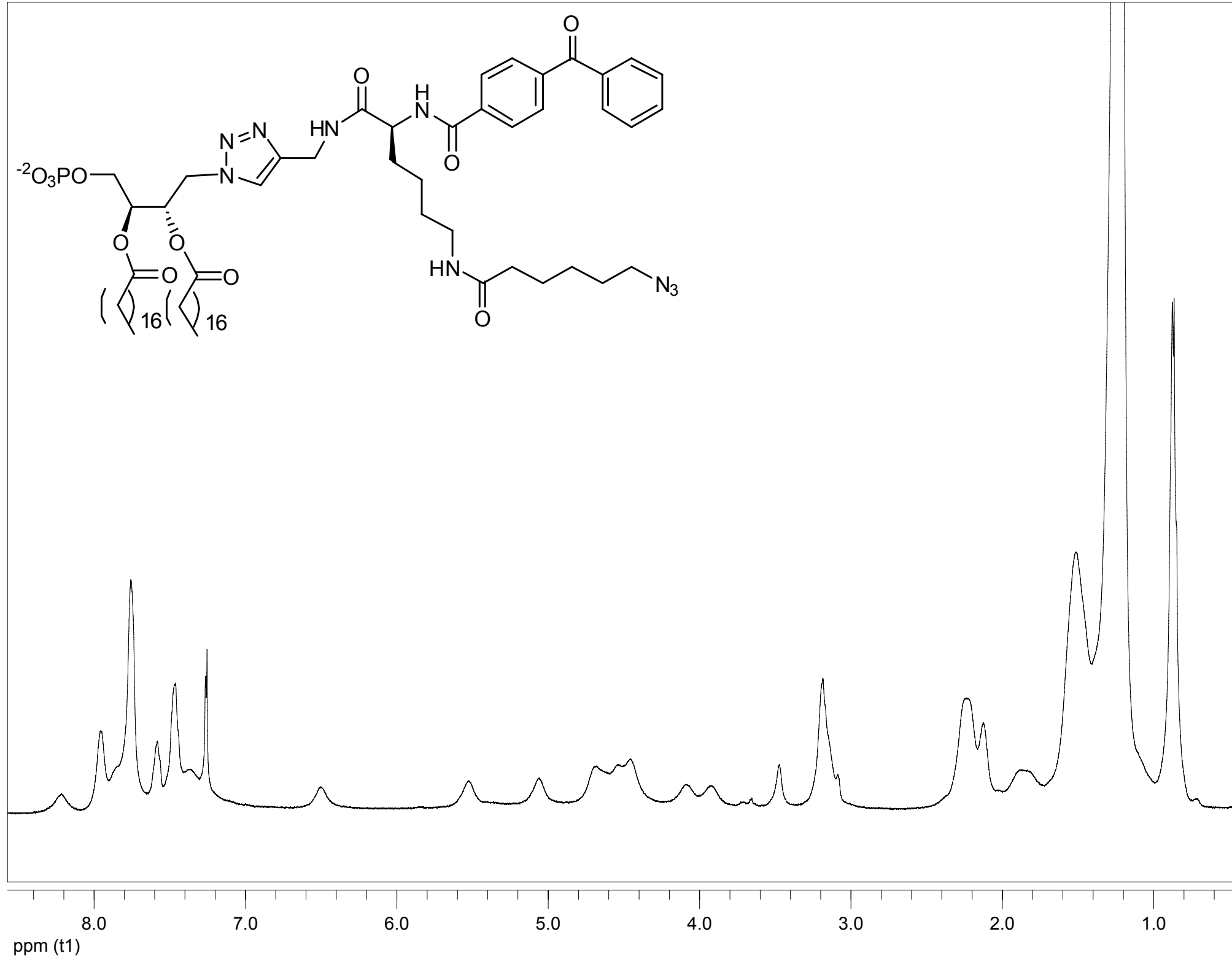
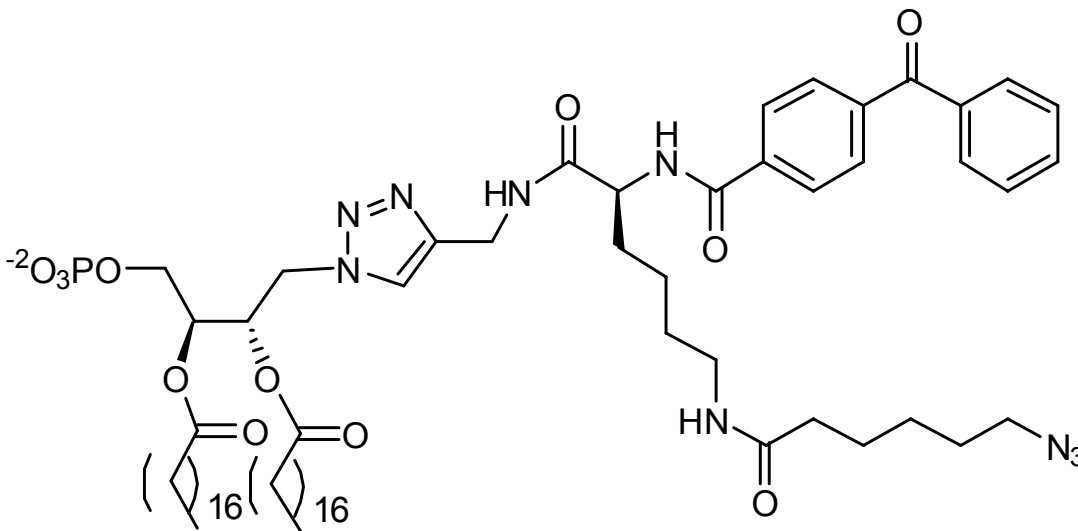




1.250

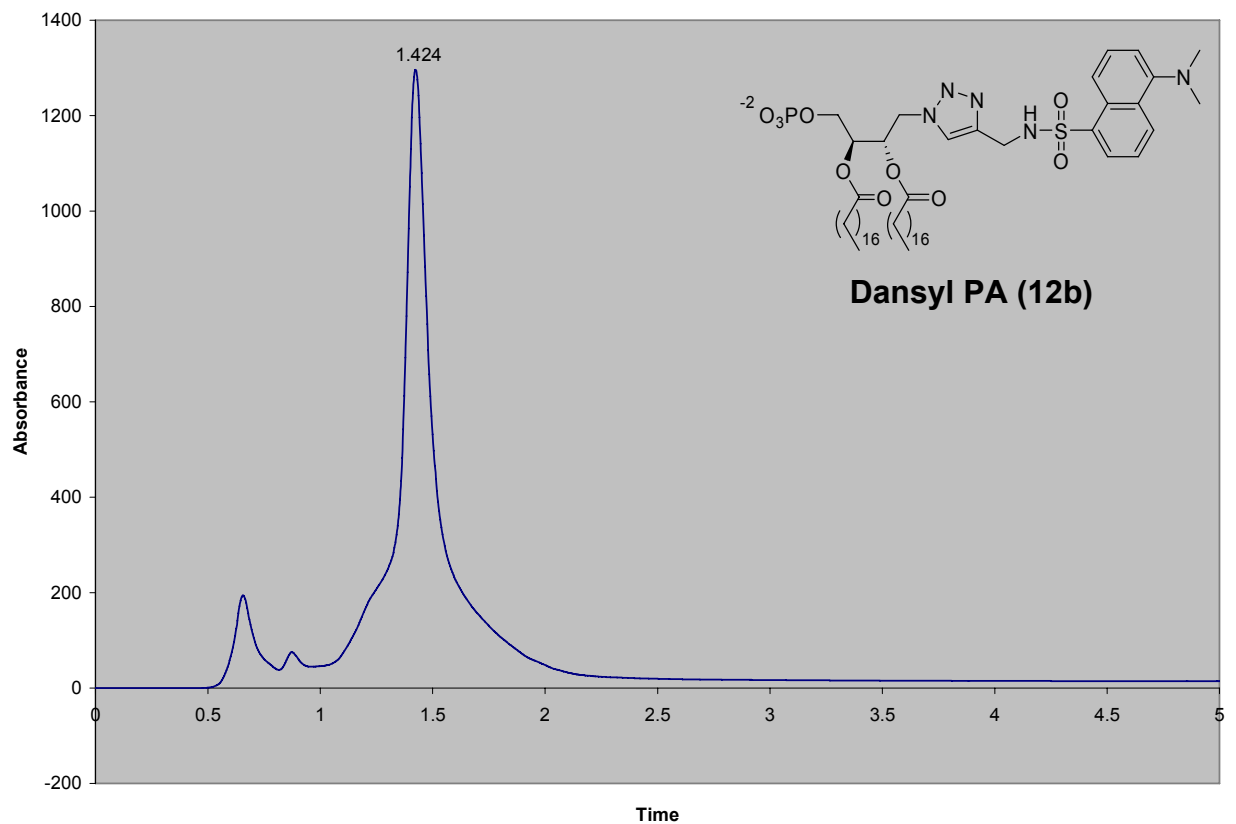
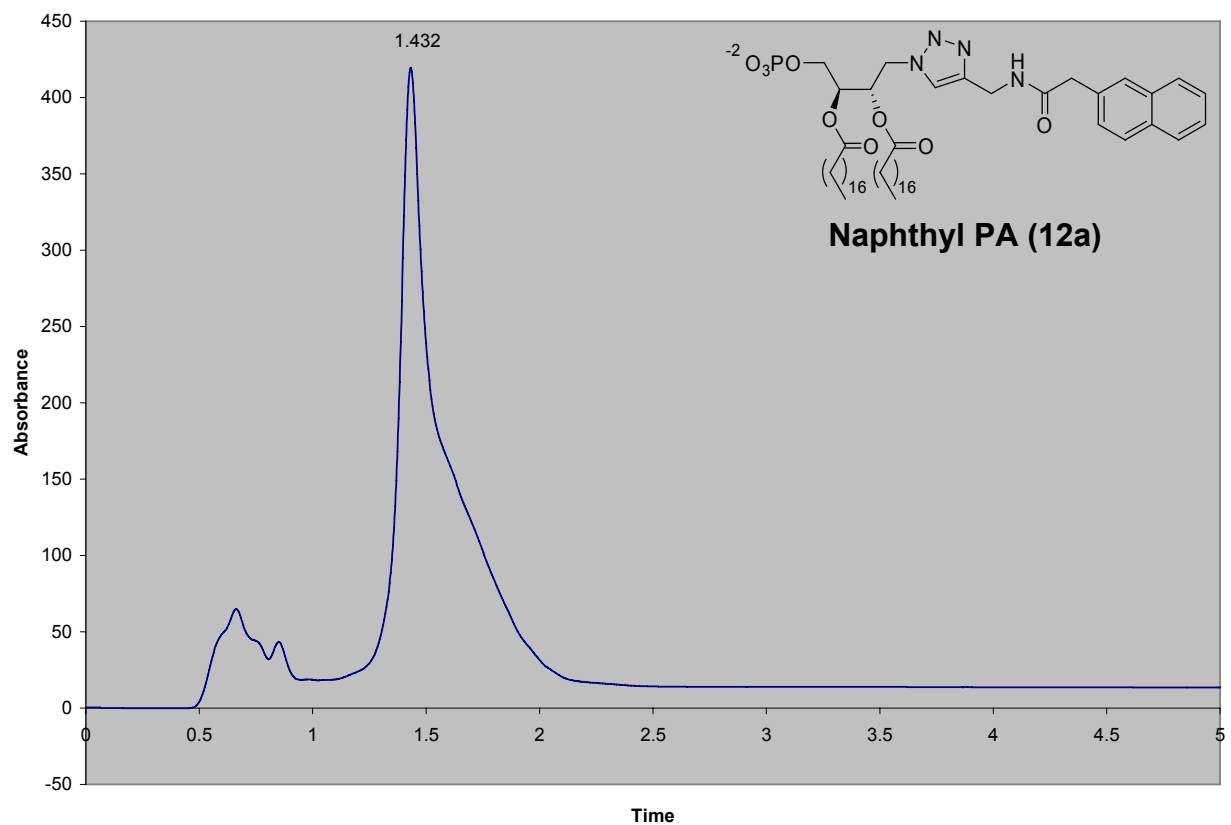


15.0 10.0 5.0 0.0 ppm (t1) S45

