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Life Sciences Reporting Summary

Nature Research wishes to improve the reproducibility of the work that we publish. This form is intended for publication with all accepted life science papers and provides structure for consistency and transparency in reporting. Every life science submission will use this form; some list items might not apply to an individual manuscript, but all fields must be completed for clarity.

For further information on the points included in this form, see Reporting Life Sciences Research. For further information on Nature Research policies, including our data availability policy, see Authors & Referees and the Editorial Policy Checklist.

Please do not complete any field with "not applicable" or n/a. Refer to the help text for what text to use if an item is not relevant to your study. <u>For final submission</u>: please carefully check your responses for accuracy; you will not be able to make changes later.

	Experimental design	
1.	Sample size	
	Describe how sample size was determined.	Not applicable to the study
2.	Data exclusions	
	Describe any data exclusions.	No data was excluded.
3.	Replication	
	Describe the measures taken to verify the reproducibility of the experimental findings.	We tested the proposed methods on 85 different simulated data sets based on a published simulation method for SV. Furthermore, we evaluated the reproducibility by using trios for Arabdidopsis and Human data and benchmarked the effect of downsizing the initial data sets for various technologies as well as healthy and diseased humans. All attempts at replication were successful,
4.	Randomization	
	Describe how samples/organisms/participants were allocated into experimental groups.	Not relevant as no selection of samples were preformed no cross validation is appropriate.
5.	Blinding	
	Describe whether the investigators were blinded to	Data were not partitioned into groups

group allocation during data collection and/or analysis.

Note: all in vivo studies must report how sample size was determined and whether blinding and randomization were used.

6. Statistical parameters

For all figures and tables that use statistical methods, confirm that the following items are present in relevant figure legends (or in the Methods section if additional space is needed).

n/a	Соі	nfirmed
\boxtimes		The exact sample size (n) for each experimental group/condition, given as a discrete number and unit of measurement (animals, litters, cultures, etc.)
	\boxtimes	A description of how samples were collected, noting whether measurements were taken from distinct samples or whether the same sample was measured repeatedly
\boxtimes		A statement indicating how many times each experiment was replicated
	\boxtimes	The statistical test(s) used and whether they are one- or two-sided Only common tests should be described solely by name; describe more complex techniques in the Methods section.
	\boxtimes	A description of any assumptions or corrections, such as an adjustment for multiple comparisons
	\boxtimes	Test values indicating whether an effect is present Provide confidence intervals or give results of significance tests (e.g. P values) as exact values whenever appropriate and with effect sizes noted.
	\boxtimes	A clear description of statistics including central tendency (e.g. median, mean) and variation (e.g. standard deviation, interquartile range)
	\boxtimes	Clearly defined error bars in <u>all</u> relevant figure captions (with explicit mention of central tendency and variation)
		See the web collection on statistics for biologists for further resources and guidance.

Policy information about availability of computer code

Describe 1	the	software	used	to	analyze	the	data	in	this
study.									

We used BWA-MEM, NGMLR (proposed method in this manuscript), Graphmap and BlasR to align reads. Lumpy, Delly, Manta, Sniffles (proposed method), PBHoney to infer structural Variations. SURVIVOR (v2) to simulate, evaluate and compare methods. All methods (apart from Sniffles (v1.0.6) and NGMLR (v0.2.6) that are the topic of this paper) are peer reviewed and published. Sniffles and NGMLR are available on Github and links are included in the manuscript.

For manuscripts utilizing custom algorithms or software that are central to the paper but not yet described in the published literature, software must be made available to editors and reviewers upon request. We strongly encourage code deposition in a community repository (e.g. GitHub). *Nature Methods* guidance for providing algorithms and software for publication provides further information on this topic.

Materials and reagents

Policy information about availability of materials

8. Materials availability

Indicate whether there are restrictions on availability of unique materials or if these materials are only available for distribution by a third party. no eukaryotic cell lines were used

9. Antibodies

Describe the antibodies used and how they were validated for use in the system under study (i.e. assay and species).

No antibodies used.

10. Eukaryotic cell lines

- a. State the source of each eukaryotic cell line used.
- b. Describe the method of cell line authentication used.
- c. Report whether the cell lines were tested for mycoplasma contamination.
- d. If any of the cell lines used are listed in the database of commonly misidentified cell lines maintained by ICLAC, provide a scientific rationale for their use.

• Animals and human research participants

Policy information about studies involving animals; when reporting animal research, follow the ARRIVE guidelines

11. Description of research animals

Provide all relevant details on animals and/or animal-derived materials used in the study.

No animals were used.

Policy information about studies involving human research participants

12. Description of human research participants

Describe the covariate-relevant population characteristics of the human research participants.

No new data were generated for this paper; human sequence data are from publicly available sources