

**Evolution of a Strategy for the Enantioselective
Total Synthesis of (+)-Psiguadial B**

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Linglin Wu, and Sarah E. Reisman*

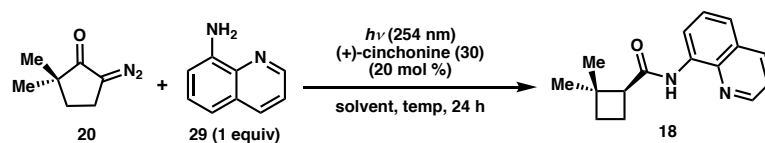
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Supporting Information

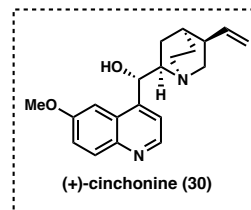
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Table S1: Optimization of the Wolff rearrangement for amide 18

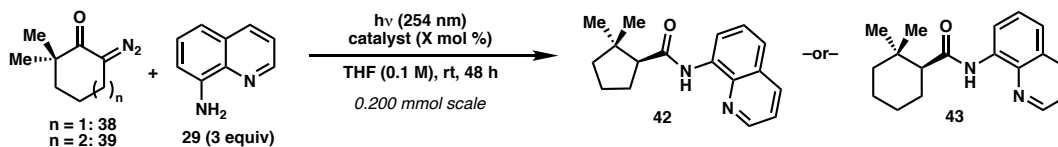


entry	solvent	temp °C	vessel ^a	atm	% yield ^b	% ee ^c
1	THF	25	quartz	N ₂	63	79
2	DMF	25	quartz	N ₂	76	50
3	CH ₂ Cl ₂	25	quartz	N ₂	62	47
4	toluene	25	quartz	N ₂	65	59
5	dioxane	25	quartz	N ₂	69	79
6	MeCN	25	quartz	N ₂	43	74
7	THF	0	quartz	N ₂	28	79
8	THF	25	quartz	air	18	75
9	THF	25	quartz	CO	33	69
10	THF	25	vykor	N ₂	29	79
11	THF	25	pyrex	N ₂	20	75

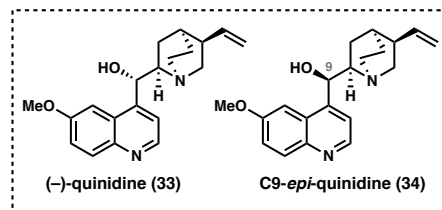


^a See reference 12 in text of article for discussion of photodecarbonylation.
^b Determined by ¹H NMR via integration relative to an added internal standard.
^c Determined by SFC using a chiral stationary phase.

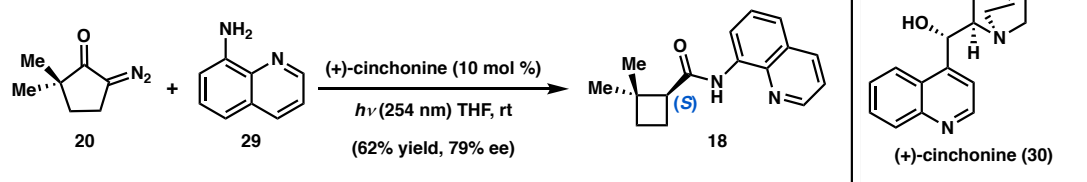
Table S2: Catalyst loading screen for amides 38 and 39



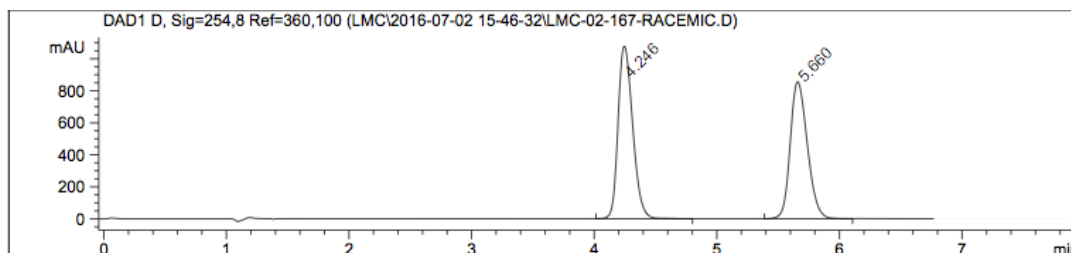
entry	diazo.	catalyst (mol %)	product	% isolated yield	% ee ^a
1	38	34 (50)	42	77	71
2	38	34 (30)	42	81	69
3	38	34 (20)	42	76	67
4	38	34 (10)	42	81	63
5	39	33 (50)	43	59	71
6	39	33 (30)	43	71	66
7	39	33 (20)	43	61	62
8	39	33 (10)	43	61	55



^a Determined by SFC using a chiral stationary phase.

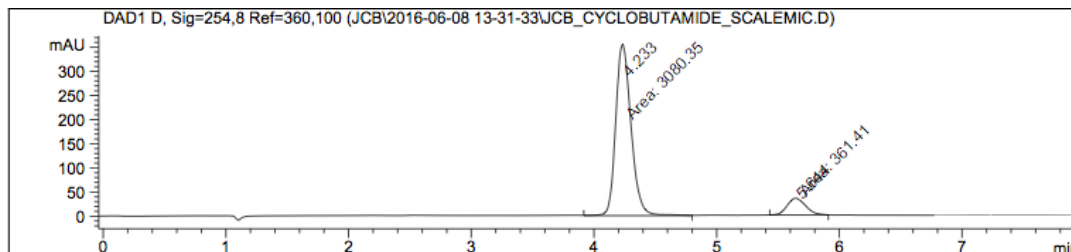


SFC data for racemic 18:



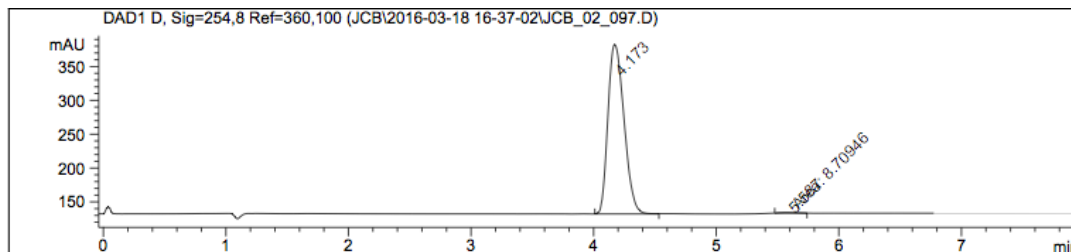
Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	4.246	BB	0.1286	8934.03320	1078.74268	51.8271
2	5.660	BB	0.1532	8304.11426	855.84570	48.1729

Enantioenriched 18 isolated directly from reaction:

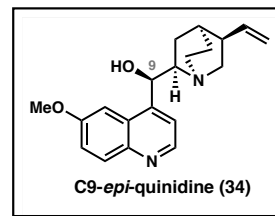
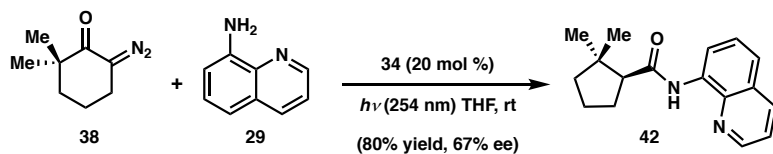


Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	4.233	MM	0.1444	3080.35034	355.58780	89.4993
2	5.644	MM	0.1739	361.41000	34.64573	10.5007

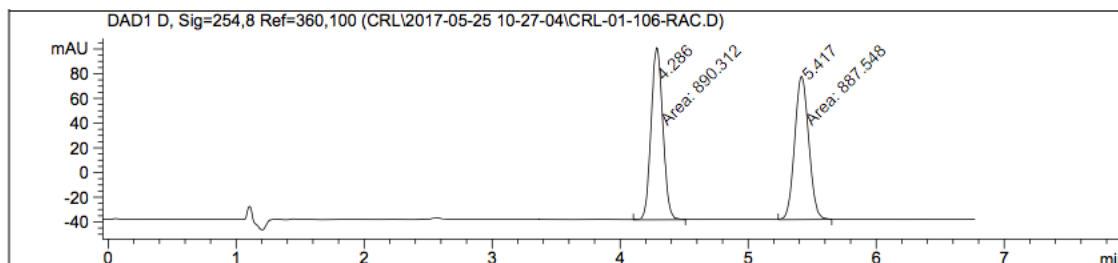
Enantiopure 18 after a single recrystallization:



Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	4.173	BB	0.1464	2285.16455	250.58836	99.6203
2	5.587	MM	0.1518	8.70946	9.55969e-1	0.3797

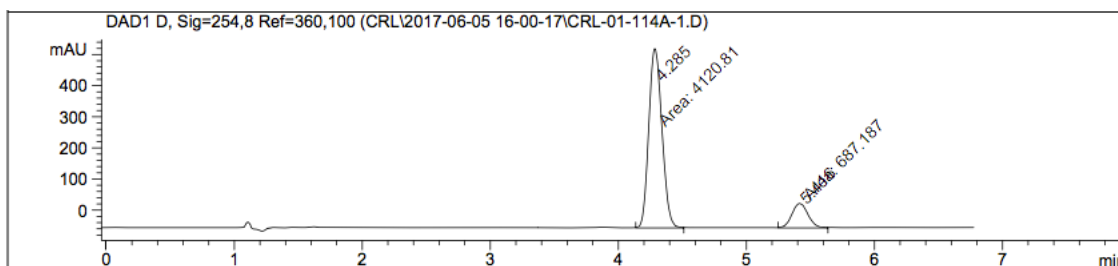


SFC data for racemic 42:



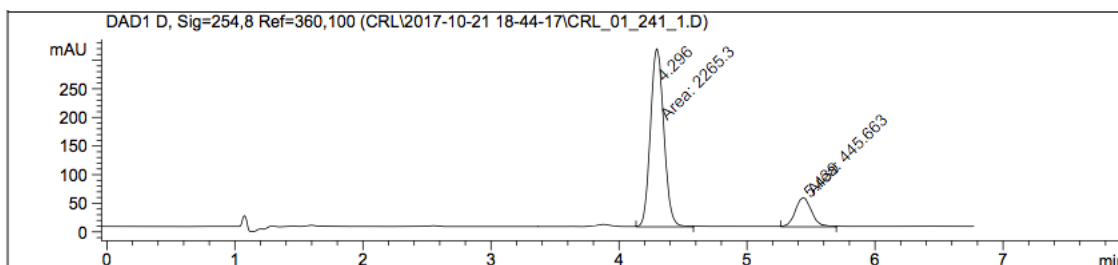
Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Area %
1	4.286	MM	0.1063	890.31165	50.0777
2	5.417	MM	0.1274	887.54810	49.9223

Enantioenriched 42:

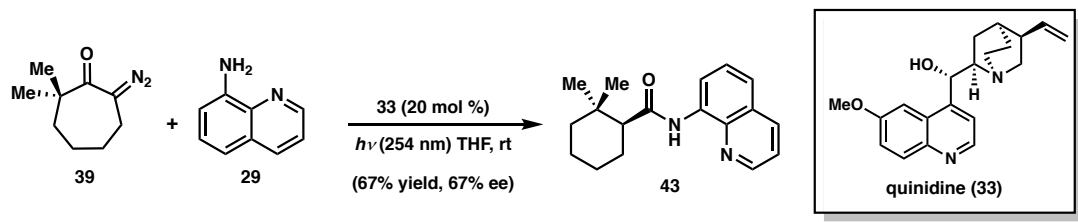


Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Area %
1	4.285	MM	0.1189	4120.80518	85.7074
2	5.416	MM	0.1449	687.18732	14.2926

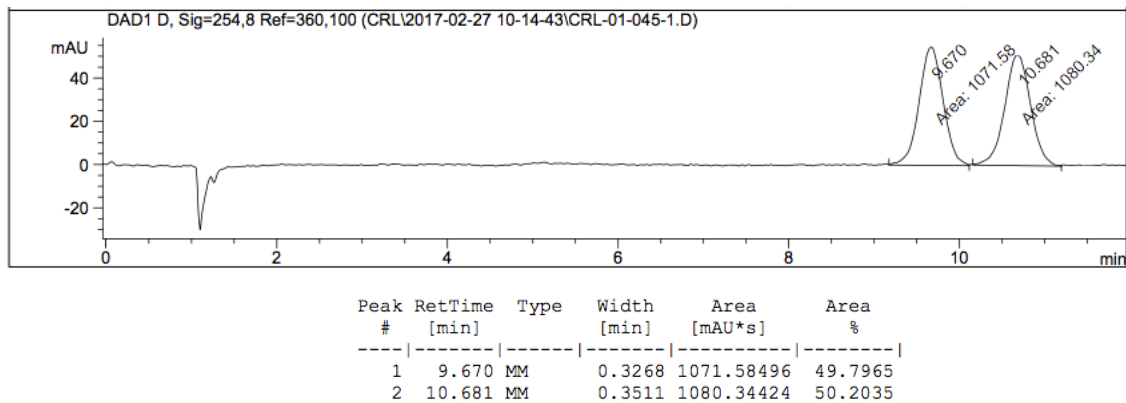
Enantioenriched 42 using 20 mol % catalyst loading:



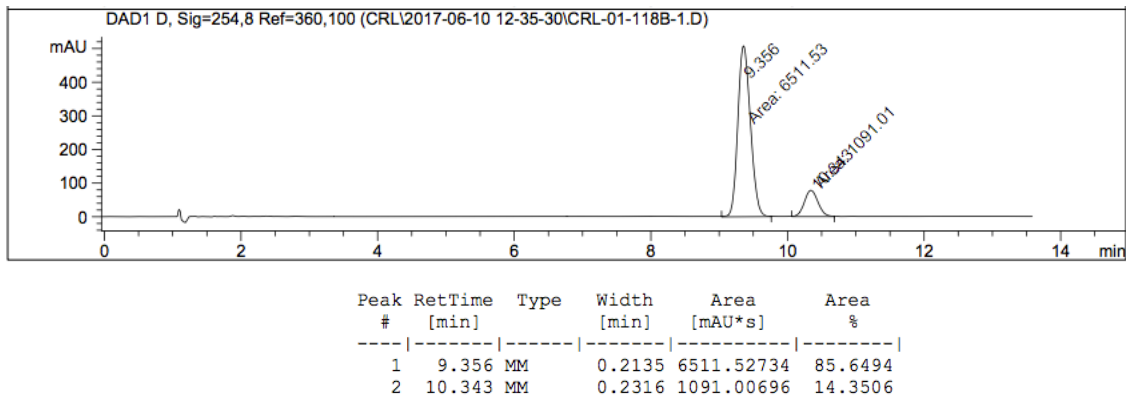
Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Area %
1	4.296	MM	0.1212	2265.30225	83.5607
2	5.439	MM	0.1461	445.66309	16.4393



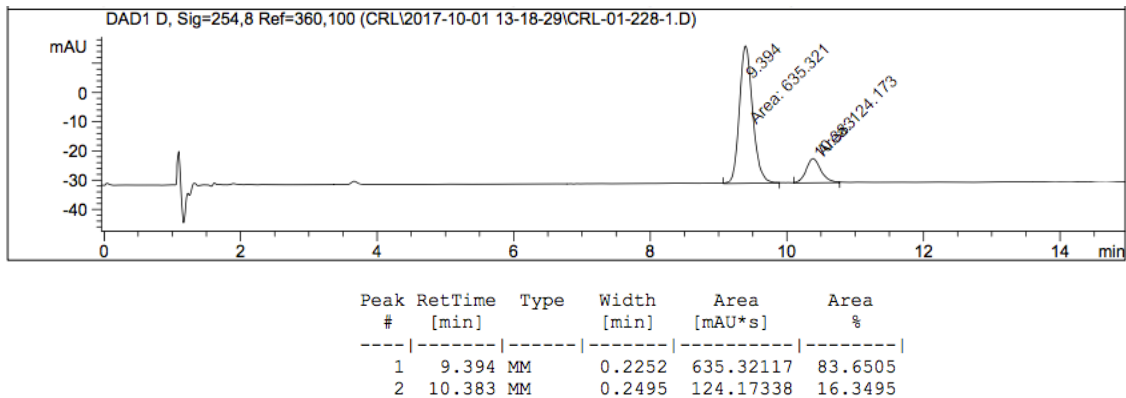
SFC data for racemic 43 (note – 12 minute run):

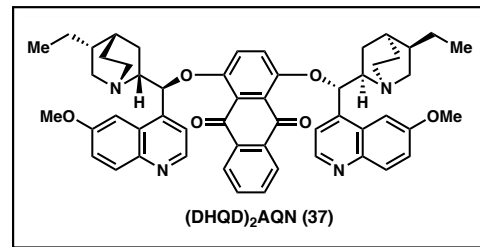
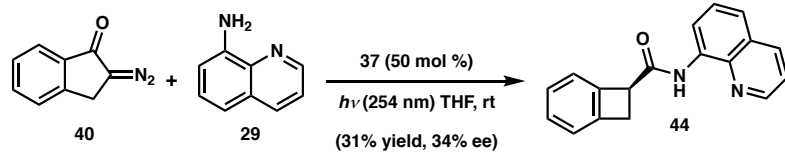


Enantioenriched 43:

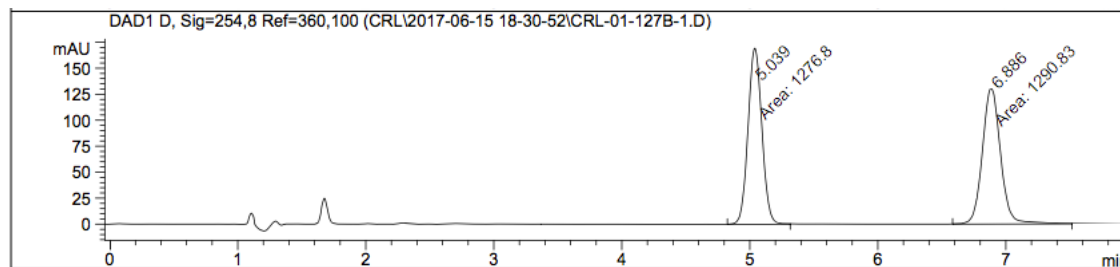


Enantioenriched 43 using 20 mol % catalyst loading:



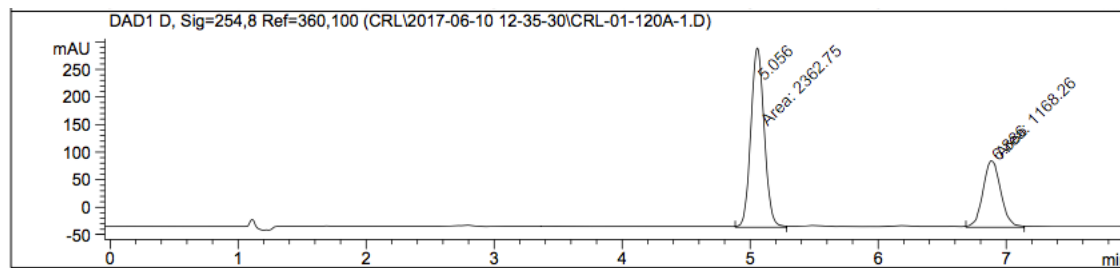


SFC data for racemic 44:

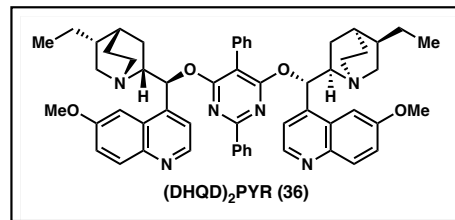
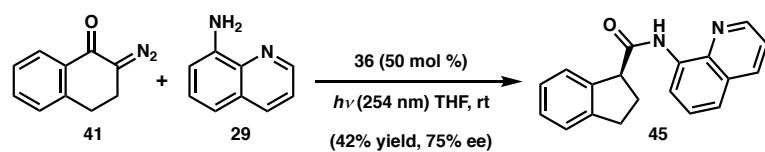


Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Area %
1	5.039	MM	0.1254	1276.80469	49.7268
2	6.886	MM	0.1641	1290.83447	50.2732

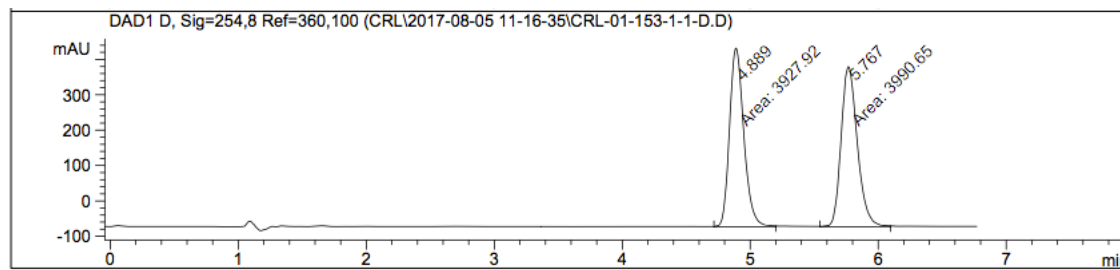
Enantioenriched 44:



Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Area %
1	5.056	MM	0.1210	2362.74805	66.9143
2	6.886	MM	0.1605	1168.25940	33.0857

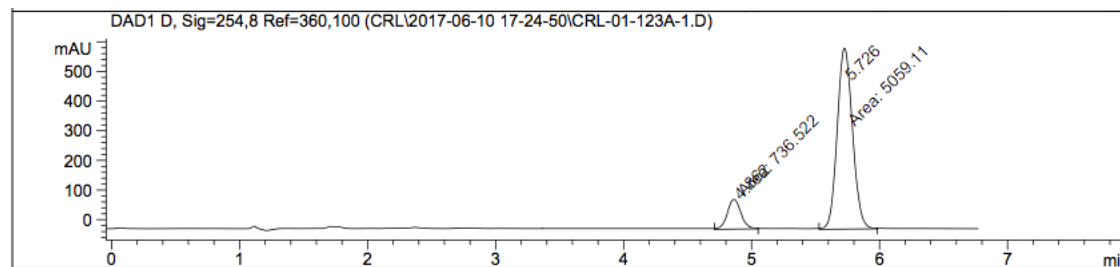


SFC data for racemic 45:



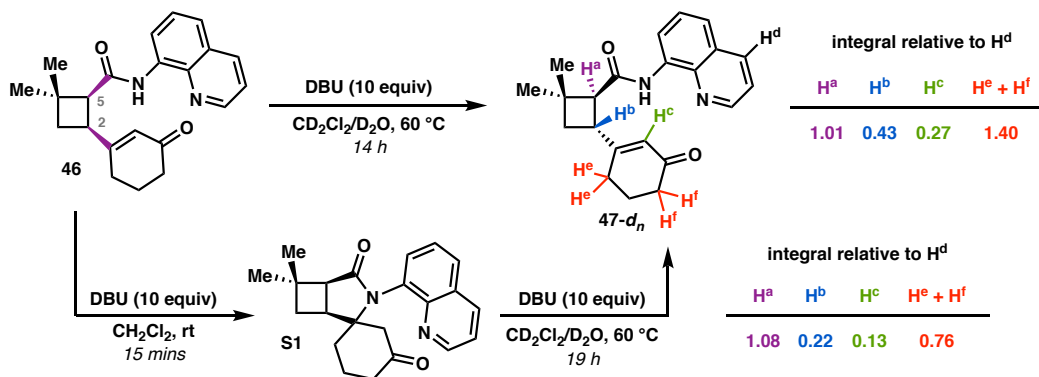
Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Area %
1	4.889	MM	0.1294	3927.92065	49.6039
2	5.767	MM	0.1468	3990.65186	50.3961

Enantioenriched 45:

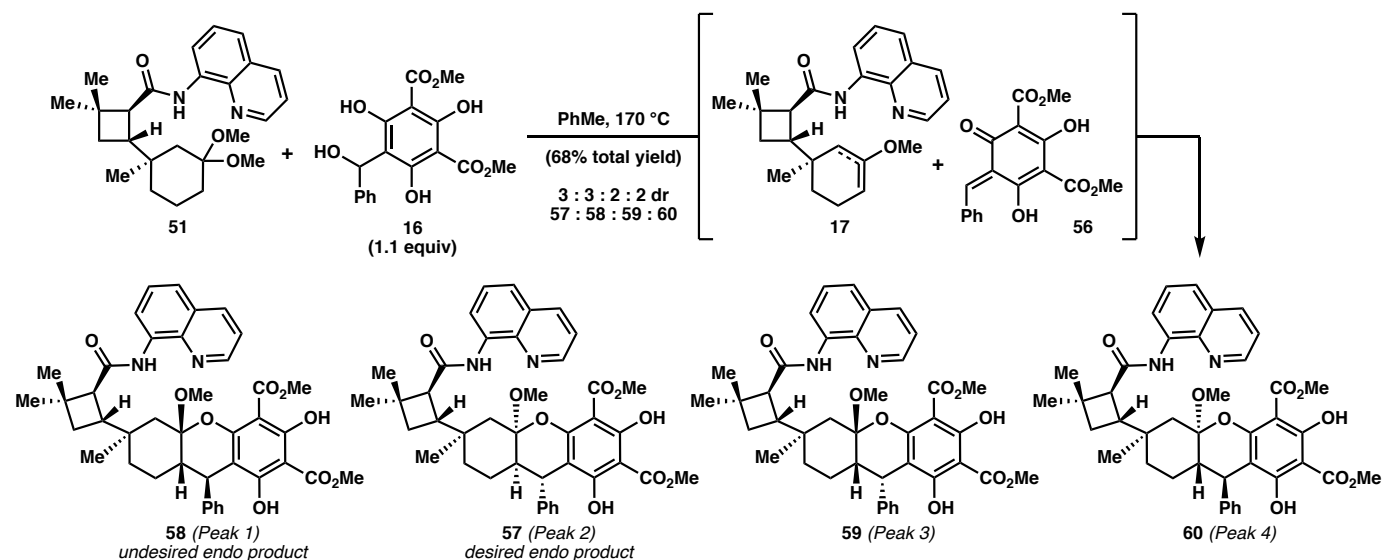


Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Area %
1	4.862	MM	0.1218	736.52246	12.7082
2	5.726	MM	0.1379	5059.11182	87.2918

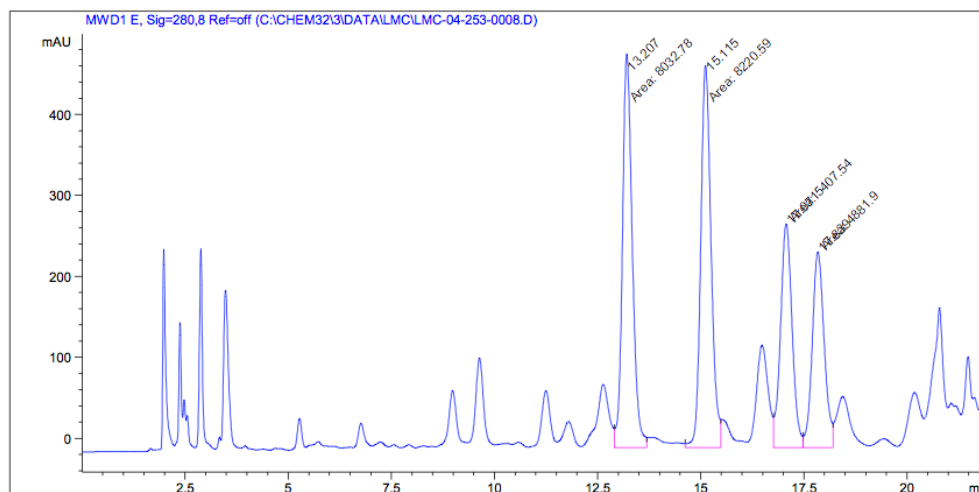
Deuterium-labeling studies to determine site of epimerization in *trans*-cyclobutane 47:



Preparation of tricyclic ketals 57–60 by thermal cycloaddition:



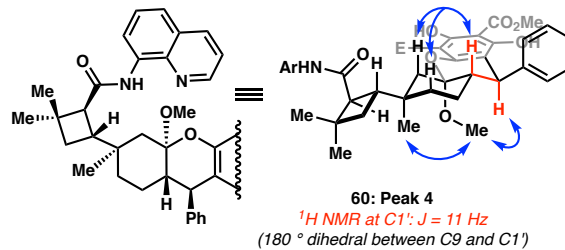
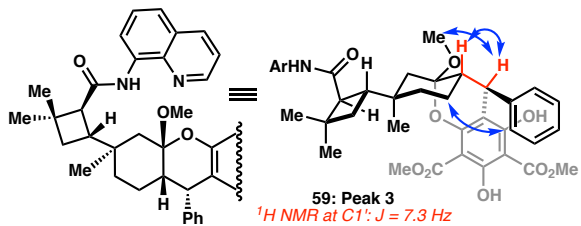
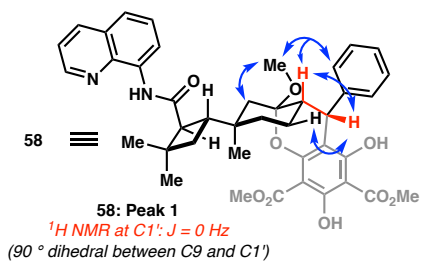
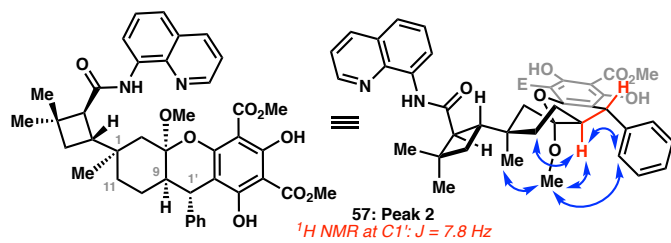
Chromatogram from HPLC separation of 57–60:



Chromatogram integration from HPLC separation of 57–60:

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	13.207	MF	0.2752	8032.78027	486.56299	30.2635
2	15.115	MF	0.2903	8220.58984	472.01566	30.9711
3	17.071	MF	0.3263	5407.54297	276.19736	20.3729
4	17.839	MF	0.3366	4881.89746	241.74495	18.3925

Summary of Key nOe correlations for stereochemical assignment of 57–60:



Regio- and stereochemical assignment of Norrish–Yang product 80:

Key HMBC and nOe correlations:

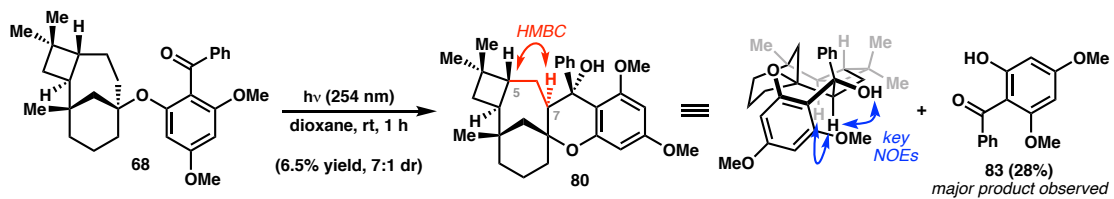
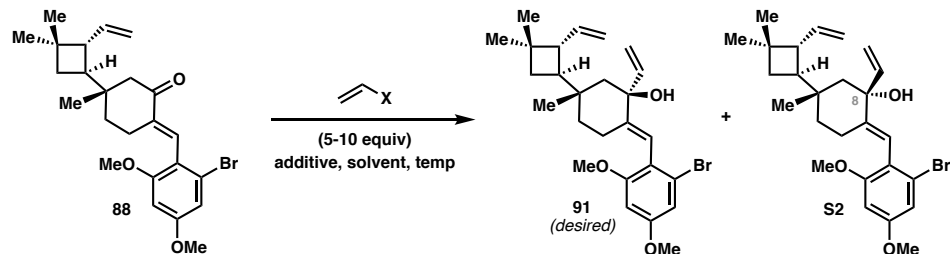


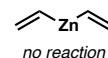
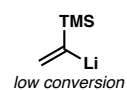
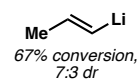
Table S3: Additional experiments toward optimization of 1,2-addition:



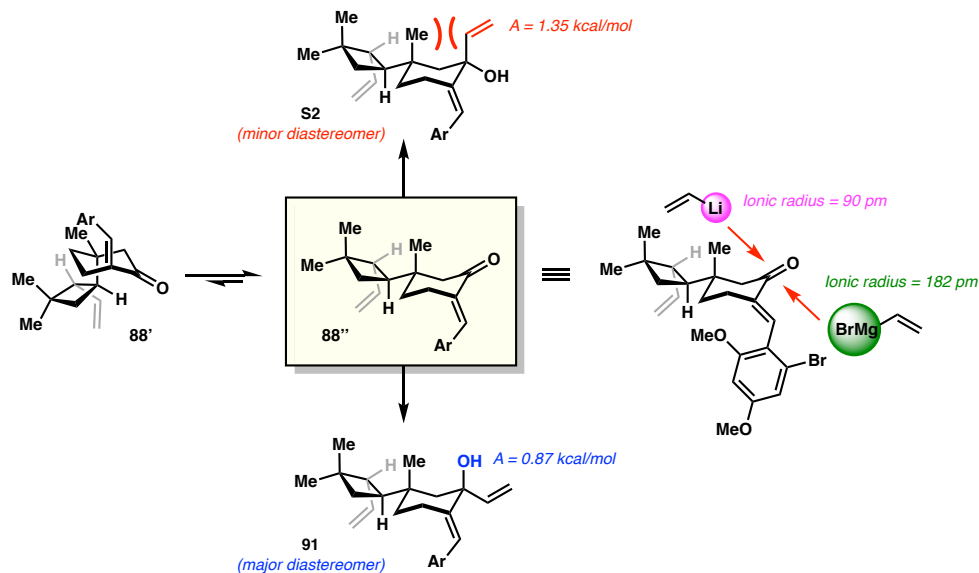
entry ^a	X	solvent	temp °C	additive	% conv. ^b	dr (91:S2)	% total yield (% yield 91)
1	MgBr	THF	-35	-	75	2.5:1	57 (41)
2	MgBr	THF	-78	-	56	4.5:1	41 (34)
3	MgBr	Et ₂ O	-35	-	ND	2.5:1	57 (41)
4	MgBr	CH ₂ Cl ₂	0	-	85	1.9:1	69 (45)
5	MgBr	THF	0	-	60	2.0:1	50 (33)
6	MgBr	THF	-78	CeCl ₃	81	1.6:1	66 (41)
7	MgBr	THF	-78	LaCl ₃ ·2LiCl	100	1.7:1	59 (37)
8	Li	THF	-78	-	86	2.1:1	77 (52)
9	Li	THF	-78	La(OTf) ₃	94	1.8:1	67 (43)
10	Li	THF	-100	-	87	1.4:1	61 (36)

^a Test reactions conducted on 0.020 mmol scale. ^b Conversion determined by relative integration of UV LC/MS peaks, detected at 254 nm.

Alternative nucleophiles tested:



Conformational analysis and diastereoselectivity model for 1,2-addition:



Overview of the successful route to (+)-psiguadial B:

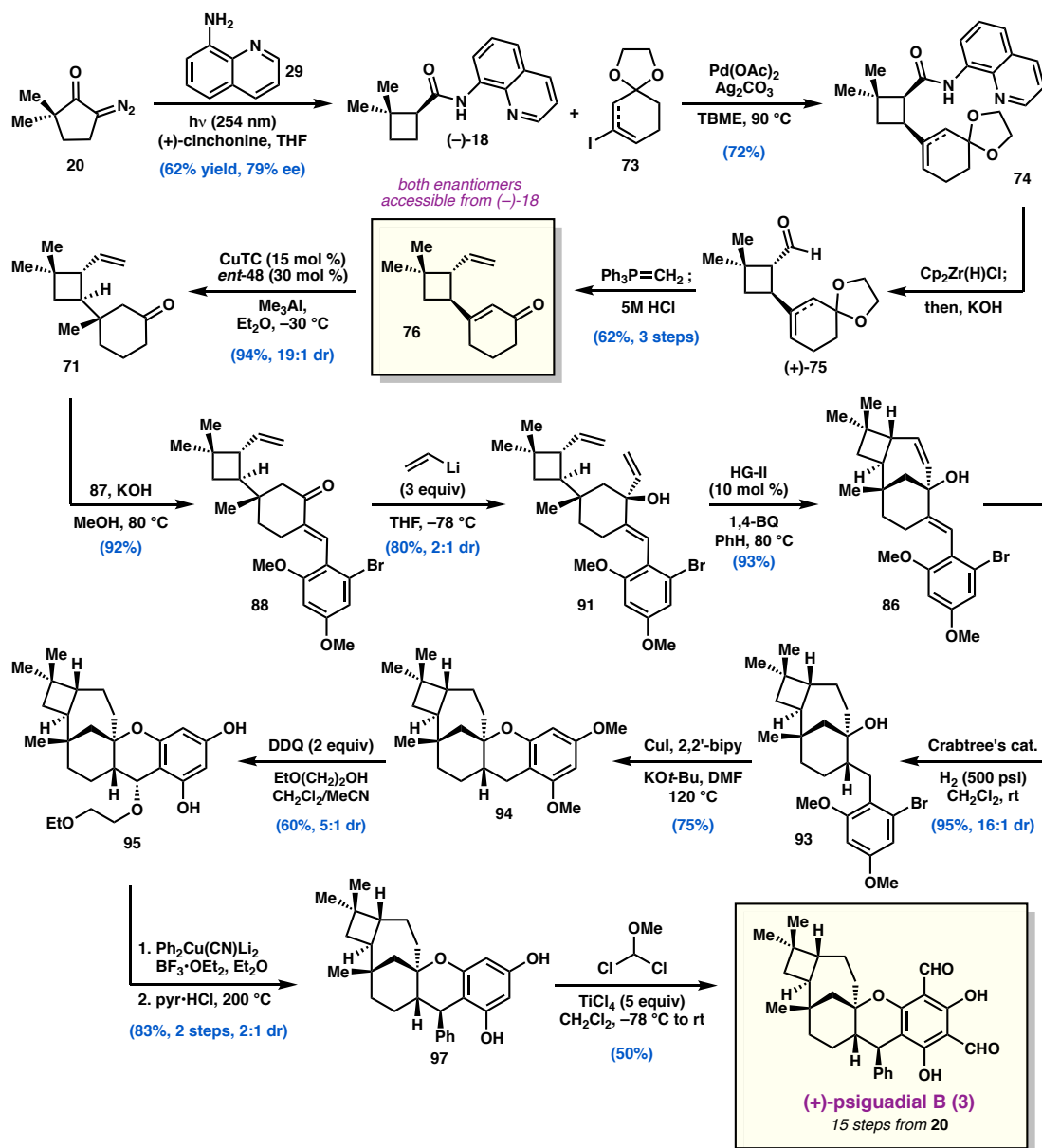
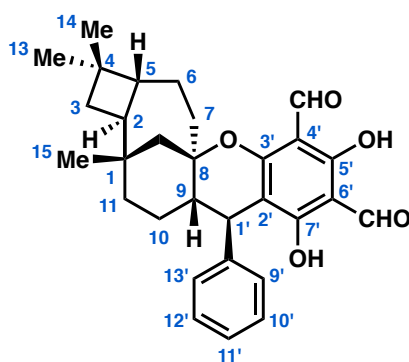


Table S4. Comparison of ¹H NMR spectroscopic data for natural and synthetic (+)-psiguadial B (3)

carbon number	Natural (+)-psiguadial B ¹ ¹ H NMR, 500 MHz, CDCl ₃	Synthetic (+)-psiguadial B ¹ H NMR, 400 MHz, CDCl ₃
5'-OH	δ 13.51 (s, 1H)	δ 13.51 (s, 1H)
7'-OH	13.04 (s, 1H)	13.04 (s, 1H)
14', 15'	10.08 (s, 2H)	10.07 (s, 2H)
9', 13' ²	7.23 (2H)	7.26 (dd, <i>J</i> = 14.6, 1.5 Hz, 2H)
11'	7.18 (3H)	7.23 – 7.17 (m, 1H)
10', 12'	–	7.10 (br m, 2H)
1'	3.49 (d, <i>J</i> = 11.5 Hz, 1H)	3.49 (d, <i>J</i> = 11.5 Hz, 1H)
2	2.16 (1H)	2.20 – 2.12 (m, 1H)
12	2.08 (1H)	2.09 (dd, <i>J</i> = 12.7, 2.4 Hz, 1H)
7	1.93 (1H)	1.92 (ddd, <i>J</i> = 14.9, 12.8, 4.2 Hz, 1H)
5	1.82 (m, 1H)	1.82 (ddd, <i>J</i> = 12.3, 8.8, 5.6 Hz, 1H)
9	1.68 (1H)	1.73 – 1.59 (m, 3H)
6	1.65 (1H)	–
7	1.58 (m, 1H)	–
3	1.52 (1H)	1.53 – 1.44 (m, 1H)
10	1.49 (m, 2H)	1.49 (ddd, <i>J</i> = 11.6, 8.1, 2.9 Hz, 2H)
6, 11	1.41 (2H)	–
3	1.37 (1H)	1.44 – 1.29 (m, 4H)
12	1.29 (1H)	–
11	1.10 (1H)	1.05 (dd, <i>J</i> = 7.6, 5.8 Hz, 1H)
13	1.02 (s, 3H)	1.02 (s, 3H)
14	1.01 (s, 3H)	1.00 (s, 3H)
15	0.86 (s, 3H)	0.85 (s, 3H)

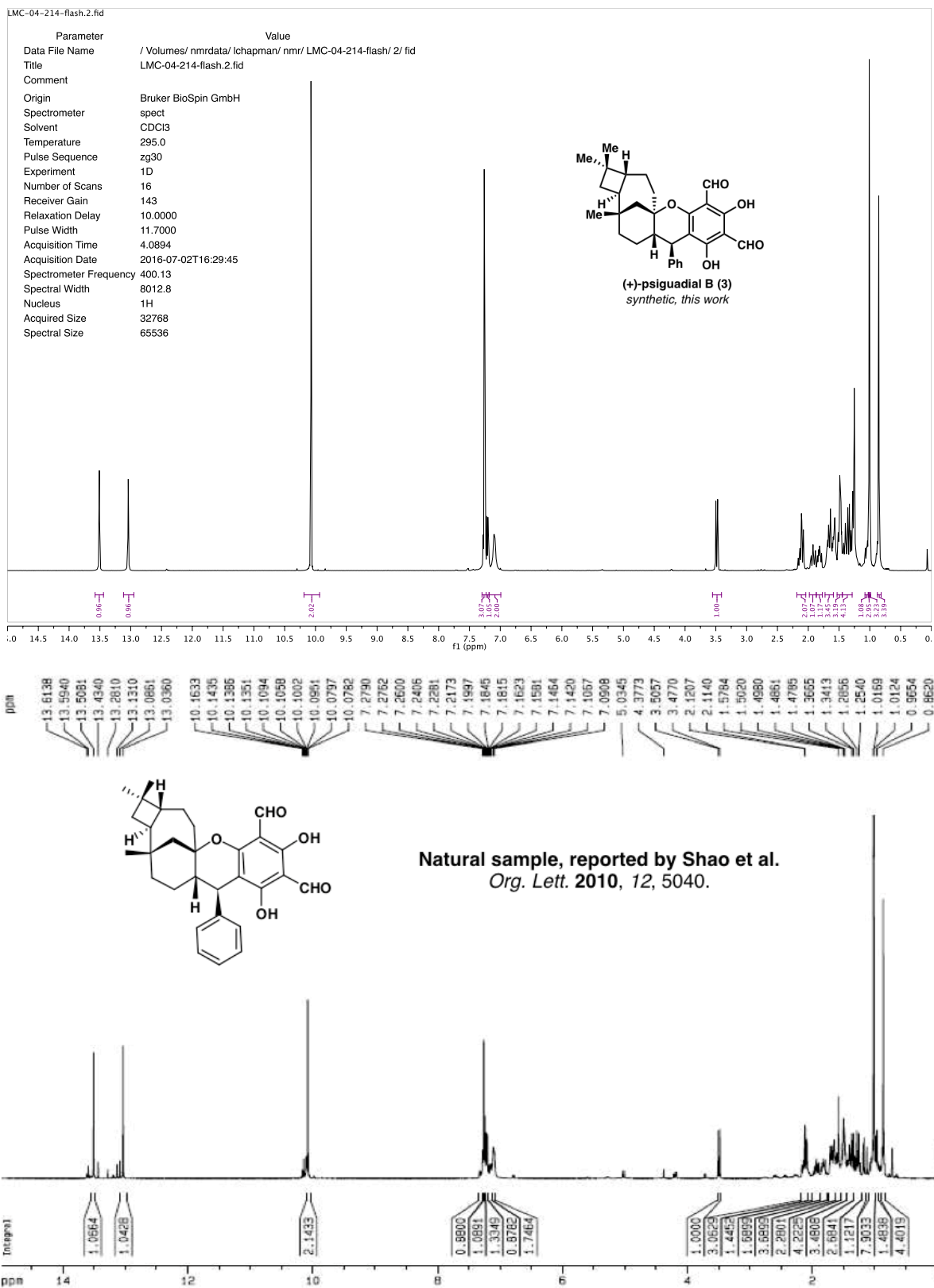


(+)-Psiguadial B (3) carbon numbering as reported by Shao et al.¹

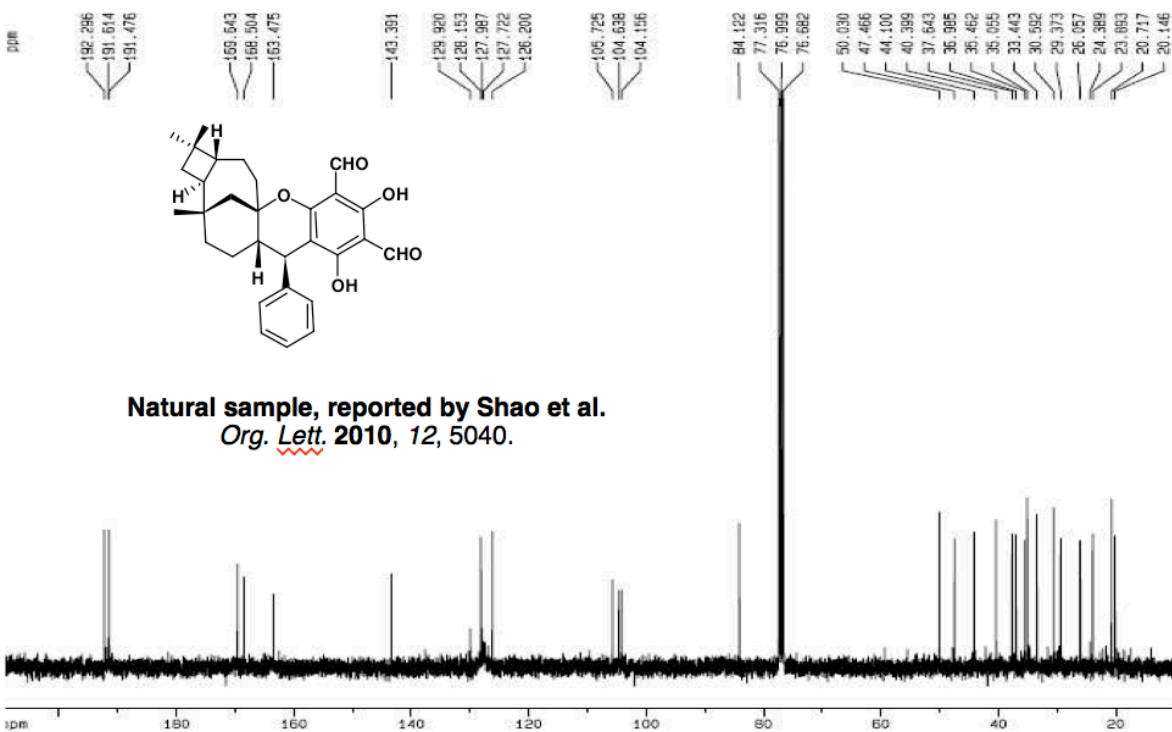
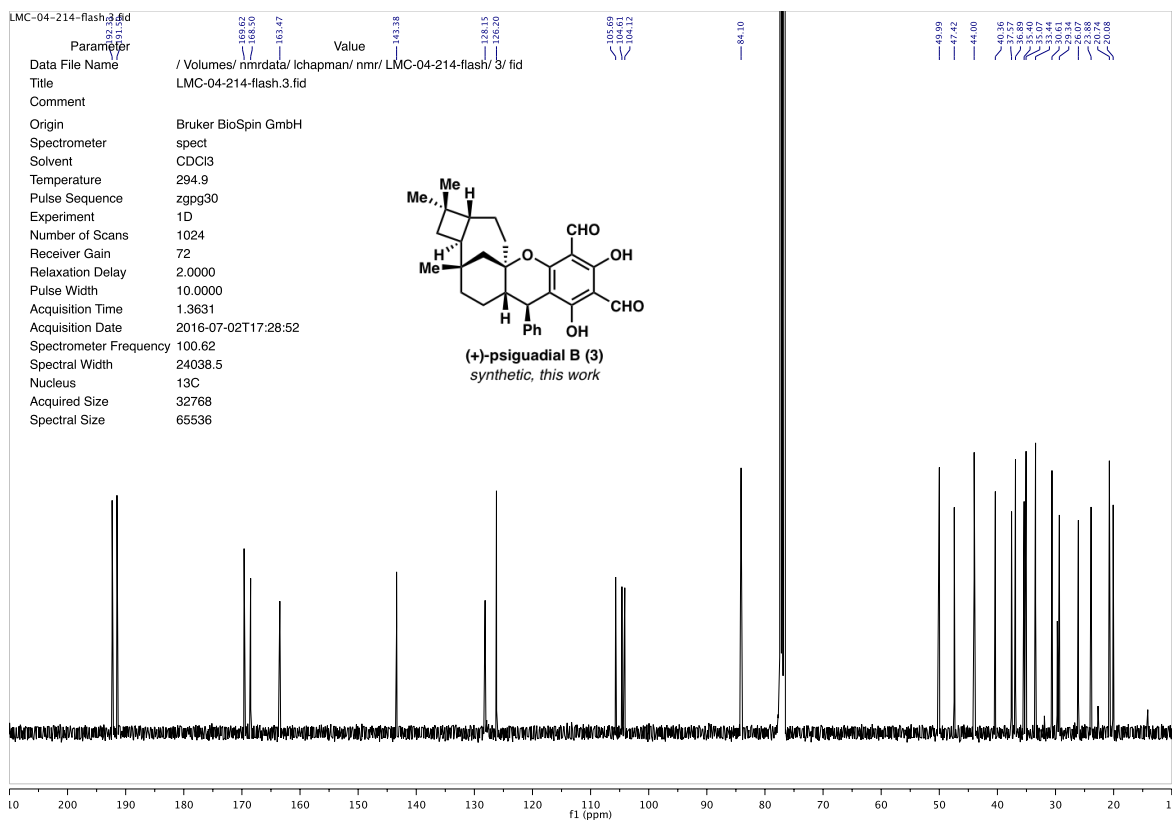
Table S5. Comparison of ^{13}C NMR spectroscopic data for natural and synthetic (+)-psiguadial B (3)

carbon number	Natural (+)-psiguadial B¹ ^{13}C NMR, 125 MHz, CDCl_3	Synthetic (+)-psiguadial B³ ^{13}C NMR, 101 MHz, CDCl_3	Δ (ppm)
15'	192.3	192.3	0.0
14'	191.4	191.5	0.1
7'	169.6	169.6	0.0
5'	168.5	168.5	0.0
3'	163.5	163.5	0.0
8'	143.4	143.4	0.0
9', 11', 13'	128.2	128.2	0.0
10', 12'	126.2	126.2	0.0
2'	105.7	105.7	0.0
4'	104.6	104.6	0.0
6'	104.2	104.1	-0.1
8	84.1	84.1	0.0
9	50.0	50.0	0.0
12	47.5	47.4	-0.1
5	44.1	44.0	-0.1
1'	40.4	40.4	0.0
11	37.6	37.6	0.0
2	37.0	36.9	-0.1
3	35.5	35.4	-0.1
4	35.1	35.1	0.0
1	33.4	33.4	0.0
13	30.6	30.6	0.0
7	29.4	29.3	-0.1
15	26.1	26.1	0.0
10	23.9	23.9	0.0
14	20.7	20.7	0.0
6	20.1	20.1	0.0

¹H NMR spectral comparison of natural and synthetic (+)-psiguadial B (3):



¹³C NMR spectral comparison of natural and synthetic (+)-psiguadial B (3):



X-Ray crystallographic data for minor ketone 50:

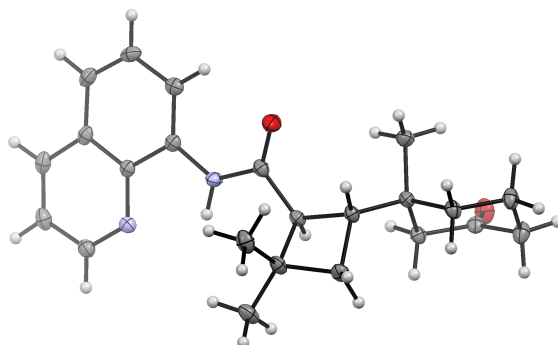


Table S6. Crystal data and structure refinement for LMC-04-038 or minor ketone **50**. Graphical representation of the structure with 50% probability thermal ellipsoids was generated using Mercury visualization software.

