

Supplementary Table S1: Bacterial strains and plasmids used in this study

Bacteria and Plasmids	Characteristics <sup>†</sup>	Source/Reference
<b>Bacteria</b>		
<i>E. coli</i> EC1000	Derivative of <i>E. coli</i> MC1000 in which <i>repA</i> is integrated in chromosome	(1)
<i>L. reuteri</i> ATCC PTA 6475	Human breast milk isolate	BioGaia AB
VPL4073	In-frame deletion of 2 kb <i>pduCDE</i> in <i>L. reuteri</i> 6475	This work
<i>L. reuteri</i> R2lc	Rat gastrointestinal isolate	(2)
VPL4192	In-frame deletion of 870 bp <i>araT</i> in <i>L. reuteri</i> R2lc	This work
<i>L. reuteri</i> DSM 17938	Plasmid-cured derivative of <i>L. reuteri</i> ATCC 55730	BioGaia AB, (3)
VPL3565	<i>L. reuteri</i> DSM 17938 harboring pVE6007	This work
VPL4171	In-frame deletion of 2.2 kb <i>sdpA</i> in <i>L. reuteri</i> DSM 17938	This work
VPL4176	In-frame deletion of 5.3 kb <i>sdpB</i> in <i>L. reuteri</i> DSM 17938	This work
<i>L. rhamnosus</i> GG	Human fecal isolate	ATCC
VPL4185	In-frame deletion of 1.5 kb <i>acs1</i> in <i>L. rhamnosus</i> GG	This work
<i>L. casei</i> BFLM218	Human fecal isolate	(4)
VPL4013	Insertion of 66 bp 3x-FLAG tag <i>prtP</i> gene in <i>L. casei</i> BFLM218	This work
<i>L. brevis</i> ATCC 8287	Fermented olives isolate	ATCC
VPL3978	<i>L. brevis</i> harboring pVPL3933	This work
<i>L. fermentum</i> ATCC 14931	Fermented beets isolate	ATCC
VPL3950	<i>L. fermentum</i> harboring pVPL3933	This work
<i>L. plantarum</i> ATCC BAA-793	Human saliva isolate	ATCC
VPL3938	<i>L. plantarum</i> harboring pVPL3933	This work
<i>L. salivarius</i> CCUG 47825	Human blood isolate	CCUG
VPL3941	<i>L. salivarius</i> harboring pVPL3933	This work
<i>L. sakei</i> ATCC 15521	Starter of sake	ATCC
VPL3952	<i>L. sakei</i> harboring pVPL3933	This work
<i>L. gasseri</i> ATCC 33323	Human intestinal isolate	ATCC
<i>Fructobacillus fructosus</i> DSM 20349	Flowers isolate	DSMZ
<i>Leuconostoc mesenteroides</i> DSM 20346	Hansen's dried starter powder	DSMZ
<i>Pediococcus acidilactici</i> DSM 20284	Barley isolate	DSMZ
<i>Weisella paramesenteroides</i> DSM 20288	Meat isolate	DSMZ
<i>Lactococcus lactis</i> subsp. <i>cremoris</i> NZ9000	Dairy starter, derivative of MG1363, <i>pepN::nisRK</i>	(5)
<b>Plasmids</b>		
pORI19	Em <sup>R</sup> , <i>repA</i> -	(6)
pSIP411	Em <sup>R</sup> , Sakacin-P based expression vector	(7)
pVPL2042	Em <sup>R</sup> , pNZ8048 derivative. Cm marker was replaced by Em marker	Van Pijkeren Lab stock
pVPL3002	pORI19 harboring <i>L. reuteri</i> derived <i>ddIF258Y</i>	This work
pVPL3010	Em <sup>R</sup> , derivative of vector pVPL3002 in which the <i>L. rhamnosus</i> GG <i>Δacs1</i> deletion cassette was cloned in the MCS site	This work
pVPL3125	Em <sup>R</sup> , derivative of vector pVPL3002 in which 2kb 5'- and 3' flanking sequence required for 3x-FLAG tag insertion in <i>prtP</i> of <i>L. casei</i> BFLM218 was cloned in the MCS site	This work
pVPL3137	Em <sup>R</sup> , derivative of pVPL3125 in which protein encodes 3x-FLAG tag was cloned in between the integration cassette of pVPL3125	This work
pVPL3478	Em <sup>R</sup> , derivative of vector pVPL3002 in which the <i>L. reuteri</i> 6475 <i>pduCDE</i> deletion cassette was cloned in the MCS site	This work
pVPL3612	Em <sup>R</sup> , derivative of vector pVPL3002 in which the <i>L. reuteri</i> 17938 <i>sdpA</i> deletion cassette was cloned in the MCS site	This work
pVPL3694	Em <sup>R</sup> , derivative of vector pVPL3002 in which the <i>L. reuteri</i> R2lc <i>araT</i> deletion cassette was cloned in the MCS site	This work
pVPL3762	Em <sup>R</sup> , derivative of vector pVPL3002 in which the <i>L. reuteri</i> 17938 <i>sdpB</i> deletion cassette was cloned in the MCS site	This work
pVPL3859	Em <sup>R</sup> , derivative of vector 2042 in which the <i>L. plantarum</i> BAA-793 <i>ddl</i> was inserted in the MSC site	This work
pVPL3862	Em <sup>R</sup> , derivative of pVPL2042 in which <i>ddIF258Y</i> was cloned in the backbone of pVPL2042	This work