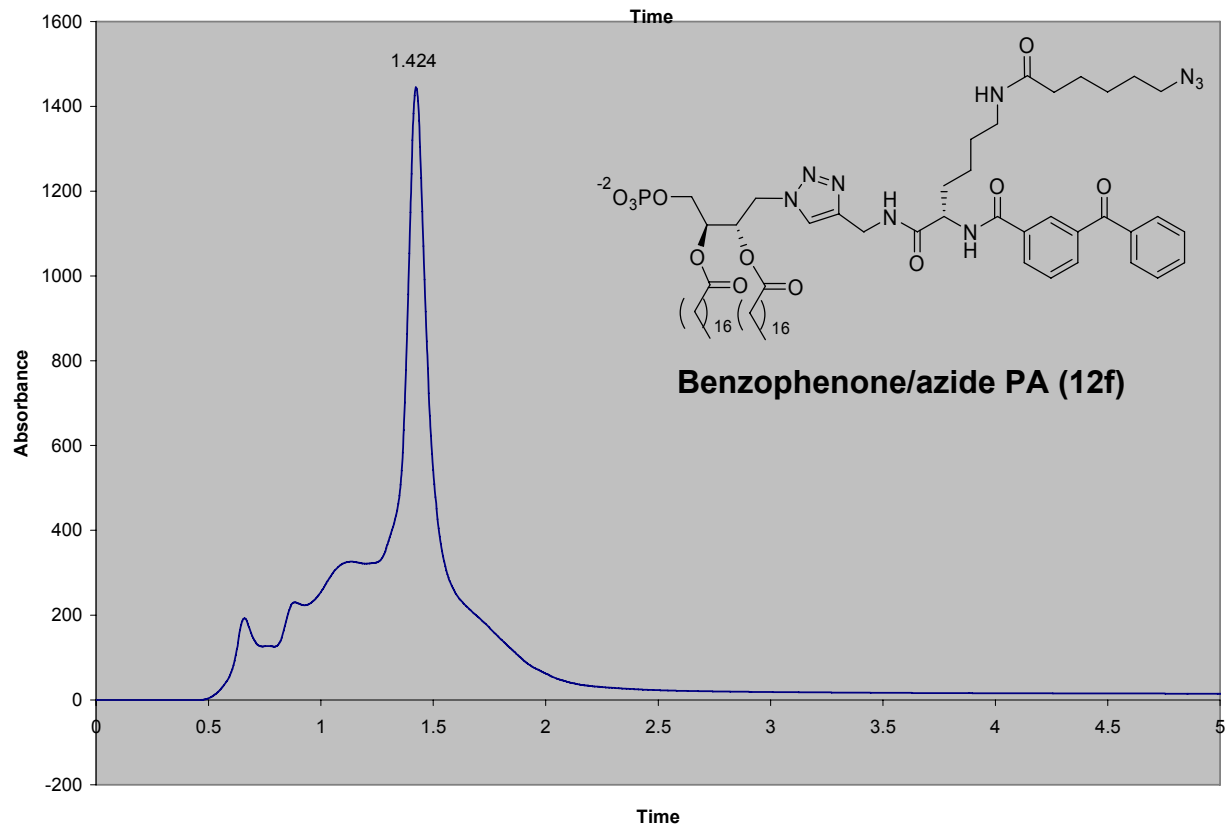
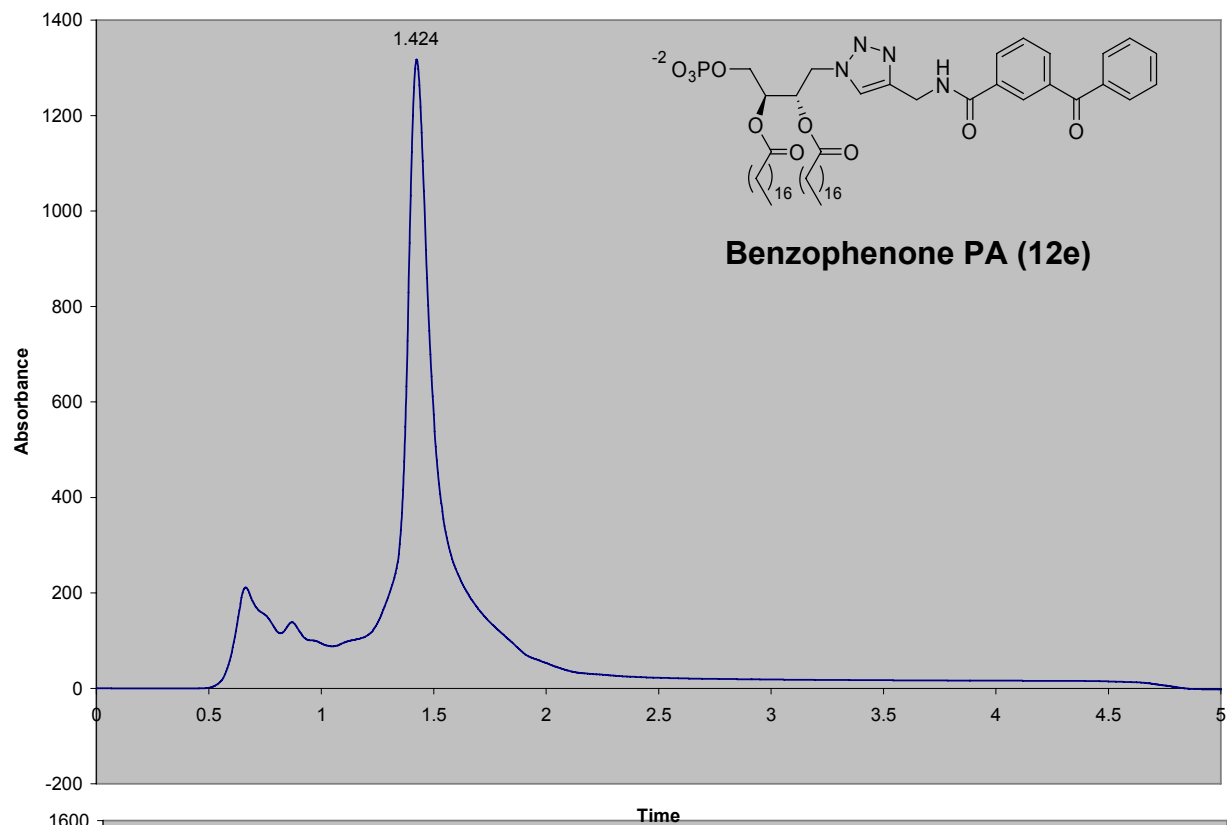




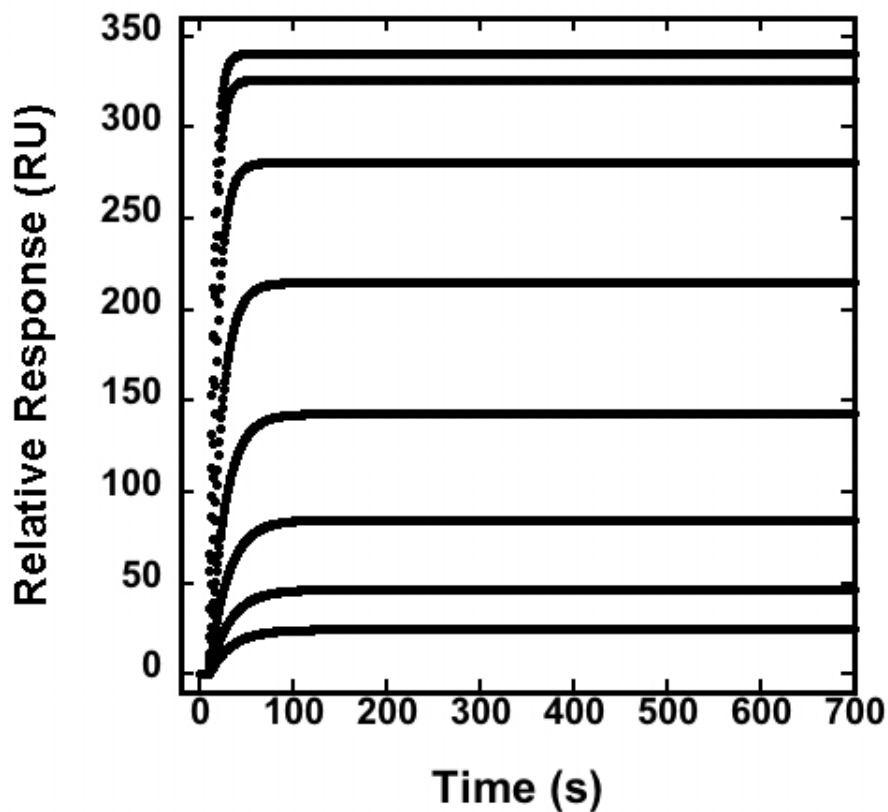
### III. HPLC Traces of Derivatized PA Probes







## Raw SPR spectrograms



**Figure S51. Equilibrium SPR analyses of PKC $\alpha$ -C2.** The PKC $\alpha$ -C2 was injected at 10  $\mu$ l/min at varying concentrations (50, 100, 200, 400, 800, 1500, 3000, and 4000 nM from bottom to top) over the POPC:POPE:Dansyl-PA (40:40:20) surface using a POPC:POPE (50:50) surface as a control. Subtraction of the POPC:POPE (50:50) response for each protein concentration was performed to yield the binding sensorgrams shown.  $R_{eq}$  values were then measured to determine  $K_d$  by a nonlinear least-squares analysis of the binding isotherm (See Figure 3).