Identification code	LMC-04-038 or minor ketone 50 .	
Empirical formula	C ₂₃ H ₂₈ N ₂ O ₂	
Formula weight	364.47	
Temperature	100 K	
Wavelength	1.54178 Å	
Crystal system	Orthorhombic	
Space group	P2 ₁ 2 ₁ 2 ₁	
Unit cell dimensions	a = 6.5496(4) Å	α = 90°.
	b = 10.8428(6) Å	β = 90°.
	c = 27.6666(15) Å	γ = 90°.
Volume	1964.77(19) Å ³	
Z	4	
Density (calculated)	1.232 Mg/m ³	
Absorption coefficient	0.619 mm ⁻¹	
F(000)	784	
Crystal size	0.25 x 0.15 x 0.1 mm ³	
Theta range for data collection	3.195 to 79.392°.	
Index ranges	-8 ≤ h ≤ 8, -13 ≤ k ≤ 13, -35 ≤ l ≤ 34	
Reflections collected	24424	
Independent reflections	4179 [R(int) = 0.0414]	
Completeness to theta = 67.679°	100.0 %	
Absorption correction	Semi-empirical from equivalents	
Max. and min. transmission	0.7542 and 0.7017	
Refinement method	Full-matrix least-squares on F ²	
Data / restraints / parameters	4179 / 0 / 251	

Goodness-of-fit on F^2	1.021
Final R indices [$I > 2\sigma(I)$]	R1 = 0.0272, wR2 = 0.0655
R indices (all data)	R1 = 0.0300, wR2 = 0.0672
Absolute structure parameter	0.08(9)
Extinction coefficient	0.0031(3)
Largest diff. peak and hole	0.187 and -0.154 e.Å ⁻³

Crystallographic analysis and refinement details:

Low-temperature diffraction data (φ - and ω -scans) were collected on a Bruker AXS D8 VENTURE KAPPA diffractometer coupled to a PHOTON 100 CMOS detector with Cu-K α radiation ($\lambda = 1.54178 \text{ \AA}$) from a I μ S HB micro-focus sealed X-ray tube. All diffractometer manipulations, including data collection, integration, and scaling were carried out using the Bruker APEXII software.⁴ Absorption corrections were applied using SADABS.⁵ The structure was solved by intrinsic phasing using SHELXT14 and refined against F^2 on all data by full-matrix least squares with SHELXL-2014⁶ using established refinement techniques.⁷ All non-hydrogen atoms were refined anisotropically. The coordinates for the hydrogen atom bound to N1 were located in the difference Fourier synthesis and refined using a riding model. All other hydrogen atoms were included into the model at geometrically calculated positions and refined using a riding model. The isotropic displacement parameters of all hydrogen atoms were fixed to 1.2 times the U value of the atoms they are linked to (1.5 times for methyl groups and hydroxyl groups). Compound **50** crystallizes in the monoclinic space group $P2_12_12_1$ with one molecule in the asymmetric unit. Absolute configuration was determined by anomalous dispersion (Flack = 0.08(9)).⁸ Graphical representation of the structure with 50% probability thermal ellipsoids was generated using Mercury visualization software.⁹

Table S7. Atomic coordinates ($\times 10^4$) and equivalent isotropic displacement parameters ($\text{\AA}^2 \times 10^3$) for minor ketone **50**. $U(\text{eq})$ is defined as one third of the trace of the orthogonalized U^{ij} tensor.

	x	y	z	$U(\text{eq})$
O(2)	1276(2)	7506(1)	2689(1)	28(1)
O(1)	8526(2)	4834(1)	3832(1)	28(1)
N(1)	6907(2)	5042(1)	4555(1)	17(1)
N(2)	6036(2)	5230(1)	5486(1)	17(1)
C(1)	6977(2)	4762(1)	4073(1)	17(1)
C(19)	9262(2)	6260(1)	5671(1)	18(1)
C(8)	4149(2)	4860(1)	2948(1)	15(1)
C(23)	7898(2)	5670(1)	5348(1)	15(1)
C(15)	8440(2)	5545(1)	4849(1)	16(1)
C(7)	4752(2)	3938(1)	3343(1)	16(1)
C(2)	4949(2)	4328(1)	3880(1)	16(1)
C(17)	11684(2)	6499(1)	5023(1)	21(1)
C(18)	11180(2)	6672(1)	5498(1)	21(1)
C(16)	10309(2)	5955(1)	4692(1)	19(1)
C(3)	4183(2)	3002(1)	4028(1)	19(1)
C(22)	5507(2)	5373(2)	5942(1)	21(1)
C(10)	1519(2)	6401(2)	2662(1)	19(1)
C(14)	5948(2)	5744(2)	2867(1)	22(1)
C(21)	6728(3)	5976(2)	6290(1)	24(1)
C(9)	2231(2)	5619(1)	3083(1)	18(1)
C(6)	3268(3)	2907(1)	3511(1)	20(1)
C(13)	3699(2)	4120(2)	2484(1)	19(1)
C(20)	8601(3)	6410(2)	6153(1)	22(1)
C(12)	2998(2)	4916(2)	2060(1)	22(1)
C(4)	2696(3)	2947(2)	4449(1)	25(1)
C(5)	5955(3)	2106(2)	4103(1)	26(1)
C(11)	1133(3)	5697(2)	2200(1)	24(1)

Table S8. Bond lengths [Å] and angles [°] for minor ketone **50**.

O(2)-C(10)	1.211(2)
O(1)-C(1)	1.2176(18)
N(1)-C(1)	1.3676(19)
N(1)-C(15)	1.402(2)
N(1)-H(1)	0.90(2)
N(2)-C(23)	1.364(2)
N(2)-C(22)	1.319(2)
C(1)-C(2)	1.507(2)
C(19)-C(23)	1.417(2)
C(19)-C(18)	1.416(2)
C(19)-C(20)	1.412(2)
C(8)-C(7)	1.534(2)
C(8)-C(14)	1.535(2)
C(8)-C(9)	1.548(2)
C(8)-C(13)	1.540(2)
C(23)-C(15)	1.431(2)
C(15)-C(16)	1.373(2)
C(7)-H(7)	1.0000
C(7)-C(2)	1.5486(19)
C(7)-C(6)	1.552(2)
C(2)-H(2)	1.0000
C(2)-C(3)	1.577(2)
C(17)-H(17)	0.9500
C(17)-C(18)	1.369(2)
C(17)-C(16)	1.414(2)
C(18)-H(18)	0.9500
C(16)-H(16)	0.9500
C(3)-C(6)	1.555(2)
C(3)-C(4)	1.520(2)
C(3)-C(5)	1.527(2)
C(22)-H(22)	0.9500
C(22)-C(21)	1.410(2)
C(10)-C(9)	1.514(2)
C(10)-C(11)	1.511(2)

C(14)-H(14A)	0.9800
C(14)-H(14B)	0.9800
C(14)-H(14C)	0.9800
C(21)-H(21)	0.9500
C(21)-C(20)	1.367(2)
C(9)-H(9A)	0.9900
C(9)-H(9B)	0.9900
C(6)-H(6A)	0.9900
C(6)-H(6B)	0.9900
C(13)-H(13A)	0.9900
C(13)-H(13B)	0.9900
C(13)-C(12)	1.527(2)
C(20)-H(20)	0.9500
C(12)-H(12A)	0.9900
C(12)-H(12B)	0.9900
C(12)-C(11)	1.535(2)
C(4)-H(4A)	0.9800
C(4)-H(4B)	0.9800
C(4)-H(4C)	0.9800
C(5)-H(5A)	0.9800
C(5)-H(5B)	0.9800
C(5)-H(5C)	0.9800
C(11)-H(11A)	0.9900
C(11)-H(11B)	0.9900
C(1)-N(1)-C(15)	128.85(13)
C(1)-N(1)-H(1)	119.7(13)
C(15)-N(1)-H(1)	111.4(13)
C(22)-N(2)-C(23)	117.51(13)
O(1)-C(1)-N(1)	123.29(14)
O(1)-C(1)-C(2)	124.01(13)
N(1)-C(1)-C(2)	112.68(12)
C(18)-C(19)-C(23)	119.31(14)
C(20)-C(19)-C(23)	117.04(14)
C(20)-C(19)-C(18)	123.65(15)
C(7)-C(8)-C(14)	108.23(12)

C(7)-C(8)-C(9)	112.51(12)
C(7)-C(8)-C(13)	107.68(12)
C(14)-C(8)-C(9)	109.05(13)
C(14)-C(8)-C(13)	110.57(12)
C(13)-C(8)-C(9)	108.79(12)
N(2)-C(23)-C(19)	123.01(13)
N(2)-C(23)-C(15)	117.29(13)
C(19)-C(23)-C(15)	119.68(14)
N(1)-C(15)-C(23)	114.75(13)
C(16)-C(15)-N(1)	125.50(14)
C(16)-C(15)-C(23)	119.69(14)
C(8)-C(7)-H(7)	107.9
C(8)-C(7)-C(2)	121.88(12)
C(8)-C(7)-C(6)	121.41(13)
C(2)-C(7)-H(7)	107.9
C(2)-C(7)-C(6)	87.86(11)
C(6)-C(7)-H(7)	107.9
C(1)-C(2)-C(7)	119.93(12)
C(1)-C(2)-H(2)	109.5
C(1)-C(2)-C(3)	118.23(13)
C(7)-C(2)-H(2)	109.5
C(7)-C(2)-C(3)	88.51(11)
C(3)-C(2)-H(2)	109.5
C(18)-C(17)-H(17)	119.1
C(18)-C(17)-C(16)	121.76(15)
C(16)-C(17)-H(17)	119.1
C(19)-C(18)-H(18)	120.2
C(17)-C(18)-C(19)	119.62(15)
C(17)-C(18)-H(18)	120.2
C(15)-C(16)-C(17)	119.85(14)
C(15)-C(16)-H(16)	120.1
C(17)-C(16)-H(16)	120.1
C(6)-C(3)-C(2)	86.77(11)
C(4)-C(3)-C(2)	116.02(12)
C(4)-C(3)-C(6)	117.16(14)
C(4)-C(3)-C(5)	110.98(13)

C(5)-C(3)-C(2)	111.89(13)
C(5)-C(3)-C(6)	112.06(13)
N(2)-C(22)-H(22)	118.0
N(2)-C(22)-C(21)	123.90(15)
C(21)-C(22)-H(22)	118.0
O(2)-C(10)-C(9)	123.15(15)
O(2)-C(10)-C(11)	122.04(15)
C(11)-C(10)-C(9)	114.80(14)
C(8)-C(14)-H(14A)	109.5
C(8)-C(14)-H(14B)	109.5
C(8)-C(14)-H(14C)	109.5
H(14A)-C(14)-H(14B)	109.5
H(14A)-C(14)-H(14C)	109.5
H(14B)-C(14)-H(14C)	109.5
C(22)-C(21)-H(21)	120.7
C(20)-C(21)-C(22)	118.69(15)
C(20)-C(21)-H(21)	120.7
C(8)-C(9)-H(9A)	109.4
C(8)-C(9)-H(9B)	109.4
C(10)-C(9)-C(8)	111.24(12)
C(10)-C(9)-H(9A)	109.4
C(10)-C(9)-H(9B)	109.4
H(9A)-C(9)-H(9B)	108.0
C(7)-C(6)-C(3)	89.19(11)
C(7)-C(6)-H(6A)	113.8
C(7)-C(6)-H(6B)	113.8
C(3)-C(6)-H(6A)	113.8
C(3)-C(6)-H(6B)	113.8
H(6A)-C(6)-H(6B)	111.0
C(8)-C(13)-H(13A)	108.8
C(8)-C(13)-H(13B)	108.8
H(13A)-C(13)-H(13B)	107.7
C(12)-C(13)-C(8)	113.71(13)
C(12)-C(13)-H(13A)	108.8
C(12)-C(13)-H(13B)	108.8
C(19)-C(20)-H(20)	120.1

C(21)-C(20)-C(19)	119.80(15)
C(21)-C(20)-H(20)	120.1
C(13)-C(12)-H(12A)	109.4
C(13)-C(12)-H(12B)	109.4
C(13)-C(12)-C(11)	111.00(13)
H(12A)-C(12)-H(12B)	108.0
C(11)-C(12)-H(12A)	109.4
C(11)-C(12)-H(12B)	109.4
C(3)-C(4)-H(4A)	109.5
C(3)-C(4)-H(4B)	109.5
C(3)-C(4)-H(4C)	109.5
H(4A)-C(4)-H(4B)	109.5
H(4A)-C(4)-H(4C)	109.5
H(4B)-C(4)-H(4C)	109.5
C(3)-C(5)-H(5A)	109.5
C(3)-C(5)-H(5B)	109.5
C(3)-C(5)-H(5C)	109.5
H(5A)-C(5)-H(5B)	109.5
H(5A)-C(5)-H(5C)	109.5
H(5B)-C(5)-H(5C)	109.5
C(10)-C(11)-C(12)	110.96(13)
C(10)-C(11)-H(11A)	109.4
C(10)-C(11)-H(11B)	109.4
C(12)-C(11)-H(11A)	109.4
C(12)-C(11)-H(11B)	109.4
H(11A)-C(11)-H(11B)	108.0

Table S9. Anisotropic displacement parameters ($\text{\AA}^2 \times 10^4$) for minor ketone **50**. The anisotropic displacement factor exponent takes the form: $-2\pi^2 [h^2 a^{*2} U^{11} + \dots + 2 h k a^* b^* U^{12}]$

	U ¹¹	U ²²	U ³³	U ²³	U ¹³	U ¹²
O(2)	36(1)	22(1)	27(1)	0(1)	-3(1)	8(1)
O(1)	19(1)	47(1)	18(1)	-3(1)	4(1)	-2(1)
N(1)	15(1)	21(1)	16(1)	-2(1)	3(1)	-3(1)
N(2)	18(1)	17(1)	17(1)	1(1)	1(1)	1(1)
C(1)	20(1)	16(1)	16(1)	1(1)	2(1)	2(1)
C(19)	20(1)	12(1)	21(1)	2(1)	-6(1)	2(1)
C(8)	17(1)	16(1)	14(1)	1(1)	2(1)	0(1)
C(23)	16(1)	12(1)	17(1)	1(1)	-1(1)	2(1)
C(15)	17(1)	13(1)	18(1)	0(1)	-1(1)	2(1)
C(7)	19(1)	15(1)	14(1)	0(1)	1(1)	1(1)
C(2)	18(1)	15(1)	14(1)	0(1)	3(1)	0(1)
C(17)	14(1)	16(1)	33(1)	5(1)	-1(1)	-1(1)
C(18)	20(1)	16(1)	28(1)	2(1)	-9(1)	-2(1)
C(16)	18(1)	16(1)	22(1)	2(1)	2(1)	2(1)
C(3)	26(1)	14(1)	16(1)	1(1)	3(1)	-2(1)
C(22)	23(1)	23(1)	18(1)	2(1)	1(1)	0(1)
C(10)	16(1)	22(1)	19(1)	-1(1)	2(1)	4(1)
C(14)	19(1)	23(1)	25(1)	6(1)	-1(1)	-3(1)
C(21)	32(1)	24(1)	15(1)	0(1)	-1(1)	2(1)
C(9)	20(1)	18(1)	15(1)	-1(1)	2(1)	1(1)
C(6)	25(1)	14(1)	19(1)	-1(1)	1(1)	-3(1)
C(13)	21(1)	20(1)	15(1)	-2(1)	2(1)	3(1)
C(20)	29(1)	17(1)	19(1)	-1(1)	-8(1)	2(1)
C(12)	24(1)	29(1)	15(1)	-3(1)	1(1)	7(1)
C(4)	34(1)	21(1)	21(1)	1(1)	8(1)	-6(1)
C(5)	36(1)	18(1)	22(1)	3(1)	0(1)	4(1)
C(11)	24(1)	30(1)	18(1)	-3(1)	-3(1)	7(1)

Table S10. Hydrogen coordinates ($\times 10^4$) and isotropic displacement parameters ($\text{\AA}^2 \times 10^3$) for minor ketone **50**.

	x	y	z	U(eq)
H(7)	6064	3539	3245	19
H(2)	3872	4950	3959	19
H(17)	12993	6751	4913	25
H(18)	12112	7066	5712	25
H(16)	10677	5874	4361	22
H(22)	4228	5051	6044	25
H(14A)	6225	6198	3166	34
H(14B)	5607	6327	2609	34
H(14C)	7162	5270	2775	34
H(21)	6258	6078	6612	28
H(9A)	1121	5052	3180	21
H(9B)	2548	6159	3361	21
H(6A)	3511	2101	3353	23
H(6B)	1810	3142	3492	23
H(13A)	4948	3668	2389	22
H(13B)	2627	3502	2556	22
H(20)	9454	6811	6382	26
H(12A)	2645	4379	1783	27
H(12B)	4127	5466	1959	27
H(4A)	3400	3189	4747	38
H(4B)	2174	2105	4483	38
H(4C)	1557	3513	4389	38
H(5A)	6918	2181	3833	38
H(5B)	5430	1261	4117	38
H(5C)	6654	2302	4406	38
H(11A)	825	6286	1936	29
H(11B)	-67	5153	2242	29
H(1)	5760(30)	4900(20)	4724(7)	29

Table S11. Torsion angles [°] for minor ketone **50**.

O(2)-C(10)-C(9)-C(8)	-125.20(17)
O(2)-C(10)-C(11)-C(12)	127.00(17)
O(1)-C(1)-C(2)-C(7)	0.1(2)
O(1)-C(1)-C(2)-C(3)	106.07(18)
N(1)-C(1)-C(2)-C(7)	-178.49(13)
N(1)-C(1)-C(2)-C(3)	-72.53(17)
N(1)-C(15)-C(16)-C(17)	176.96(14)
N(2)-C(23)-C(15)-N(1)	3.66(19)
N(2)-C(23)-C(15)-C(16)	-179.03(13)
N(2)-C(22)-C(21)-C(20)	-2.0(3)
C(1)-N(1)-C(15)-C(23)	-177.11(14)
C(1)-N(1)-C(15)-C(16)	5.8(3)
C(1)-C(2)-C(3)-C(6)	-144.20(13)
C(1)-C(2)-C(3)-C(4)	97.03(17)
C(1)-C(2)-C(3)-C(5)	-31.70(18)
C(19)-C(23)-C(15)-N(1)	-174.83(13)
C(19)-C(23)-C(15)-C(16)	2.5(2)
C(8)-C(7)-C(2)-C(1)	-91.02(17)
C(8)-C(7)-C(2)-C(3)	146.90(14)
C(8)-C(7)-C(6)-C(3)	-147.60(13)
C(8)-C(13)-C(12)-C(11)	-55.22(18)
C(23)-N(2)-C(22)-C(21)	1.6(2)
C(23)-C(19)-C(18)-C(17)	0.3(2)
C(23)-C(19)-C(20)-C(21)	0.9(2)
C(23)-C(15)-C(16)-C(17)	0.0(2)
C(15)-N(1)-C(1)-O(1)	6.0(3)
C(15)-N(1)-C(1)-C(2)	-175.39(14)
C(7)-C(8)-C(9)-C(10)	-172.85(13)
C(7)-C(8)-C(13)-C(12)	177.89(13)
C(7)-C(2)-C(3)-C(6)	-20.67(11)
C(7)-C(2)-C(3)-C(4)	-139.44(14)
C(7)-C(2)-C(3)-C(5)	91.84(14)
C(2)-C(7)-C(6)-C(3)	-20.99(11)
C(2)-C(3)-C(6)-C(7)	20.61(11)

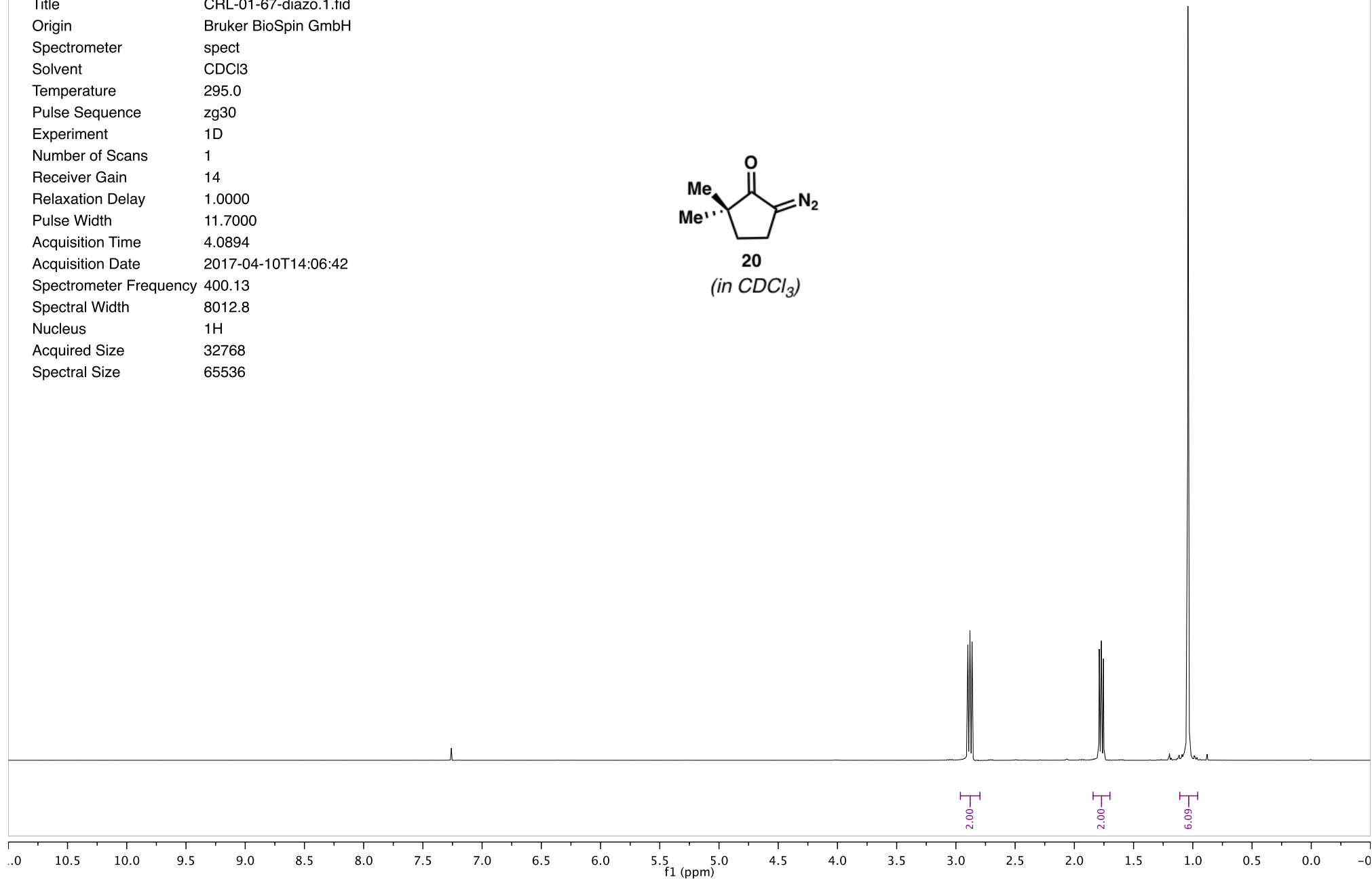
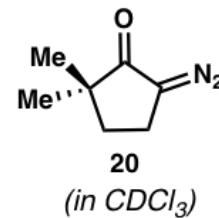
C(18)-C(19)-C(23)-N(2)	179.01(14)
C(18)-C(19)-C(23)-C(15)	-2.6(2)
C(18)-C(19)-C(20)-C(21)	-179.54(15)
C(18)-C(17)-C(16)-C(15)	-2.4(2)
C(16)-C(17)-C(18)-C(19)	2.3(2)
C(22)-N(2)-C(23)-C(19)	0.2(2)
C(22)-N(2)-C(23)-C(15)	-178.22(14)
C(22)-C(21)-C(20)-C(19)	0.7(2)
C(14)-C(8)-C(7)-C(2)	71.18(17)
C(14)-C(8)-C(7)-C(6)	-179.66(13)
C(14)-C(8)-C(9)-C(10)	67.06(16)
C(14)-C(8)-C(13)-C(12)	-64.05(17)
C(9)-C(8)-C(7)-C(2)	-49.38(18)
C(9)-C(8)-C(7)-C(6)	59.77(18)
C(9)-C(8)-C(13)-C(12)	55.69(16)
C(9)-C(10)-C(11)-C(12)	-52.89(19)
C(6)-C(7)-C(2)-C(1)	142.76(14)
C(6)-C(7)-C(2)-C(3)	20.68(11)
C(13)-C(8)-C(7)-C(2)	-169.26(13)
C(13)-C(8)-C(7)-C(6)	-60.10(17)
C(13)-C(8)-C(9)-C(10)	-53.62(16)
C(13)-C(12)-C(11)-C(10)	51.44(19)
C(20)-C(19)-C(23)-N(2)	-1.4(2)
C(20)-C(19)-C(23)-C(15)	176.96(13)
C(20)-C(19)-C(18)-C(17)	-179.26(15)
C(4)-C(3)-C(6)-C(7)	138.32(14)
C(5)-C(3)-C(6)-C(7)	-91.73(14)
C(11)-C(10)-C(9)-C(8)	54.70(18)

References

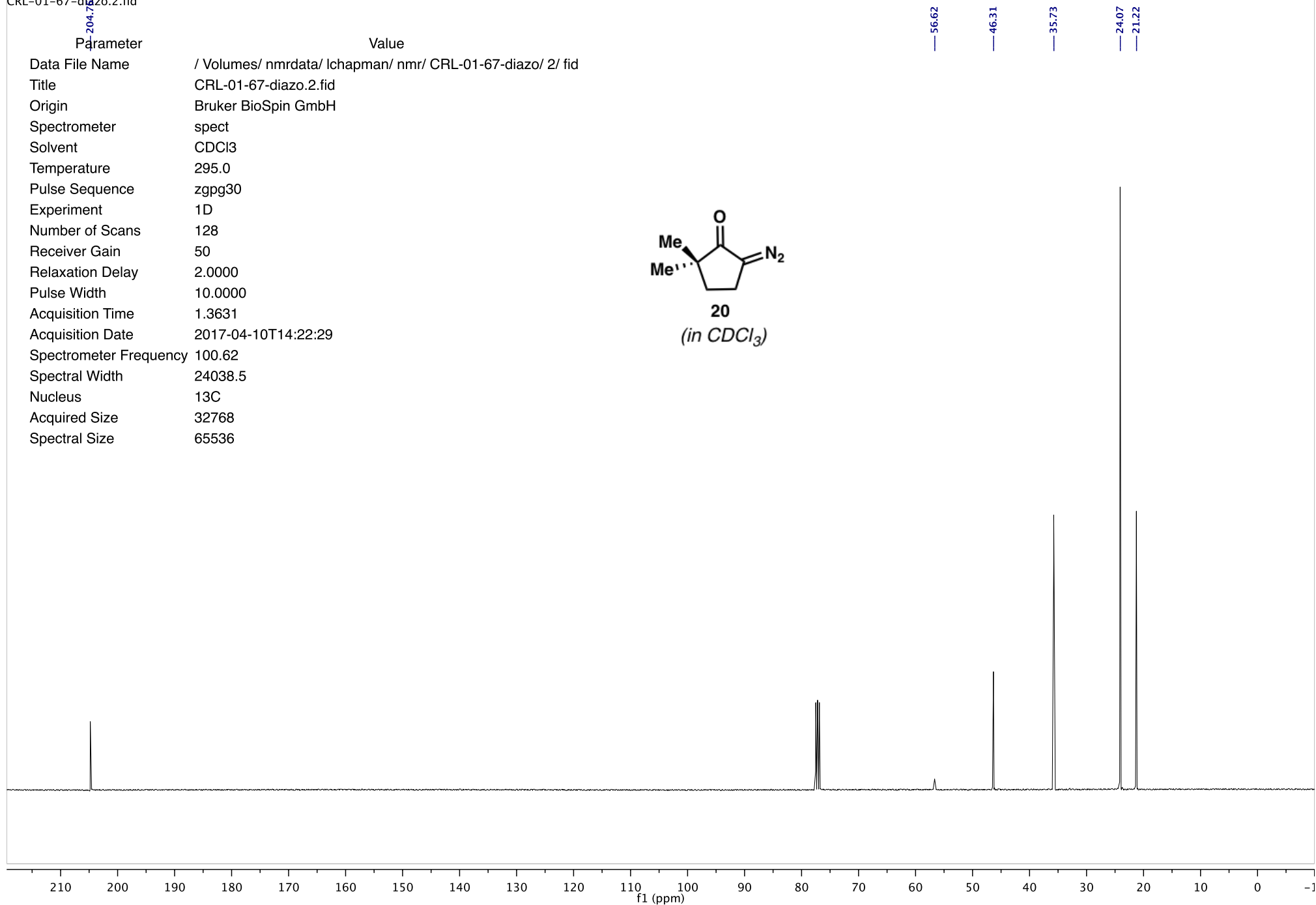
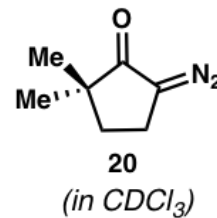
- (1) Isolation of psiguadials A and B: Shao, M.; Wang, Y.; Liu, Z.; Zhang, D.-M.; Cao, H.-H.; Jiang, R.-W.; Fan, C.-L.; Zhang, X.-Q.; Chen, H.-R.; Yao, X.-S.; Ye, W.-C. *Org. Lett.* **2010**, *12*, 5040. Except where designated, multiplicities are not specified.
- (2) Shao et al. report the two protons at 7.23 ppm as corresponding to the carbon signal at 126.2 ppm (carbons 10' and 12'). This is assumed to be a typographical error based on the reported plot of their HSQC spectrum and our own data showing a correlation between the proton signal at 7.26 ppm and the more downfield carbon shift at 128.2 ppm, which corresponds to carbons 9' and 13'. The remaining three aromatic proton signals are adjusted accordingly in the comparison list.
- (3) The center peak of CDCl₃ is referenced to 76.999 ppm, as shown in the Shao et al. report, see ¹³C spectral comparison on page S18.
- (4) APEX2, Version 2 User Manual, M86-E01078, Bruker Analytical X-ray Systems, Madison, WI, June 2006.
- (5) Sheldrick, G.M. "*SADABS (version 2008/1): Program for Absorption Correction for Data from Area Detector Frames*", University of Göttingen, 2008.
- (6) Sheldrick, G. M. *Acta Crystallogr A* **2008**, *64*, 112.
- (7) Müller, P. *Crystallogr. Rev.* **2009**, *15*, 57.
- (8) Parsons, S.; Flack, H. D.; Wagner, T. *Acta Cryst.* **2013**, *69*, 249.
- (9) Macrae, C. F.; Edgington, P. R.; McCabe, P.; Pidcock, E.; Shields, G. P.; Taylor, R.; Towler M.; van de Streek, J. *J. Appl. Cryst.* **2006**, *39*, 453.

CRL-01-67-diazo.1.fid

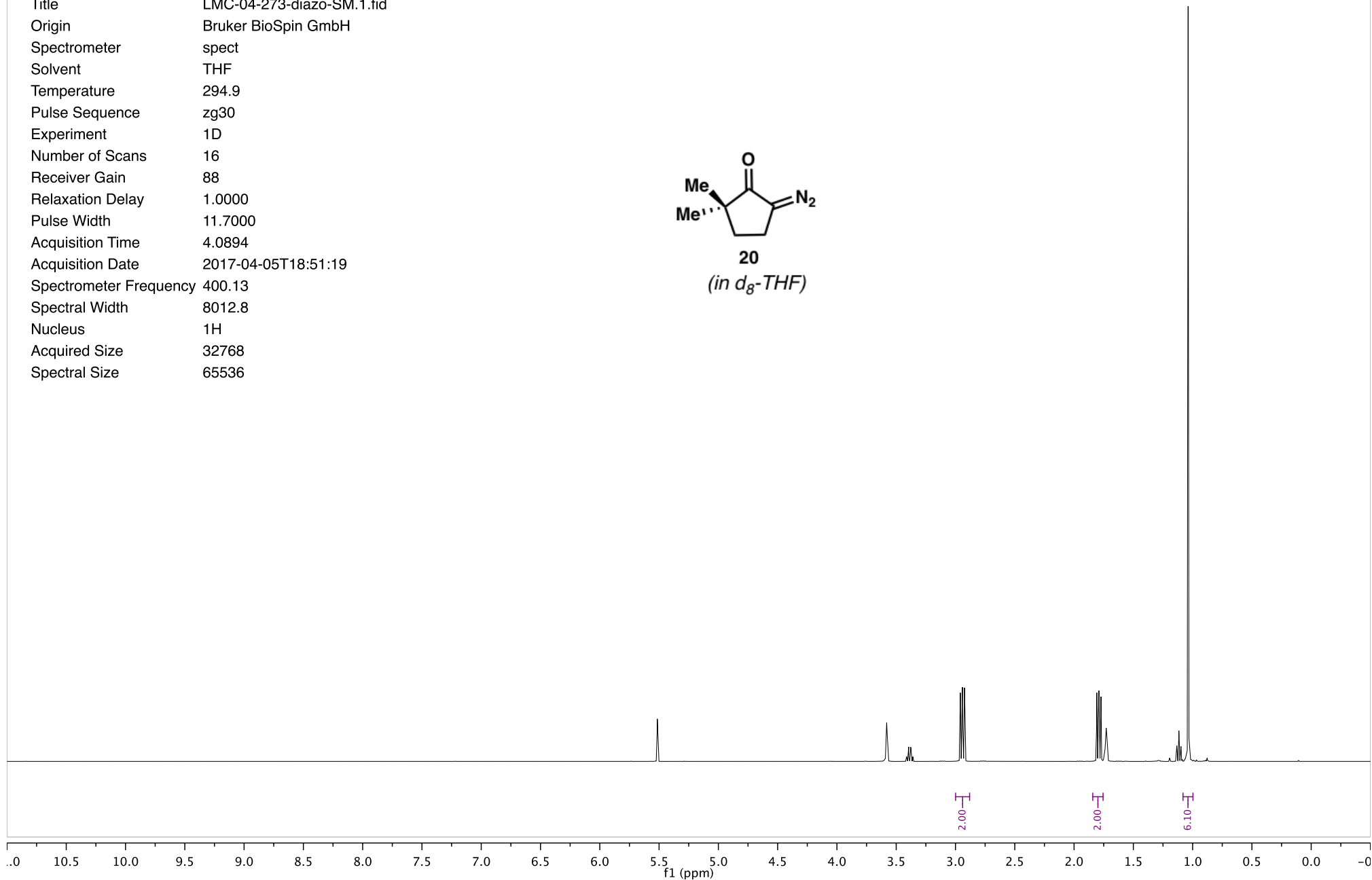
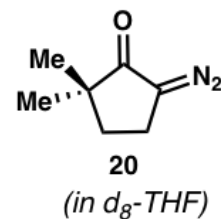
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Origin	Bruker BioSpin GmbH
Spectrometer	spect
Solvent	CDCl ₃
Temperature	295.0
Pulse Sequence	zg30
Experiment	1D
Number of Scans	1
Receiver Gain	14
Relaxation Delay	1.0000
Pulse Width	11.7000
Acquisition Time	4.0894
Acquisition Date	2017-04-10T14:06:42
Spectrometer Frequency	400.13
Spectral Width	8012.8
Nucleus	¹ H
Acquired Size	32768
Spectral Size	65536



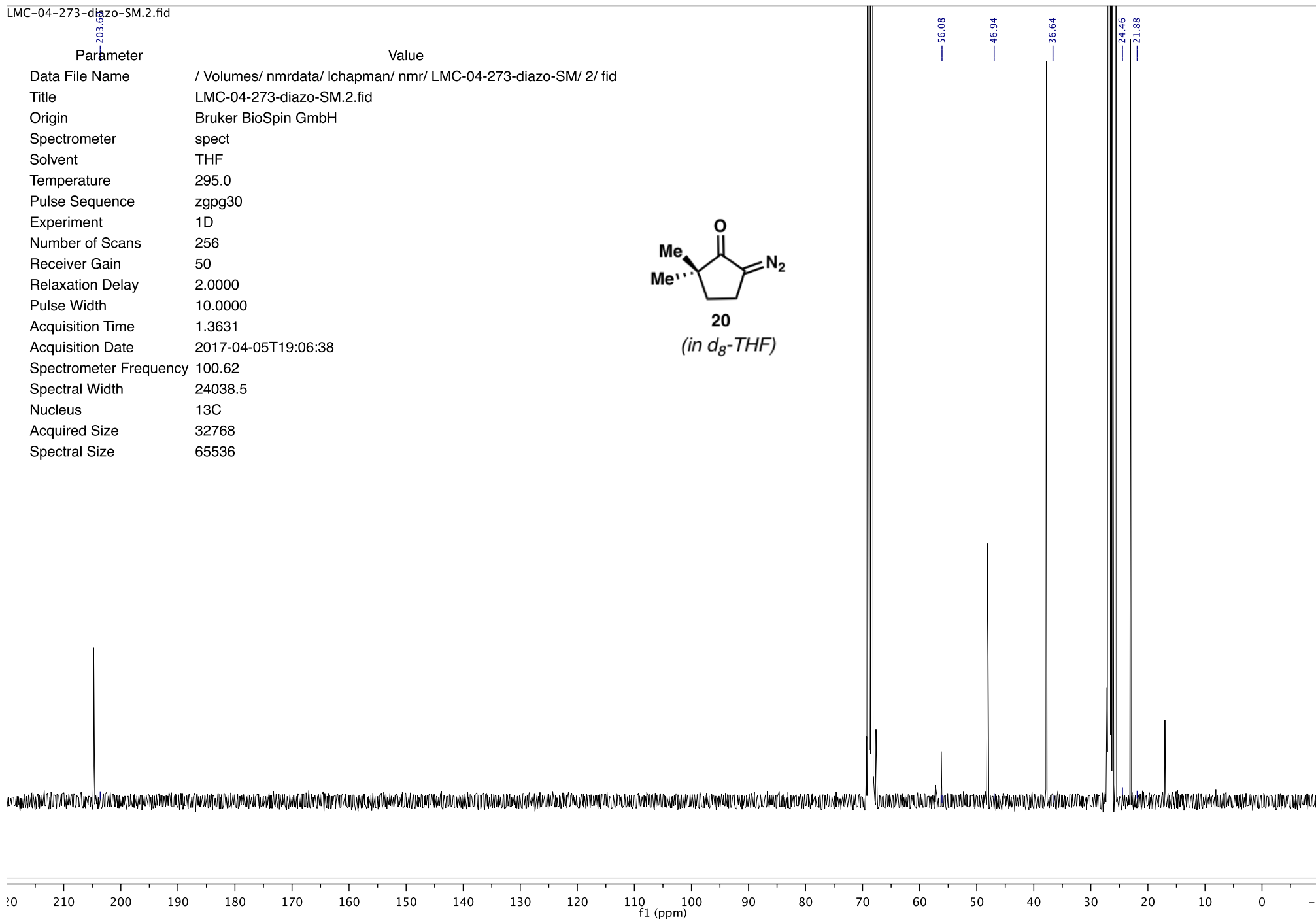
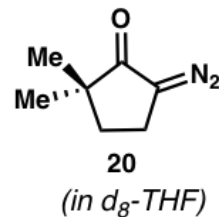
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Title	CRL-01-67-diazo.2.fid
Origin	Bruker BioSpin GmbH
Spectrometer	spect
Solvent	CDCl ₃
Temperature	295.0
Pulse Sequence	zgpg30
Experiment	1D
Number of Scans	128
Receiver Gain	50
Relaxation Delay	2.0000
Pulse Width	10.0000
Acquisition Time	1.3631
Acquisition Date	2017-04-10T14:22:29
Spectrometer Frequency	100.62
Spectral Width	24038.5
Nucleus	¹³ C
Acquired Size	32768
Spectral Size	65536



