## **Supplementary Figure Legends:**

### **Supplementary Figure 1**

Pedigree of the family with THRA mutation, showing their thyroid function tests and genotype.

### **Supplementary Figure 2**

Amplicons encompassing the mutant codon (A263V) in TR $\alpha$ 1 or  $\alpha$ 2 cDNAs, synthesized from PBMC RNA in the patient (P1) or control subject, were generated using a common forward ( $\alpha$ -FOR) and specific ( $\alpha$ 1-REV,  $\alpha$ 2-REV) reverse primers (upper panel). The location of Cac8I restriction sites (vertical line) together with expected size of DNA products following such enzymatic digestion of either WT or A263V mutant TR $\alpha$ 1 or  $\alpha$ 2 cDNAs is shown in the middle panel. Gel electrophoresis indicates three fragments (223bp, 174bp, 102bp) from Cac8I digestion of WT TR $\alpha$ 1 cDNA (Control), whereas loss of one Cac8I site in the A263V mutant cDNA from the patient (P1), yields a novel band (276bp) together with lower intensity fragments (174, 102bp) bands, with the 223bp band (asterisked) common to both cDNA species being unchanged. Likewise RFLP analysis of  $\alpha$ 2 cDNAs yields 174 and 102bp bands derived from WT alleles in the control subject, whereas a high intensity 276bp band

in P1 reflects loss of a Cac8I site derived from the mutant allele. Additional bands (asterisked 153bp, 122bp) present in both subjects, confirm that the digested cDNAs are  $\alpha$ 2-derived. Electropherograms (lower panel) show sequencing of  $\alpha$ 1 and  $\alpha$ 2 cDNAs and heterozygosity for the single nucleotide substitution (T/C) in P1 or homozygosity for the wild type nucleotide (C) in the control subject.

## **Supplementary Figure 3**

Radiolabelled T3 binding to in vitro translated WT TR $\alpha$ 1, A263V mutant TR $\alpha$ 1, WT TR $\beta$ 1 and A317V mutant TR $\beta$ 1 proteins, in the absence (open bars) or presence (black bars) of excess (100nM) unlabelled hormone, is shown. The inset shows comparable parallel translation of <sup>35</sup>S-methionine labelled wild type and mutant proteins.

## **Supplementary Figure 4**

Protein-protein interaction assays in JEG-3 cells cotransfected with a reporter gene (UASTKLUC), vectors expressing GAL4-coactivator (TRAP220) or GAL4-corepressor (NCoR) fusion proteins and constructs expressing a heterologous activation domain linked to TRs (VP16-WT, VP16-A263V). Western blotting (inset) confirmed comparable VP16-receptor fusion expression.

#### **Supplementary Table**

Thyroid function tests, reverse T3 and thyroglobulin levels, together with *THRB* genotype, in family with A317V TR $\beta$  mutation. Abbreviation: ND: not done.

### **Supplementary Figure 5**

JEG-3 cells were transfected with empty, WT or A317V TR $\beta$  expression vectors together with a thyroid hormone responsive reporter gene, assaying T3-dependent activation.

### **Supplementary Figure 6**

Dominant negative inhibition by A317V mutant TR $\beta$  was tested in JEG-3 cells cotransfected with reporter gene and equal combinations of expression vectors as indicated. \*\*p<0.01.

## **Supplementary Figure 7**

Crystallographic modelling of the TRβ1 ligand binding domain (LBD) bound to T3 (blue), highlighting the normal amino acid (alanine 317, green), with substitution of the larger value residue (red) predicting steric hindrance to T3 binding.

# **Supplementary Figure 8**

Clinical photographs of skin tags from other patients with *THRA* mutations. Affected father (panel A) and proband (panel B) with F397fs406X mutation and child (panel C) with E403X mutation in TRα1.

# **Supplementary Figure 9**

Dominant negative inhibition of WT TR $\beta$ 1 by A263V mutant TR $\alpha$ 1 was tested in cells cotransfected with reporter gene and equal combinations of expression vectors as indicated. \* p<0.05; \*\*\*p<0.001,