Plasmid	Characteristics <sup>†</sup>	Source/Reference
pVPL3925	Em <sup>R</sup> , derivative of vector 2042 in which the <i>L. plantarum</i> BAA-793 <i>ddlF260Y</i> was inserted in the MSC site	This work
pVPL3933	Em <sup>R</sup> , derivative of pSIP411 in which <i>ddlF260Y</i> <sub><i>plantarum</i></sub> was cloned under the control of sakacin-based inducible promoter	This work

VPL: Van Pijkeren Lab strain identification number. pVPL: Van Pijkeren Lab plasmid identification number

<sup>†</sup>: *pduCDE*: glycerol dehydratase large subunit PduC, glycerol dehydratase medium subunit PduD, glycerol dehydratase small subunit PduE (BAG26151, BAG26150.1, BAG26149.1); *araT*: aromatic amino acid aminotransferase

(MG822655); *sdpA*: bifunctional phosphodiesterase (WP\_003672341.1); *sdpB*: hypothetical protein and LPXTG-motif cell wall anchor domain protein (WP\_013923860.1 , AEI56985.1); *acs1*: acyl-CoA synthetase (WP\_014570113.1);

Em<sup>R</sup>: erythromycin-resistant; Cm<sup>R</sup>: chloramphenicol resistant. The accession number listed can be found on <https://www.ncbi.nlm.nih.gov>. \*: ATCC: American Type Culture Collection; DSMZ: German Collection of Microorganisms and Cell Cultures; CCUG: Culture Collection University of Gothenburg.