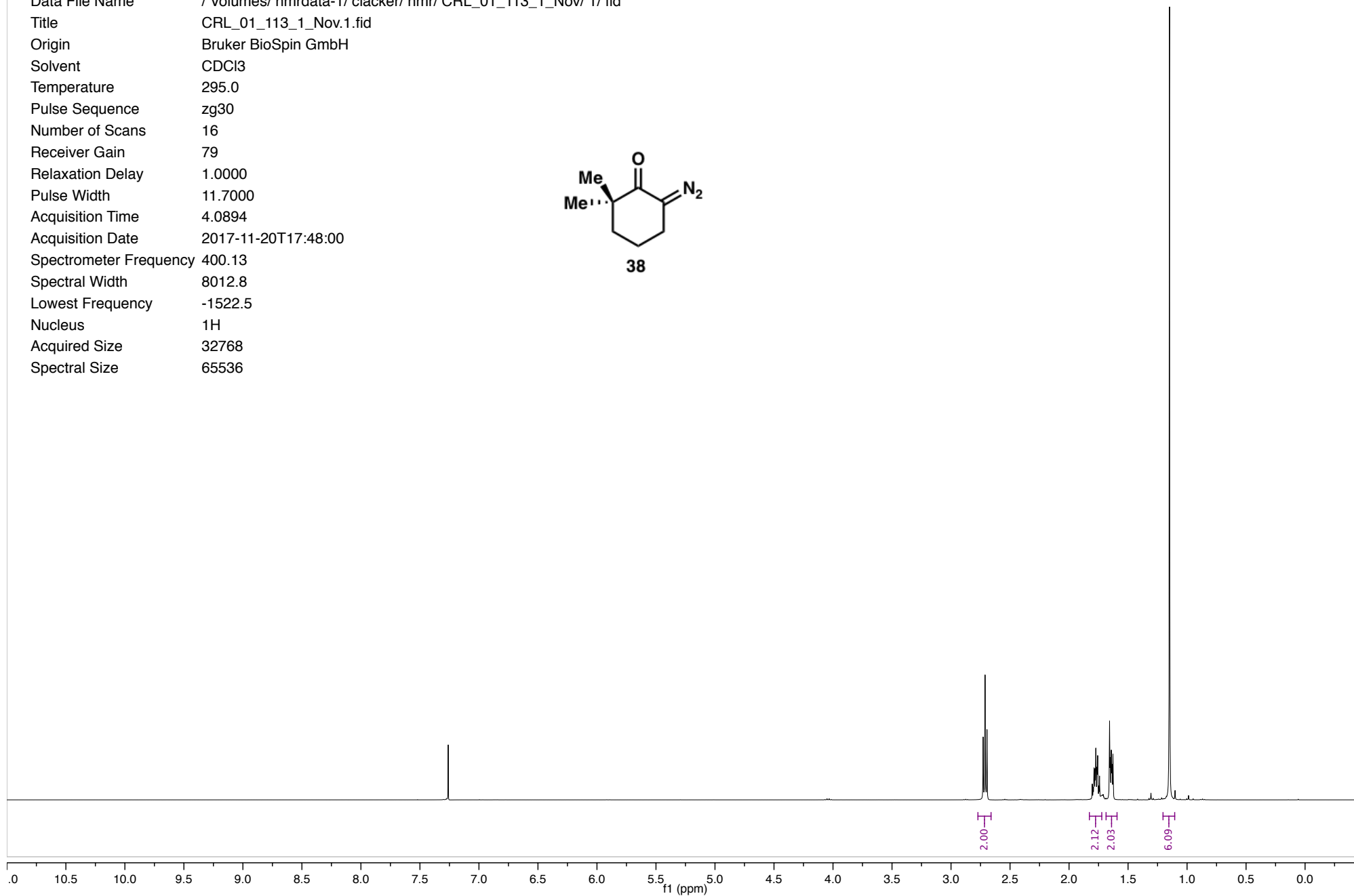
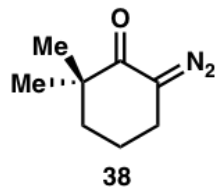
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Origin	Bruker BioSpin GmbH
Spectrometer	spect
Solvent	THF
Temperature	294.9
Pulse Sequence	zg30
Experiment	1D
Number of Scans	16
Receiver Gain	88
Relaxation Delay	1.0000
Pulse Width	11.7000
Acquisition Time	4.0894
Acquisition Date	2017-04-05T18:51:19
Spectrometer Frequency	400.13
Spectral Width	8012.8
Nucleus	¹ H
Acquired Size	32768
Spectral Size	65536



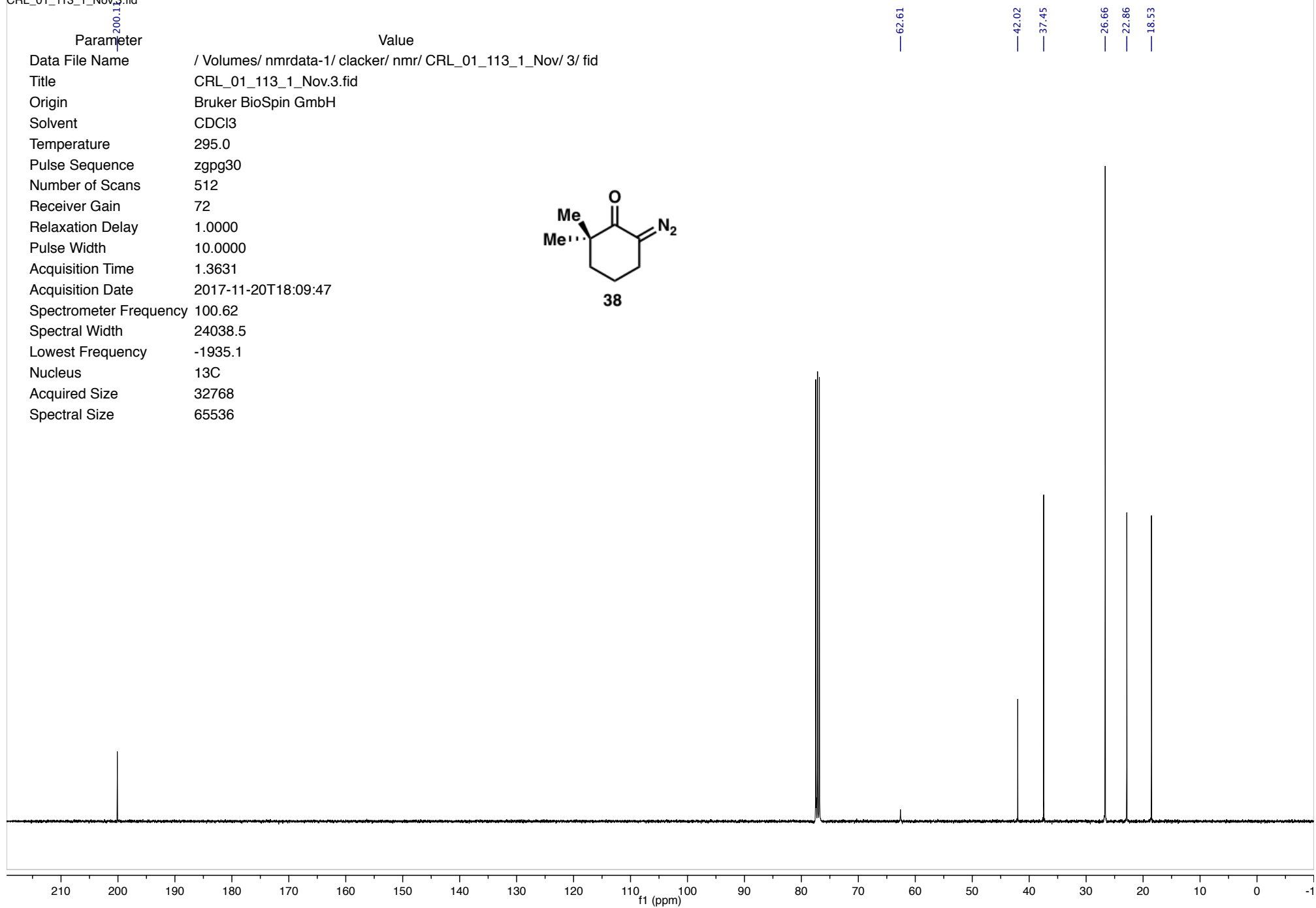
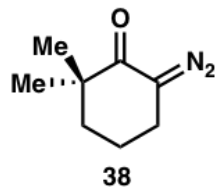
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Origin	Bruker BioSpin GmbH
Spectrometer	spect
Solvent	THF
Temperature	295.0
Pulse Sequence	zgpg30
Experiment	1D
Number of Scans	256
Receiver Gain	50
Relaxation Delay	2.0000
Pulse Width	10.0000
Acquisition Time	1.3631
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Spectrometer Frequency	100.62
Spectral Width	24038.5
Nucleus	13C
Acquired Size	32768
Spectral Size	65536



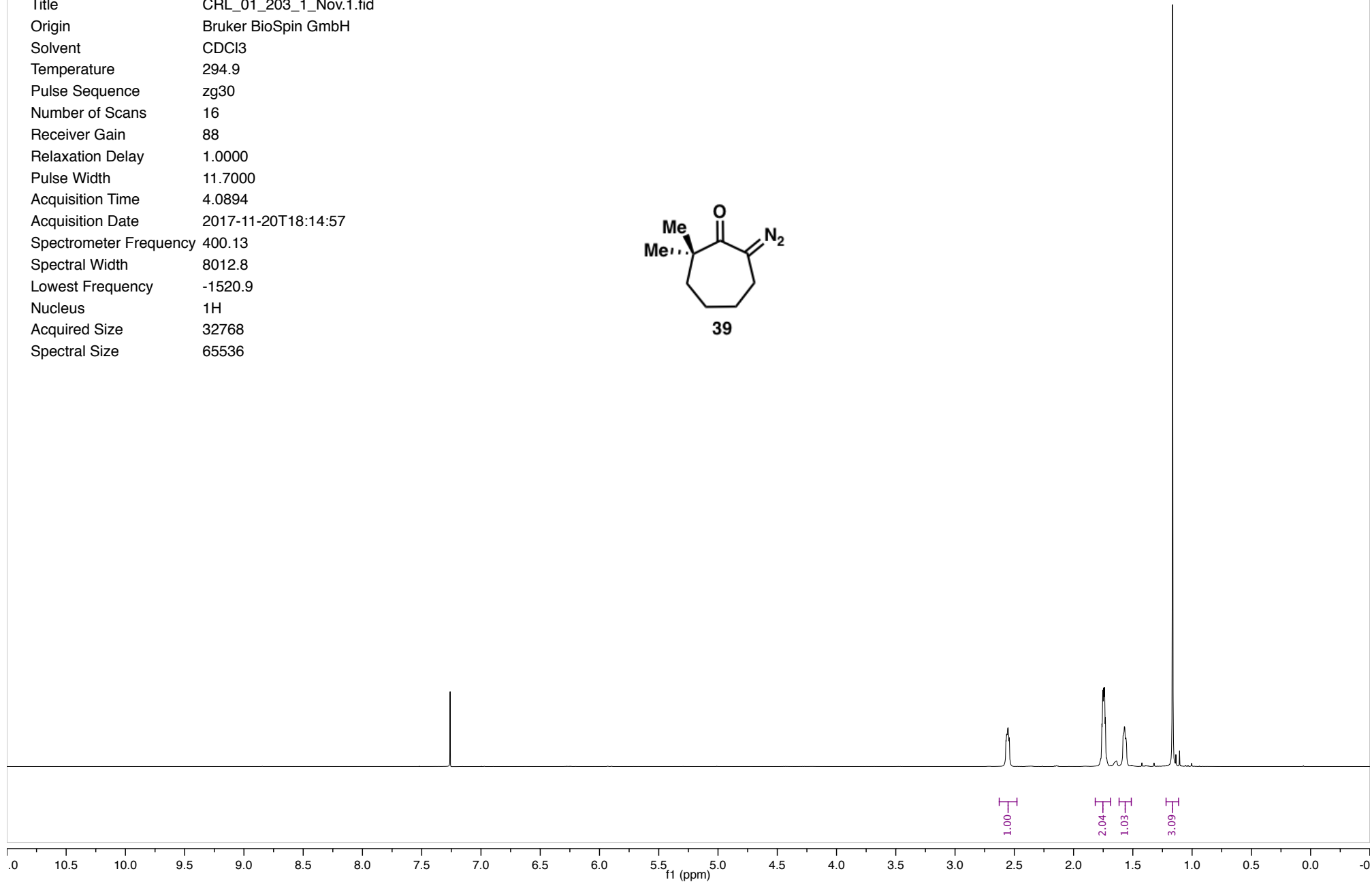
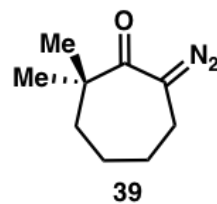
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Origin	Bruker BioSpin GmbH
Solvent	CDCl3
Temperature	295.0
Pulse Sequence	zg30
Number of Scans	16
Receiver Gain	79
Relaxation Delay	1.0000
Pulse Width	11.7000
Acquisition Time	4.0894
Acquisition Date	2017-11-20T17:48:00
Spectrometer Frequency	400.13
Spectral Width	8012.8
Lowest Frequency	-1522.5
Nucleus	1H
Acquired Size	32768
Spectral Size	65536



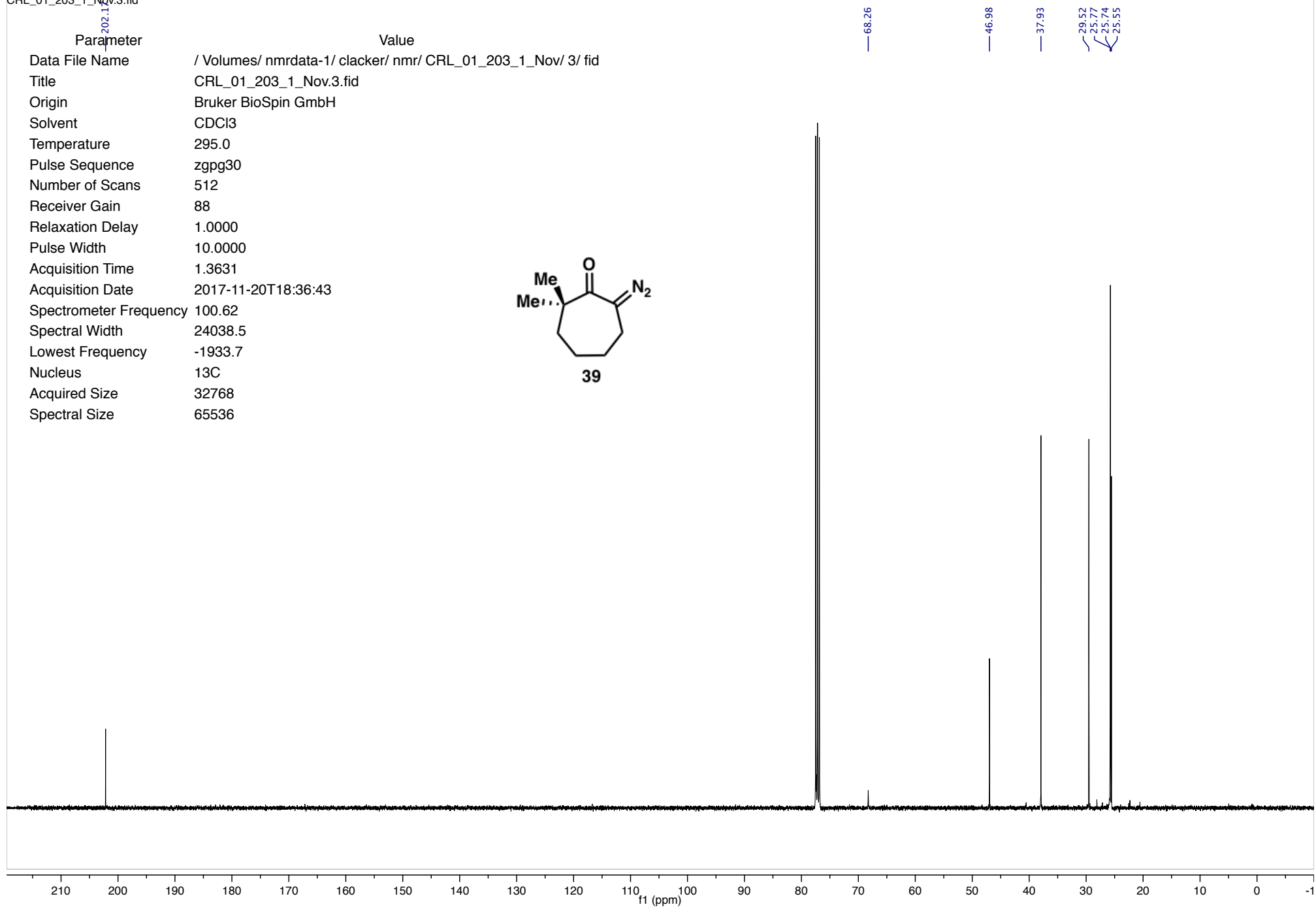
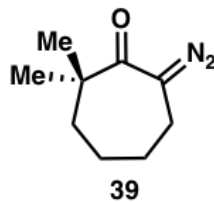
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Origin	Bruker BioSpin GmbH
Solvent	CDCl3
Temperature	295.0
Pulse Sequence	zgpg30
Number of Scans	512
Receiver Gain	72
Relaxation Delay	1.0000
Pulse Width	10.0000
Acquisition Time	1.3631
Acquisition Date	2017-11-20T18:09:47
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Lowest Frequency	-1935.1
Nucleus	13C
Acquired Size	32768
Spectral Size	65536



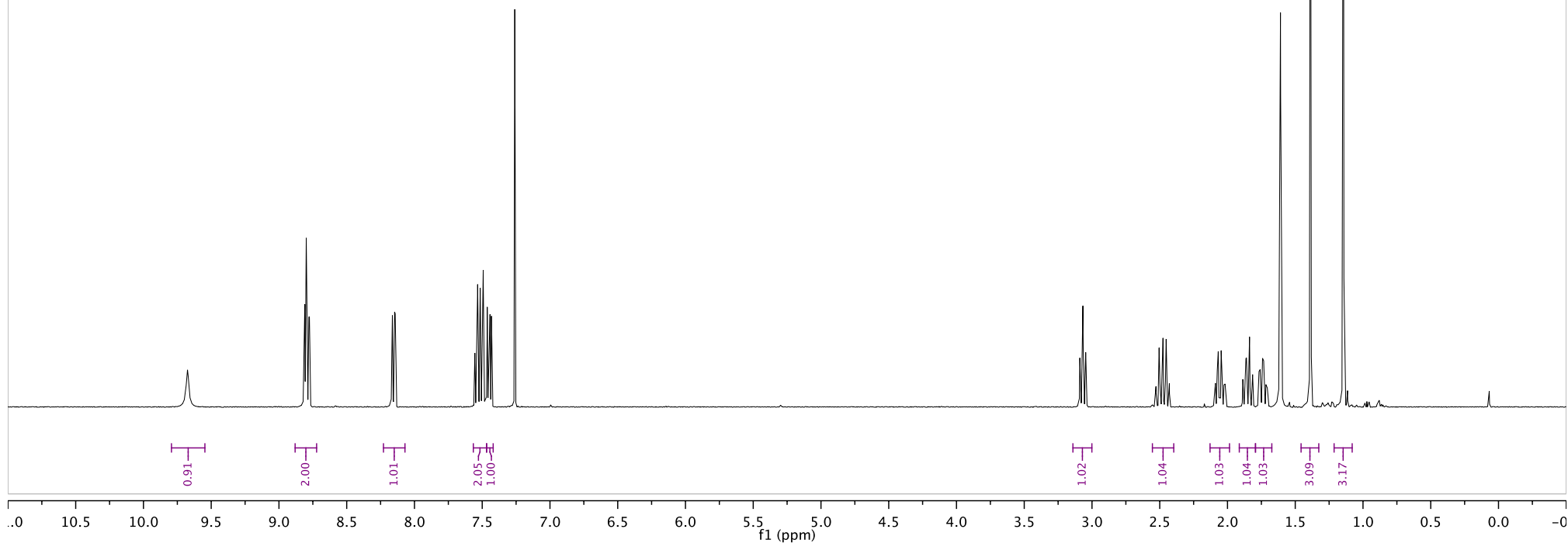
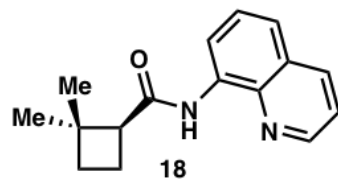
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Origin	Bruker BioSpin GmbH
Solvent	CDCl3
Temperature	294.9
Pulse Sequence	zg30
Number of Scans	16
Receiver Gain	88
Relaxation Delay	1.0000
Pulse Width	11.7000
Acquisition Time	4.0894
Acquisition Date	2017-11-20T18:14:57
Spectrometer Frequency	400.13
Spectral Width	8012.8
Lowest Frequency	-1520.9
Nucleus	1H
Acquired Size	32768
Spectral Size	65536



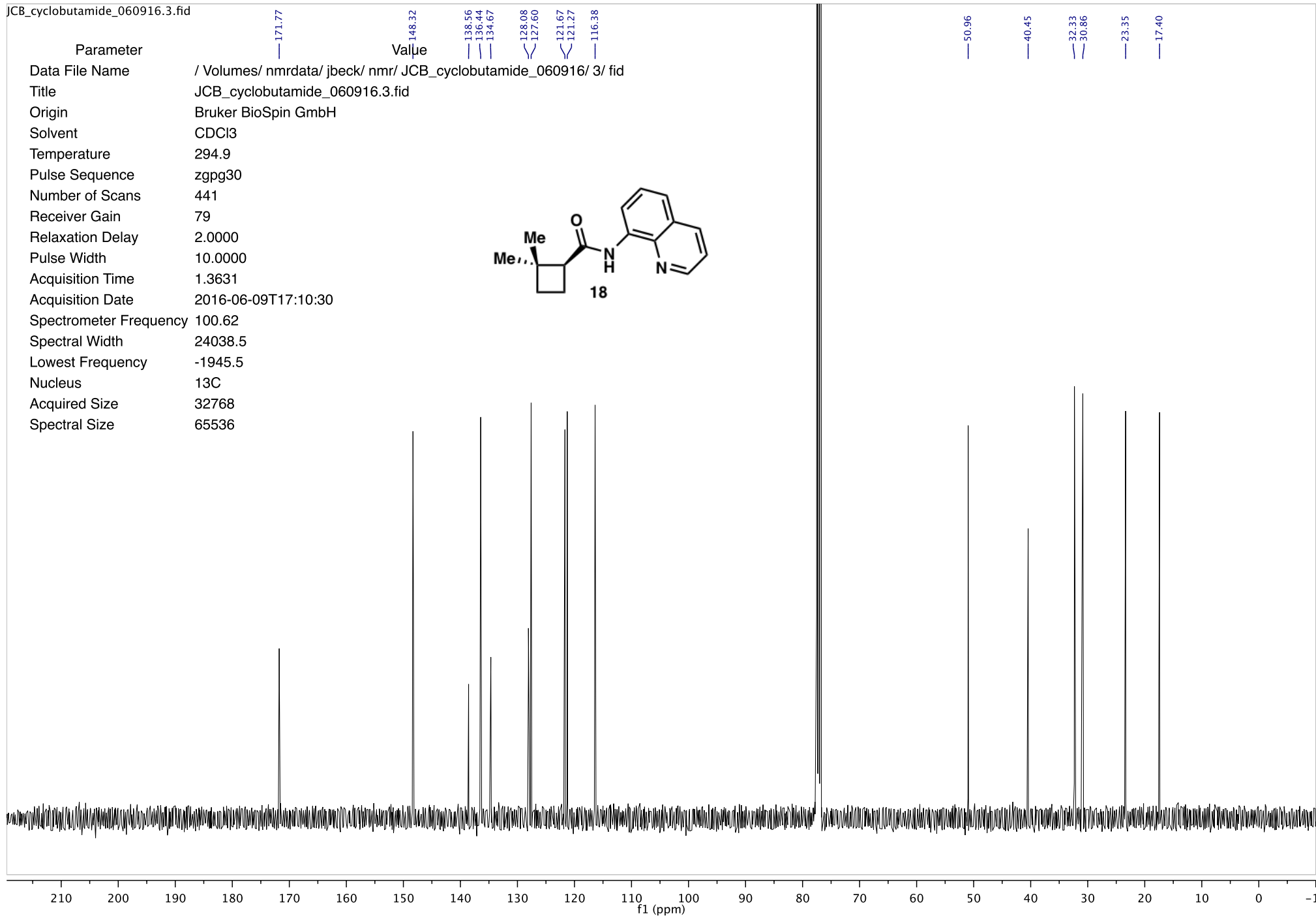
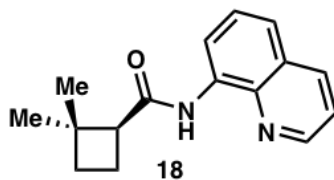
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Origin	Bruker BioSpin GmbH
Solvent	CDCl ₃
Temperature	295.0
Pulse Sequence	zgpg30
Number of Scans	512
Receiver Gain	88
Relaxation Delay	1.0000
Pulse Width	10.0000
Acquisition Time	1.3631
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Spectral Width	24038.5
Lowest Frequency	-1933.7
Nucleus	¹³ C
Acquired Size	32768
Spectral Size	65536



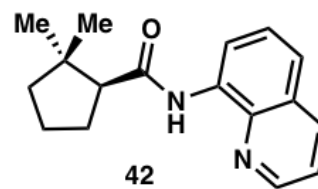
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Origin	Bruker BioSpin GmbH
Solvent	CDCl3
Temperature	295.0
Pulse Sequence	zg30
Number of Scans	1
Receiver Gain	197
Relaxation Delay	1.0000
Pulse Width	11.7000
Acquisition Time	4.0894
Acquisition Date	2016-06-09T16:38:44
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Lowest Frequency	-1545.6
Nucleus	1H
Acquired Size	32768
Spectral Size	65536



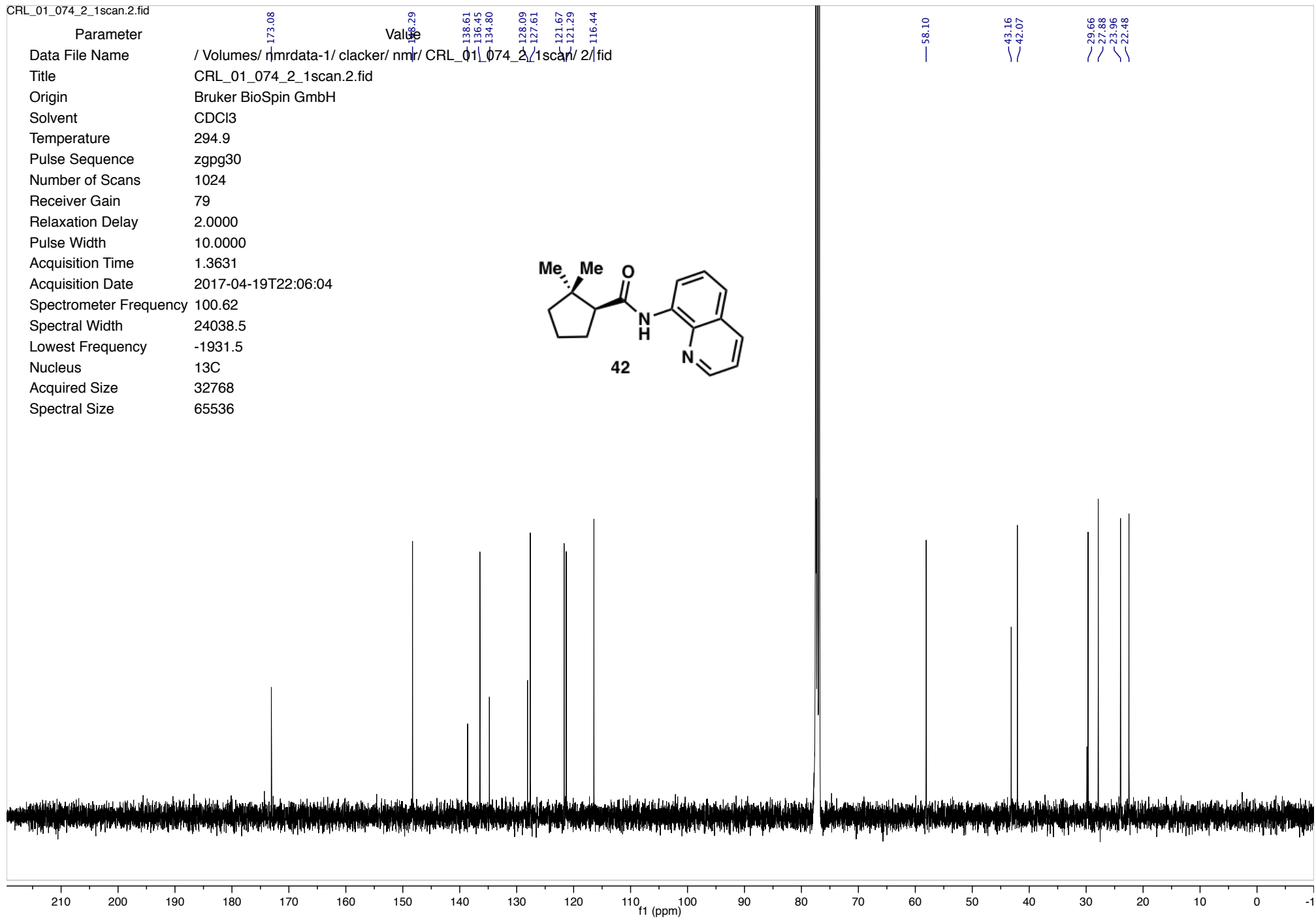
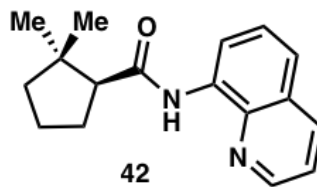
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Solvent	CDCl3
Temperature	294.9
Pulse Sequence	zgpg30
Number of Scans	441
Receiver Gain	79
Relaxation Delay	2.0000
Pulse Width	10.0000
Acquisition Time	1.3631
Acquisition Date	2016-06-09T17:10:30
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Spectral Width	24038.5
Lowest Frequency	-1945.5
Nucleus	13C
Acquired Size	32768
Spectral Size	65536



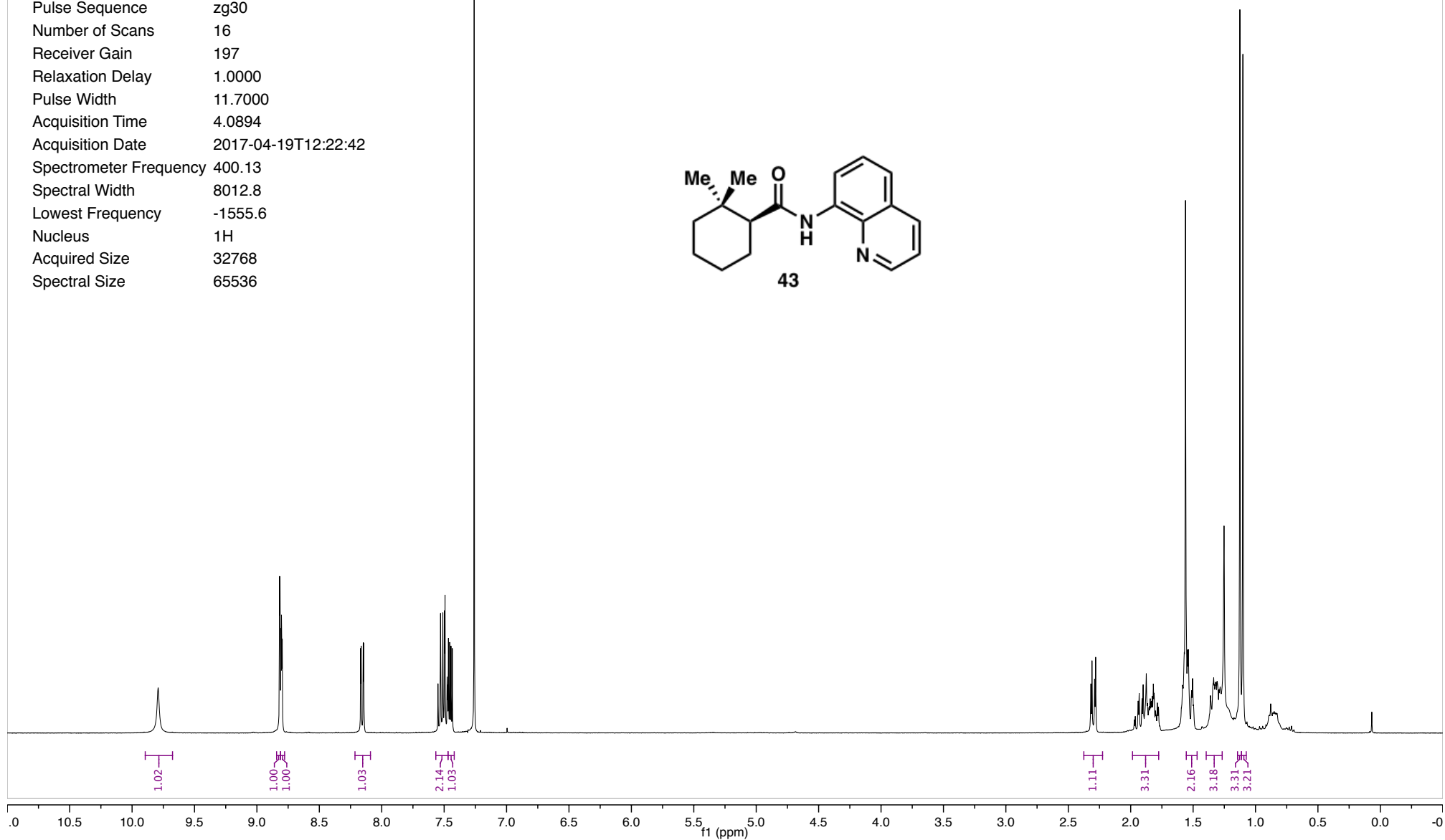
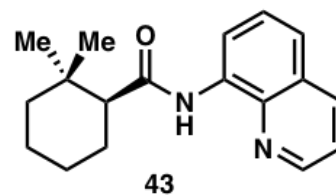
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Origin	Bruker BioSpin GmbH
Solvent	CDCl3
Temperature	294.9
Pulse Sequence	zg30
Number of Scans	1
Receiver Gain	197
Relaxation Delay	1.0000
Pulse Width	11.7000
Acquisition Time	4.0894
Acquisition Date	2017-04-19T21:04:33
Spectrometer Frequency	400.13
Spectral Width	8012.8
Lowest Frequency	-1555.6
Nucleus	1H
Acquired Size	32768
Spectral Size	65536



Parameter	Value
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Solvent	CDCl3
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Pulse Sequence	zgpg30
Number of Scans	1024
Receiver Gain	79
Relaxation Delay	2.0000
Pulse Width	10.0000
Acquisition Time	1.3631
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Lowest Frequency	-1931.5
Nucleus	¹³ C
Acquired Size	32768
Spectral Size	65536

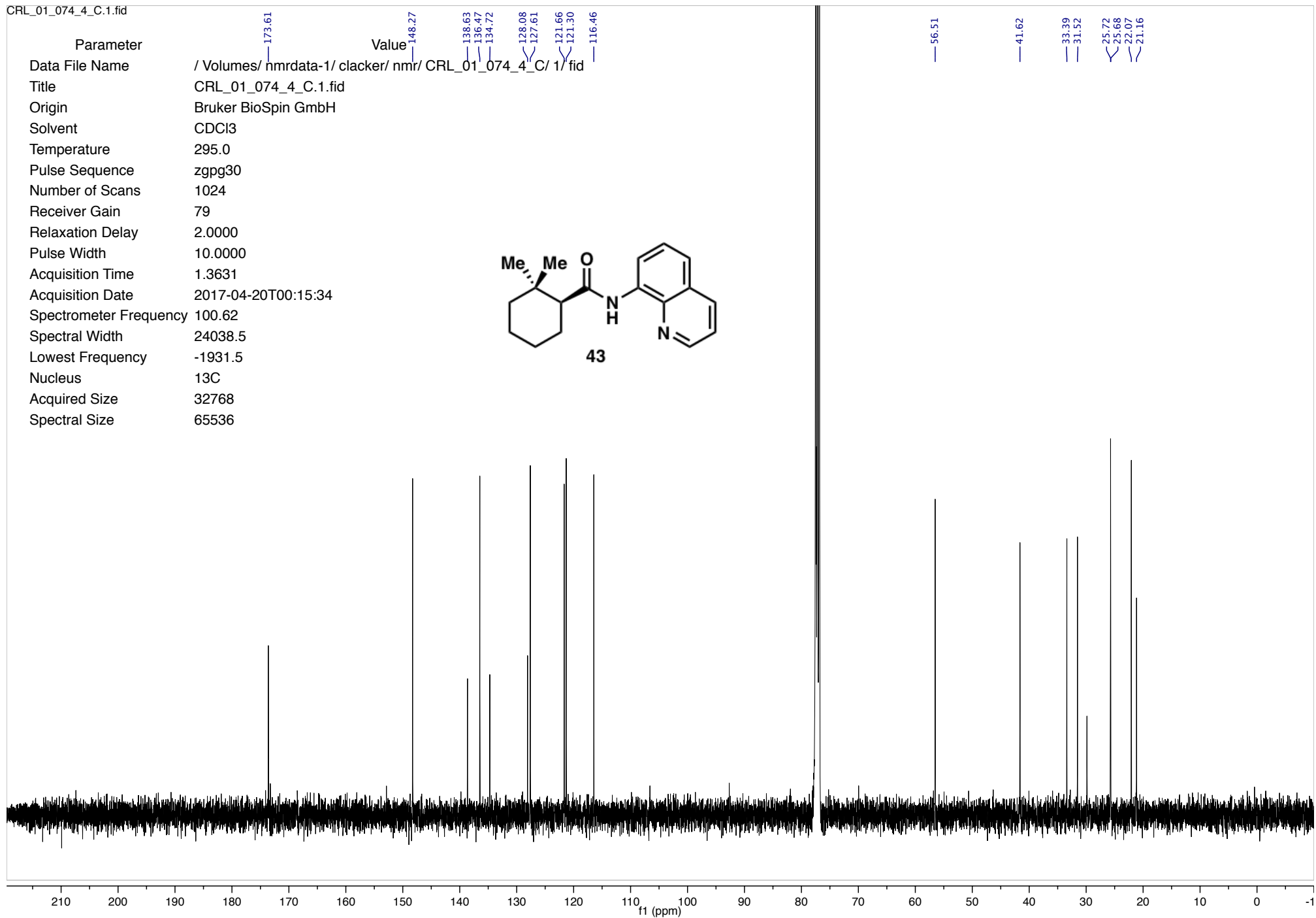
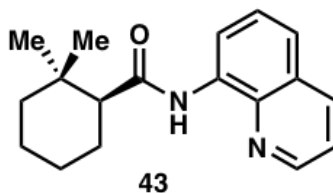


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Origin	Bruker BioSpin GmbH
Solvent	CDCl3
Temperature	295.0
Pulse Sequence	zg30
Number of Scans	16
Receiver Gain	197
Relaxation Delay	1.0000
Pulse Width	11.7000
Acquisition Time	4.0894
Acquisition Date	2017-04-19T12:22:42
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Spectral Width	8012.8
Lowest Frequency	-1555.6
Nucleus	1H
Acquired Size	32768
Spectral Size	65536

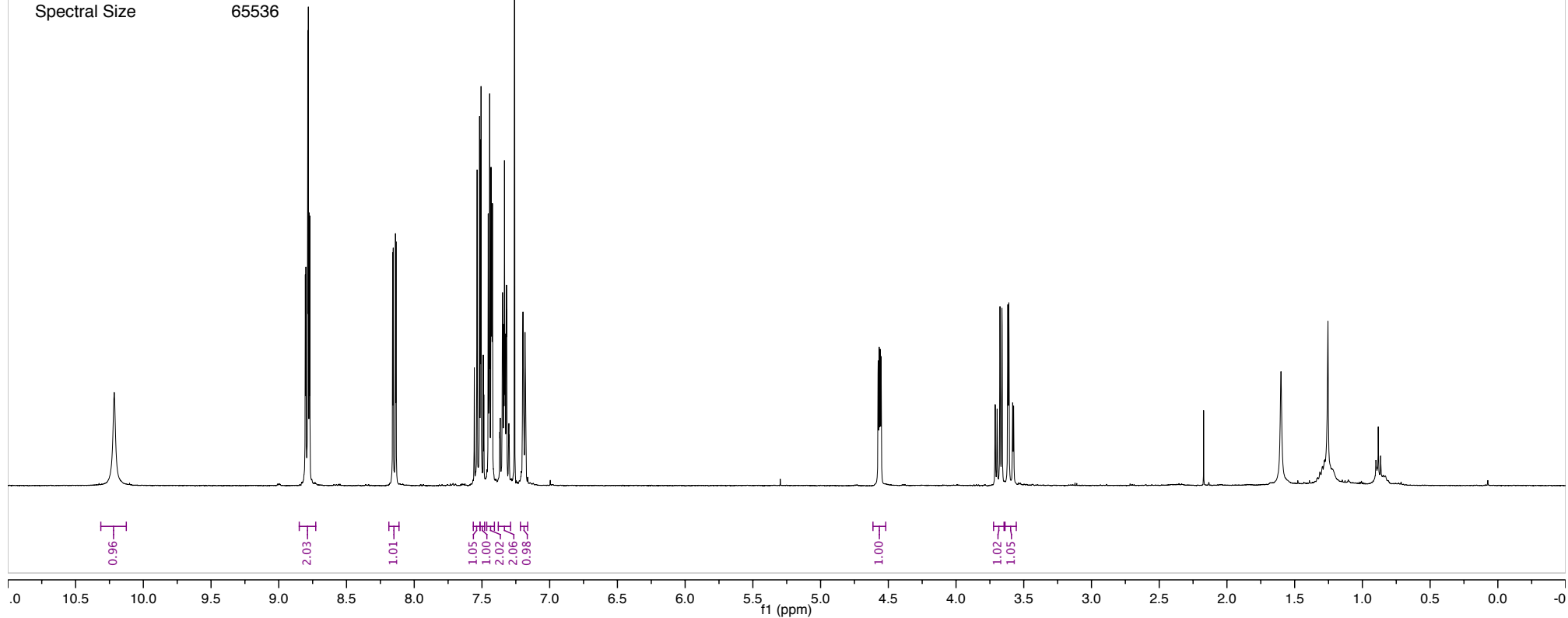
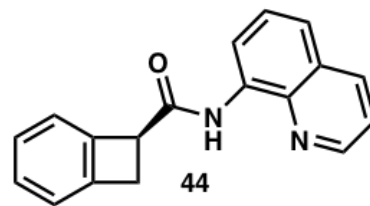


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Solvent	CDCl ₃
Temperature	295.0
Pulse Sequence	zgpg30
Number of Scans	1024
Receiver Gain	79
Relaxation Delay	2.0000
Pulse Width	10.0000
Acquisition Time	1.3631
Acquisition Date	2017-04-20T00:15:34
Spectrometer Frequency	100.62
Spectral Width	24038.5
Lowest Frequency	-1931.5
Nucleus	¹³ C
Acquired Size	32768
Spectral Size	65536



Parameter	Value
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Origin	Bruker BioSpin GmbH
Solvent	CDCl3
Temperature	295.0
Pulse Sequence	zg30
Number of Scans	1
Receiver Gain	197
Relaxation Delay	1.0000
Pulse Width	11.7000
Acquisition Time	4.0894
Acquisition Date	2017-06-12T17:08:01
Spectrometer Frequency	400.13
Spectral Width	8012.8
Lowest Frequency	-1555.6
Nucleus	¹ H
Acquired Size	32768
Spectral Size	65536



Parameter

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Solvent	CDCl3
Temperature	295.0
Pulse Sequence	zgpg30
Number of Scans	1024
Receiver Gain	72
Relaxation Delay	2.0000
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Acquisition Time	1.3631
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Lowest Frequency	-1933.9
Nucleus	¹³ C
Acquired Size	32768
Spectral Size	65536

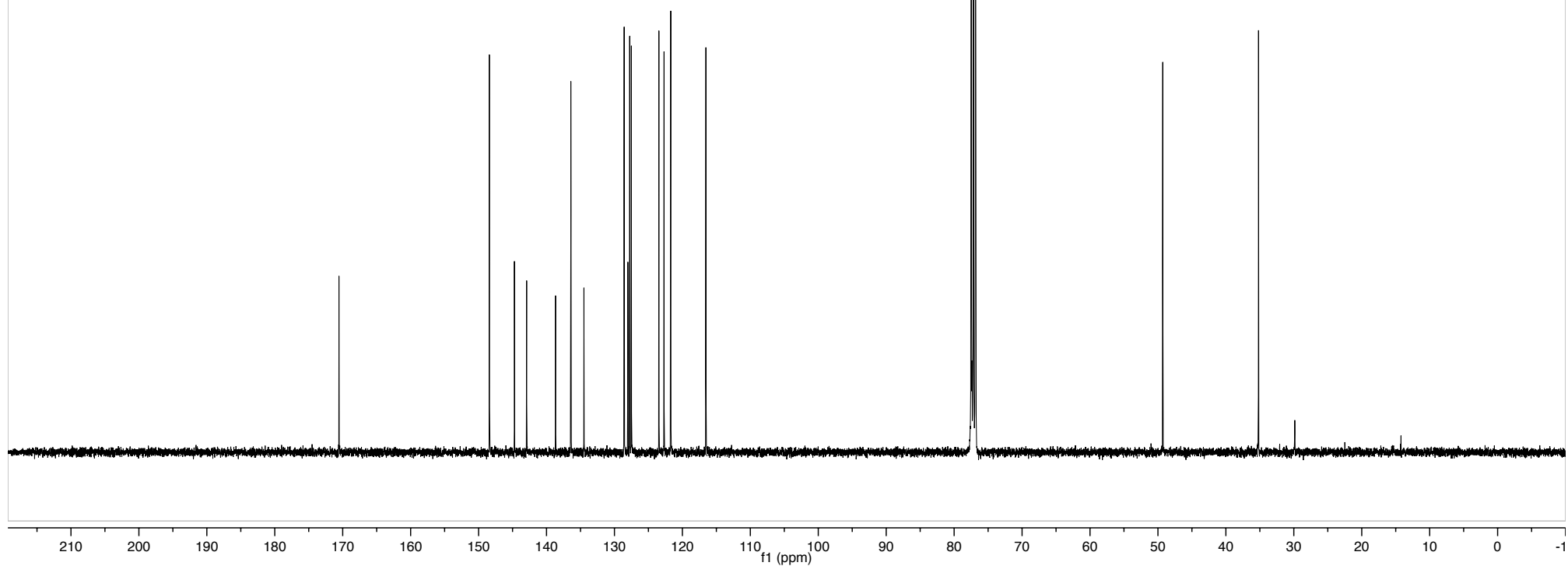
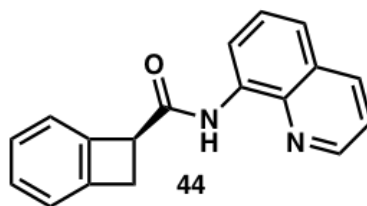
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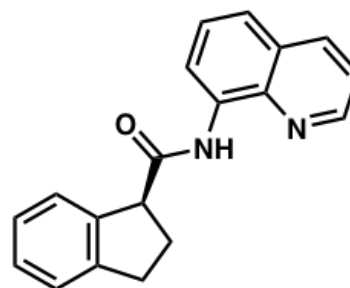
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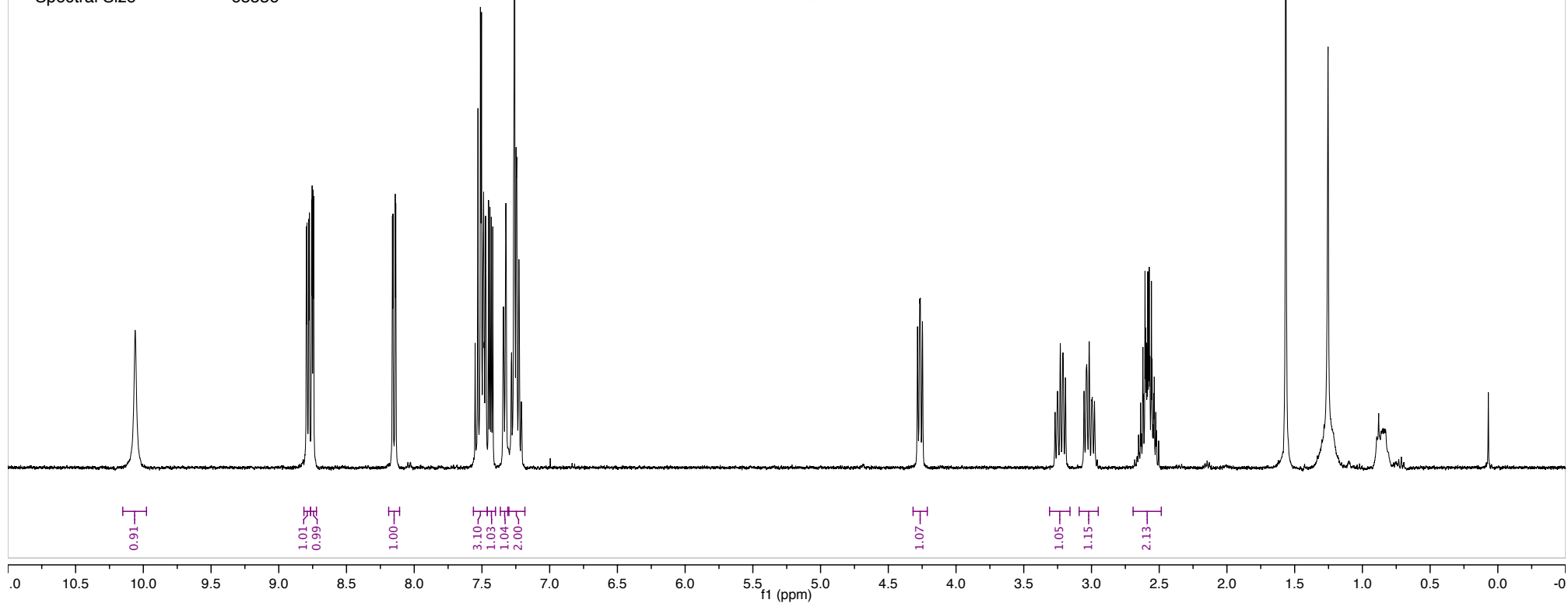


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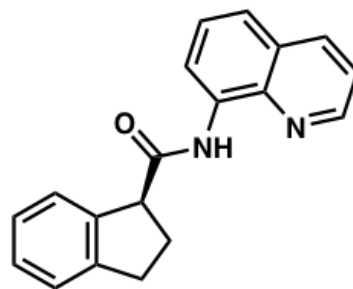
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Origin	Bruker BioSpin GmbH
Solvent	CDCl3
Temperature	295.0
Pulse Sequence	zg30
Number of Scans	1
Receiver Gain	197
Relaxation Delay	1.0000
Pulse Width	11.7000
Acquisition Time	4.0894
Acquisition Date	2017-04-19T22:10:20
Spectrometer Frequency	400.13
Spectral Width	8012.8
Lowest Frequency	-1555.6
Nucleus	1H
Acquired Size	32768
Spectral Size	65536



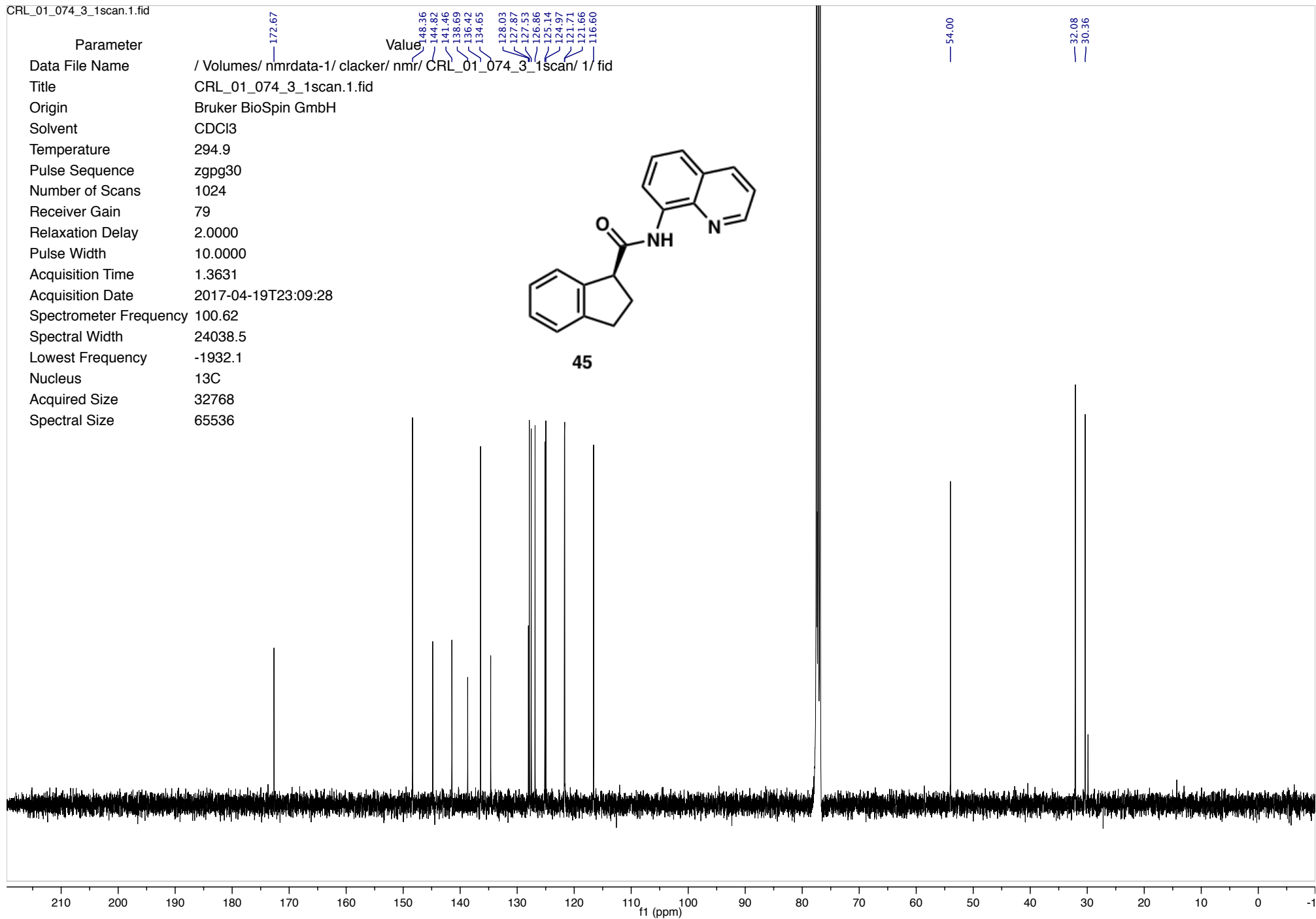
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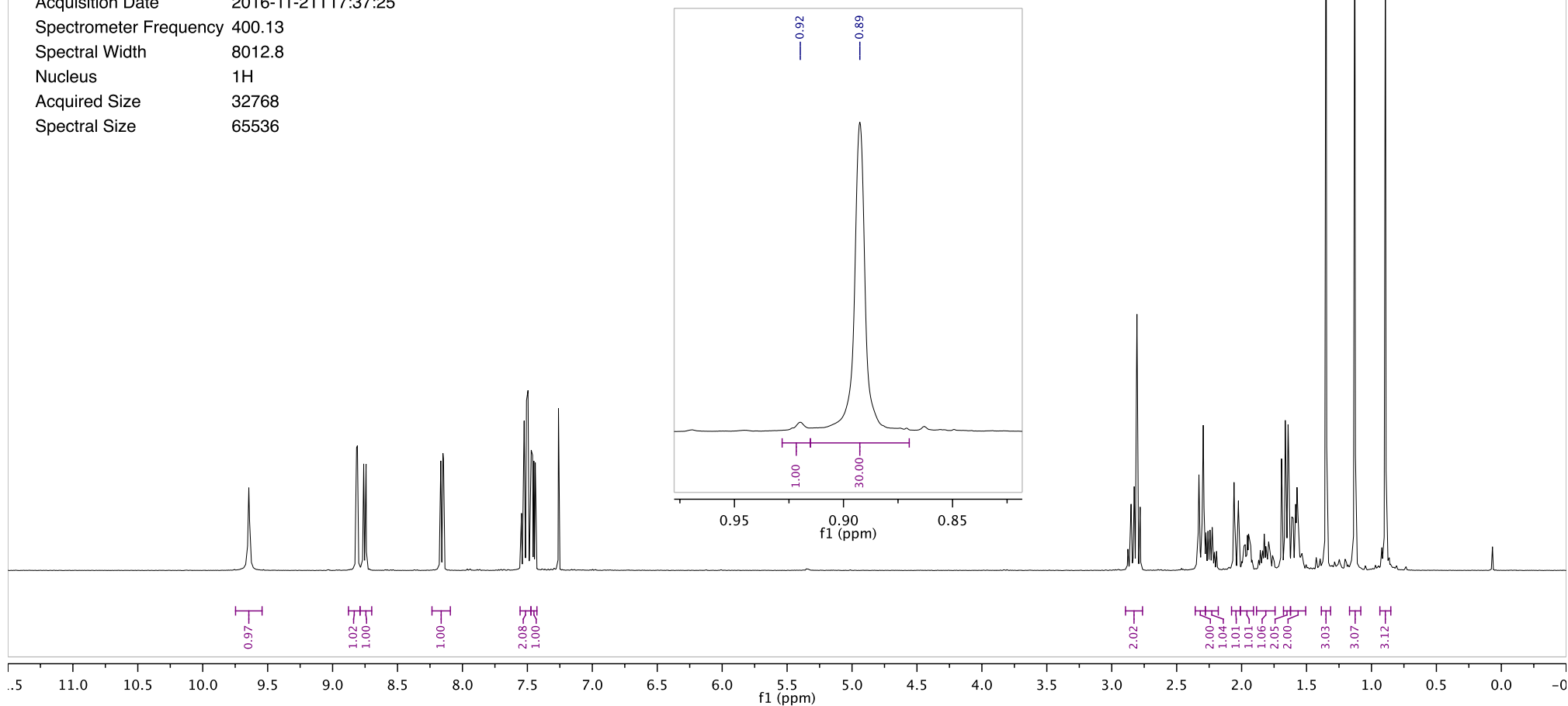
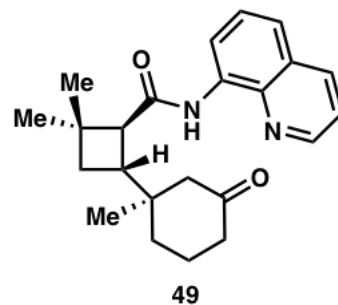
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Origin	Bruker BioSpin GmbH
Solvent	CDCl3
Temperature	294.9
Pulse Sequence	zgpg30
Number of Scans	1024
Receiver Gain	79
Relaxation Delay	2.0000
Pulse Width	10.0000
Acquisition Time	1.3631
Acquisition Date	2017-04-19T23:09:28
Spectrometer Frequency	100.62
Spectral Width	24038.5
Lowest Frequency	-1932.1
Nucleus	¹³ C
Acquired Size	32768
Spectral Size	65536



45

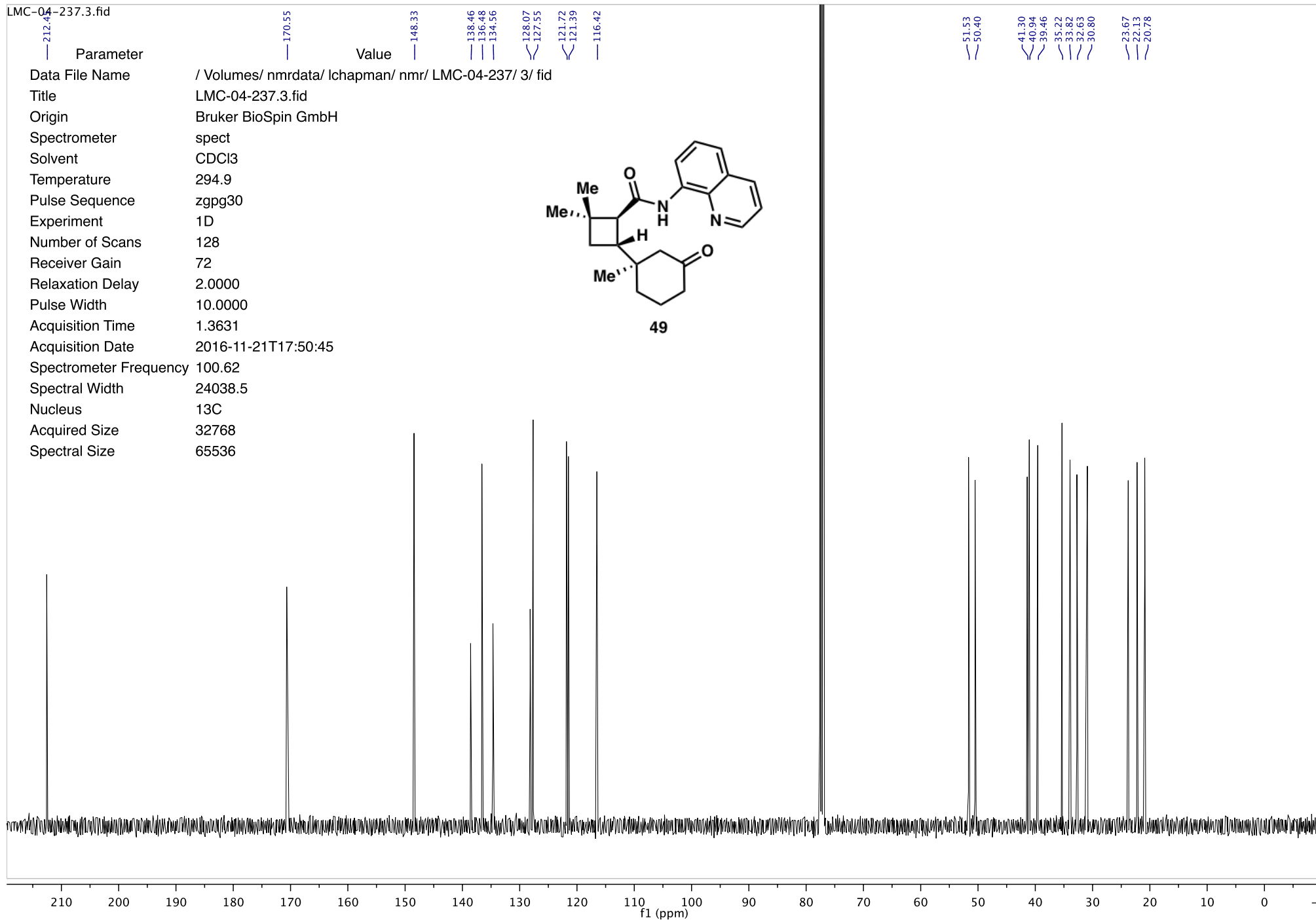
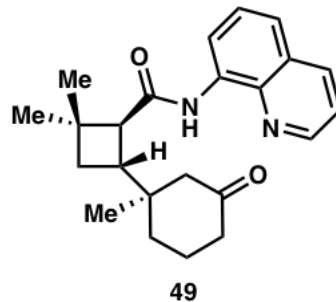


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Data File Name	/ Volumes/ nmrdata/ lchapman/ nmr/ LMC-04-237/ 1/ fid
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Spectrometer	spect
Solvent	CDCl3
Temperature	294.9
Pulse Sequence	zg30
Experiment	1D
Number of Scans	1
Receiver Gain	79
Relaxation Delay	1.0000
Pulse Width	11.7000
Acquisition Time	4.0894
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Spectrometer Frequency	400.13
Spectral Width	8012.8
Nucleus	1H
Acquired Size	32768
Spectral Size	65536

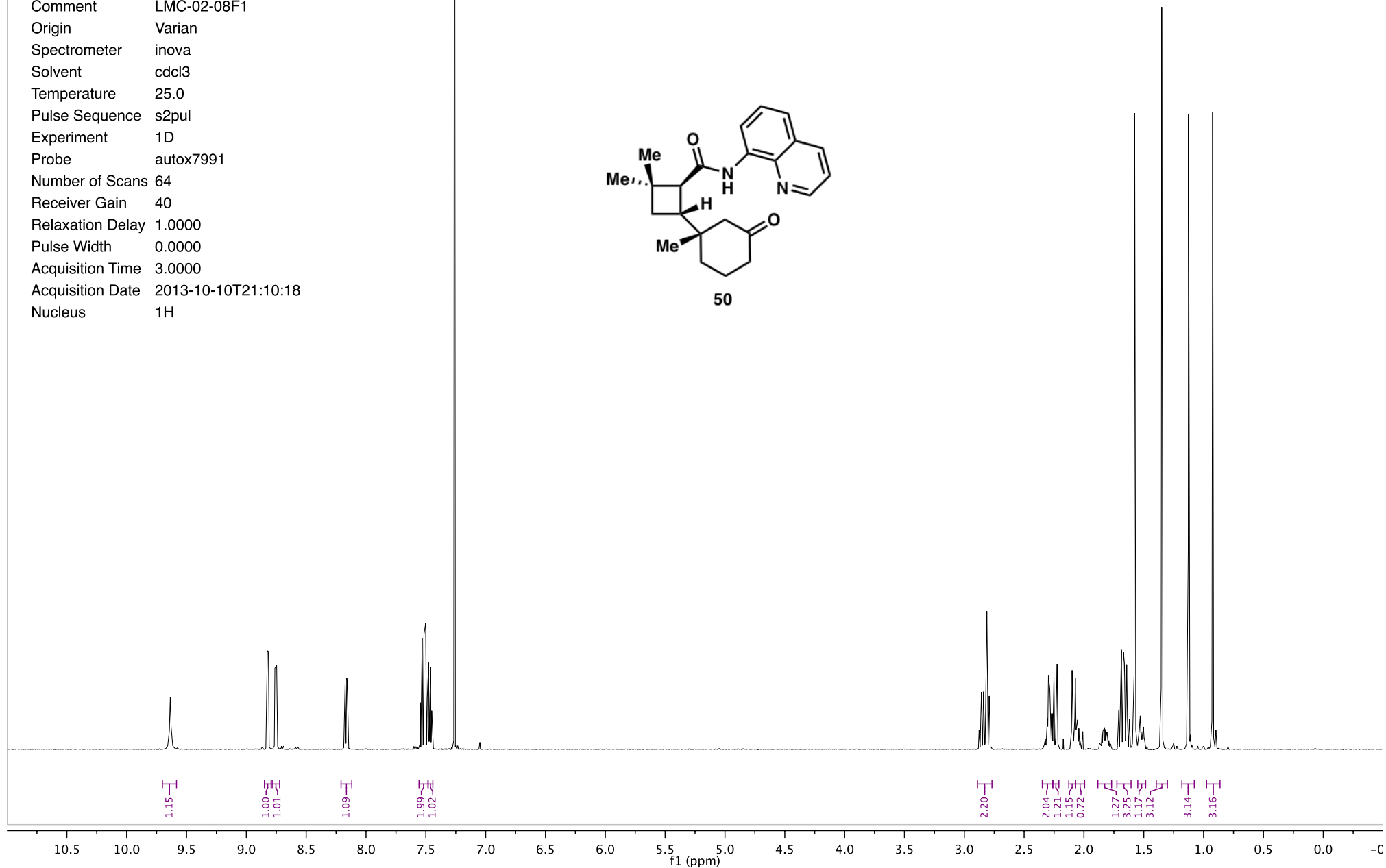
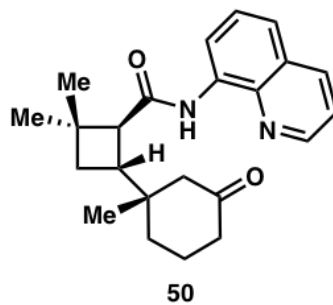


LMC-04-237.3.fid

Parameter	Value
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Origin	Bruker BioSpin GmbH
Spectrometer	spect
Solvent	CDCl3
Temperature	294.9
Pulse Sequence	zgpg30
Experiment	1D
Number of Scans	128
Receiver Gain	72
Relaxation Delay	2.0000
Pulse Width	10.0000
Acquisition Time	1.3631
Acquisition Date	2016-11-21T17:50:45
Spectrometer Frequency	100.62
Spectral Width	24038.5
Nucleus	13C
Acquired Size	32768
Spectral Size	65536



PROTON01
 LMC-02-08F1
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 Comment LMC-02-08F1
 Origin Varian
 Spectrometer inova
 Solvent cdcl3
 Temperature 25.0
 Pulse Sequence s2pul
 Experiment 1D
 Probe autox7991
 Number of Scans 64
 Receiver Gain 40
 Relaxation Delay 1.0000
 Pulse Width 0.0000
 Acquisition Time 3.0000
 Acquisition Date 2013-10-10T21:10:18
 Nucleus 1H



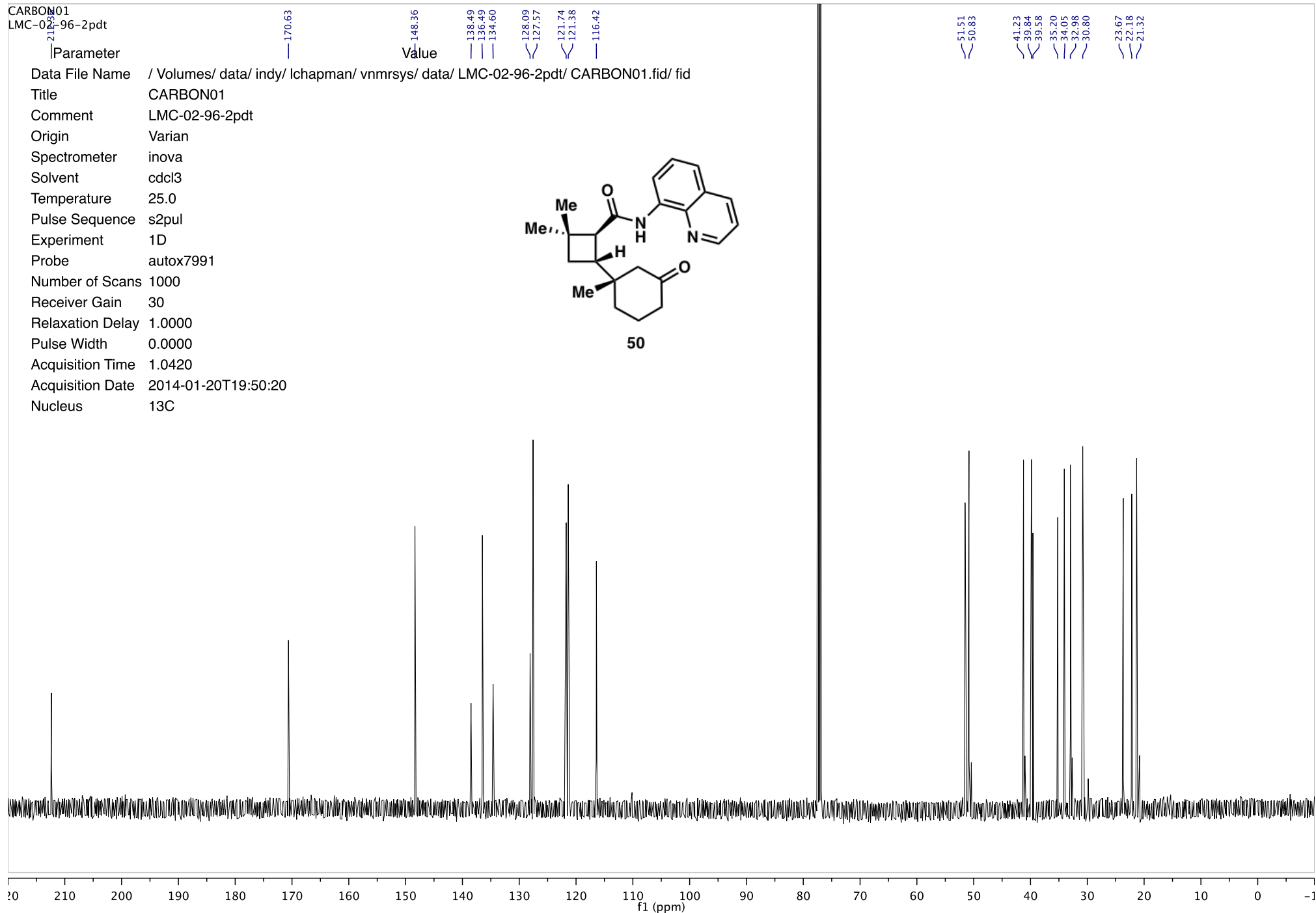
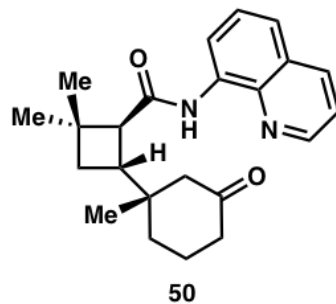
CARBON01

LMC-02-96-2pdt

Parameter

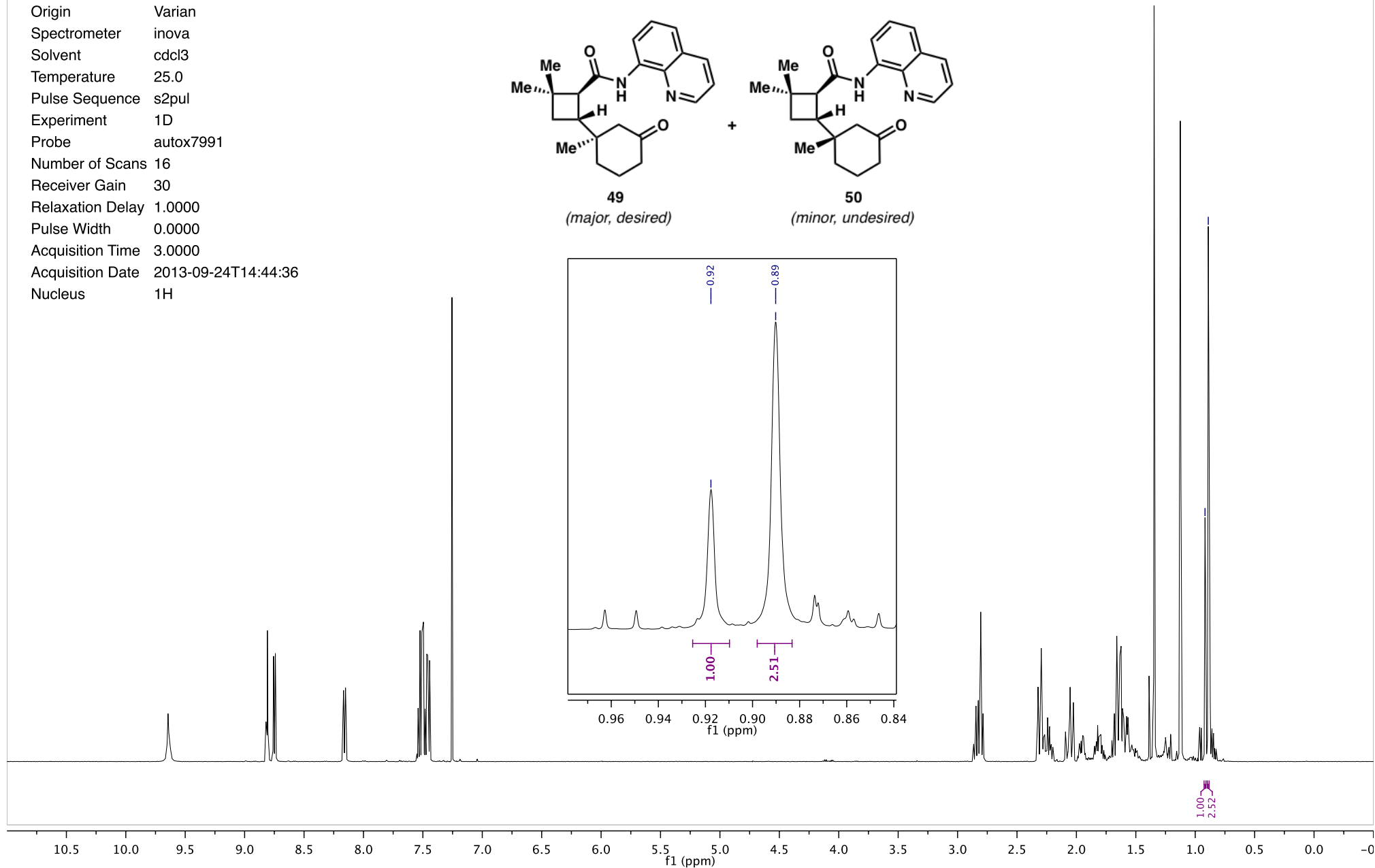
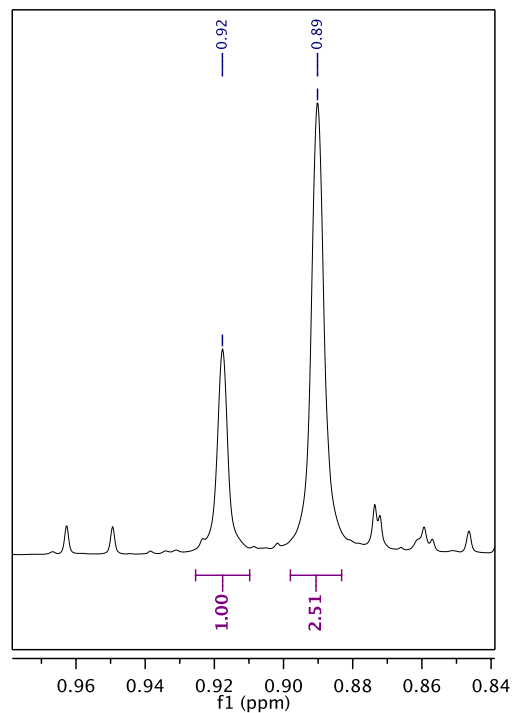
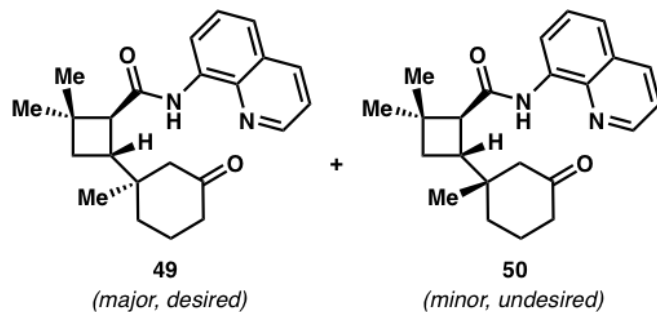
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Title CARBON01
Comment LMC-02-96-2pdt
Origin Varian
Spectrometer inova
Solvent cdcl3
Temperature 25.0
Pulse Sequence s2pul
Experiment 1D
Probe autox7991
Number of Scans 1000
Receiver Gain 30
Relaxation Delay 1.0000
Pulse Width 0.0000
Acquisition Time 1.0420
Acquisition Date 2014-01-20T19:50:20
Nucleus 13C

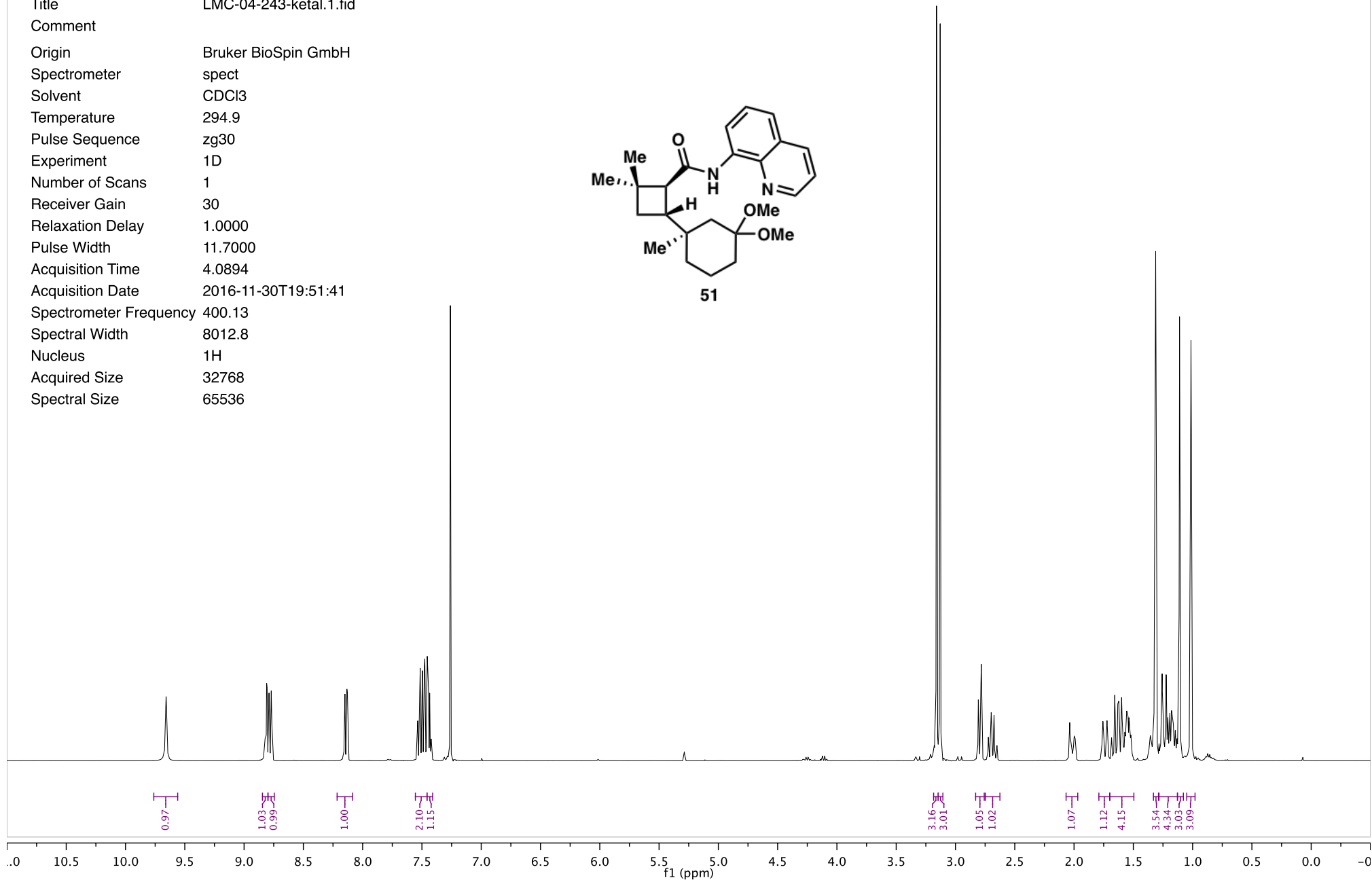
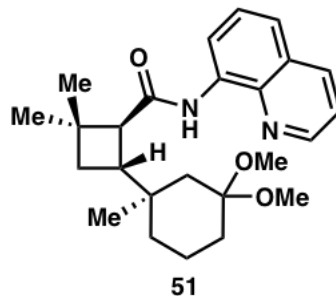


PROTON01

Parameter	Value
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Title	PROTON01
Origin	Varian
Spectrometer	inova
Solvent	cdcl3
Temperature	25.0
Pulse Sequence	s2pul
Experiment	1D
Probe	autox7991
Number of Scans	16
Receiver Gain	30
Relaxation Delay	1.0000
Pulse Width	0.0000
Acquisition Time	3.0000
Acquisition Date	2013-09-24T14:44:36
Nucleus	1H

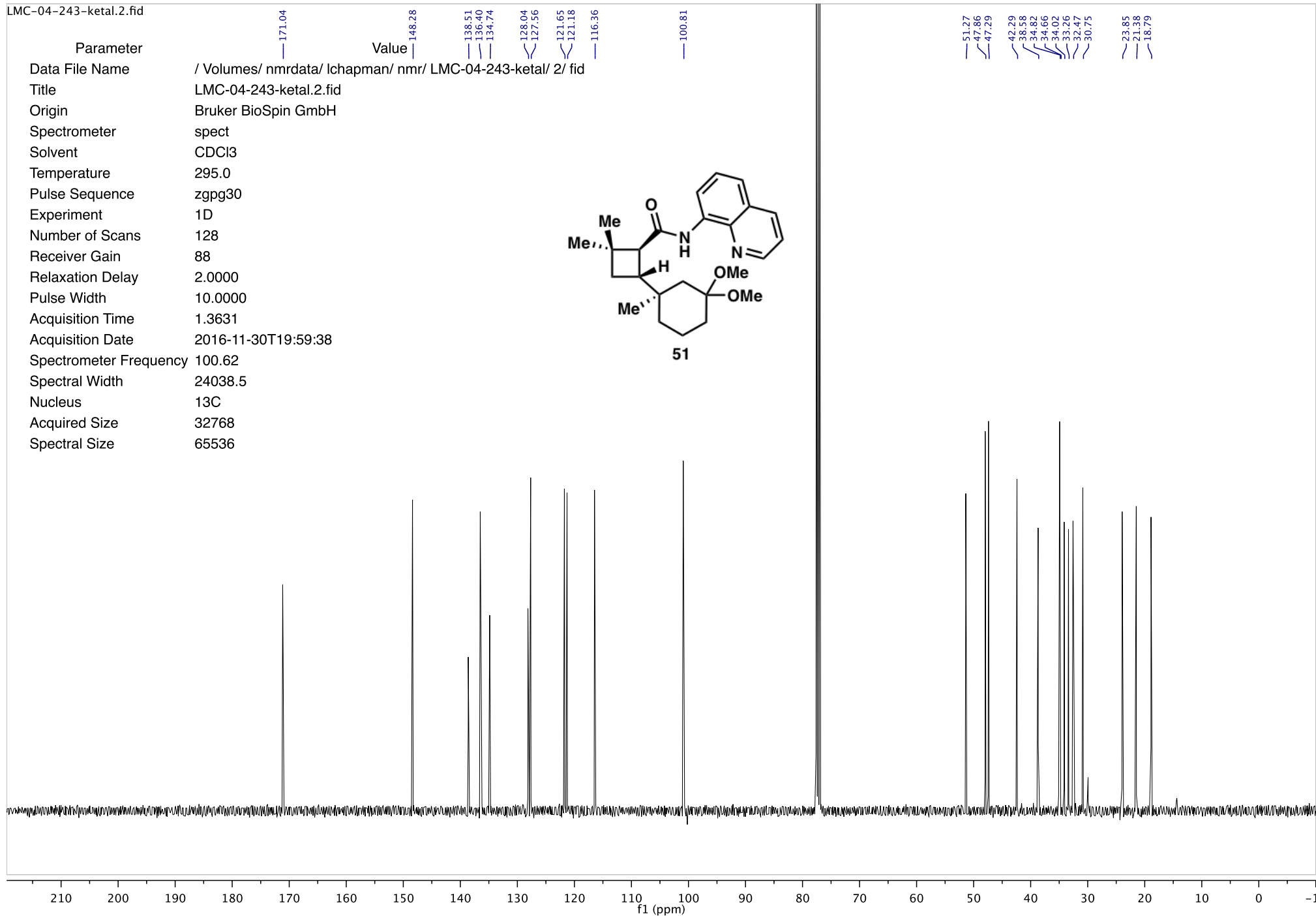
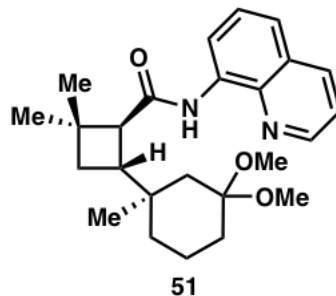


Parameter	Value
Data File Name	/ Volumes/ nmrdata/ lchapman/ nmr/ LMC-04-243-ketal/ 1/ fid
Title	LMC-04-243-ketal.1.fid
Comment	
Origin	Bruker BioSpin GmbH
Spectrometer	spect
Solvent	CDCl3
Temperature	294.9
Pulse Sequence	zg30
Experiment	1D
Number of Scans	1
Receiver Gain	30
Relaxation Delay	1.0000
Pulse Width	11.7000
Acquisition Time	4.0894
Acquisition Date	2016-11-30T19:51:41
Spectrometer Frequency	400.13
Spectral Width	8012.8
Nucleus	1H
Acquired Size	32768
Spectral Size	65536

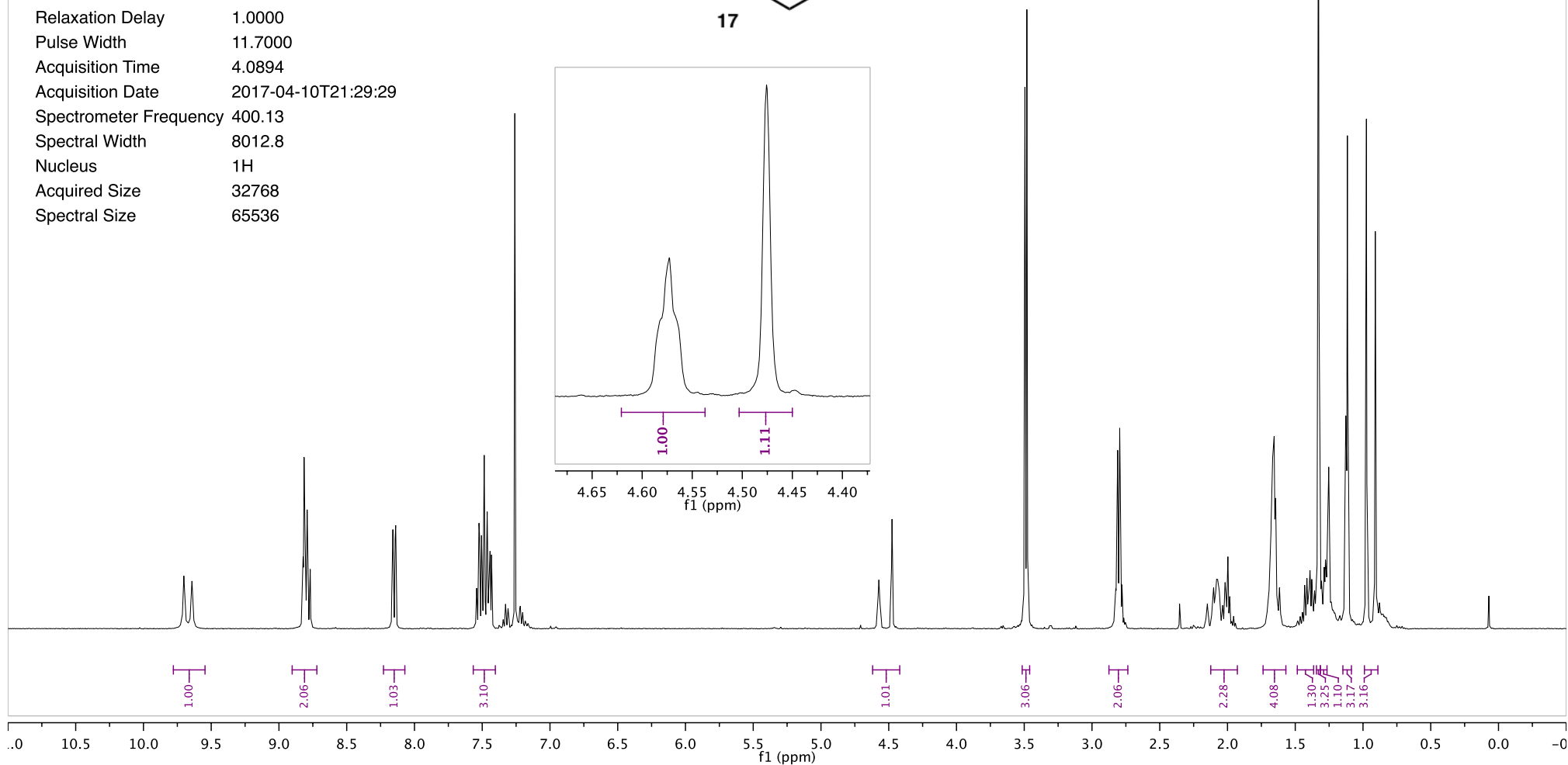
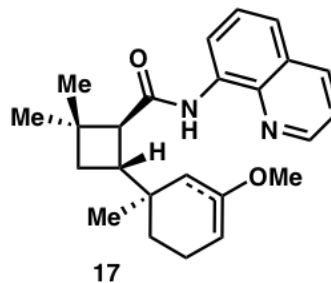


LMC-04-243-ketal.2.fid

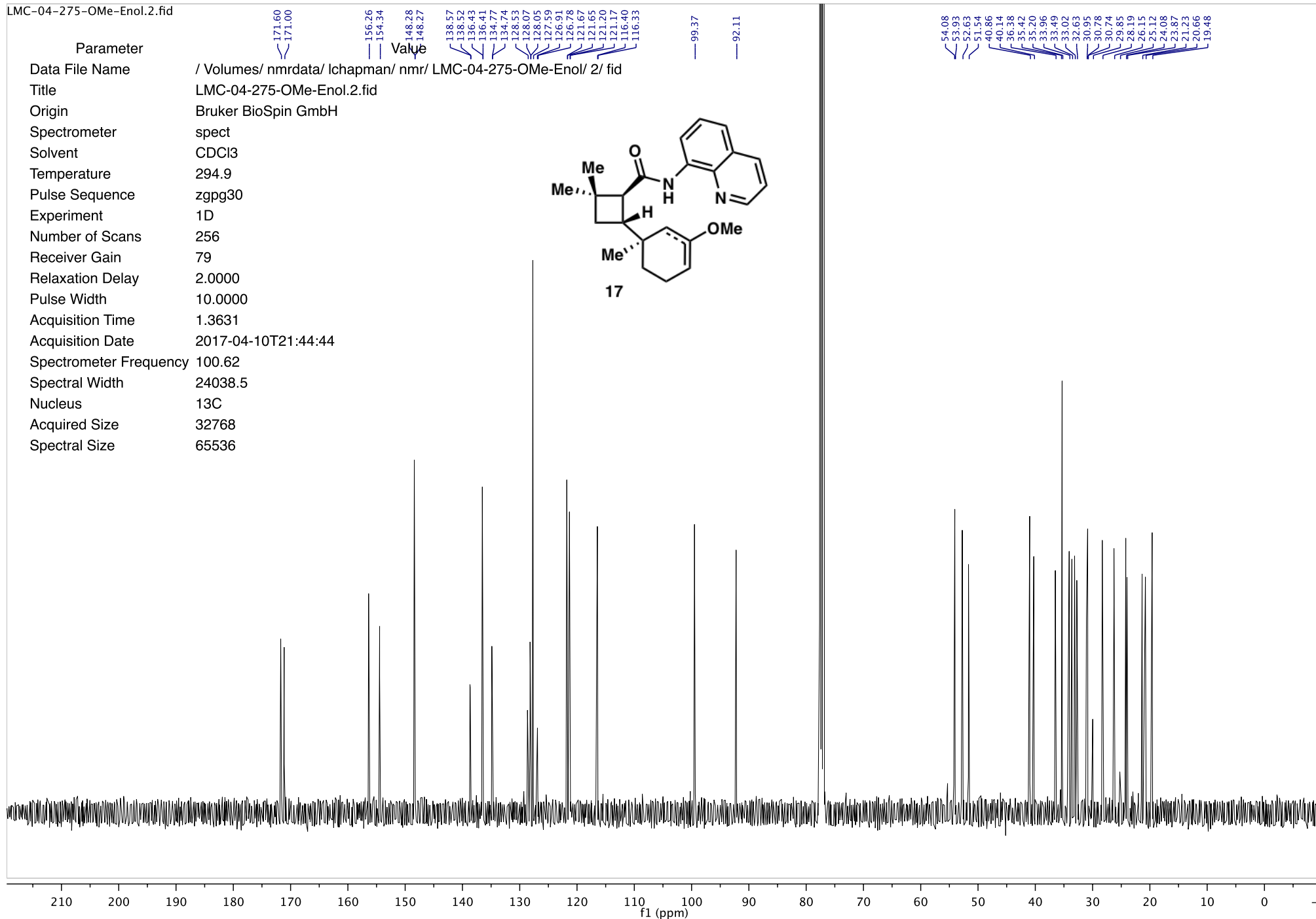
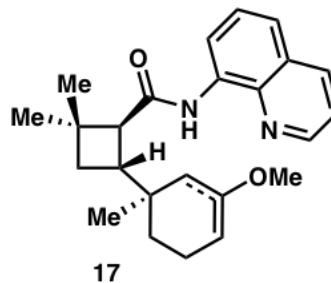
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Title	LMC-04-243-ketal.2.fid
Origin	Bruker BioSpin GmbH
Spectrometer	spect
Solvent	CDCl3
Temperature	295.0
Pulse Sequence	zgpg30
Experiment	1D
Number of Scans	128
Receiver Gain	88
Relaxation Delay	2.0000
Pulse Width	10.0000
Acquisition Time	1.3631
Acquisition Date	2016-11-30T19:59:38
Spectrometer Frequency	100.62
Spectral Width	24038.5
Nucleus	13C
Acquired Size	32768
Spectral Size	65536