**Table S2. Oligonucleotides used in this study**



oligo name	sequence (5'-3') <sup>†</sup>	Target/comment <sup>#</sup>
oVPL1555	agtcataaagccaacccaaaagg	Fwd, 5' u/s flanking region of <i>L. reuteri</i> DSM 17938 <i>sdpA</i>
oVPL1556	tgcataatcgaaaaatgaaatagac	Rev, 5' u/s flanking region of <i>L. reuteri</i> DSM 17938 <i>sdpA</i>
oVPL1557	tgcataaaaaaaactcctctataatatac	Fwd, 3' u/s flanking region of <i>L. reuteri</i> DSM 17938 <i>sdpA</i>
oVPL1558	tcataccggccgtcgccg	Rev, 3' u/s flanking region of <i>L. reuteri</i> DSM 17938 <i>sdpA</i>
oVPL1559	aaacgacggccaggtaattcgagctcgtaagtataaagccaacccaaaaggaaatttat	LCR bridging oligo for pORI-ddl/F258Y:: <i>sdpA</i> deletion cassette
oVPL1560	agtctattcattttaaacgattaaggcatgcataaaaaaaactcctctataatatacataat	LCR bridging oligo for pORI-ddl/F258Y:: <i>sdpA</i> deletion cassette
oVPL1561	gtaatatgcgaaccgatcaggccgtatgaatcctctagactgcacctgcaggcatgcaa	LCR bridging oligo for pORI-ddl/F258Y:: <i>sdpA</i> deletion cassette
oVPL2065	aaaaacaactaacaattatctaacagaacc	Fwd, 5' u/s flanking region of <i>L. reuteri</i> DSM 17938 <i>sdpB</i>
oVPL2066	gaagactaattcttagacaagtg	Rev, 5' u/s flanking region of <i>L. reuteri</i> DSM 17938 <i>sdpB</i>
oVPL2067	taacgttggattccttattcattcc	Fwd, 3' u/s flanking region of <i>L. reuteri</i> DSM 17938 <i>sdpB</i>
oVPL2068	ttcatgtcatgcgtaattcaagg	Rev, 3' u/s flanking region of <i>L. reuteri</i> DSM 17938 <i>sdpB</i>
oVPL2069	aaacgacggccaggtaattcgagctcgtaaaaaacaactaacaattatctaacagtaac	LCR bridging oligo for pORI-ddl/F258Y:: <i>sdpB</i> deletion cassette
oVPL2070	taactattcactgtctagaatttagtctctaacgttggaaattccattattcctct	LCR bridging oligo for pORI-ddl/F258Y:: <i>sdpB</i> deletion cassette
oVPL2071	cgagttccgtgaaattacgcgtacatgacatggaaatcccttagactgcacctgcaggcatgcaa	LCR bridging oligo for pORI-ddl/F258Y:: <i>sdpB</i> deletion cassette
oVPL2072	aacccaccaatcccaagctatg	Fwd, for screening of $\Delta sdpB$ in <i>L. reuteri</i> DSM 17938
oVPL2073	tgcttatcaacaaagttagattgcc	Rev, for screening of $\Delta sdpB$ in <i>L. reuteri</i> DSM 17938
oVPL2589	ctatcaggccatcgatggcgcacg	Fwd, amplifies promoter region of <i>L. plantarum</i> BAA-793 Ddl
oVPL2590	ttatccgcgtctaacgcaacgaaa	Rev, amplifies <i>L. plantarum</i> BAA-793 Ddl
oVPL2591	aaaataaattataaggaggcactcaccatgctatcgttgccatcgatggcgcacgcagg	Bridging oligo for <i>L. plantarum</i> BAA-793 Ddl and pNZ8048 backbone.
oVPL2592	tatgatttcgttgcgttagacgcggaaataatagtcttagagagactcaaggctttttgaa	Bridging oligo for <i>L. plantarum</i> BAA-793 Ddl and pNZ8048 backbone.
oVPL2593	/5phos/tcacaccactcgcatccacgtacttattatagtcgttagaacgc	Rev, modifies F260Y of <i>ddl</i> of <i>L. plantarum</i> BAA-793
oVPL2594	/5phos/cgttgcattaccaggatcgaaattacc	Fwd, amplifies the <i>ddl</i> of <i>L. plantarum</i> BAA-793
oVPL2694	taatcgttgcgttgcacggcccc	Combined with oVPL2072-oVPL2073 for screening of $\Delta sdpB$ in <i>L. reuteri</i> DSM 17938
oVPL2841	aaacgacgtgcataatcacag	Fwd, amplifies GAPDH gene of <i>L. salivarius</i> CCUG 47825. For qPCR.
oVPL2842	cagcaccagttgttaggtgagga	Rev, amplifies GAPDH gene of <i>L. salivarius</i> CCUG 47825. For qPCR.
oVPL2845	cctgctcgtaagggtcttc	Fwd, amplifies native <i>ddl</i> of <i>L. salivarius</i> CCUG 47825. For qPCR.
oVPL2846	acggattgcacctaacttgg	Rev, amplifies native <i>ddl</i> of <i>L. salivarius</i> CCUG 47825. For qPCR.
oVPL2849	tggaatgcgtcgatcttaggc	Fwd, amplifies <i>ddl</i> /F260Y of pVPL3933. For qPCR.
oVPL2850	aaacgcgtcgacatct	Rev, amplifies <i>ddl</i> /F260Y of pVPL3933. For qPCR.