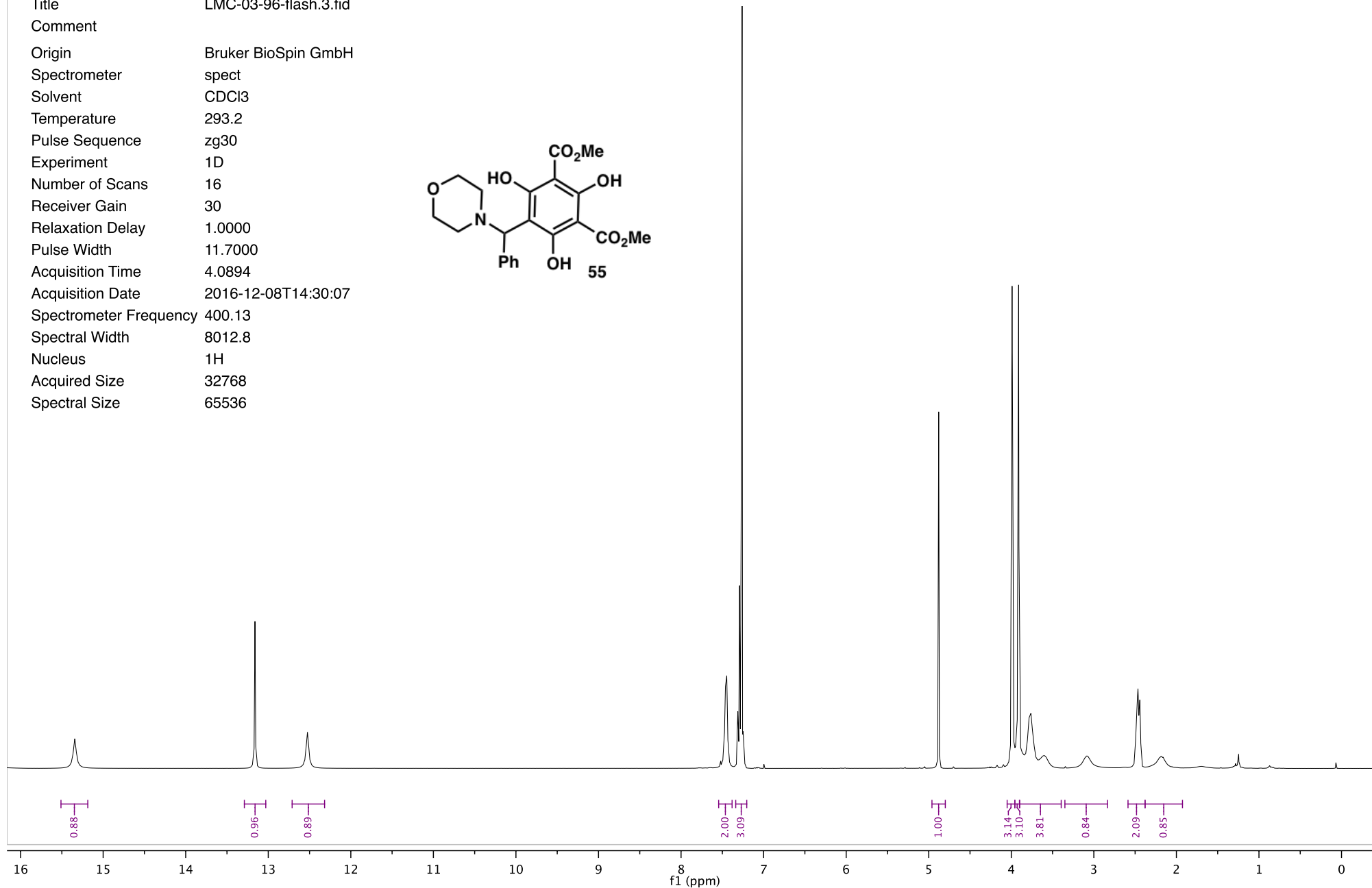
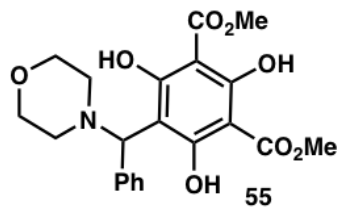
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Title	LMC-04-275-OMe-Enol.1.fid
Comment	
Origin	Bruker BioSpin GmbH
Spectrometer	spect
Solvent	CDCl3
Temperature	295.0
Pulse Sequence	zg30
Experiment	1D
Number of Scans	1
Receiver Gain	99
Relaxation Delay	1.0000
Pulse Width	11.7000
Acquisition Time	4.0894
Acquisition Date	2017-04-10T21:29:29
Spectrometer Frequency	400.13
Spectral Width	8012.8
Nucleus	1H
Acquired Size	32768
Spectral Size	65536



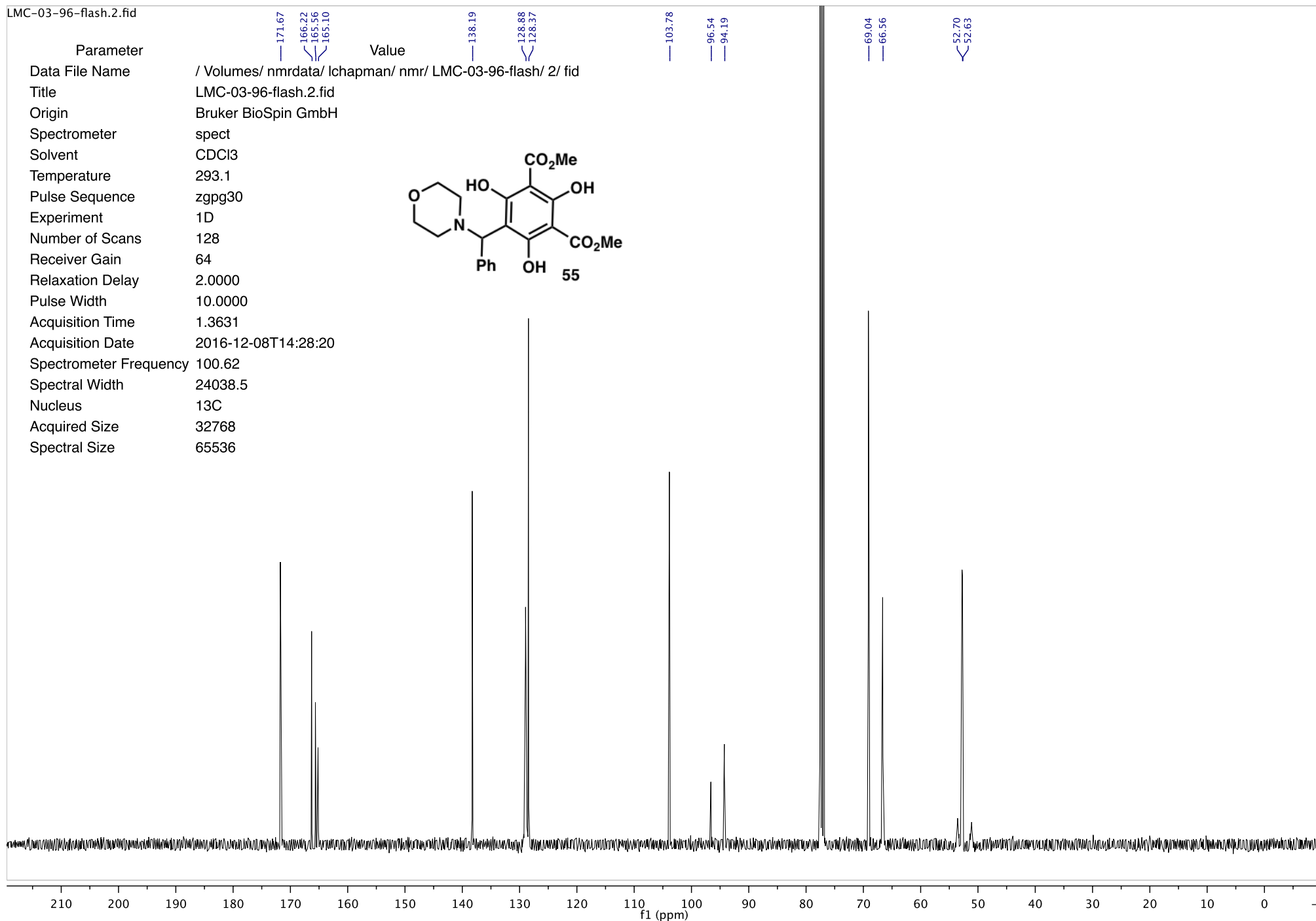
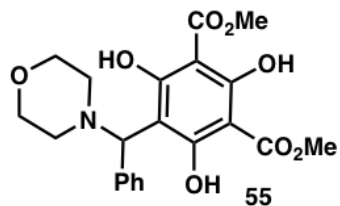
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Data File Name	/ Volumes/ nmrdata/ lchapman/ nmr/ LMC-04-275-OMe-Enol/ 2/ fid
Title	LMC-04-275-OMe-Enol.2.fid
Origin	Bruker BioSpin GmbH
Spectrometer	spect
Solvent	CDCl3
Temperature	294.9
Pulse Sequence	zgpg30
Experiment	1D
Number of Scans	256
Receiver Gain	79
Relaxation Delay	2.0000
Pulse Width	10.0000
Acquisition Time	1.3631
Acquisition Date	2017-04-10T21:44:44
Spectrometer Frequency	100.62
Spectral Width	24038.5
Nucleus	13C
Acquired Size	32768
Spectral Size	65536



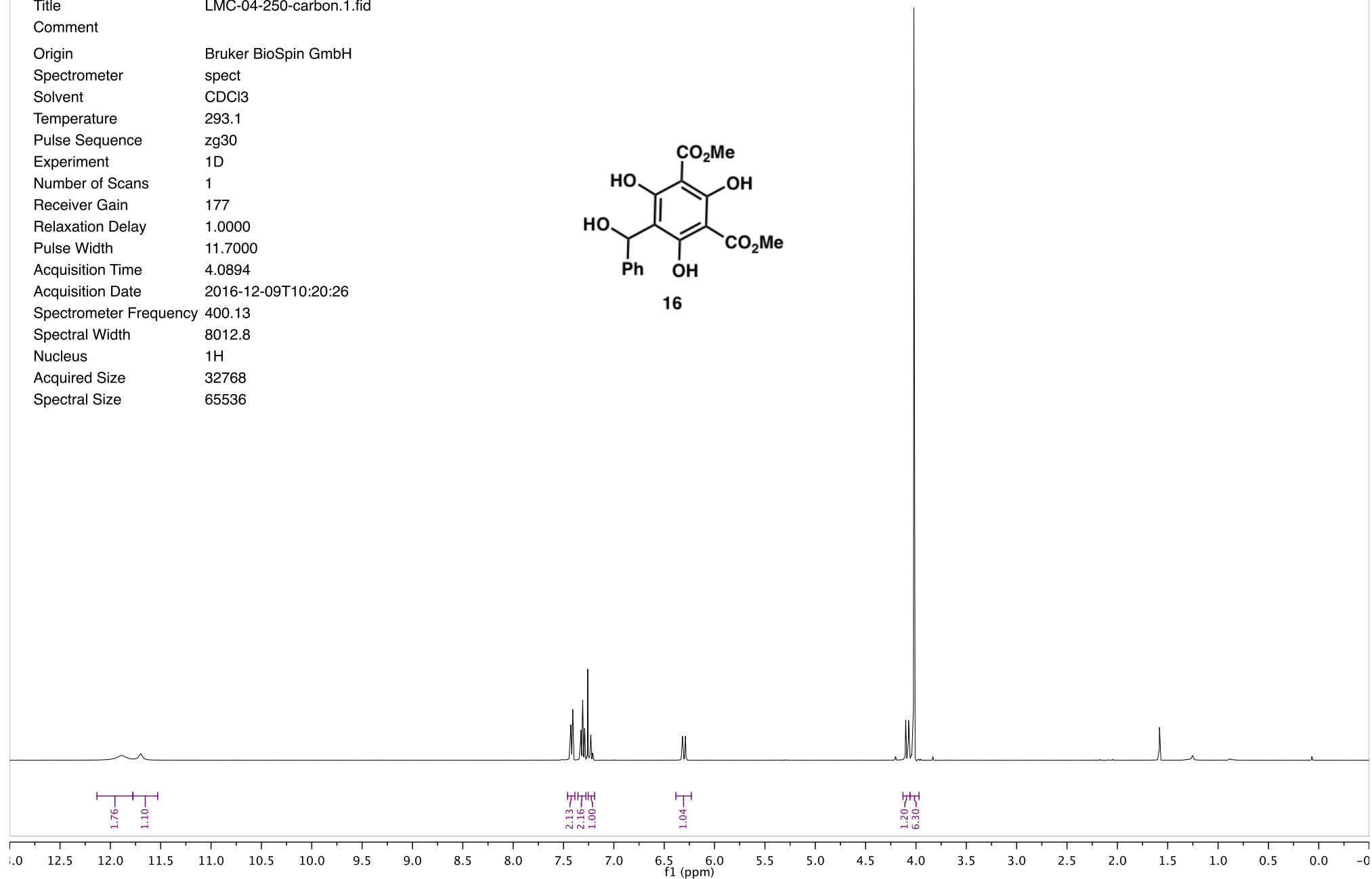
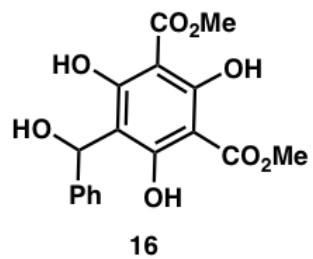
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Data File Name	/ Volumes/ nmrdata/ lchapman/ nmr/ LMC-03-96-flash/ 3/ fid
Title	LMC-03-96-flash.3.fid
Comment	
Origin	Bruker BioSpin GmbH
Spectrometer	spect
Solvent	CDCl3
Temperature	293.2
Pulse Sequence	zg30
Experiment	1D
Number of Scans	16
Receiver Gain	30
Relaxation Delay	1.0000
Pulse Width	11.7000
Acquisition Time	4.0894
Acquisition Date	2016-12-08T14:30:07
Spectrometer Frequency	400.13
Spectral Width	8012.8
Nucleus	1H
Acquired Size	32768
Spectral Size	65536



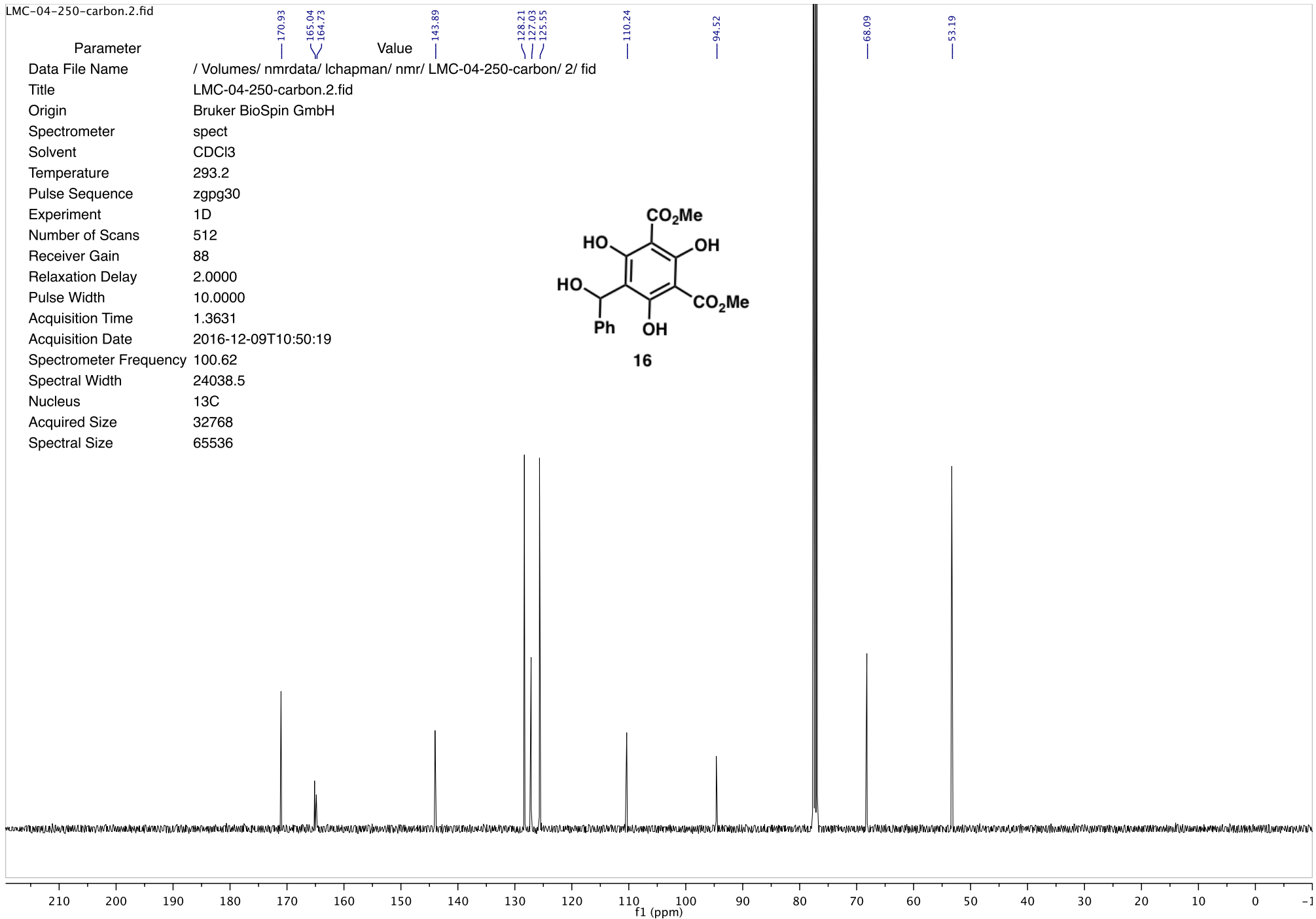
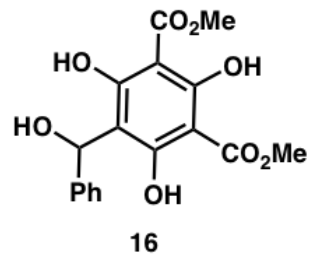
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Data File Name	/ Volumes/ nmrdata/ lchapman/ nmr/ LMC-03-96-flash/ 2/ fid
Title	LMC-03-96-flash.2.fid
Origin	Bruker BioSpin GmbH
Spectrometer	spect
Solvent	CDCl3
Temperature	293.1
Pulse Sequence	zgpg30
Experiment	1D
Number of Scans	128
Receiver Gain	64
Relaxation Delay	2.0000
Pulse Width	10.0000
Acquisition Time	1.3631
Acquisition Date	2016-12-08T14:28:20
Spectrometer Frequency	100.62
Spectral Width	24038.5
Nucleus	13C
Acquired Size	32768
Spectral Size	65536



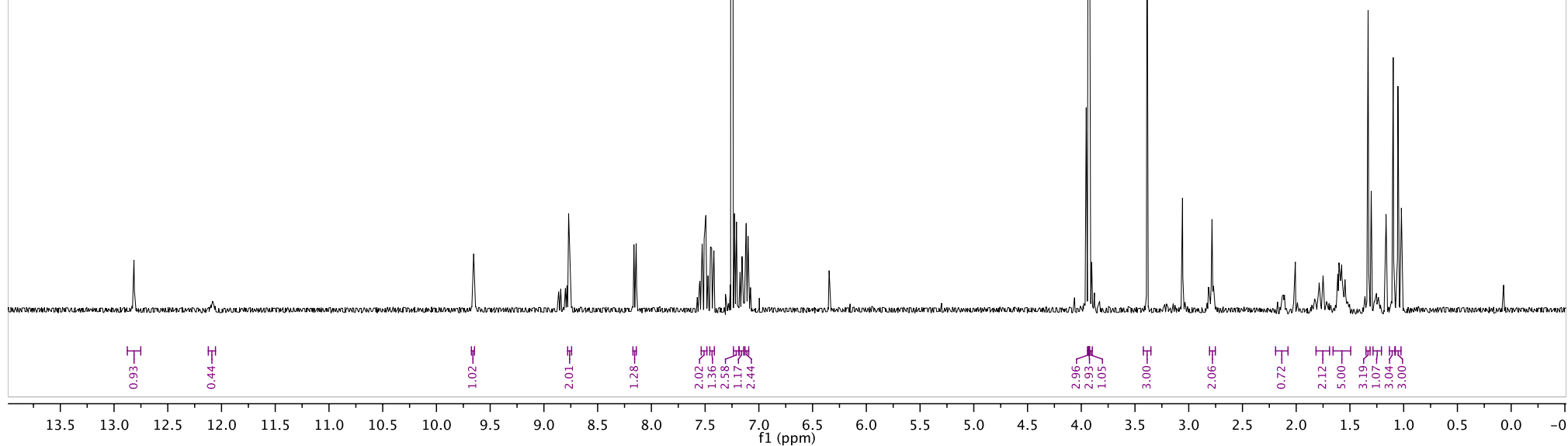
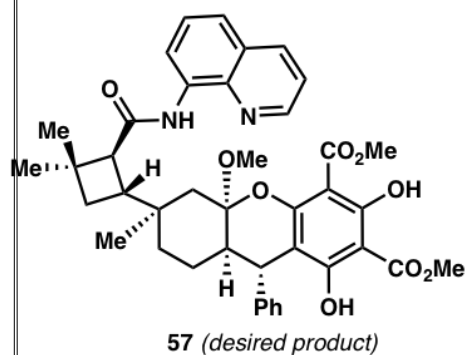
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Data File Name	/ Volumes/ nmrdata/ lchapman/ nmr/ LMC-04-250-carbon/ 1/ fid
Title	LMC-04-250-carbon.1.fid
Comment	
Origin	Bruker BioSpin GmbH
Spectrometer	spect
Solvent	CDCl3
Temperature	293.1
Pulse Sequence	zg30
Experiment	1D
Number of Scans	1
Receiver Gain	177
Relaxation Delay	1.0000
Pulse Width	11.7000
Acquisition Time	4.0894
Acquisition Date	2016-12-09T10:20:26
Spectrometer Frequency	400.13
Spectral Width	8012.8
Nucleus	1H
Acquired Size	32768
Spectral Size	65536



Parameter	Value
Data File Name	/ Volumes/ nmrdata/ lchapman/ nmr/ LMC-04-250-carbon/ 2/ fid
Title	LMC-04-250-carbon.2.fid
Origin	Bruker BioSpin GmbH
Spectrometer	spect
Solvent	CDCl3
Temperature	293.2
Pulse Sequence	zgpg30
Experiment	1D
Number of Scans	512
Receiver Gain	88
Relaxation Delay	2.0000
Pulse Width	10.0000
Acquisition Time	1.3631
Acquisition Date	2016-12-09T10:50:19
Spectrometer Frequency	100.62
Spectral Width	24038.5
Nucleus	¹³ C
Acquired Size	32768
Spectral Size	65536



Parameter	Value
Data File Name	/ Volumes/ nmrdata/ lchapman/ nmr/ LMC-04-253-HPLC-P2/ 1/ fid
Title	LMC-04-253-HPLC-P2.1.fid
Origin	Bruker BioSpin GmbH
Spectrometer	spect
Solvent	CDCl3
Temperature	295.0
Pulse Sequence	zg30
Experiment	1D
Number of Scans	1
Receiver Gain	197
Relaxation Delay	1.0000
Pulse Width	11.7000
Acquisition Time	4.0894
Acquisition Date	2016-12-21T04:34:12
Spectrometer Frequency	400.13
Spectral Width	8012.8
Nucleus	1H
Acquired Size	32768
Spectral Size	65536

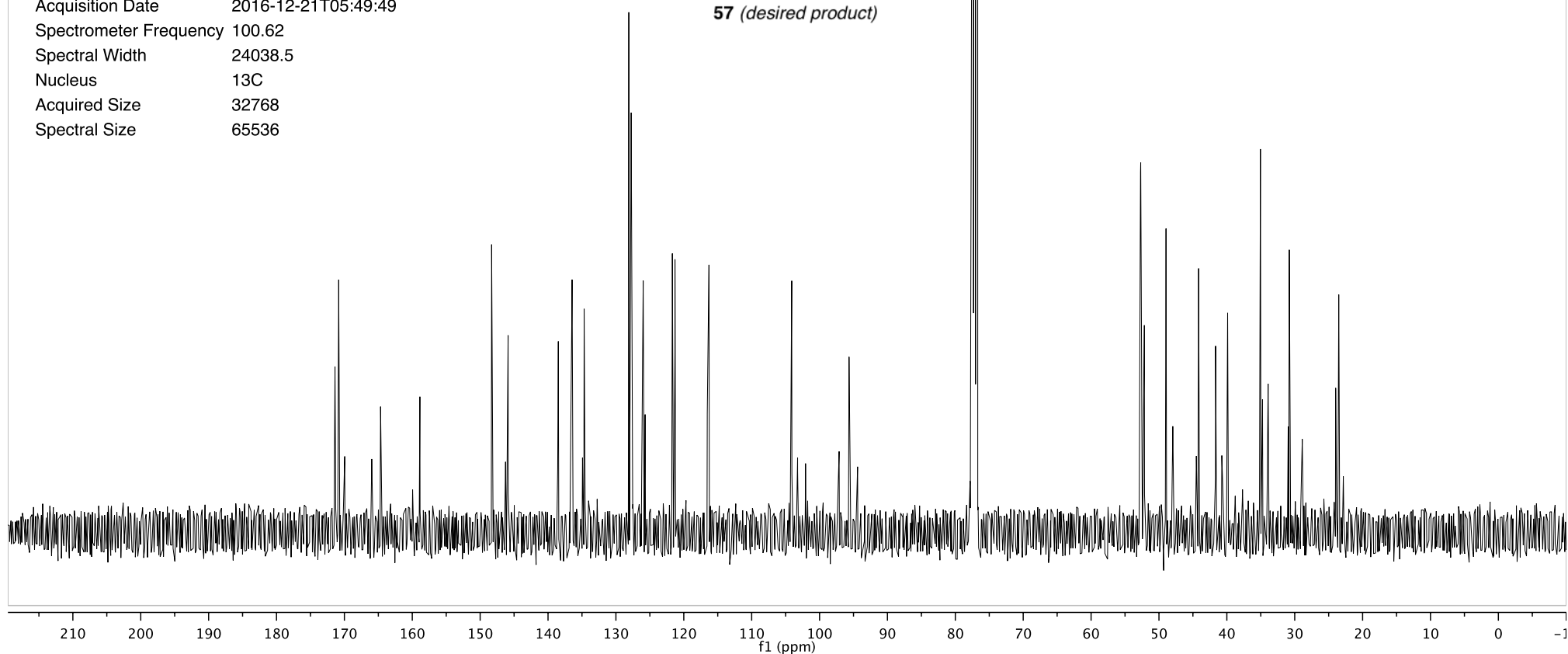
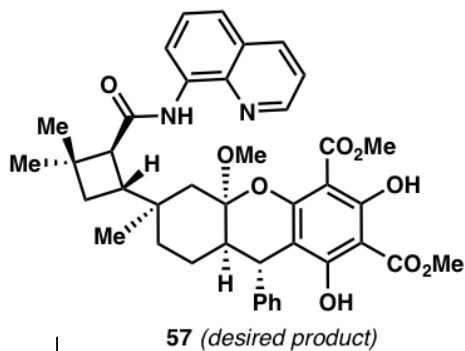


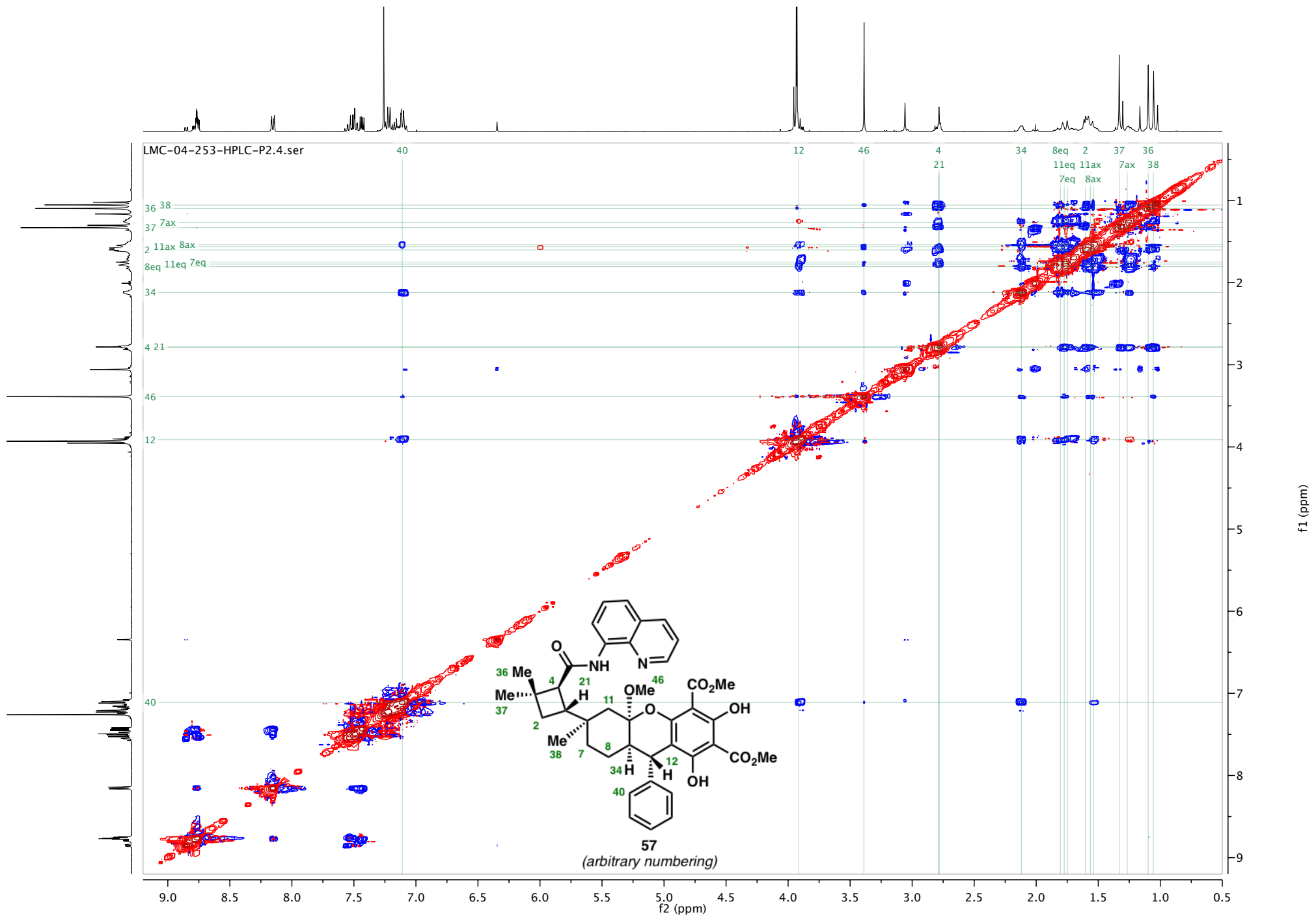
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 Title LMC-04-253-HPLC-P2.3.fid
 Comment
 Origin Bruker BioSpin GmbH
 Spectrometer spect
 Solvent CDCl3
 Temperature 294.9
 Pulse Sequence zgpg30
 Experiment 1D
 Number of Scans 1024
 Receiver Gain 64
 Relaxation Delay 2.0000
 Pulse Width 10.0000
 Acquisition Time 1.3631
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 Spectrometer Frequency 100.62
 Spectral Width 24038.5
 Nucleus 13C
 Acquired Size 32768
 Spectral Size 65536

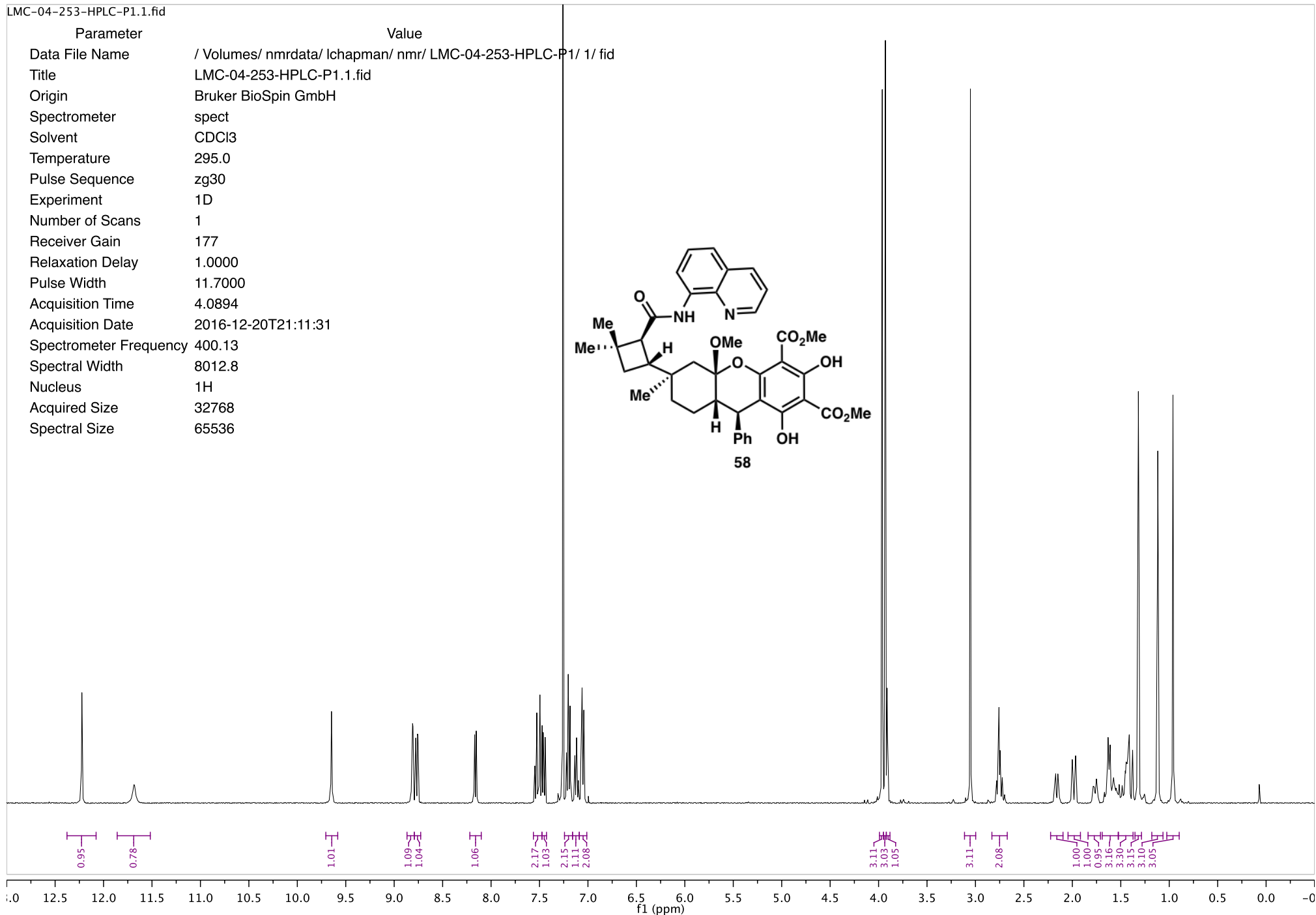
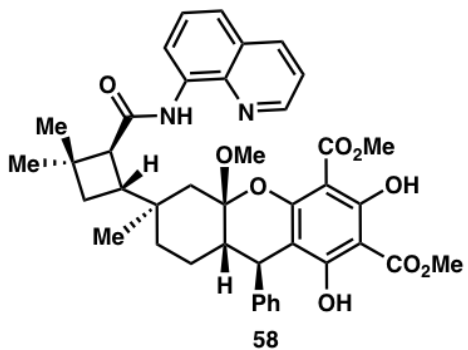
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 104.10
 97.12
 95.65

52.72
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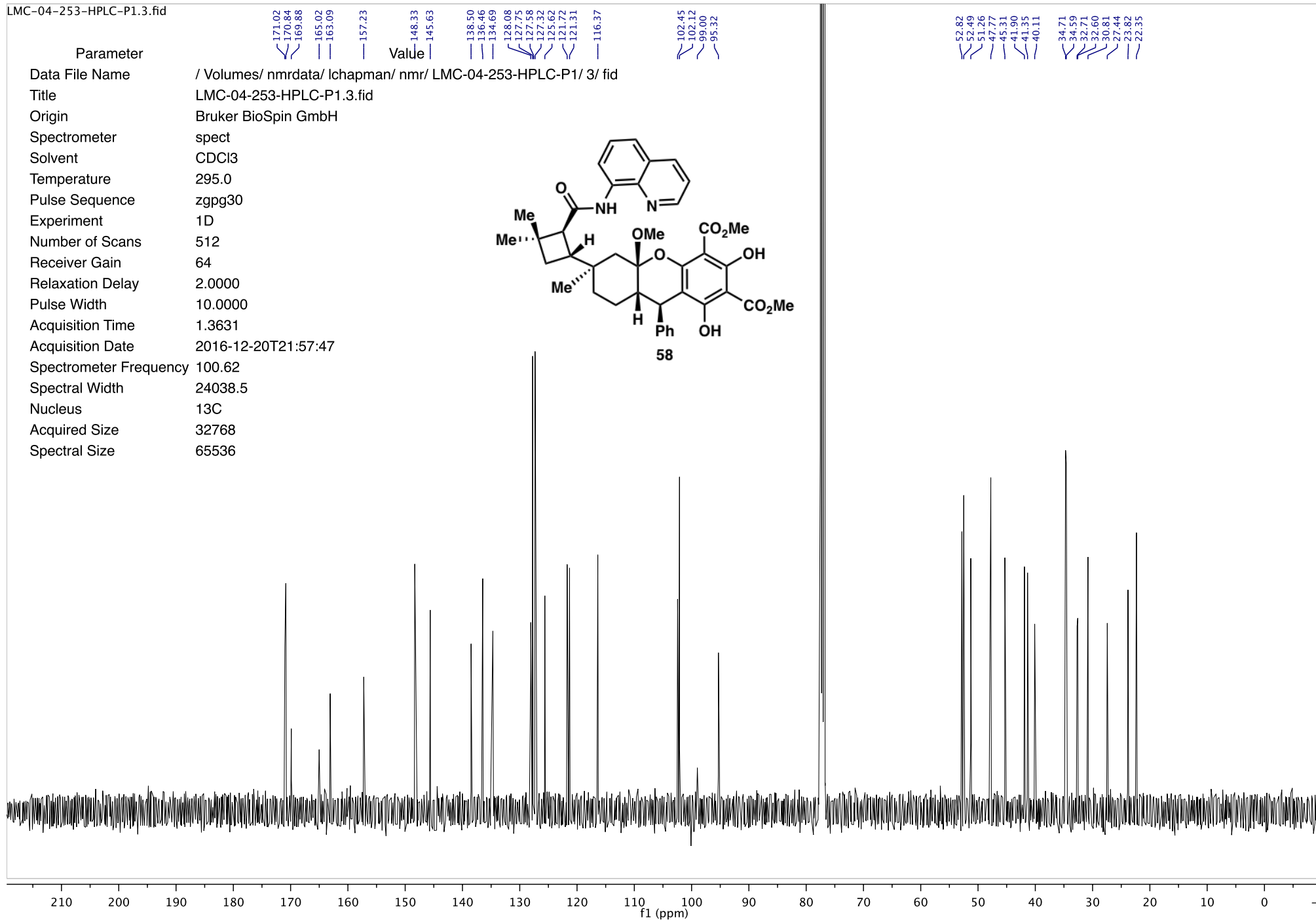
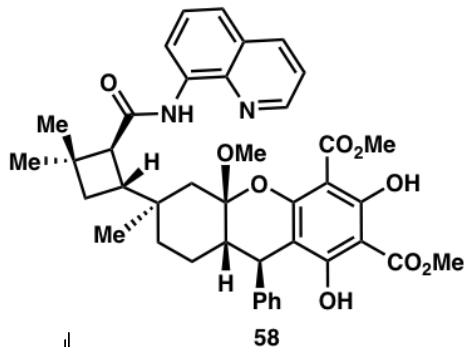


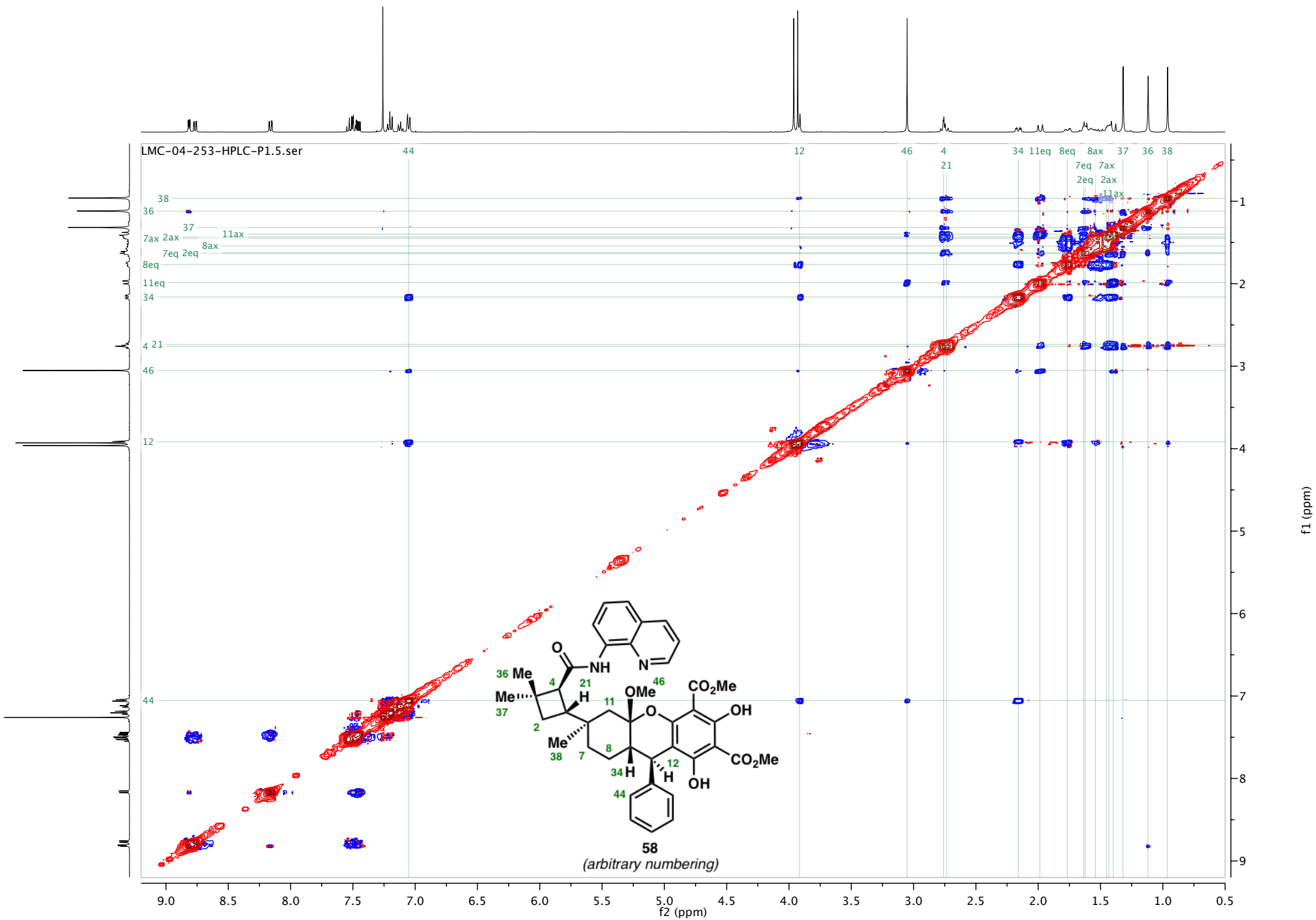


Parameter	Value
Data File Name	/ Volumes/ nmrdata/ lchapman/ nmr/ LMC-04-253-HPLC-P1/ 1/ fid
Title	LMC-04-253-HPLC-P1.1.fid
Origin	Bruker BioSpin GmbH
Spectrometer	spect
Solvent	CDCl3
Temperature	295.0
Pulse Sequence	zg30
Experiment	1D
Number of Scans	1
Receiver Gain	177
Relaxation Delay	1.0000
Pulse Width	11.7000
Acquisition Time	4.0894
Acquisition Date	2016-12-20T21:11:31
Spectrometer Frequency	400.13
Spectral Width	8012.8
Nucleus	1H
Acquired Size	32768
Spectral Size	65536

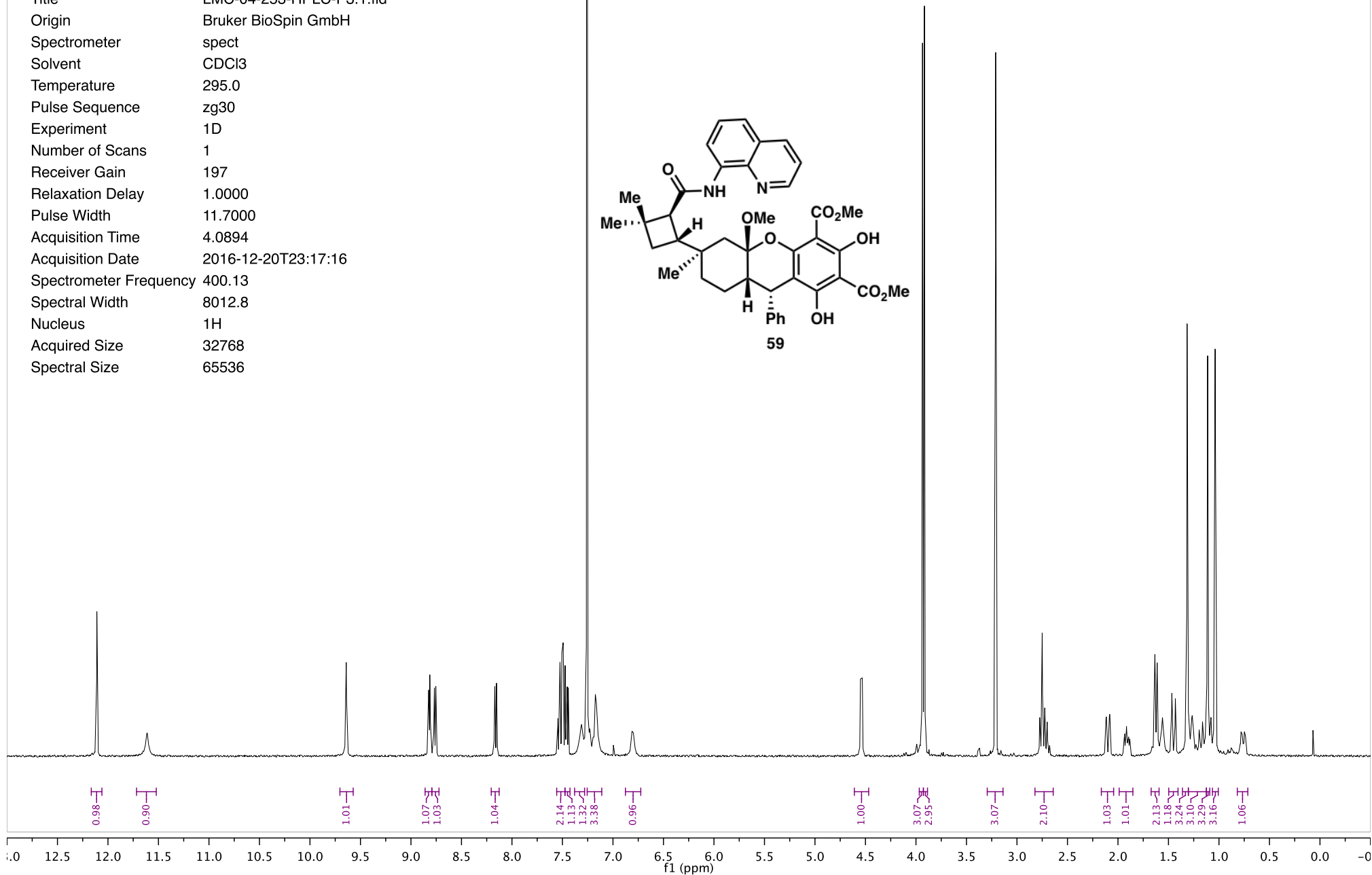
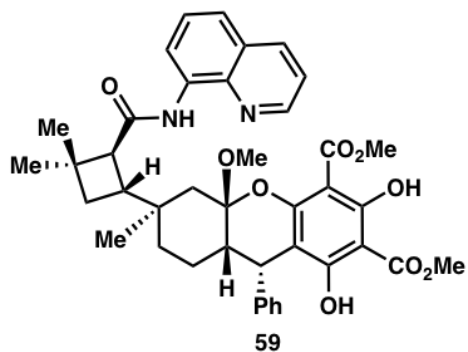


Parameter	Value
Data File Name	/ Volumes/ nmrdata/ lchapman/ nmr/ LMC-04-253-HPLC-P1/ 3/ fid
Title	LMC-04-253-HPLC-P1.3.fid
Origin	Bruker BioSpin GmbH
Spectrometer	spect
Solvent	CDCl3
Temperature	295.0
Pulse Sequence	zgpg30
Experiment	1D
Number of Scans	512
Receiver Gain	64
Relaxation Delay	2.0000
Pulse Width	10.0000
Acquisition Time	1.3631
Acquisition Date	2016-12-20T21:57:47
Spectrometer Frequency	100.62
Spectral Width	24038.5
Nucleus	13C
Acquired Size	32768
Spectral Size	65536

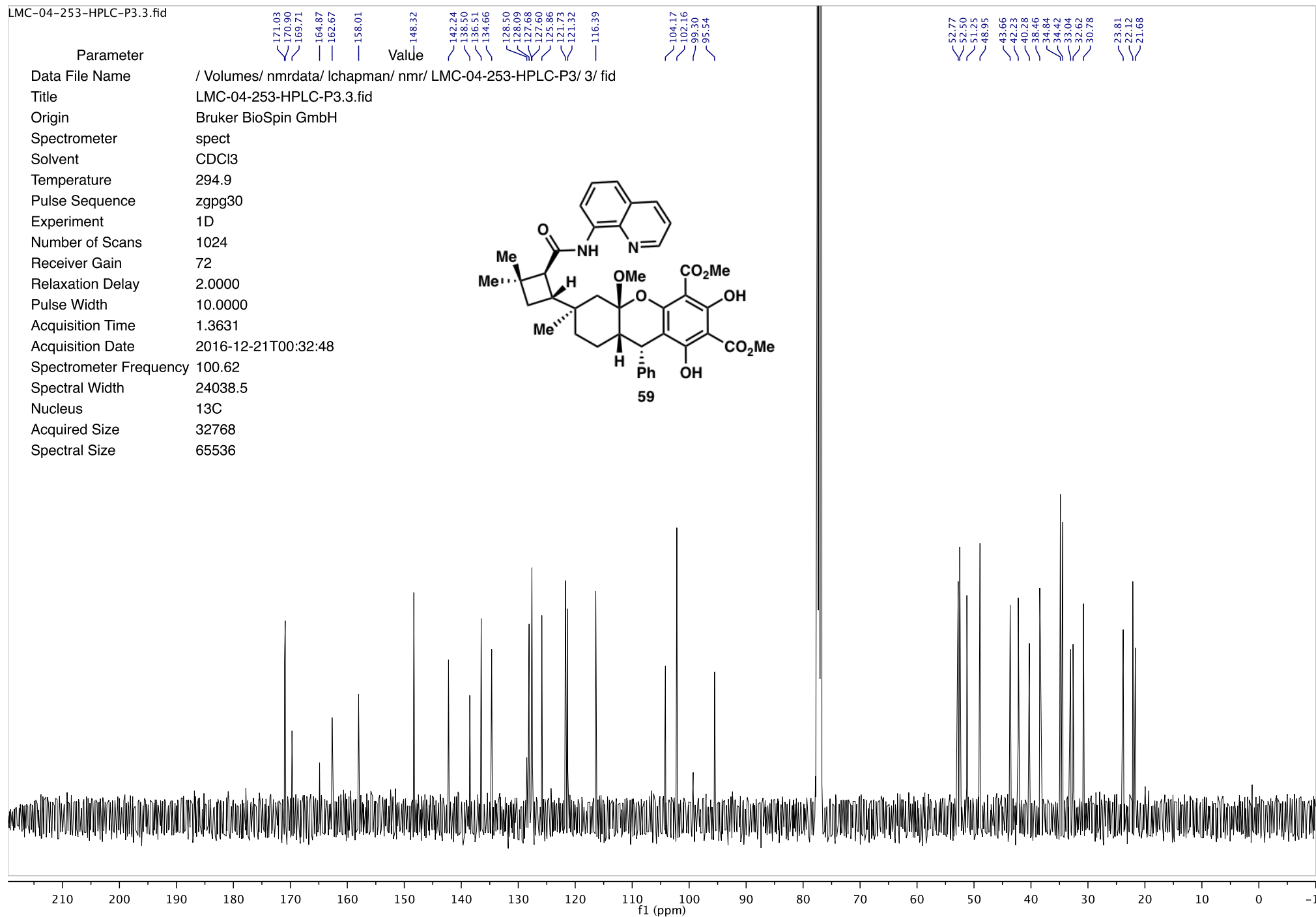
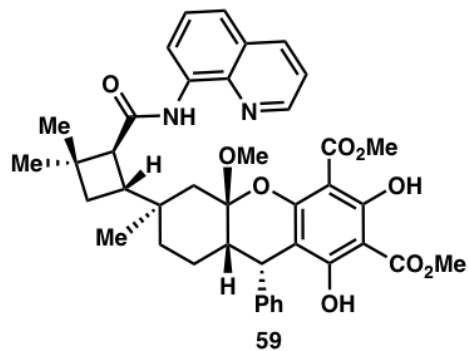


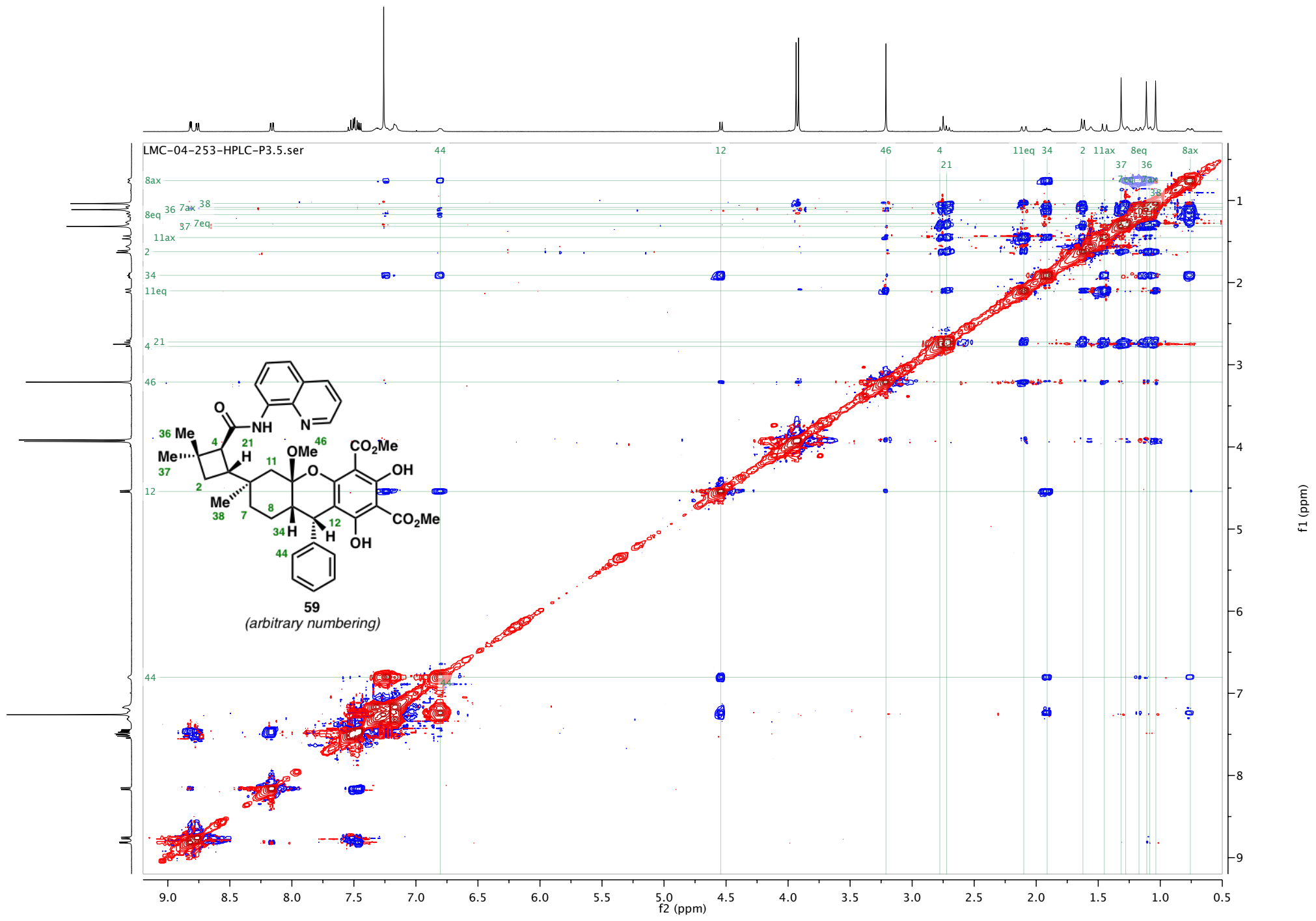


Parameter	Value
Data File Name	/ Volumes/ nmrdata/ lchapman/ nmr/ LMC-04-253-HPLC-P3/ 1/ fid
Title	LMC-04-253-HPLC-P3.1.fid
Origin	Bruker BioSpin GmbH
Spectrometer	spect
Solvent	CDCl3
Temperature	295.0
Pulse Sequence	zg30
Experiment	1D
Number of Scans	1
Receiver Gain	197
Relaxation Delay	1.0000
Pulse Width	11.7000
Acquisition Time	4.0894
Acquisition Date	2016-12-20T23:17:16
Spectrometer Frequency	400.13
Spectral Width	8012.8
Nucleus	1H
Acquired Size	32768
Spectral Size	65536

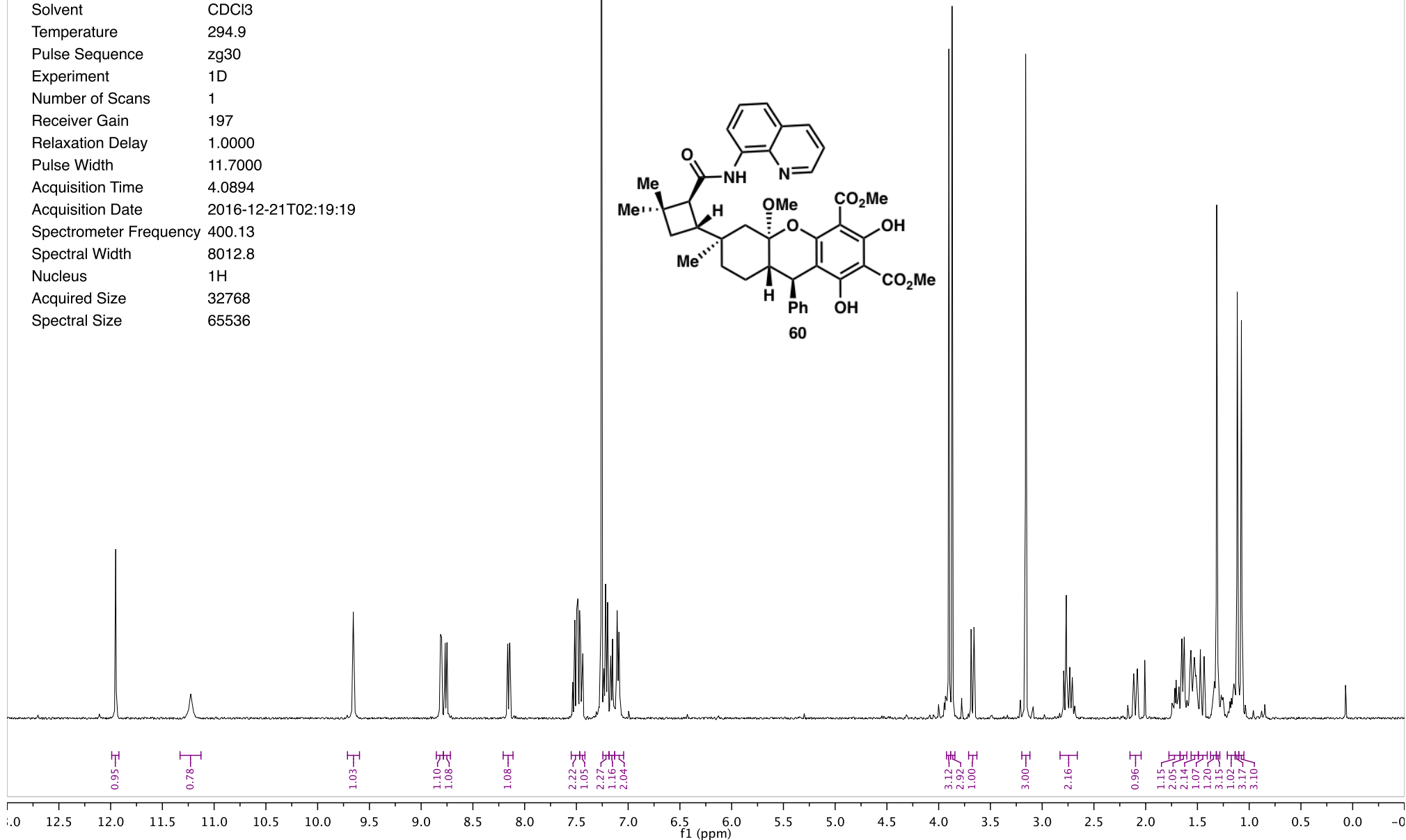
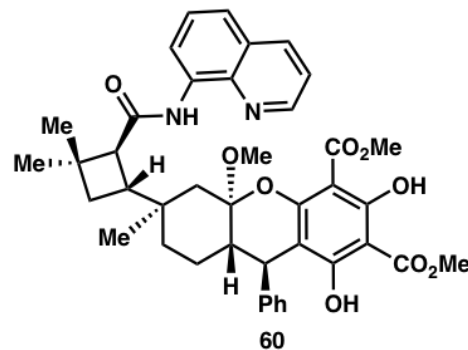


Parameter	Value
Data File Name	/ Volumes/ nmrdata/ lchapman/ nmr/ LMC-04-253-HPLC-P3/ 3/ fid
Title	LMC-04-253-HPLC-P3.3.fid
Origin	Bruker BioSpin GmbH
Spectrometer	spect
Solvent	CDCl3
Temperature	294.9
Pulse Sequence	zgpg30
Experiment	1D
Number of Scans	1024
Receiver Gain	72
Relaxation Delay	2.0000
Pulse Width	10.0000
Acquisition Time	1.3631
Acquisition Date	2016-12-21T00:32:48
Spectrometer Frequency	100.62
Spectral Width	24038.5
Nucleus	13C
Acquired Size	32768
Spectral Size	65536





Parameter	Value
Data File Name	/ Volumes/ nmrdata/ lchapman/ nmr/ LMC-04-253-HPLC-P4/ 1/ fid
Title	LMC-04-253-HPLC-P4.1.fid
Origin	Bruker BioSpin GmbH
Spectrometer	spect
Solvent	CDCl3
Temperature	294.9
Pulse Sequence	zg30
Experiment	1D
Number of Scans	1
Receiver Gain	197
Relaxation Delay	1.0000
Pulse Width	11.7000
Acquisition Time	4.0894
Acquisition Date	2016-12-21T02:19:19
Spectrometer Frequency	400.13
Spectral Width	8012.8
Nucleus	1H
Acquired Size	32768
Spectral Size	65536

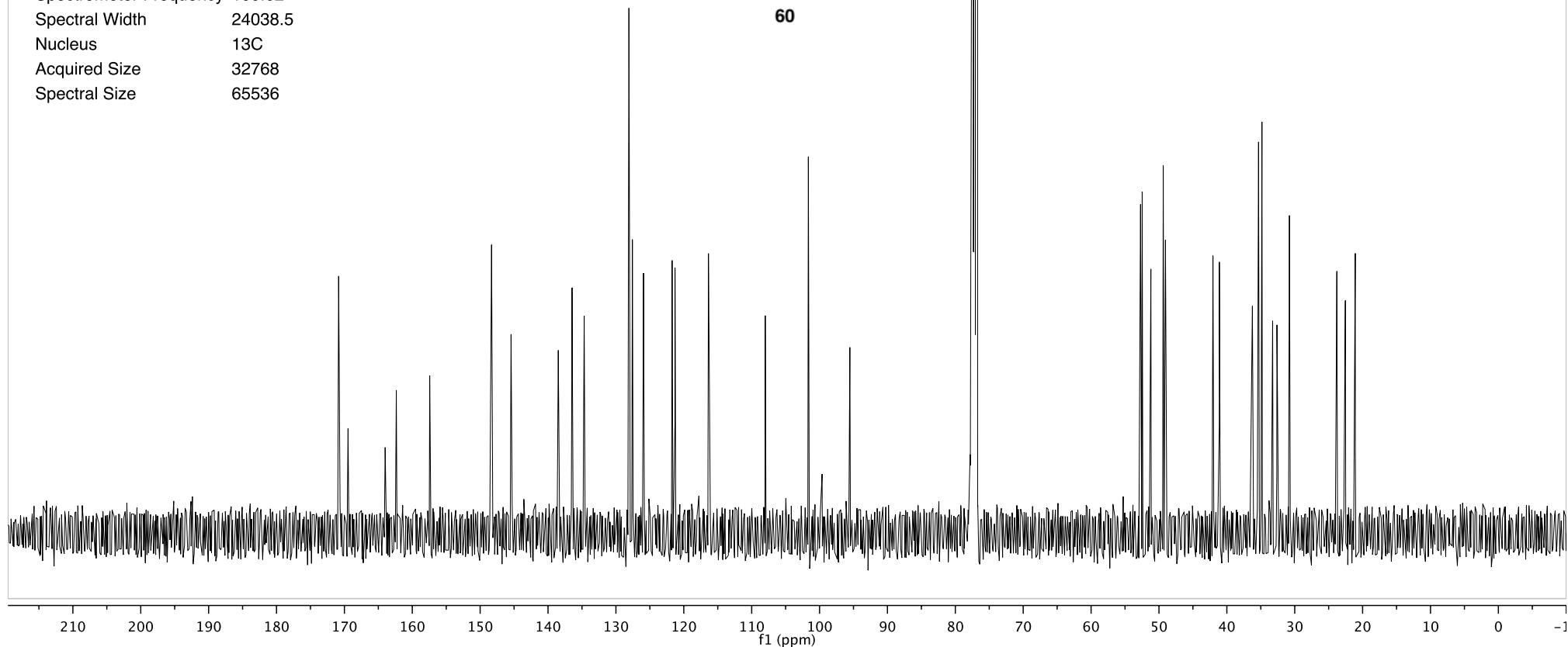
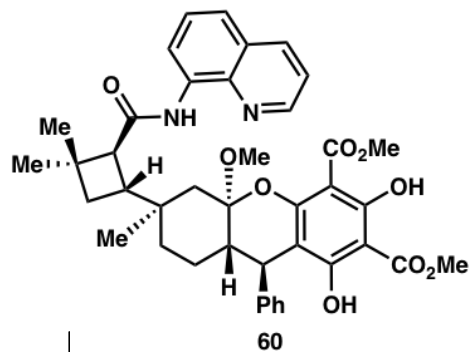


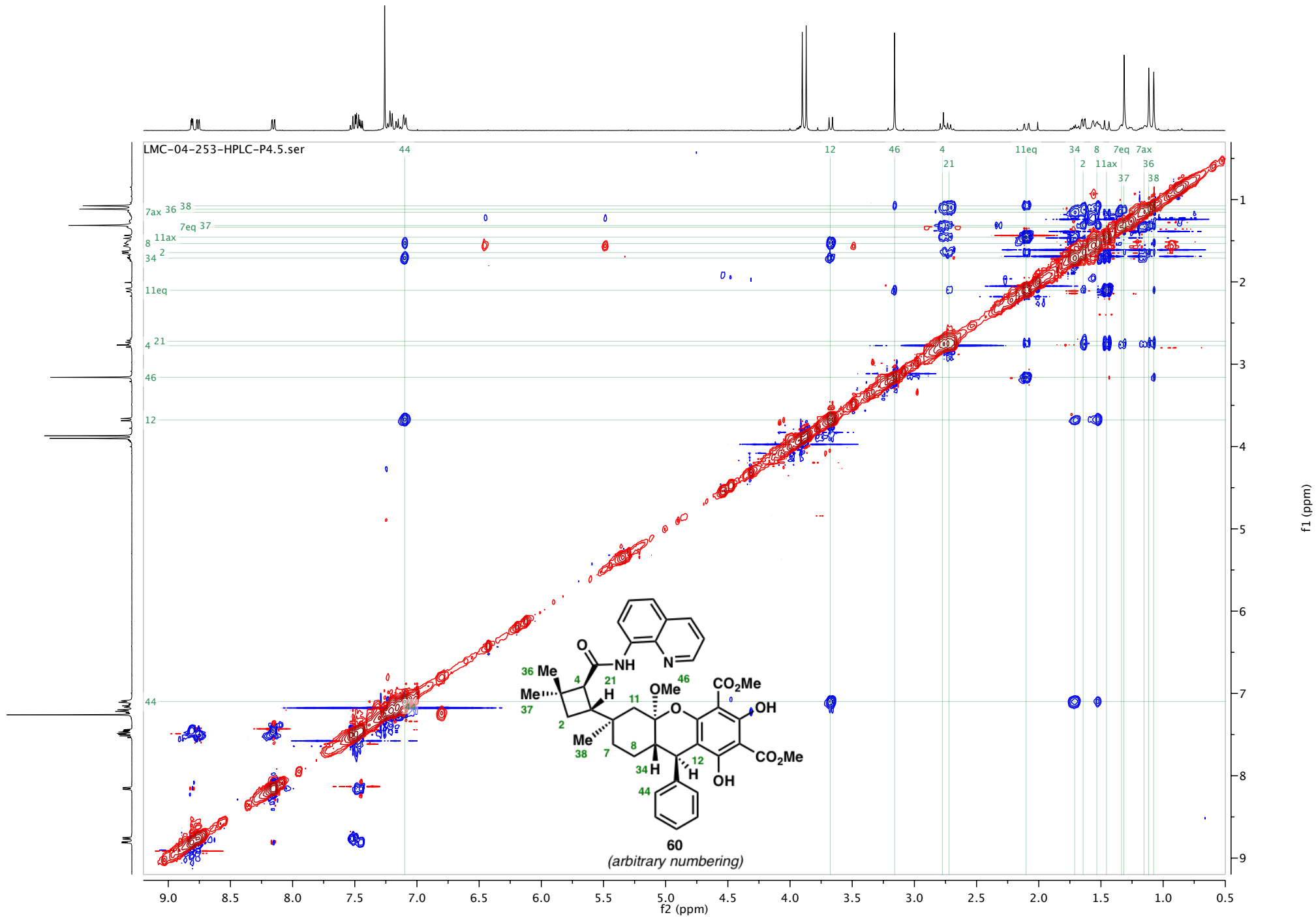
Parameter

Data File Name / Volumes/ nmrdata/ lchapman/ nmr/ LMC-04-253-HPLC-P4/ 3/ fid
 Title LMC-04-253-HPLC-P4.3.fid
 Origin Bruker BioSpin GmbH
 Spectrometer spect
 Solvent CDCl3
 Temperature 294.9
 Pulse Sequence zgpg30
 Experiment 1D
 Number of Scans 1024
 Receiver Gain 72
 Relaxation Delay 2.0000
 Pulse Width 10.0000
 Acquisition Time 1.3631
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 Spectrometer Frequency 100.62
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 Nucleus 13C
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 Spectral Size 65536

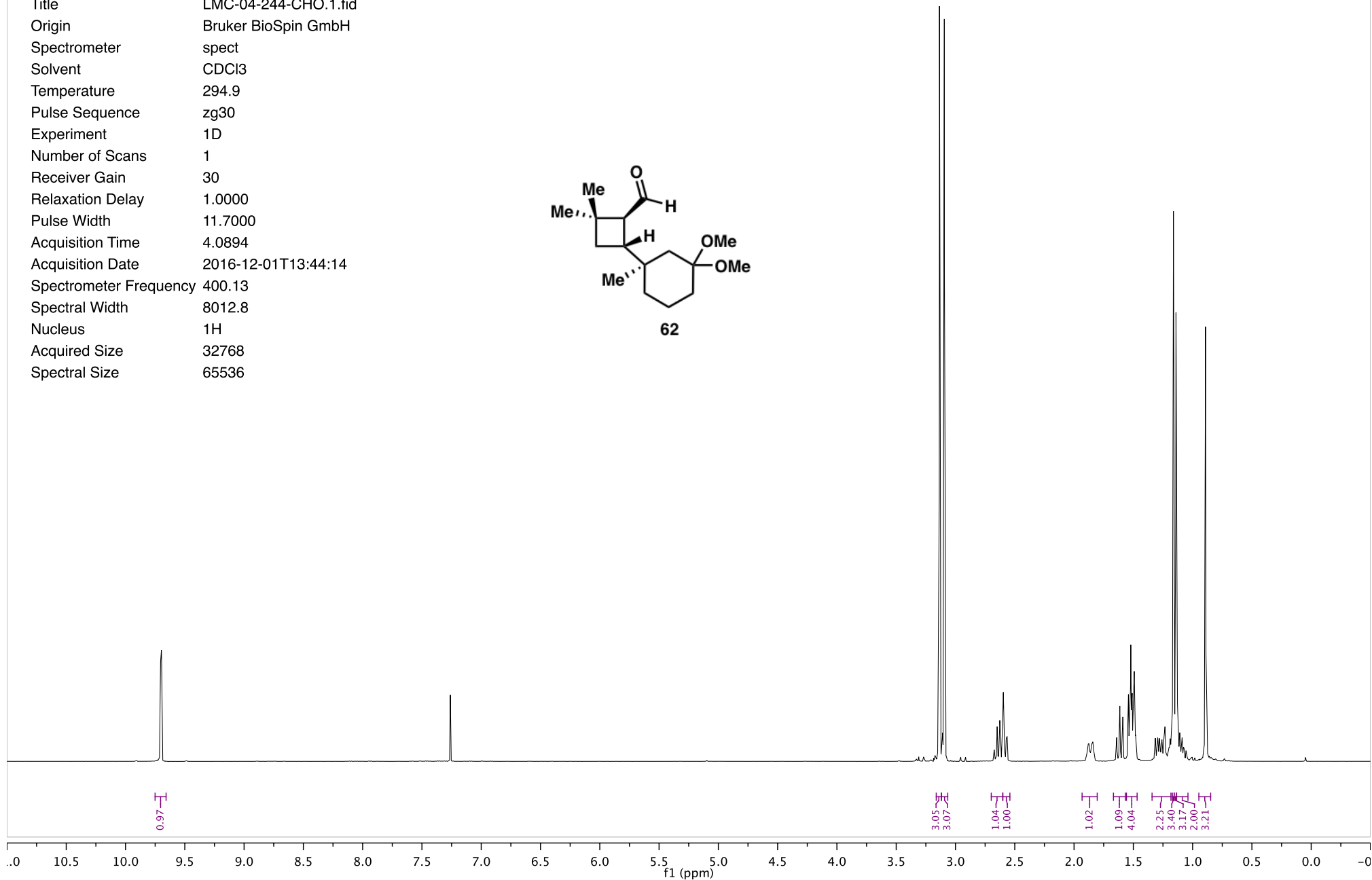
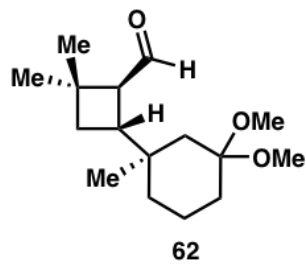
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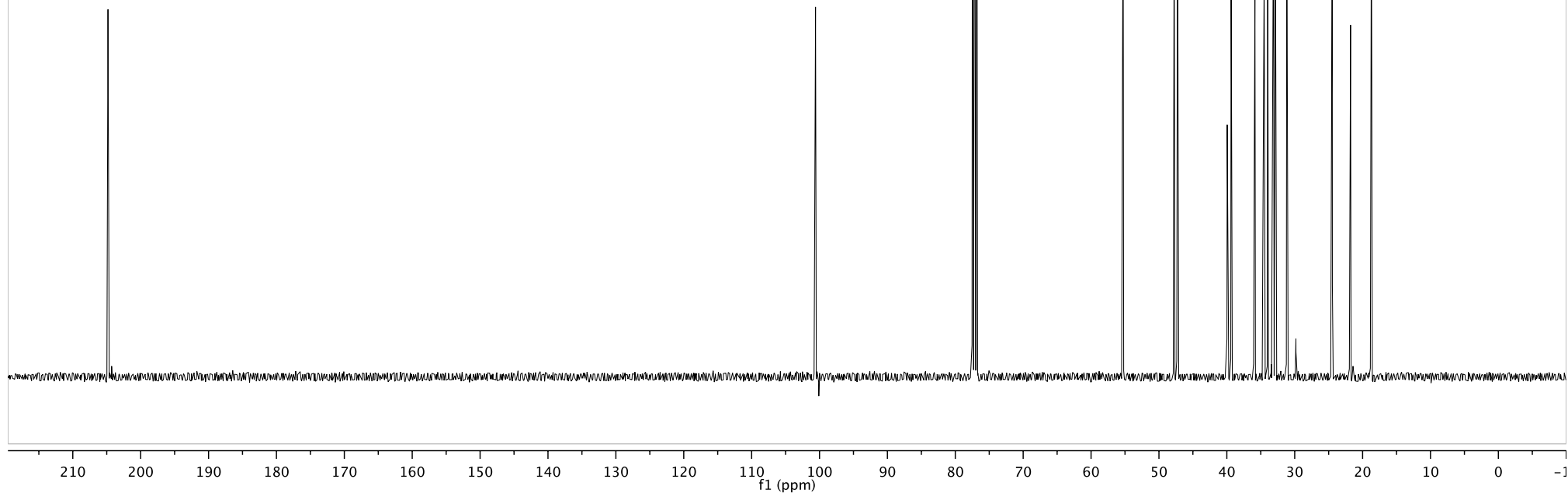
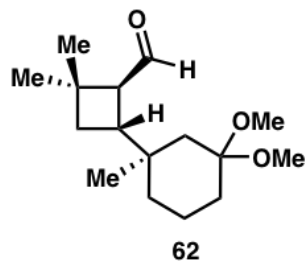




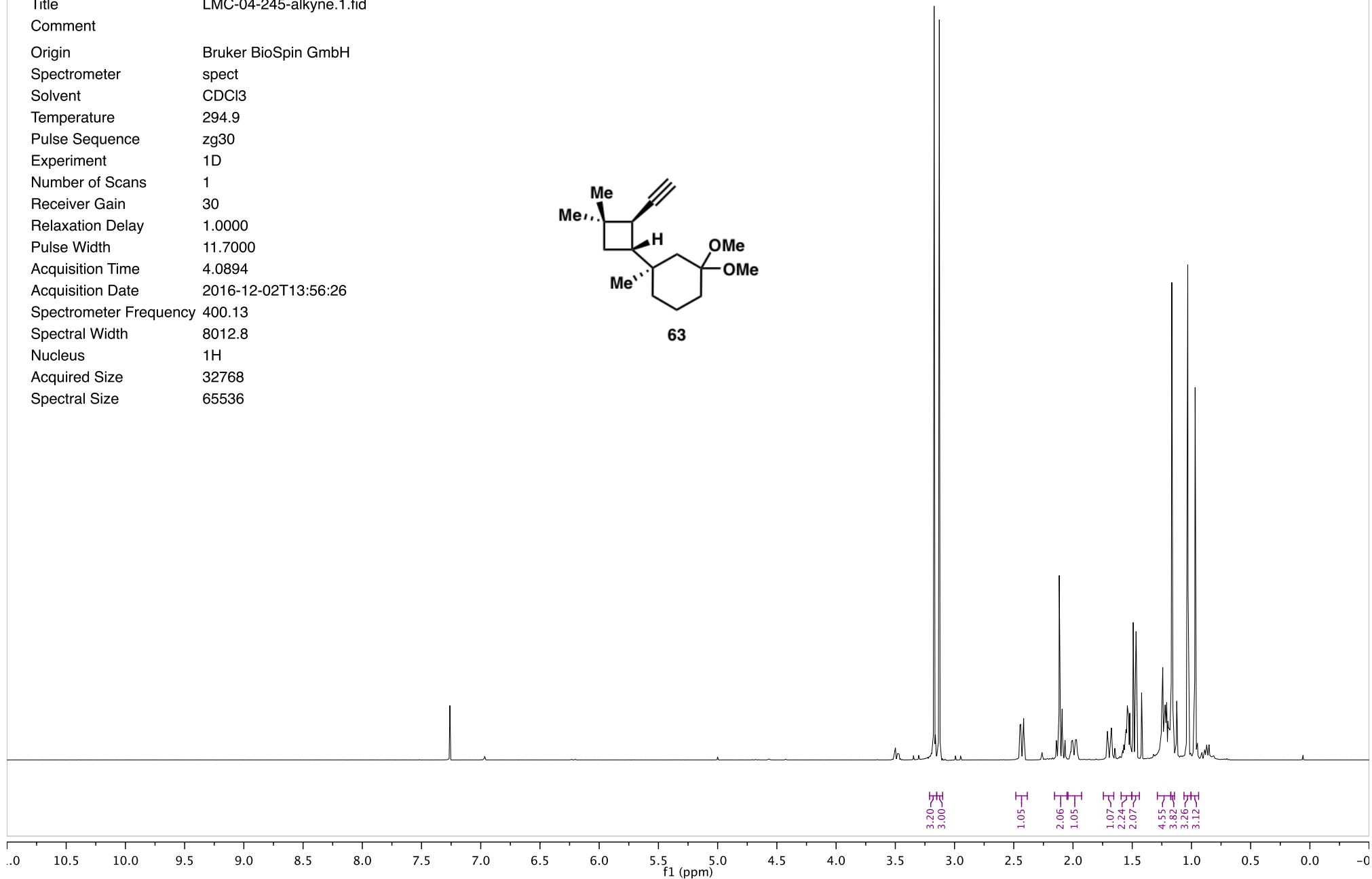
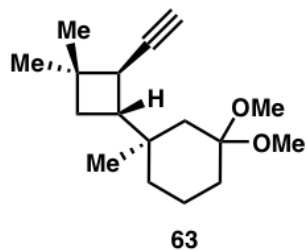
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Data File Name	/ Volumes/ nmrdata/ lchapman/ nmr/ LMC-04-244-CHO/ 1/ fid
Title	LMC-04-244-CHO.1.fid
Origin	Bruker BioSpin GmbH
Spectrometer	spect
Solvent	CDCl3
Temperature	294.9
Pulse Sequence	zg30
Experiment	1D
Number of Scans	1
Receiver Gain	30
Relaxation Delay	1.0000
Pulse Width	11.7000
Acquisition Time	4.0894
Acquisition Date	2016-12-01T13:44:14
Spectrometer Frequency	400.13
Spectral Width	8012.8
Nucleus	1H
Acquired Size	32768
Spectral Size	65536



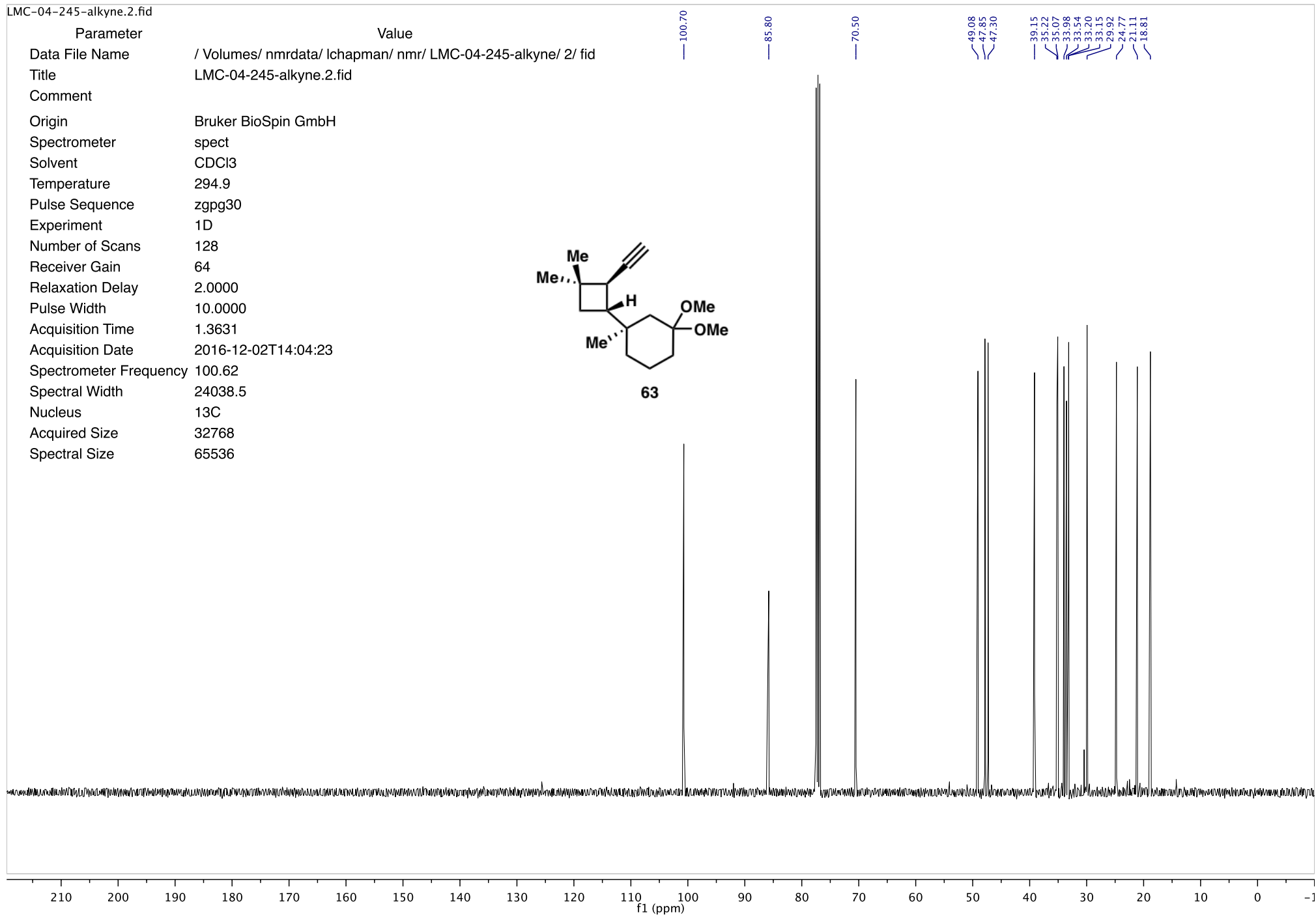
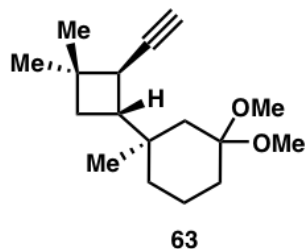
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Data File Name	/ Volumes/ nmrdata/ lchapman/ nmr/ LMC-04-244-CHO/ 2/ fid
Title	LMC-04-244-CHO.2.fid
Origin	Bruker BioSpin GmbH
Spectrometer	spect
Solvent	CDCl3
Temperature	295.0
Pulse Sequence	zgpg30
Experiment	1D
Number of Scans	128
Receiver Gain	79
Relaxation Delay	2.0000
Pulse Width	10.0000
Acquisition Time	1.3631
Acquisition Date	2016-12-01T13:52:11
Spectrometer Frequency	100.62
Spectral Width	24038.5
Nucleus	13C
Acquired Size	32768
Spectral Size	65536



Parameter	Value
Data File Name	/ Volumes/ nmrdata/ lchapman/ nmr/ LMC-04-245-alkyne/ 1/ fid
Title	LMC-04-245-alkyne.1.fid
Comment	
Origin	Bruker BioSpin GmbH
Spectrometer	spect
Solvent	CDCl3
Temperature	294.9
Pulse Sequence	zg30
Experiment	1D
Number of Scans	1
Receiver Gain	30
Relaxation Delay	1.0000
Pulse Width	11.7000
Acquisition Time	4.0894
Acquisition Date	2016-12-02T13:56:26
Spectrometer Frequency	400.13
Spectral Width	8012.8
Nucleus	1H
Acquired Size	32768
Spectral Size	65536

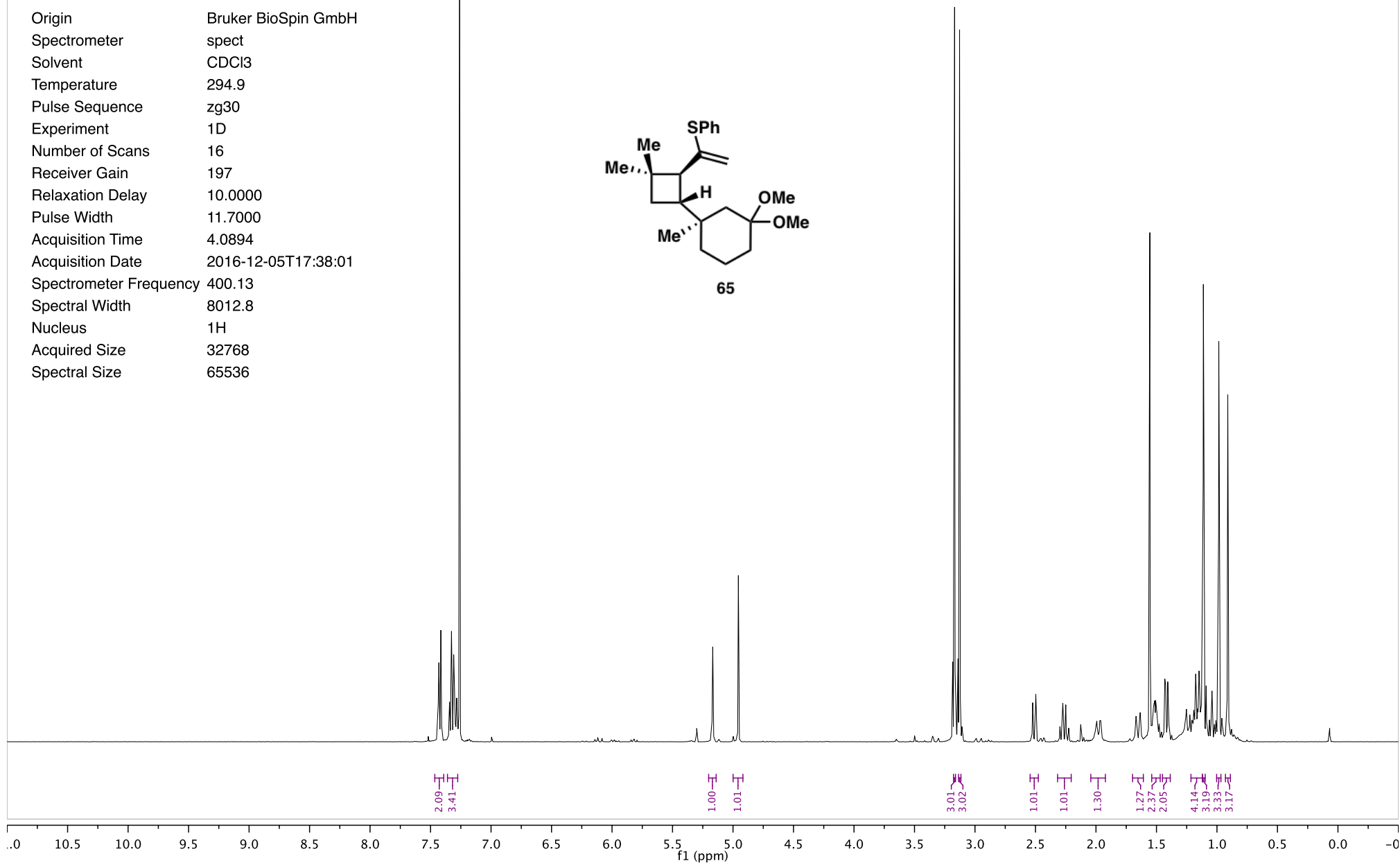
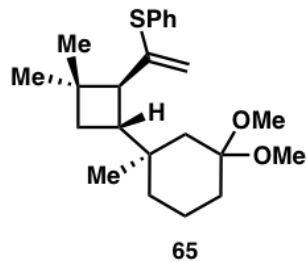


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Data File Name	/ Volumes/ nmrdata/ lchapman/ nmr/ LMC-04-245-alkyne/ 2/ fid
Title	LMC-04-245-alkyne.2.fid
Comment	
Origin	Bruker BioSpin GmbH
Spectrometer	spect
Solvent	CDCl3
Temperature	294.9
Pulse Sequence	zgpg30
Experiment	1D
Number of Scans	128
Receiver Gain	64
Relaxation Delay	2.0000
Pulse Width	10.0000
Acquisition Time	1.3631
Acquisition Date	2016-12-02T14:04:23
Spectrometer Frequency	100.62
Spectral Width	24038.5
Nucleus	13C
Acquired Size	32768
Spectral Size	65536