oVPL: Van Pijkeren Lab oligonucleotide identification number.

<sup>†</sup>: /5phos/: indicates oligonucleotides is phosphorylated on 5'-end;

<sup>#</sup>: LCR: ligase cycling reaction (8). Fwd: forward; Rev: reverse; GAPDH: glyceraldehyde-3-phosphate dehydrogenase (YP\_536057.1). *pduCDE*: glycerol dehydratase large subunit PduC, glycerol dehydratase medium subunit PduD, glycerol dehydratase small subunit PduE (BAG26151, BAG26150.1, BAG26149.1); *araT*: aromatic amino acid aminotransferase (MG822655); *sdpA*: bifunctional phosphodiesterase (WP\_003672341.1); *sdpB*: hypothetical protein and LPXTG-motif cell wall anchor domain protein (WP\_013923860.1 , AEI56985.1); *acs1*: acyl-CoA synthetase (WP\_014570113.1); The accession number listed can be found on <https://www.ncbi.nlm.nih.gov>.

## References

1. Leenhouts K, Buist G, Bolhuis A, ten Berge A, Kiel J, Mierau I, Dabrowska M, Venema G, Kok J. 1996. A general system for generating unlabelled gene replacements in bacterial chromosomes. *Mol Gen Genet* 253:217-24.
2. Molin G, Andersson R, Ahrne S, Lonner C, Marklinder I, Johansson ML, Jeppsson B, Bengmark S. 1992. Effect of fermented oatmeal soup on the cholesterol level and the *Lactobacillus* colonization of rat intestinal mucosa. *Antonie Van Leeuwenhoek* 61:167-73.
3. Rosander A, Connolly E, Roos S. 2008. Removal of antibiotic resistance gene-carrying plasmids from *Lactobacillus reuteri* ATCC 55730 and characterization of the resulting daughter strain, *L. reuteri* DSM 17938. *Appl Environ Microbiol* 74:6032-40.
4. von Schillde MA, Hormannsperger G, Weiher M, Alpert CA, Hahne H, Bauerl C, van Huynegem K, Steidler L, Hrncir T, Perez-Martinez G, Kuster B, Haller D. 2012. Lactocepin secreted by *Lactobacillus* exerts anti-inflammatory effects by selectively degrading proinflammatory chemokines. *Cell Host Microbe* 11:387-96.
5. Kuipers OP, de Ruyter PGGA, Kleerebezem M, de Vos WM. 1998. Quorum sensing-controlled gene expression in lactic acid bacteria. *Journal of Biotechnology* 64:15-21.
6. Law J, Buist G, Haandrikman A, Kok J, Venema G, Leenhouts K. 1995. A System to Generate Chromosomal Mutations in *Lactococcus-Lactis* Which Allows Fast Analysis of Targeted Genes. *Journal of Bacteriology* 177:7011-7018.
7. Sorvig E, Mathiesen G, Naterstad K, Eijsink VG, Axelsson L. 2005. High-level, inducible gene expression in *Lactobacillus sakei* and *Lactobacillus plantarum* using versatile expression vectors. *Microbiology* 151:2439-49.
8. de Kok S, Stanton LH, Slaby T, Durot M, Holmes VF, Patel KG, Platt D, Shapland EB, Serber Z, Dean J, Newman JD, Chandran SS. 2014. Rapid and reliable DNA assembly via ligase cycling reaction. *ACS Synth Biol* 3:97-106.