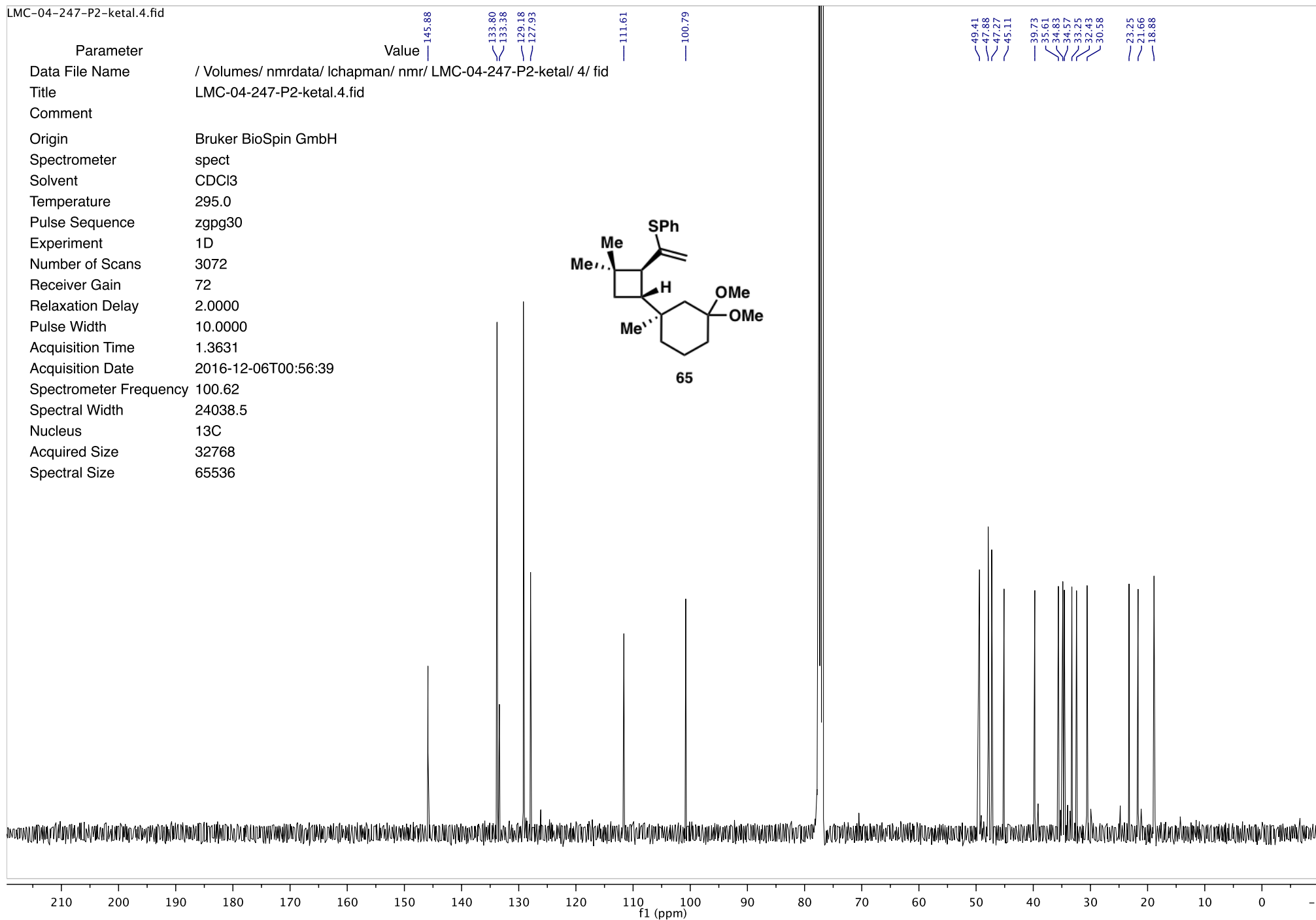
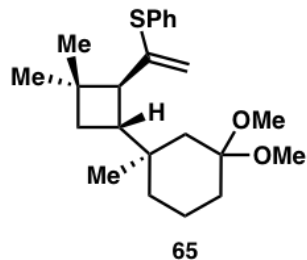


LMC-04-247-P2-ketal.2.fid

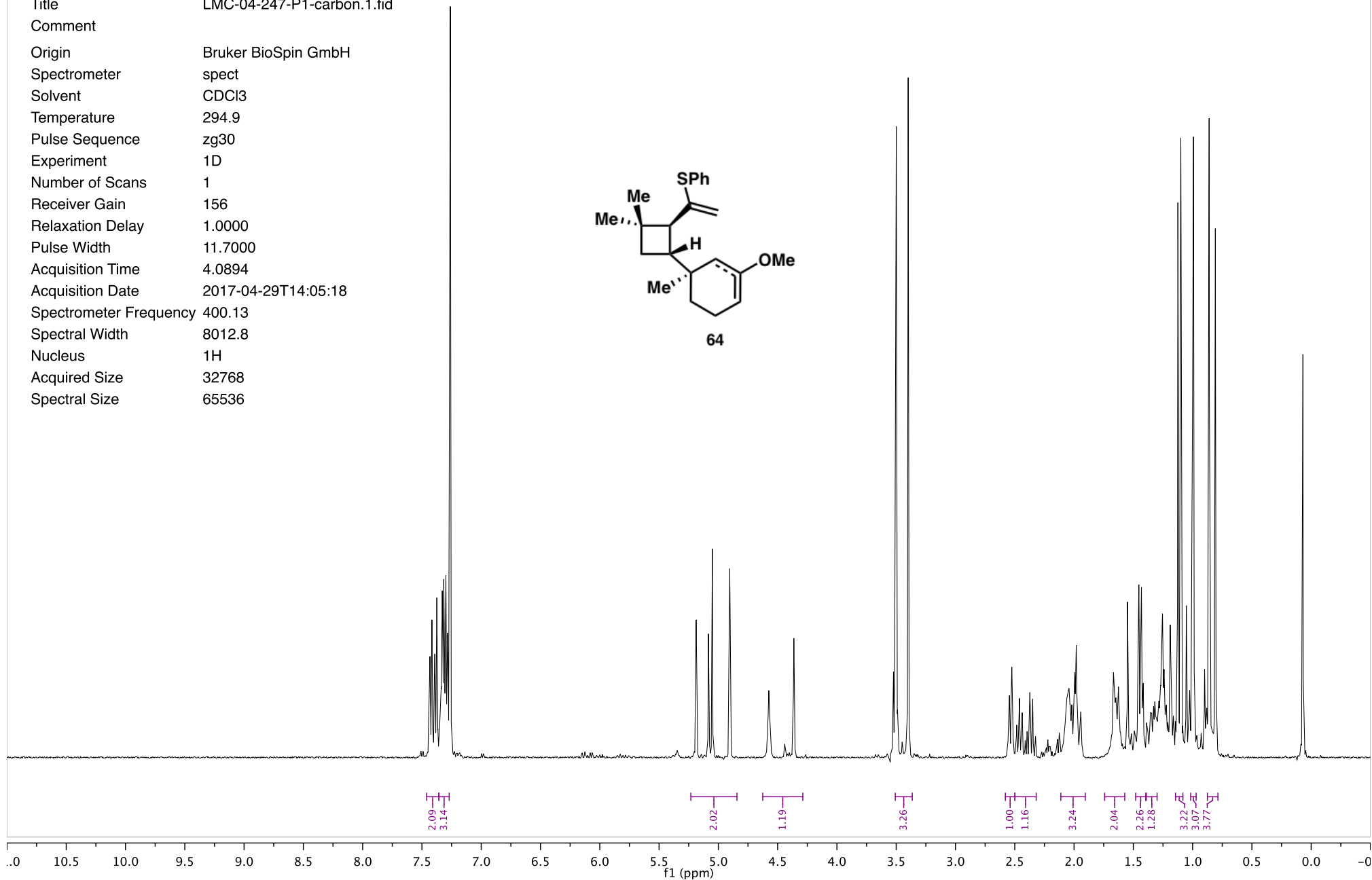
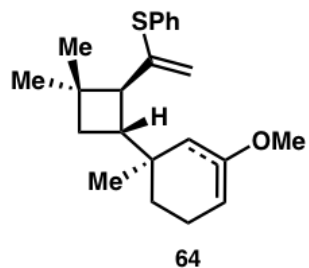
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Data File Name	/ Volumes/ nmrdata/ lchapman/ nmr/ LMC-04-247-P2-ketal/ 2/ fid
Title	LMC-04-247-P2-ketal.2.fid
Comment	
Origin	Bruker BioSpin GmbH
Spectrometer	spect
Solvent	CDCl3
Temperature	294.9
Pulse Sequence	zg30
Experiment	1D
Number of Scans	16
Receiver Gain	197
Relaxation Delay	10.0000
Pulse Width	11.7000
Acquisition Time	4.0894
Acquisition Date	2016-12-05T17:38:01
Spectrometer Frequency	400.13
Spectral Width	8012.8
Nucleus	1H
Acquired Size	32768
Spectral Size	65536



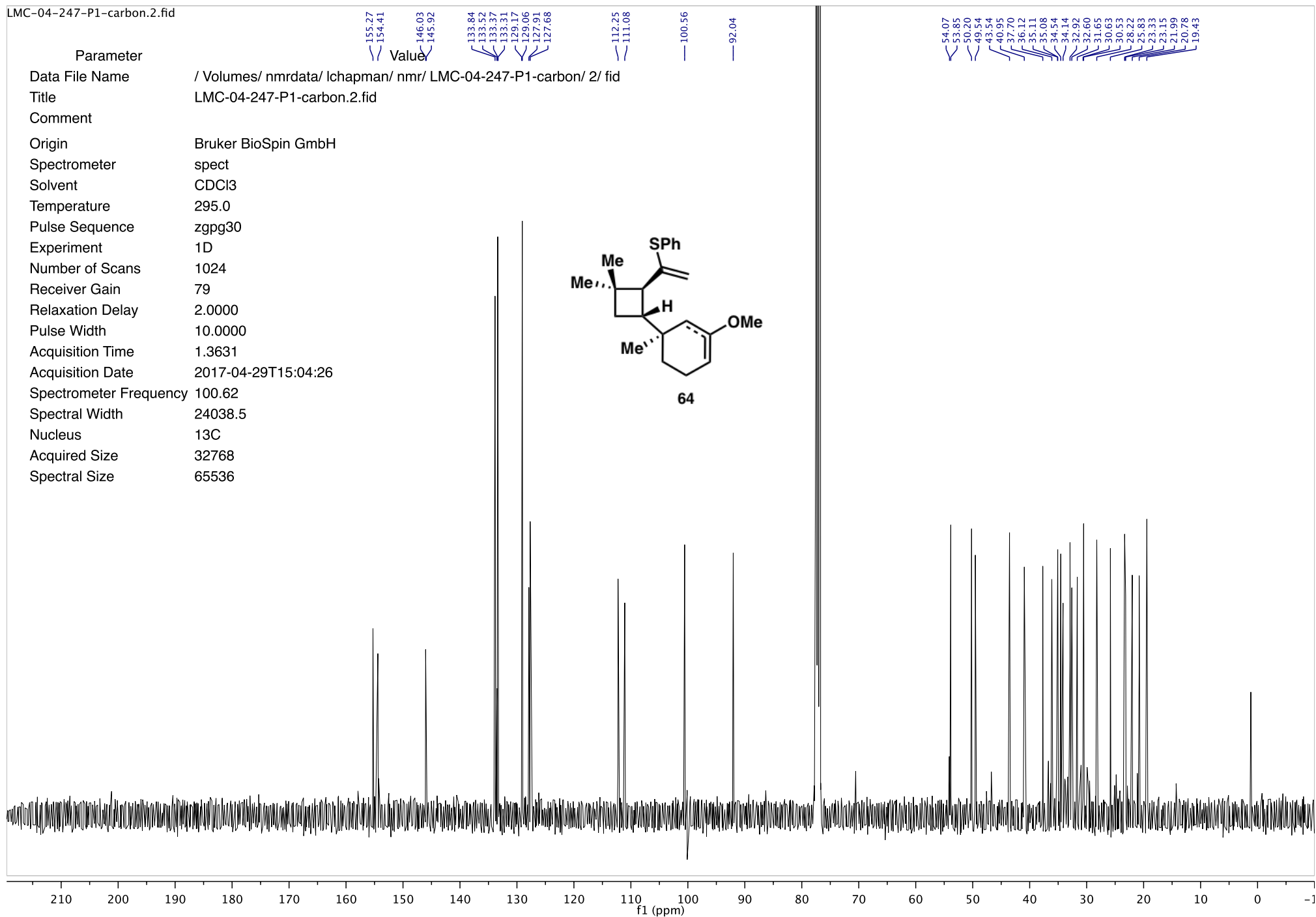
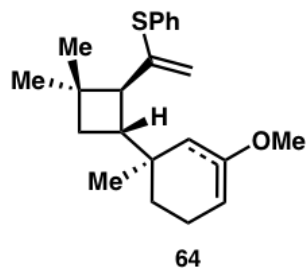
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Data File Name	/ Volumes/ nmrdata/ lchapman/ nmr/ LMC-04-247-P2-ketal/ 4/ fid
Title	LMC-04-247-P2-ketal.4.fid
Comment	
Origin	Bruker BioSpin GmbH
Spectrometer	spect
Solvent	CDCl3
Temperature	295.0
Pulse Sequence	zgpg30
Experiment	1D
Number of Scans	3072
Receiver Gain	72
Relaxation Delay	2.0000
Pulse Width	10.0000
Acquisition Time	1.3631
Acquisition Date	2016-12-06T00:56:39
Spectrometer Frequency	100.62
Spectral Width	24038.5
Nucleus	13C
Acquired Size	32768
Spectral Size	65536



Parameter	Value
Data File Name	/ Volumes/ nmrdata/ lchapman/ nmr/ LMC-04-247-P1-carbon/ 1/ fid
Title	LMC-04-247-P1-carbon.1.fid
Comment	
Origin	Bruker BioSpin GmbH
Spectrometer	spect
Solvent	CDCl3
Temperature	294.9
Pulse Sequence	zg30
Experiment	1D
Number of Scans	1
Receiver Gain	156
Relaxation Delay	1.0000
Pulse Width	11.7000
Acquisition Time	4.0894
Acquisition Date	2017-04-29T14:05:18
Spectrometer Frequency	400.13
Spectral Width	8012.8
Nucleus	1H
Acquired Size	32768
Spectral Size	65536

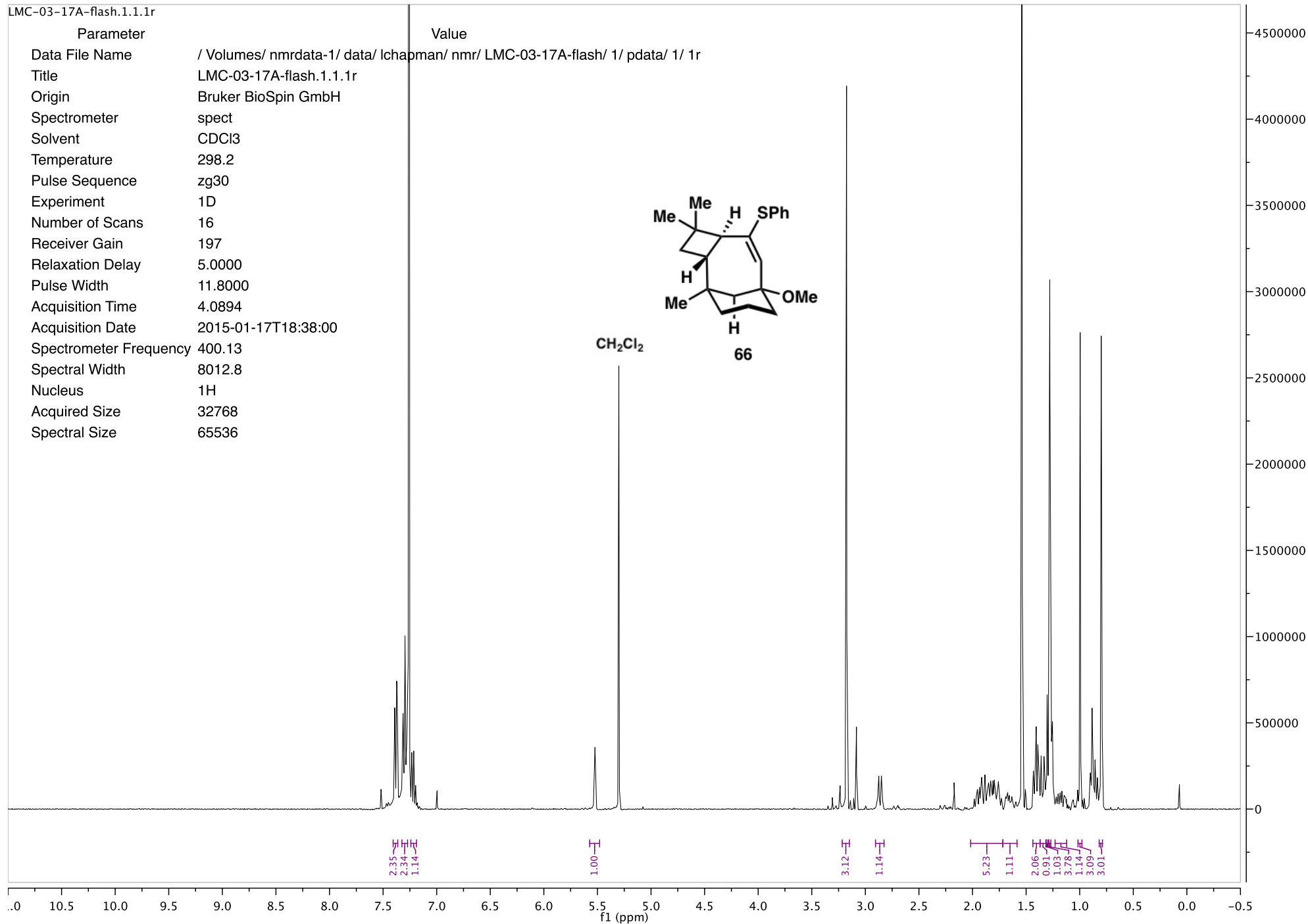
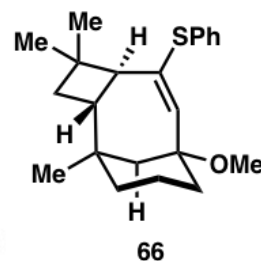


Parameter	Value
Data File Name	/ Volumes/ nmrdata/ lchapman/ nmr/ LMC-04-247-P1-carbon/ 2/ fid
Title	LMC-04-247-P1-carbon.2.fid
Comment	
Origin	Bruker BioSpin GmbH
Spectrometer	spect
Solvent	CDCl3
Temperature	295.0
Pulse Sequence	zgpg30
Experiment	1D
Number of Scans	1024
Receiver Gain	79
Relaxation Delay	2.0000
Pulse Width	10.0000
Acquisition Time	1.3631
Acquisition Date	2017-04-29T15:04:26
Spectrometer Frequency	100.62
Spectral Width	24038.5
Nucleus	13C
Acquired Size	32768
Spectral Size	65536

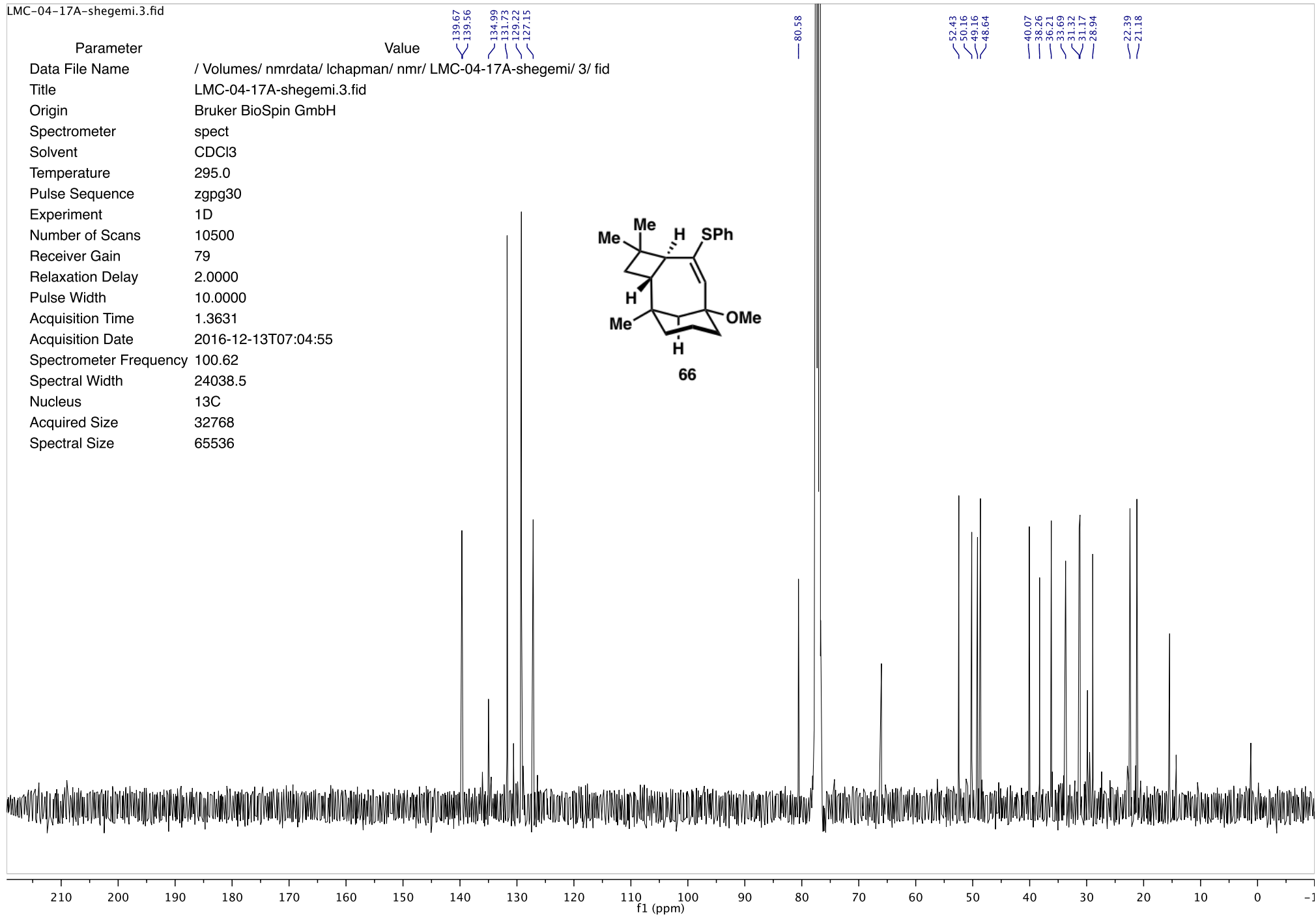
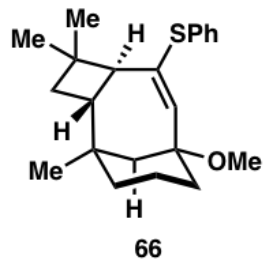


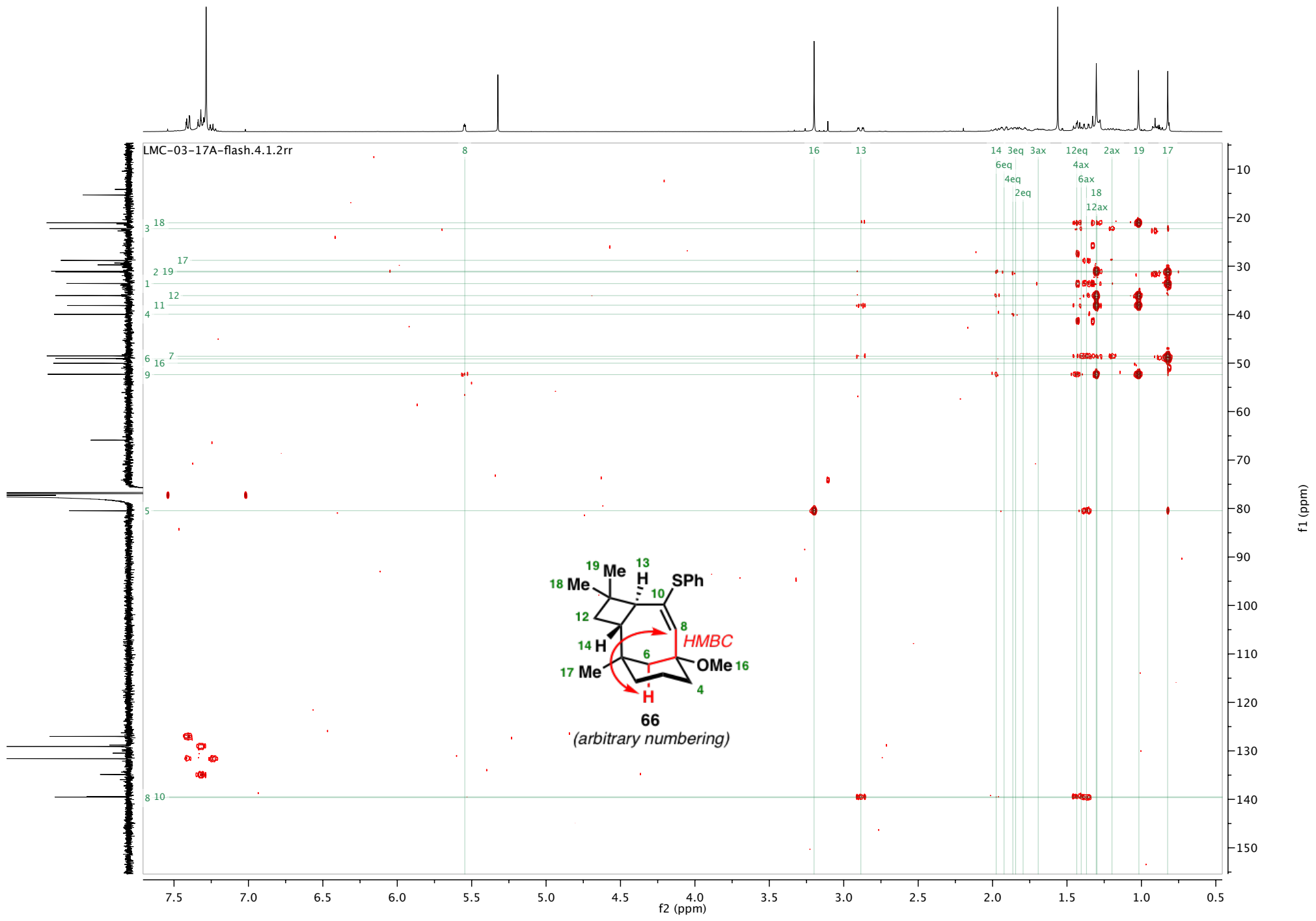
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Parameter	Value
Data File Name	/Volumes/nmrdata-1/data/lchapman/nmr/LMC-03-17A-flash/1/pdata/1/1r
Title	LMC-03-17A-flash.1.1.1r
Origin	Bruker BioSpin GmbH
Spectrometer	spect
Solvent	CDCl3
Temperature	298.2
Pulse Sequence	zg30
Experiment	1D
Number of Scans	16
Receiver Gain	197
Relaxation Delay	5.0000
Pulse Width	11.8000
Acquisition Time	4.0894
Acquisition Date	2015-01-17T18:38:00
Spectrometer Frequency	400.13
Spectral Width	8012.8
Nucleus	1H
Acquired Size	32768
Spectral Size	65536



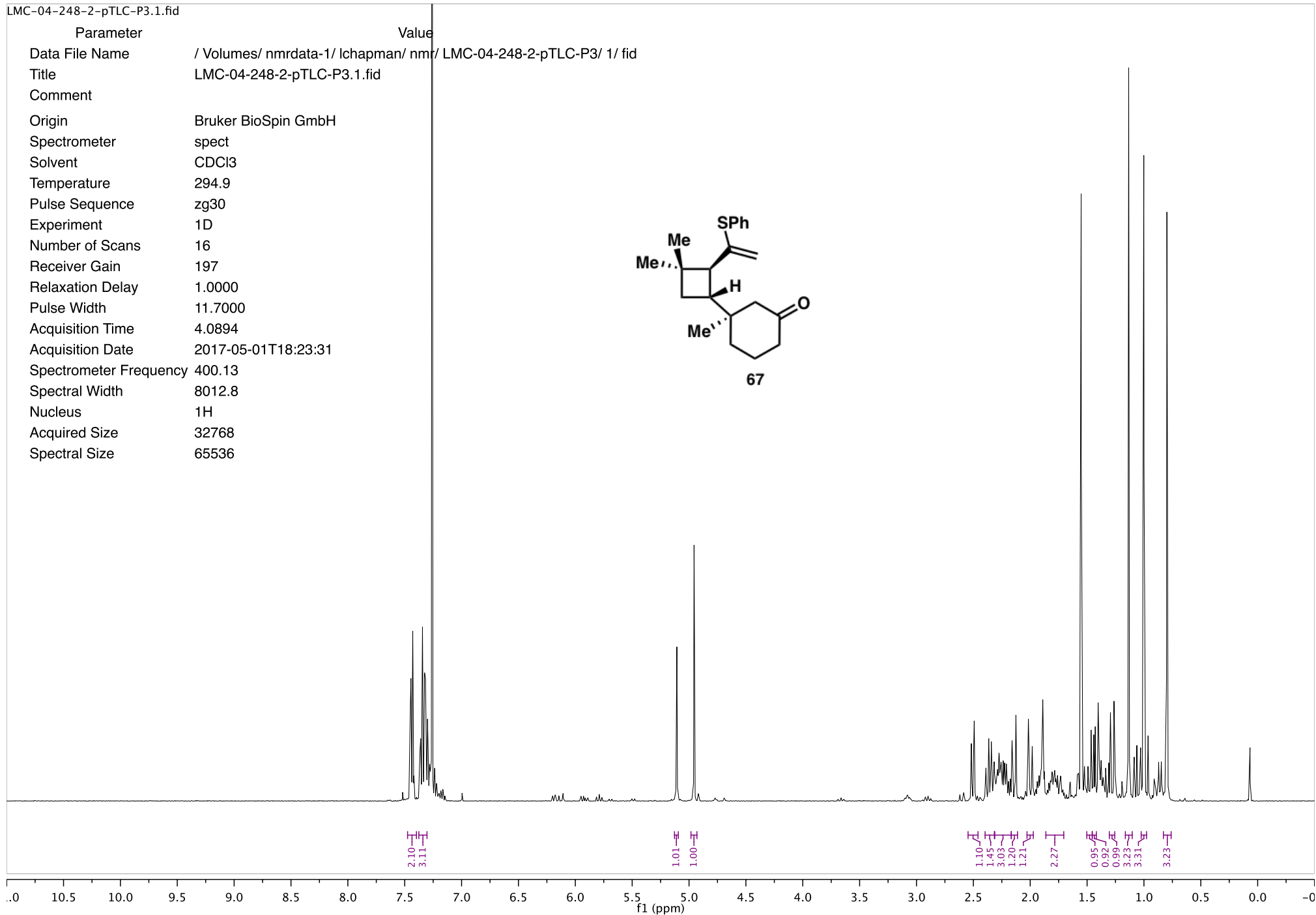
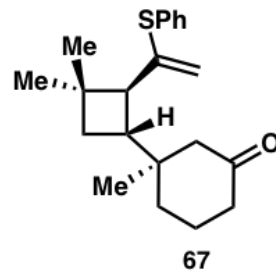
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Data File Name	/ Volumes/ nmrdata/ lchapman/ nmr/ LMC-04-17A-shegemi/ 3/ fid
Title	LMC-04-17A-shegemi.3.fid
Origin	Bruker BioSpin GmbH
Spectrometer	spect
Solvent	CDCl3
Temperature	295.0
Pulse Sequence	zgpg30
Experiment	1D
Number of Scans	10500
Receiver Gain	79
Relaxation Delay	2.0000
Pulse Width	10.0000
Acquisition Time	1.3631
Acquisition Date	2016-12-13T07:04:55
Spectrometer Frequency	100.62
Spectral Width	24038.5
Nucleus	13C
Acquired Size	32768
Spectral Size	65536





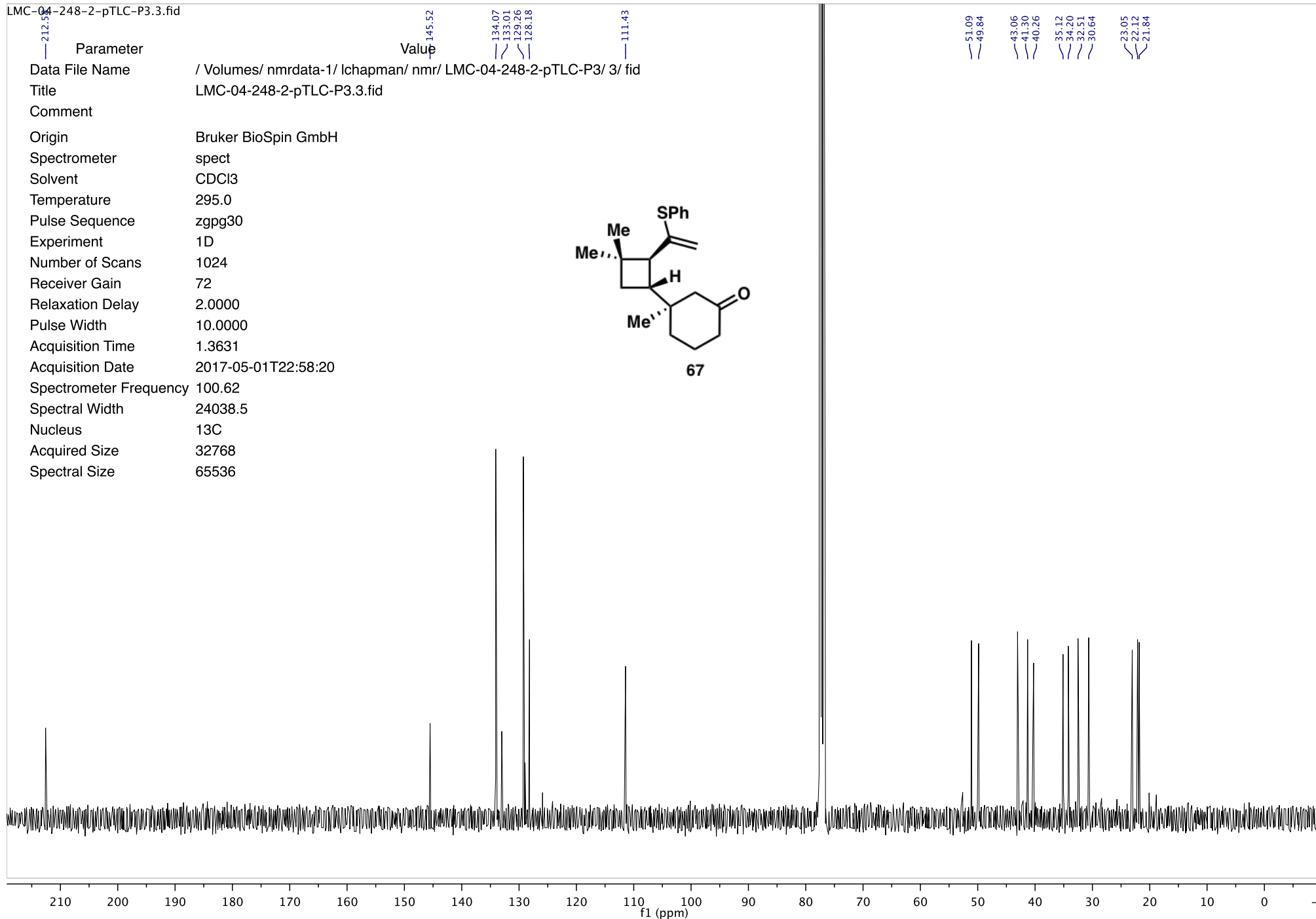
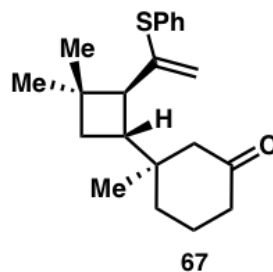
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Parameter	Value
Data File Name	/Volumes/nmrdata-1/Ichapman/nmr/ LMC-04-248-2-pTLC-P3/ 1/ fid
Title	LMC-04-248-2-pTLC-P3.1.fid
Comment	
Origin	Bruker BioSpin GmbH
Spectrometer	spect
Solvent	CDCl3
Temperature	294.9
Pulse Sequence	zg30
Experiment	1D
Number of Scans	16
Receiver Gain	197
Relaxation Delay	1.0000
Pulse Width	11.7000
Acquisition Time	4.0894
Acquisition Date	2017-05-01T18:23:31
Spectrometer Frequency	400.13
Spectral Width	8012.8
Nucleus	1H
Acquired Size	32768
Spectral Size	65536

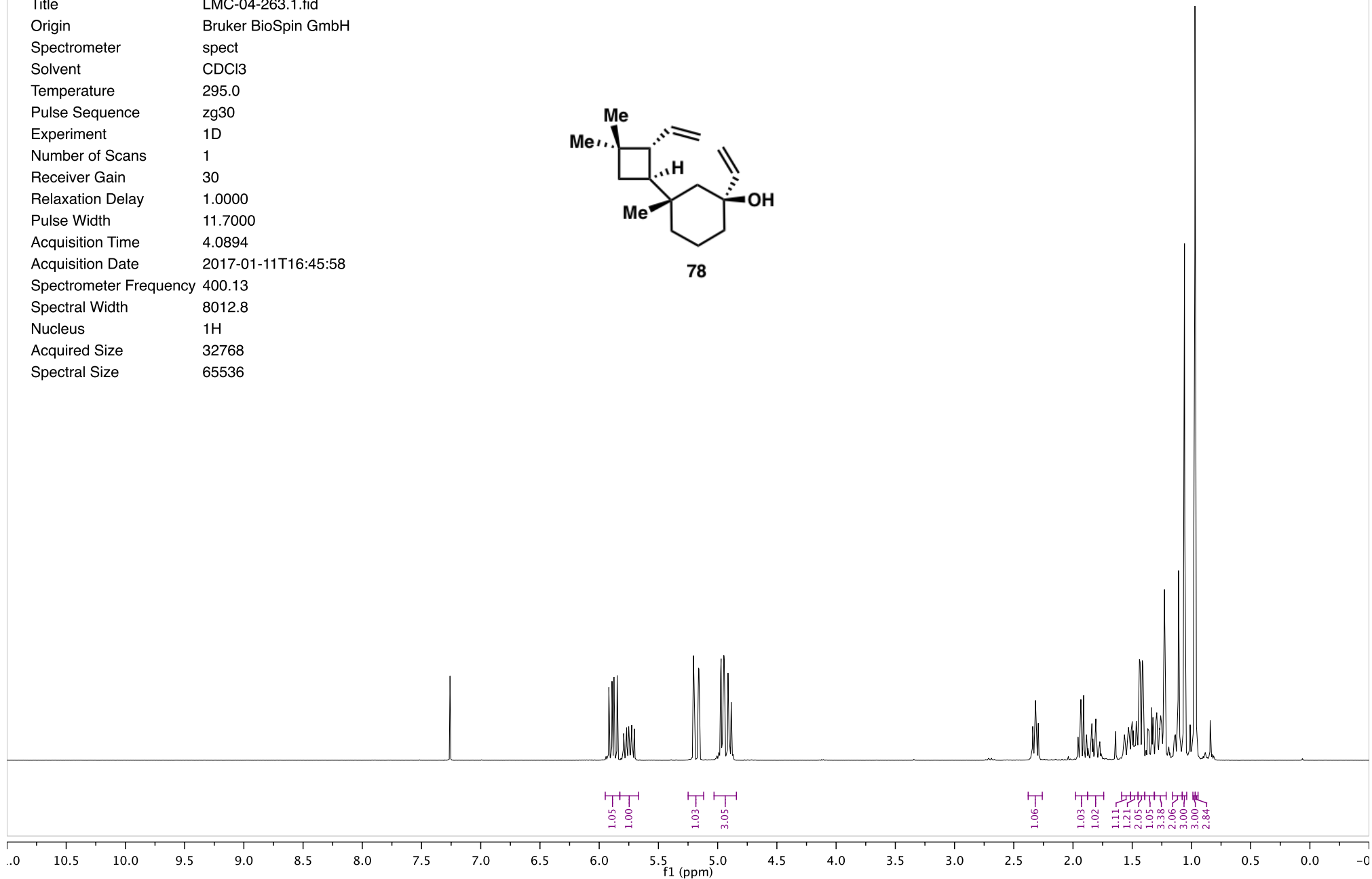
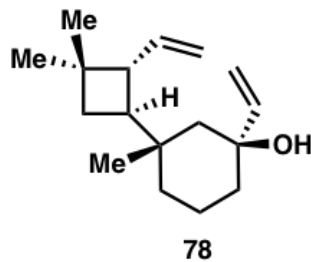


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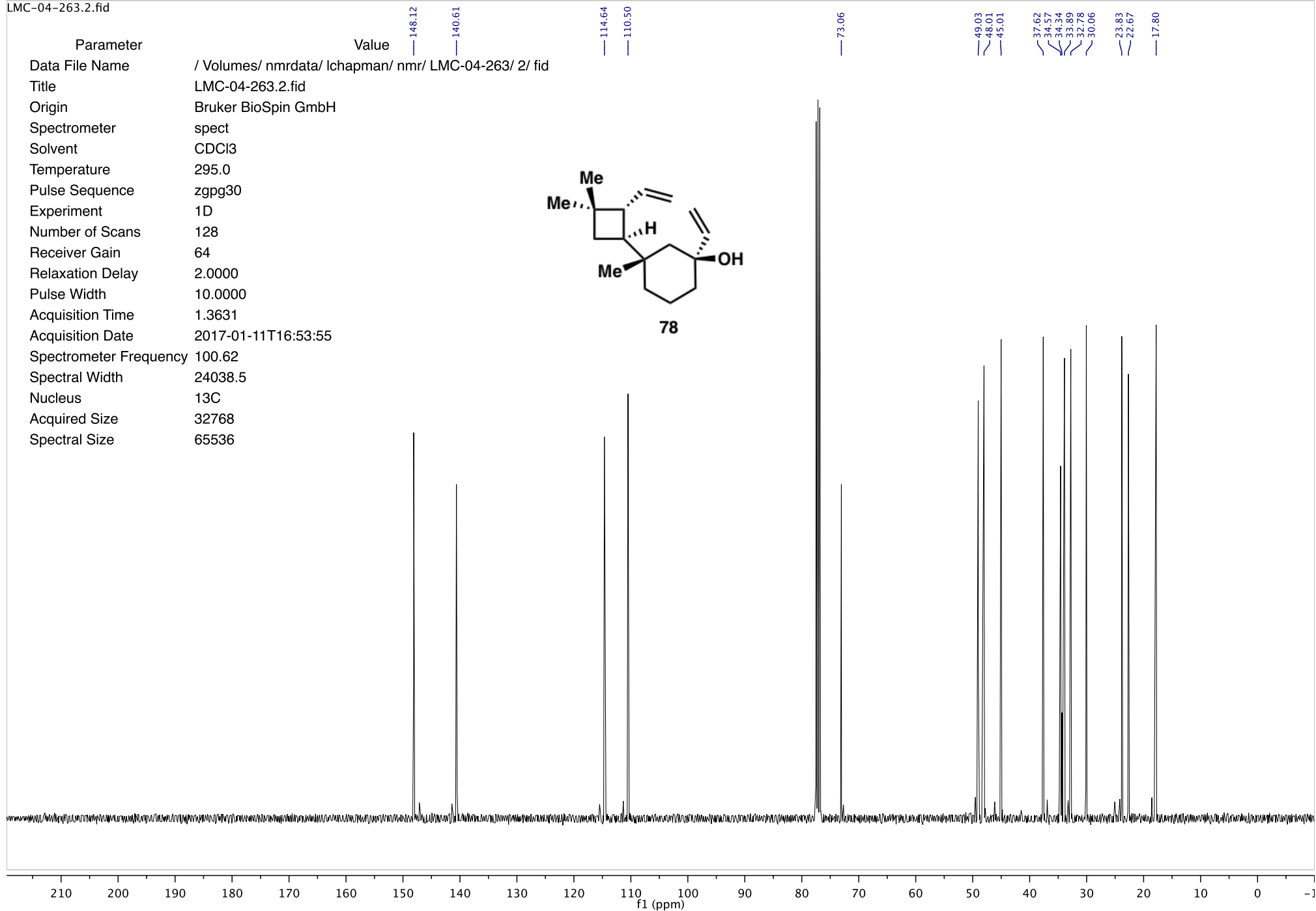
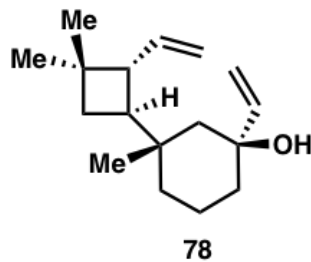
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Data File Name	/ Volumes/ nmrdata-1/ lchapman/ nmr/ LMC-04-248-2-pTLC-P3/ 3/ fid
Title	LMC-04-248-2-pTLC-P3.3.fid
Comment	
Origin	Bruker BioSpin GmbH
Spectrometer	spect
Solvent	CDCl3
Temperature	295.0
Pulse Sequence	zgpg30
Experiment	1D
Number of Scans	1024
Receiver Gain	72
Relaxation Delay	2.0000
Pulse Width	10.0000
Acquisition Time	1.3631
Acquisition Date	2017-05-01T22:58:20
Spectrometer Frequency	100.62
Spectral Width	24038.5
Nucleus	13C
Acquired Size	32768
Spectral Size	65536



Parameter	Value
Data File Name	/ Volumes/ nmrdata/ lchapman/ nmr/ LMC-04-263/ 1/ fid
Title	LMC-04-263.1.fid
Origin	Bruker BioSpin GmbH
Spectrometer	spect
Solvent	CDCl3
Temperature	295.0
Pulse Sequence	zg30
Experiment	1D
Number of Scans	1
Receiver Gain	30
Relaxation Delay	1.0000
Pulse Width	11.7000
Acquisition Time	4.0894
Acquisition Date	2017-01-11T16:45:58
Spectrometer Frequency	400.13
Spectral Width	8012.8
Nucleus	1H
Acquired Size	32768
Spectral Size	65536

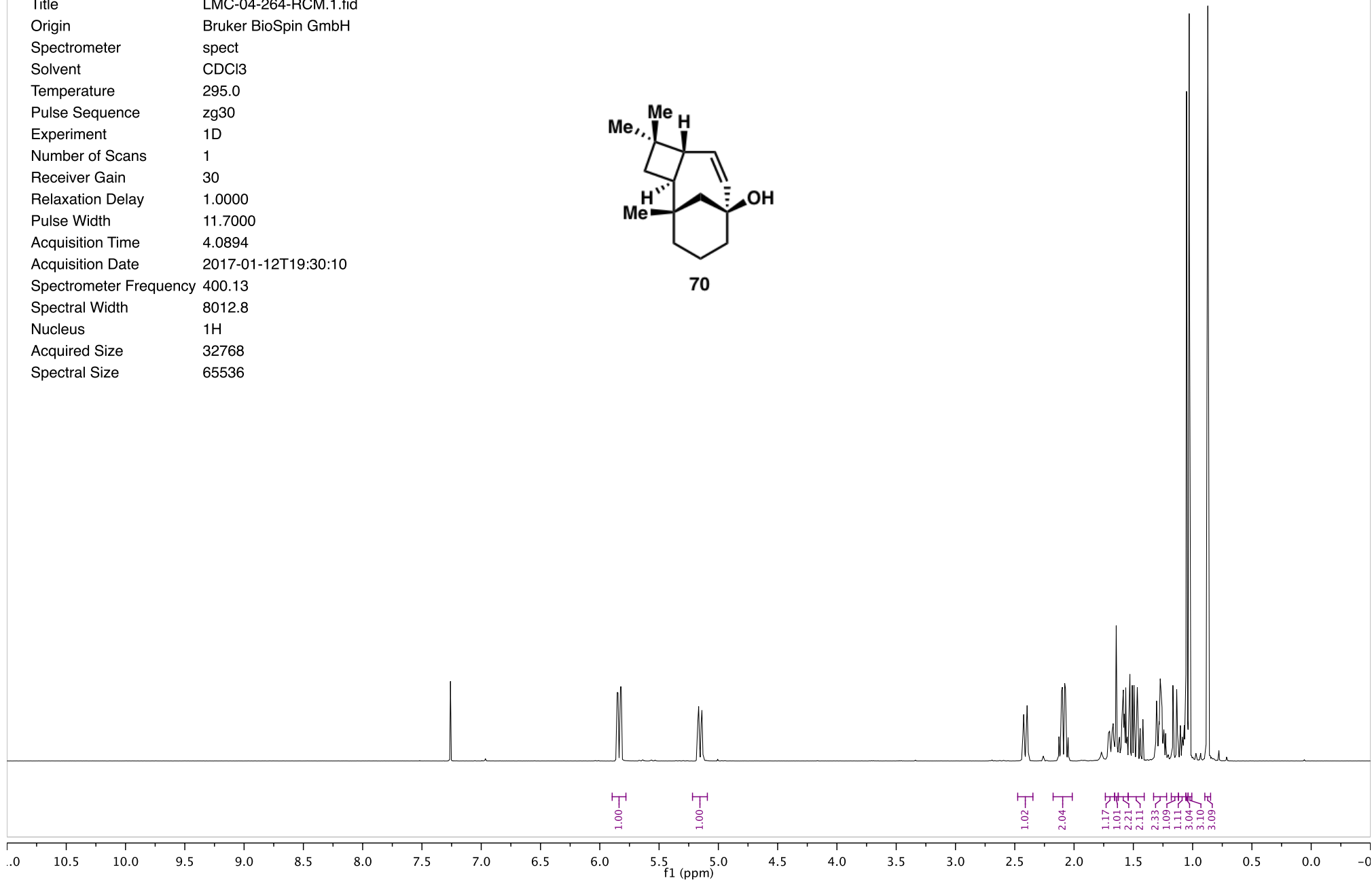
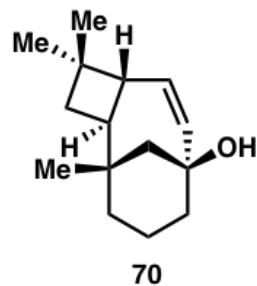


Parameter	Value
Data File Name	/ Volumes/ nmrdata/ lchapman/ nmr/ LMC-04-263/ 2/ fid
Title	LMC-04-263.2.fid
Origin	Bruker BioSpin GmbH
Spectrometer	spect
Solvent	CDCl3
Temperature	295.0
Pulse Sequence	zgpg30
Experiment	1D
Number of Scans	128
Receiver Gain	64
Relaxation Delay	2.0000
Pulse Width	10.0000
Acquisition Time	1.3631
Acquisition Date	2017-01-11T16:53:55
Spectrometer Frequency	100.62
Spectral Width	24038.5
Nucleus	13C
Acquired Size	32768
Spectral Size	65536

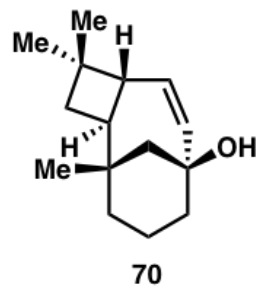


LMC-04-264-RCM.1.fid

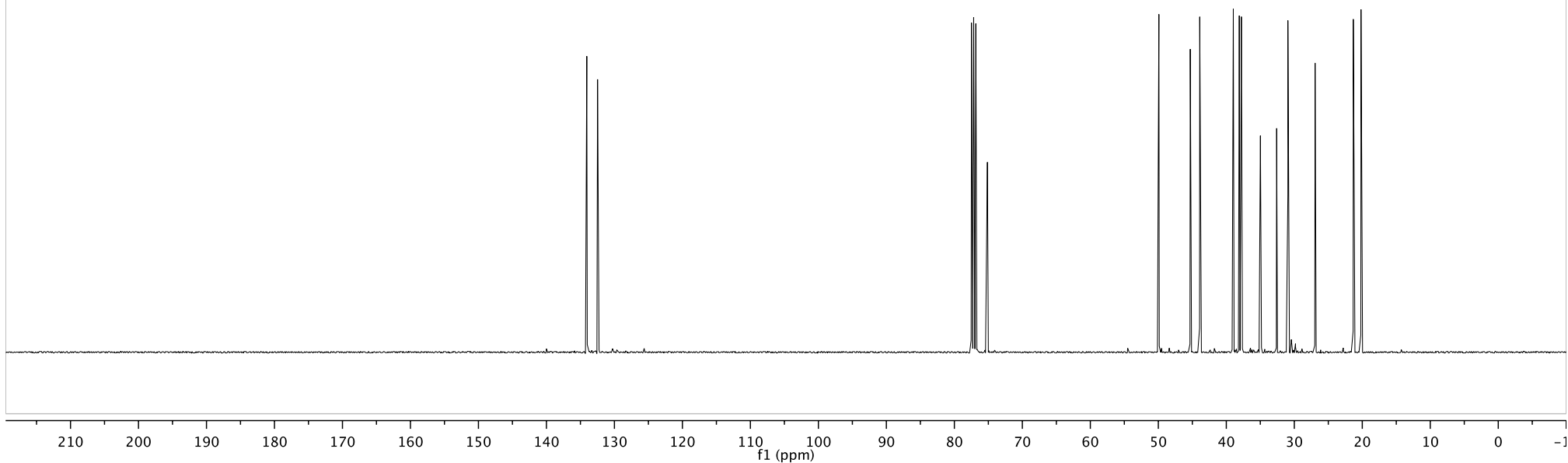
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Data File Name	/ Volumes/ nmrdata/ lchapman/ nmr/ LMC-04-264-RCM/ 1/ fid
Title	LMC-04-264-RCM.1.fid
Origin	Bruker BioSpin GmbH
Spectrometer	spect
Solvent	CDCl3
Temperature	295.0
Pulse Sequence	zg30
Experiment	1D
Number of Scans	1
Receiver Gain	30
Relaxation Delay	1.0000
Pulse Width	11.7000
Acquisition Time	4.0894
Acquisition Date	2017-01-12T19:30:10
Spectrometer Frequency	400.13
Spectral Width	8012.8
Nucleus	1H
Acquired Size	32768
Spectral Size	65536



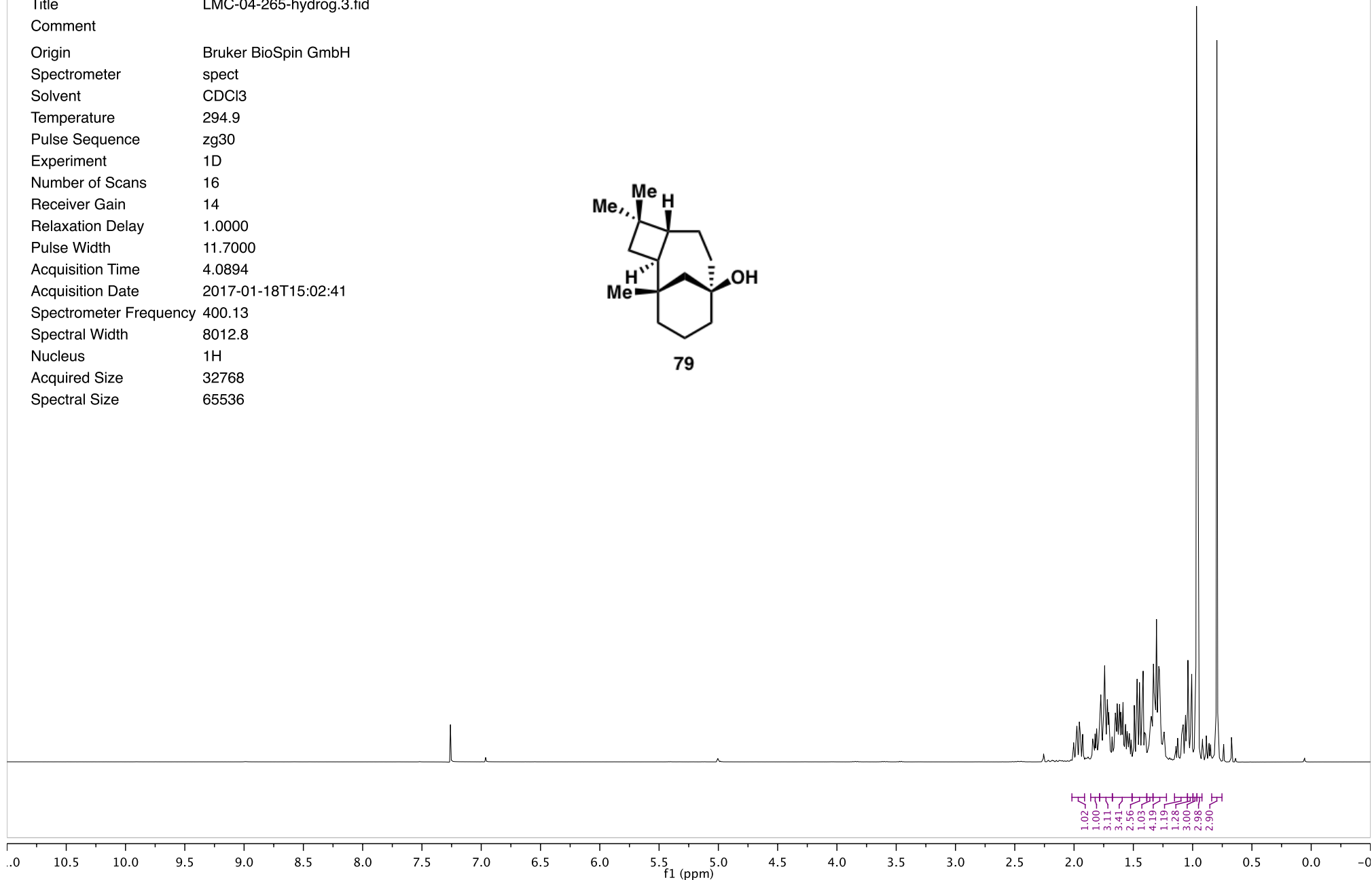
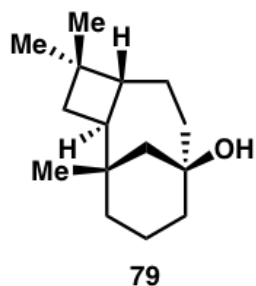
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Data File Name	/ Volumes/ nmrdata/ lchapman/ nmr/ LMC-04-264-RCM/ 2/ fid
Title	LMC-04-264-RCM.2.fid
Origin	Bruker BioSpin GmbH
Spectrometer	spect
Solvent	CDCl3
Temperature	295.0
Pulse Sequence	zgpg30
Experiment	1D
Number of Scans	1024
Receiver Gain	88
Relaxation Delay	2.0000
Pulse Width	10.0000
Acquisition Time	1.3631
Acquisition Date	2017-01-12T19:29:40
Spectrometer Frequency	100.62
Spectral Width	24038.5
Nucleus	13C
Acquired Size	32768
Spectral Size	65536



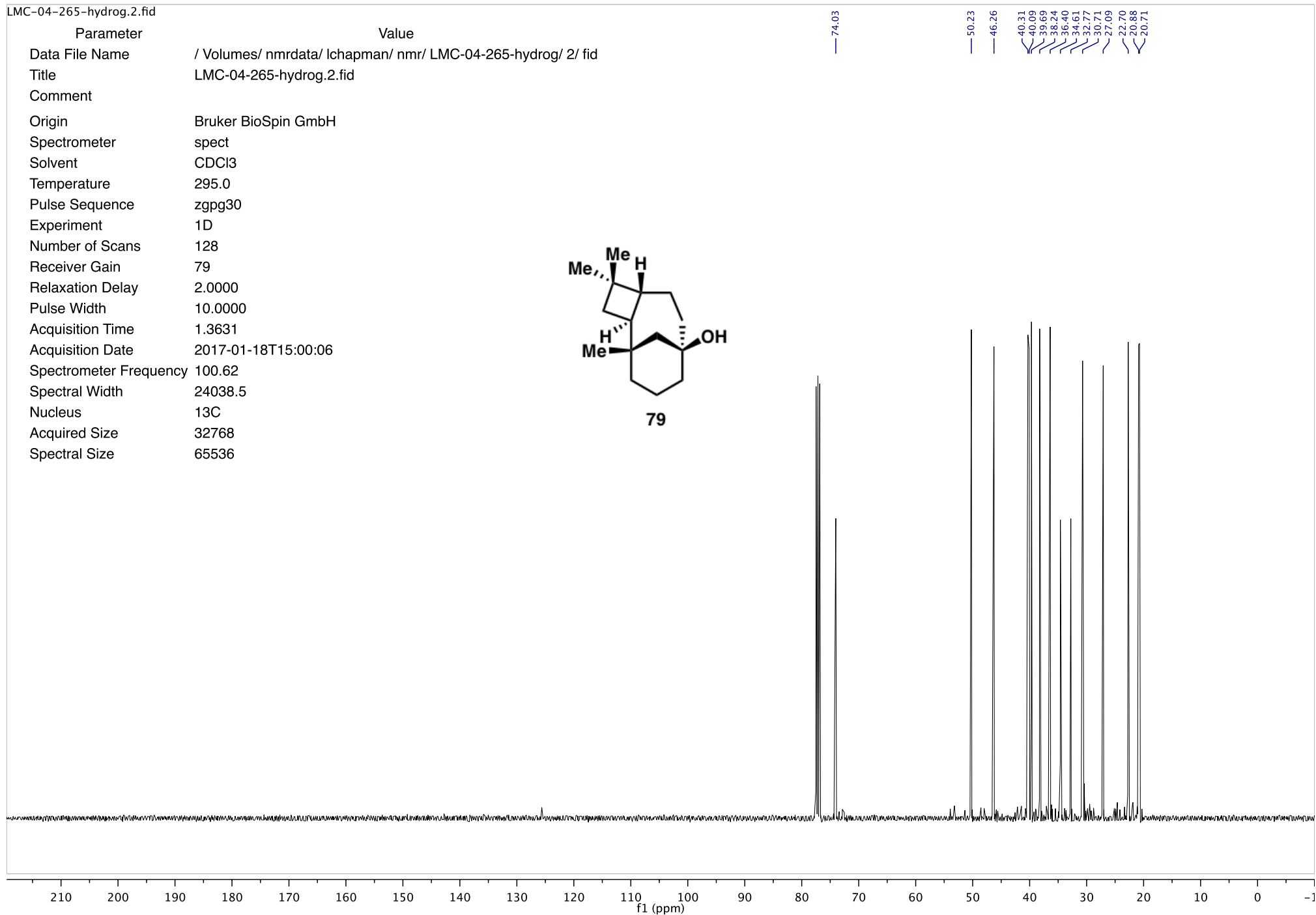
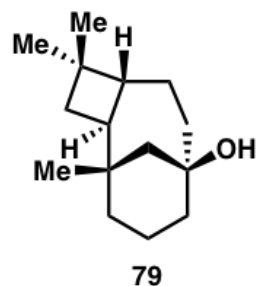
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38.97
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34.99
32.60
30.94
26.94
21.32
20.19



Parameter	Value
Data File Name	/ Volumes/ nmrdata/ lchapman/ nmr/ LMC-04-265-hydrog/ 3/ fid
Title	LMC-04-265-hydrog.3.fid
Comment	
Origin	Bruker BioSpin GmbH
Spectrometer	spect
Solvent	CDCl3
Temperature	294.9
Pulse Sequence	zg30
Experiment	1D
Number of Scans	16
Receiver Gain	14
Relaxation Delay	1.0000
Pulse Width	11.7000
Acquisition Time	4.0894
Acquisition Date	2017-01-18T15:02:41
Spectrometer Frequency	400.13
Spectral Width	8012.8
Nucleus	1H
Acquired Size	32768
Spectral Size	65536

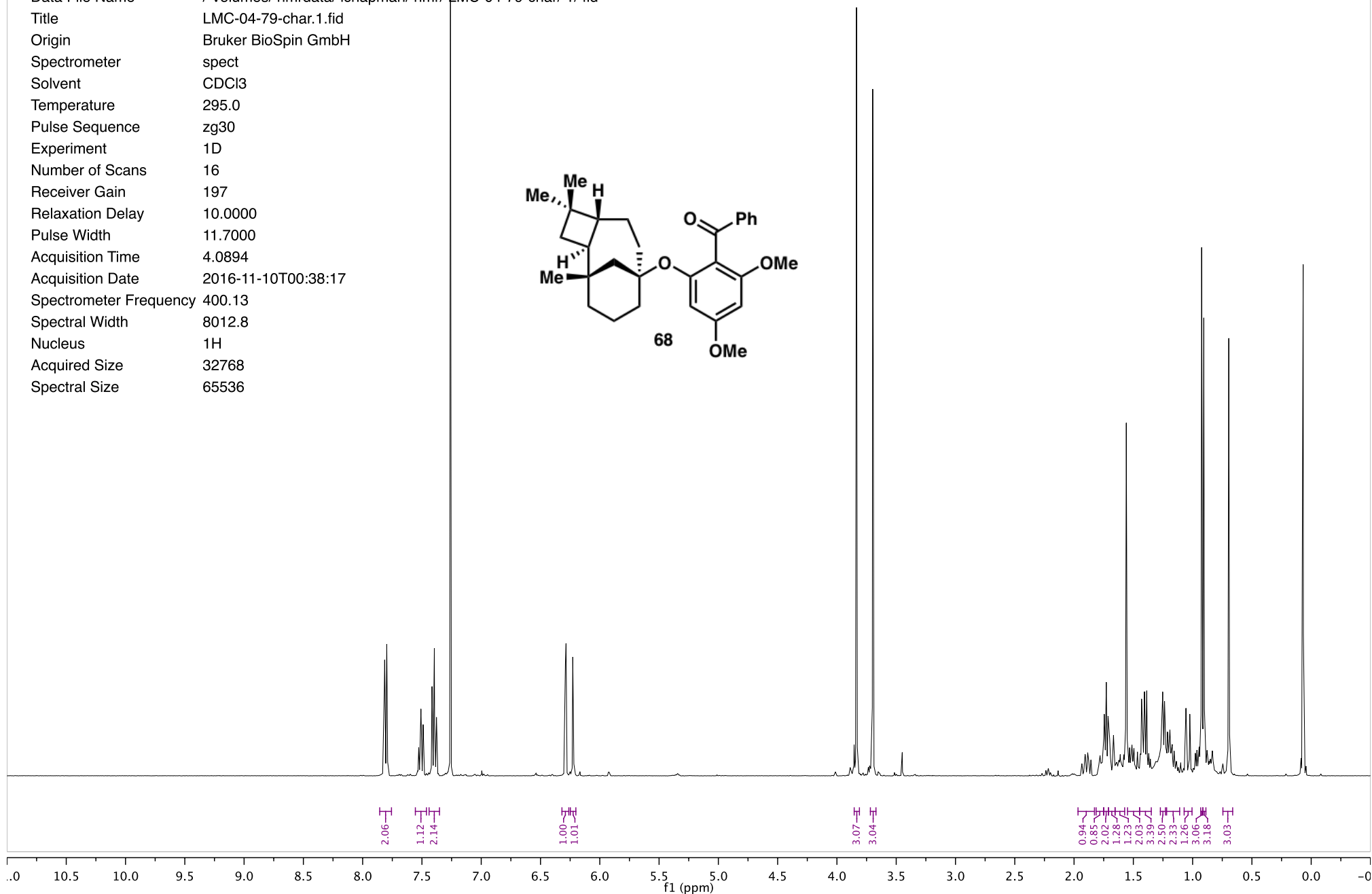
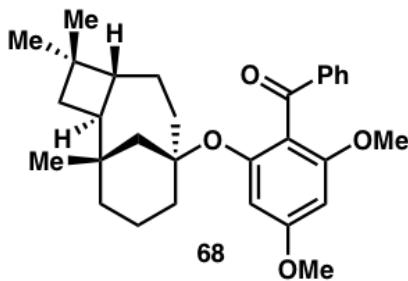


Parameter	Value
Data File Name	/ Volumes/ nmrdata/ lchapman/ nmr/ LMC-04-265-hydrog/ 2/ fid
Title	LMC-04-265-hydrog.2.fid
Comment	
Origin	Bruker BioSpin GmbH
Spectrometer	spect
Solvent	CDCl3
Temperature	295.0
Pulse Sequence	zgpg30
Experiment	1D
Number of Scans	128
Receiver Gain	79
Relaxation Delay	2.0000
Pulse Width	10.0000
Acquisition Time	1.3631
Acquisition Date	2017-01-18T15:00:06
Spectrometer Frequency	100.62
Spectral Width	24038.5
Nucleus	13C
Acquired Size	32768
Spectral Size	65536



LMC-04-79-char.1.fid

Parameter	Value
Data File Name	/ Volumes/ nmrdata/ lchapman/ nmr/ LMC-04-79-char/ 1/ fid
Title	LMC-04-79-char.1.fid
Origin	Bruker BioSpin GmbH
Spectrometer	spect
Solvent	CDCl3
Temperature	295.0
Pulse Sequence	zg30
Experiment	1D
Number of Scans	16
Receiver Gain	197
Relaxation Delay	10.0000
Pulse Width	11.7000
Acquisition Time	4.0894
Acquisition Date	2016-11-10T00:38:17
Spectrometer Frequency	400.13
Spectral Width	8012.8
Nucleus	1H
Acquired Size	32768
Spectral Size	65536



Parameter Value

Data File Name / Volumes/ nmrdata/ lchapman/ nmr/ LMC-04-267-pTLC/ 3/ fid

Title LMC-04-267-pTLC.3.fid

Origin Bruker BioSpin GmbH

Spectrometer spect

Solvent CDCl3

Temperature 295.0

Pulse Sequence zgpg30

Experiment 1D

Number of Scans 1024

Receiver Gain 64

Relaxation Delay 2.0000

Pulse Width 10.0000

Acquisition Time 1.3631

Acquisition Date 2017-01-21T02:30:53

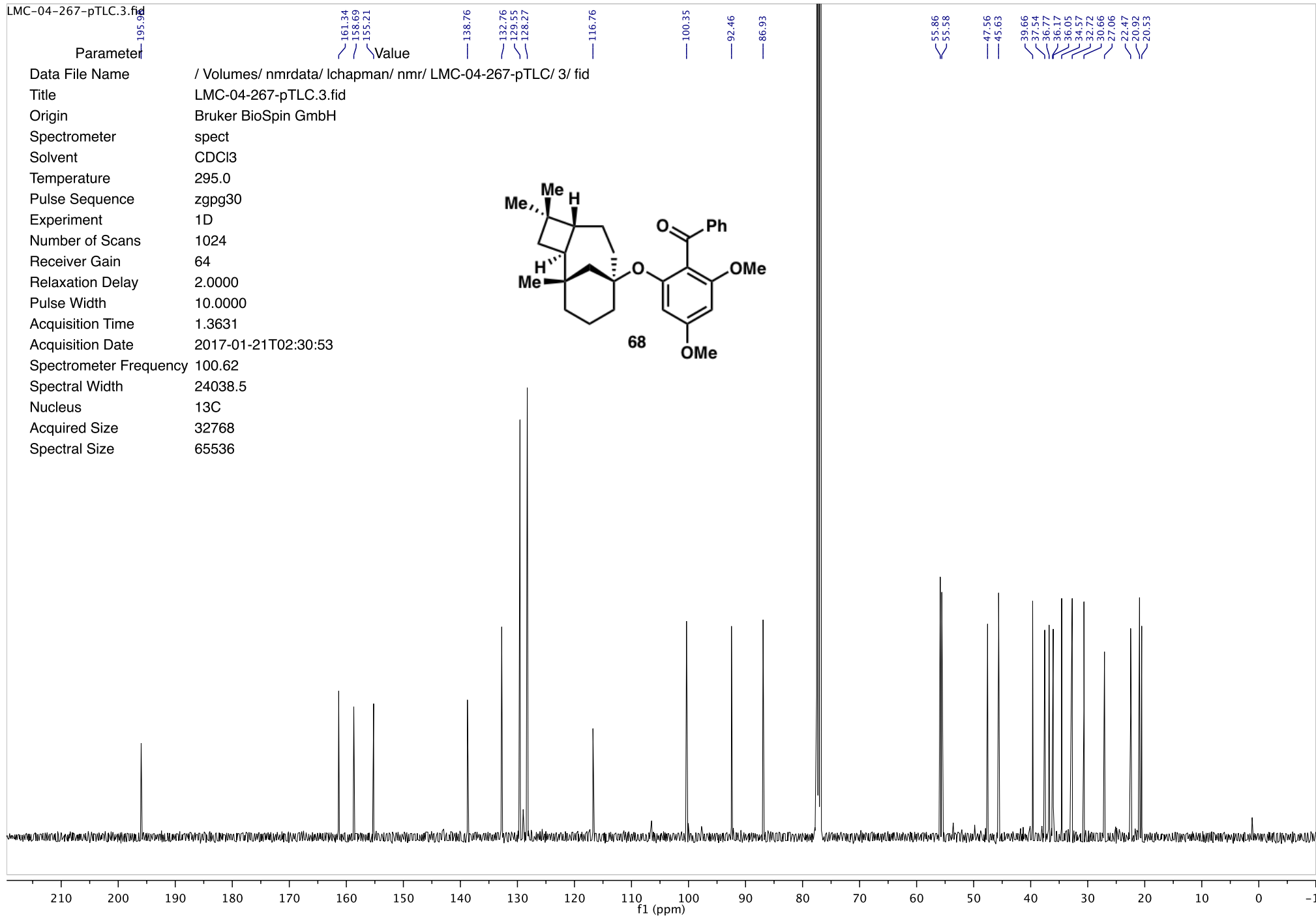
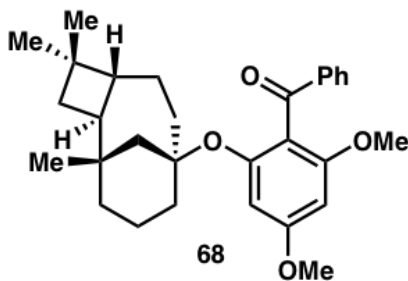
Spectrometer Frequency 100.62

Spectral Width 24038.5

Nucleus 13C

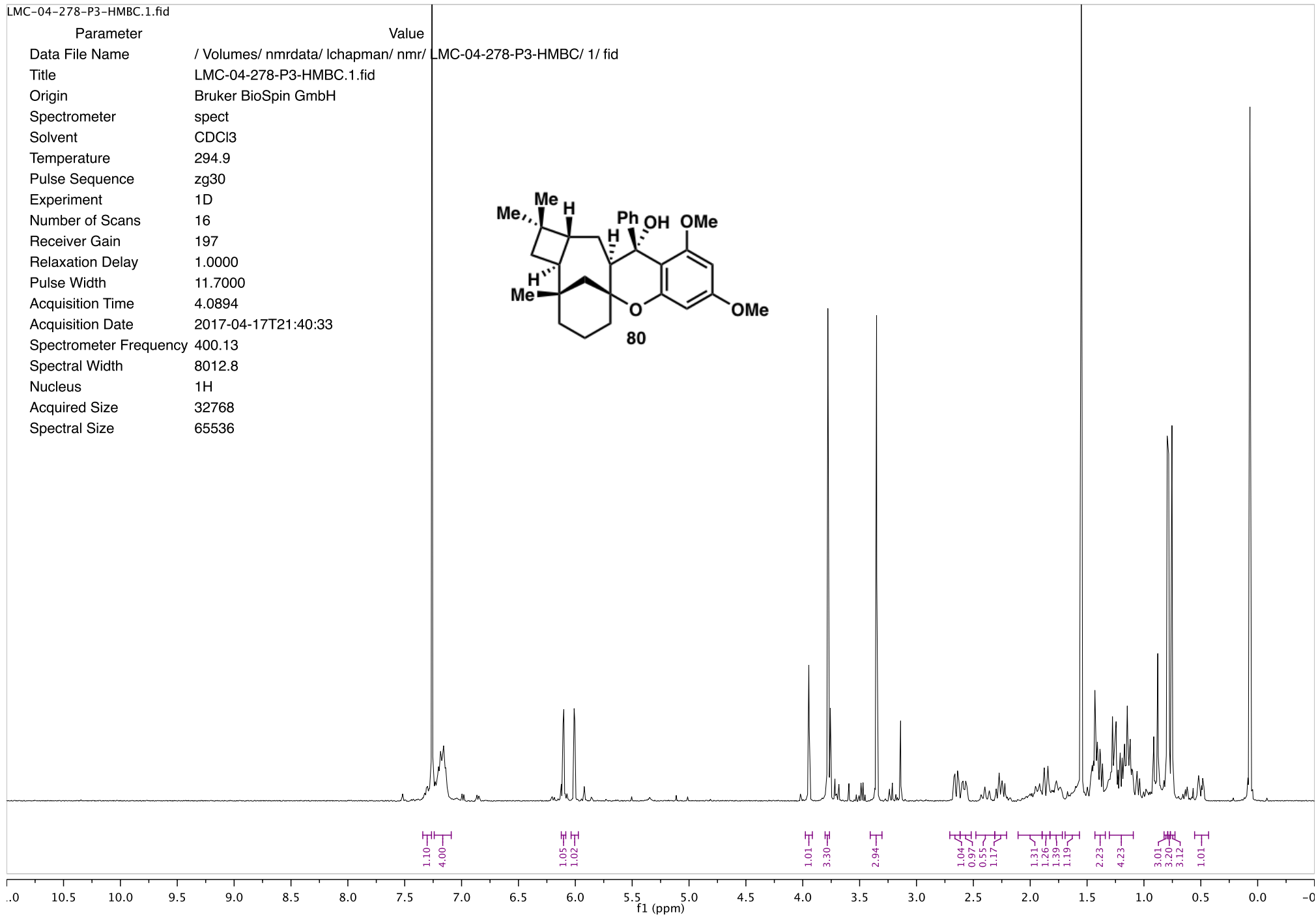
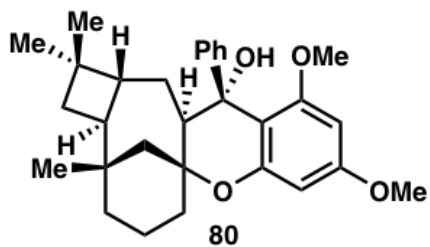
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Spectral Size 65536



LMC-04-278-P3-HMBC.1.fid

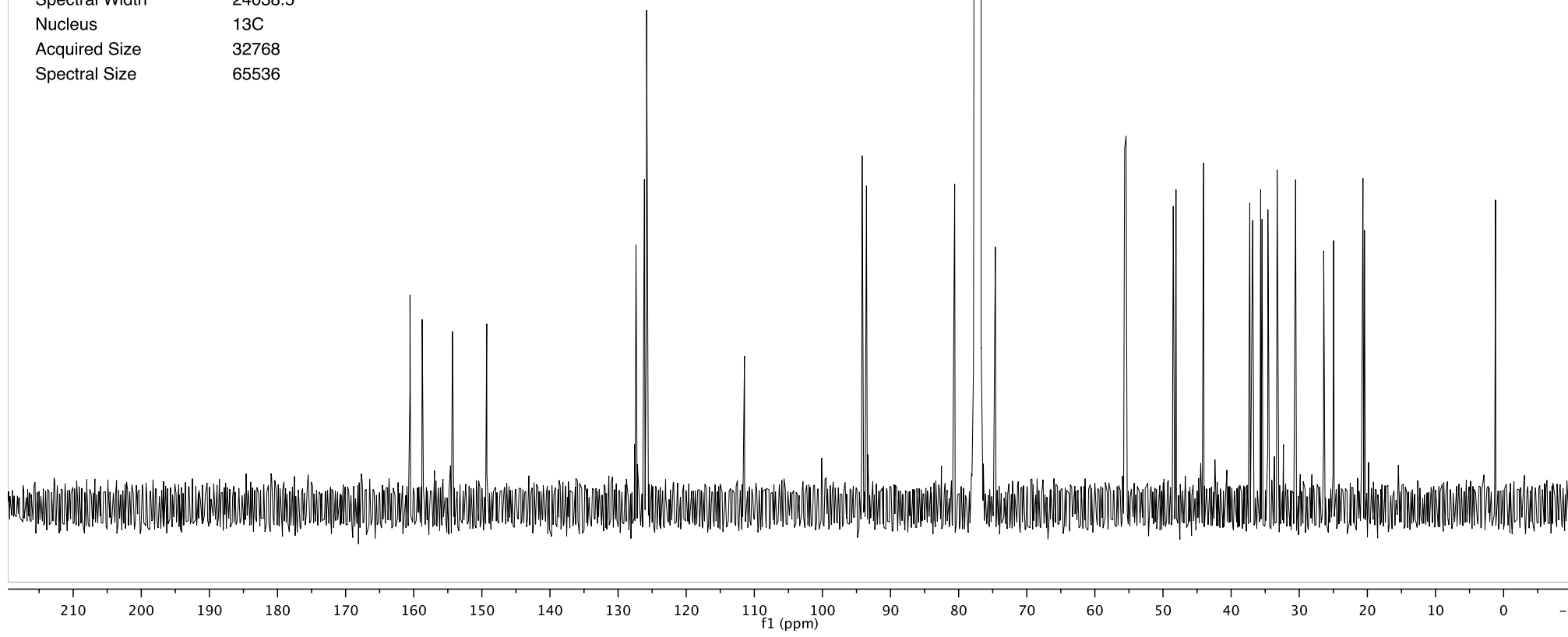
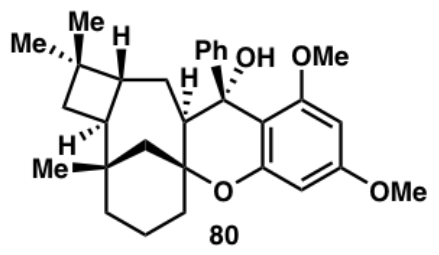
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Data File Name	/ Volumes/ nmrdata/ lchapman/ nmr/ LMC-04-278-P3-HMBC/ 1/ fid
Title	LMC-04-278-P3-HMBC.1.fid
Origin	Bruker BioSpin GmbH
Spectrometer	spect
Solvent	CDCl3
Temperature	294.9
Pulse Sequence	zg30
Experiment	1D
Number of Scans	16
Receiver Gain	197
Relaxation Delay	1.0000
Pulse Width	11.7000
Acquisition Time	4.0894
Acquisition Date	2017-04-17T21:40:33
Spectrometer Frequency	400.13
Spectral Width	8012.8
Nucleus	1H
Acquired Size	32768
Spectral Size	65536

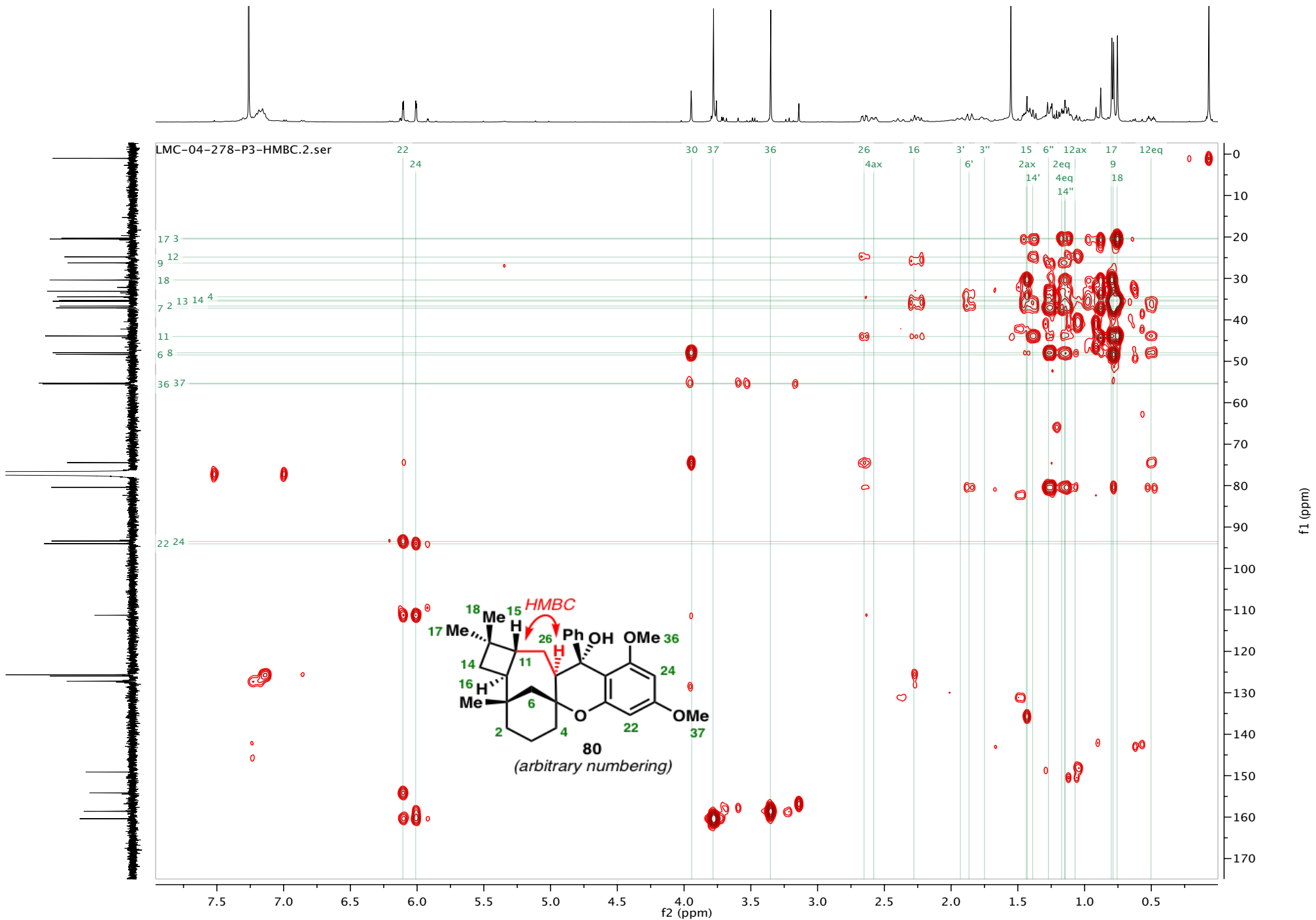


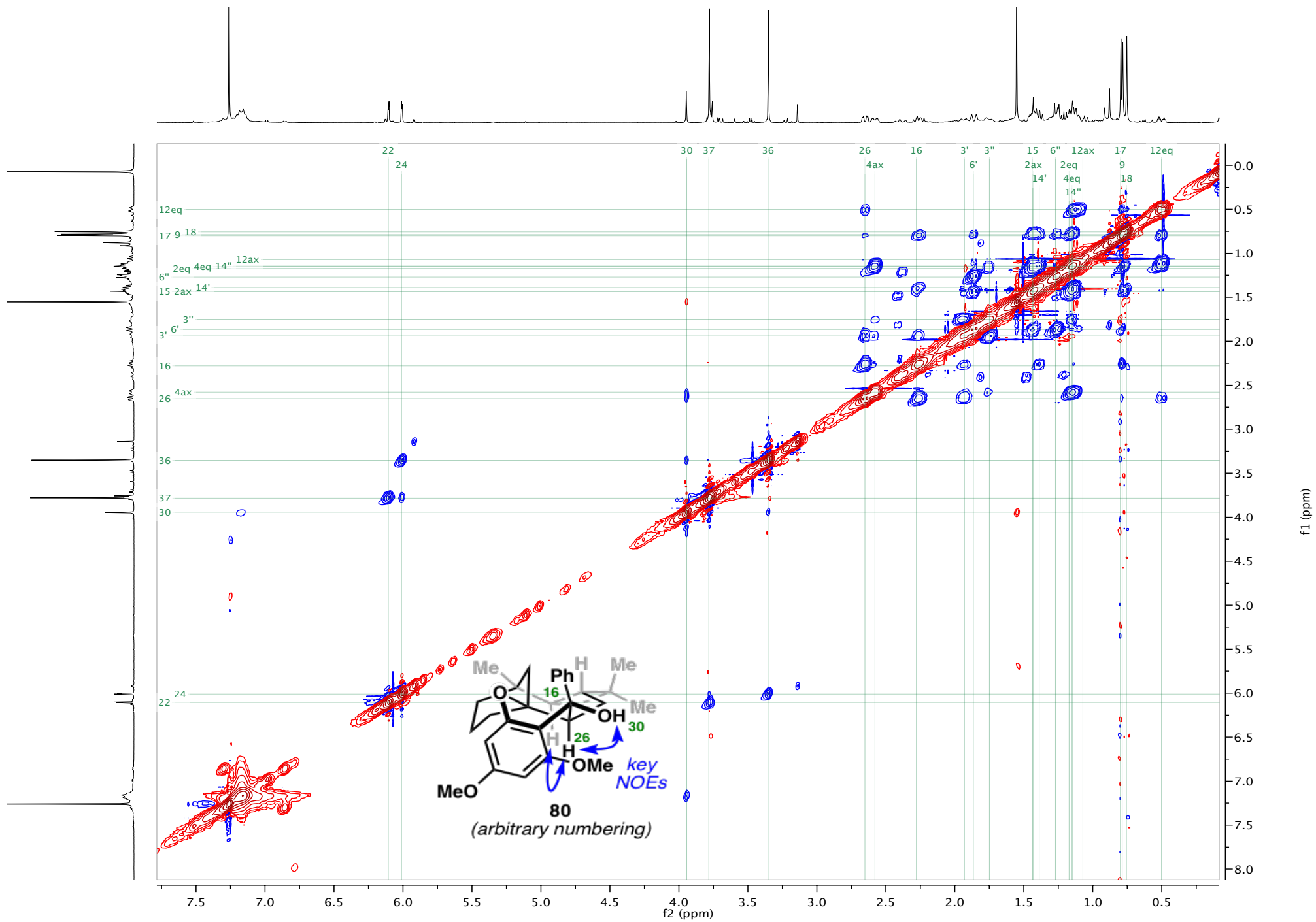
Parameter

Data File Name	/ Volumes/ nmrdata/ lchapman/ nmr/ LMC-04-278-P3-2D/ 1/ fid
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Origin	Bruker BioSpin GmbH
Spectrometer	spect
Solvent	CDCl3
Temperature	294.9
Pulse Sequence	zgpg30
Experiment	1D
Number of Scans	5200
Receiver Gain	55
Relaxation Delay	2.0000
Pulse Width	10.0000
Acquisition Time	1.3631
Acquisition Date	2017-04-16T04:03:34
Spectrometer Frequency	100.62
Spectral Width	24038.5
Nucleus	13C
Acquired Size	32768
Spectral Size	65536

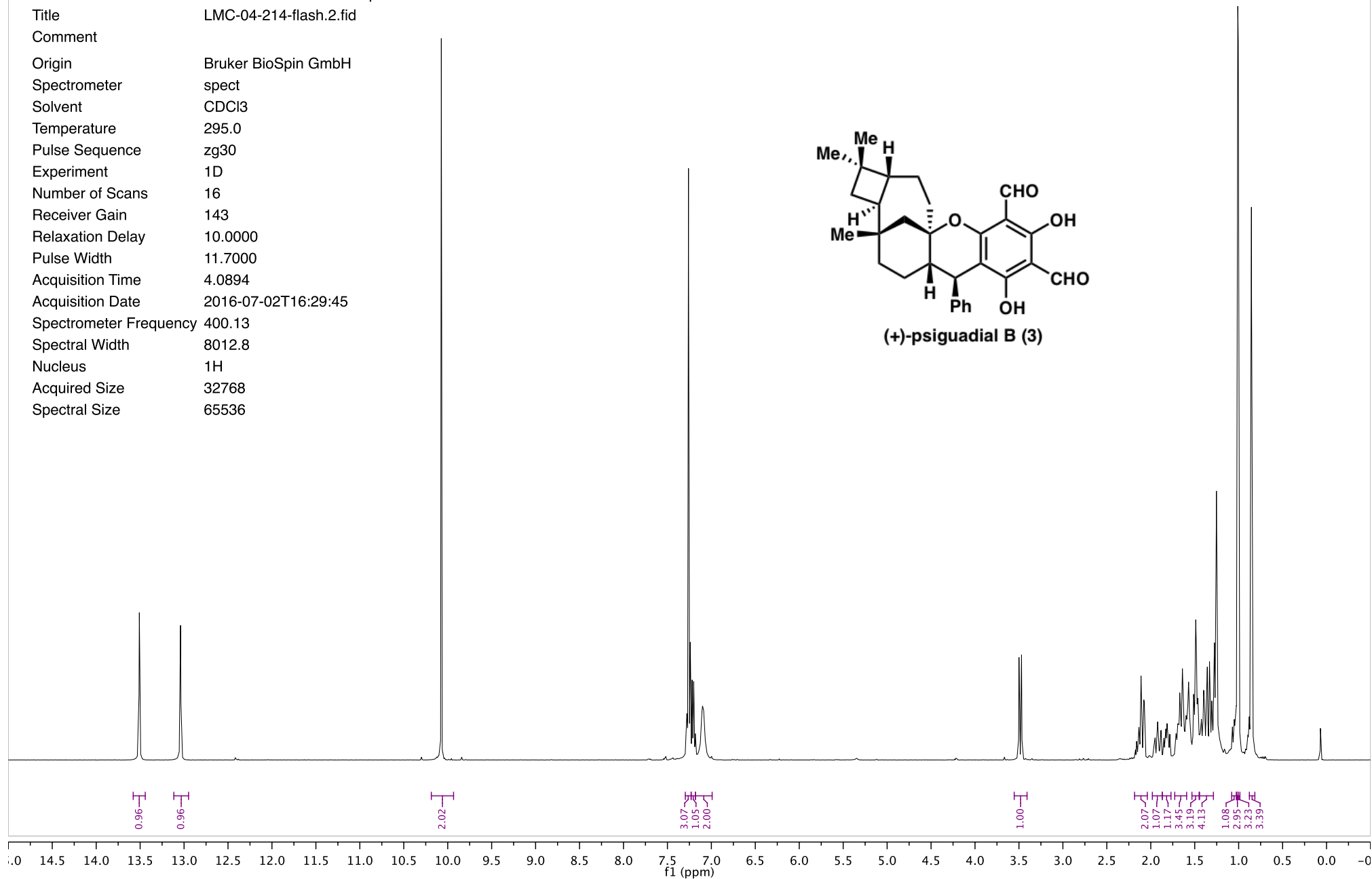
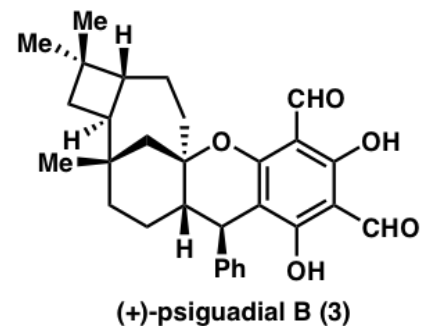
160.55	158.76	154.31	149.28	127.36	126.13	123.81	111.44	94.17	93.54	80.58	74.61	55.64	55.42	48.50	48.09	44.04	37.26	36.83	35.66	35.46	34.60	33.24	30.54	26.41	24.96	20.65	20.42
		Value																									







Parameter	Value
Data File Name	/ Volumes/ nmrdata/ lchapman/ nmr/ LMC-04-214-flash/ 2/ fid
Title	LMC-04-214-flash.2.fid
Comment	
Origin	Bruker BioSpin GmbH
Spectrometer	spect
Solvent	CDCl3
Temperature	295.0
Pulse Sequence	zg30
Experiment	1D
Number of Scans	16
Receiver Gain	143
Relaxation Delay	10.0000
Pulse Width	11.7000
Acquisition Time	4.0894
Acquisition Date	2016-07-02T16:29:45
Spectrometer Frequency	400.13
Spectral Width	8012.8
Nucleus	1H
Acquired Size	32768
Spectral Size	65536



LMC-04-214-flash.3.fid

Parameter	Value
Data File Name	/ Volumes/ nmrdata/ lchapman/ nmr/ LMC-04-214-flash/ 3/ fid
Title	LMC-04-214-flash.3.fid
Comment	
Origin	Bruker BioSpin GmbH
Spectrometer	spect
Solvent	CDCl3
Temperature	294.9
Pulse Sequence	zgpg30
Experiment	1D
Number of Scans	1024
Receiver Gain	72
Relaxation Delay	2.0000
Pulse Width	10.0000
Acquisition Time	1.3631
Acquisition Date	2016-07-02T17:28:52
Spectrometer Frequency	100.62
Spectral Width	24038.5
Nucleus	13C
Acquired Size	32768
Spectral Size	65536

