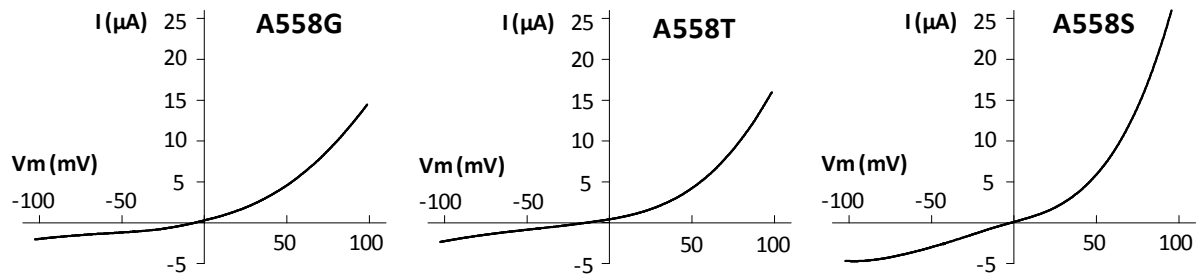


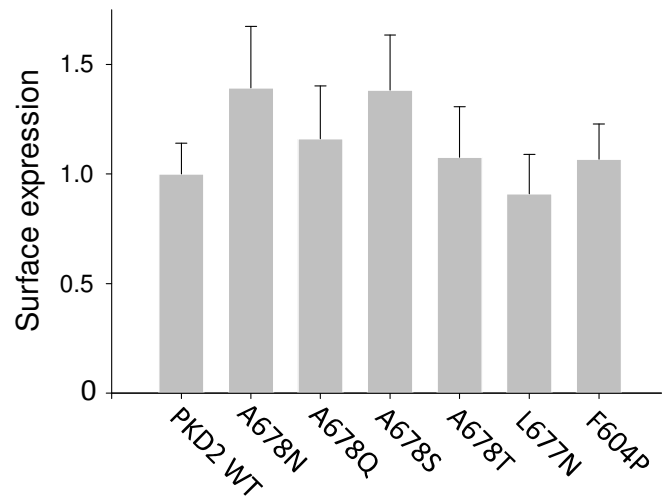
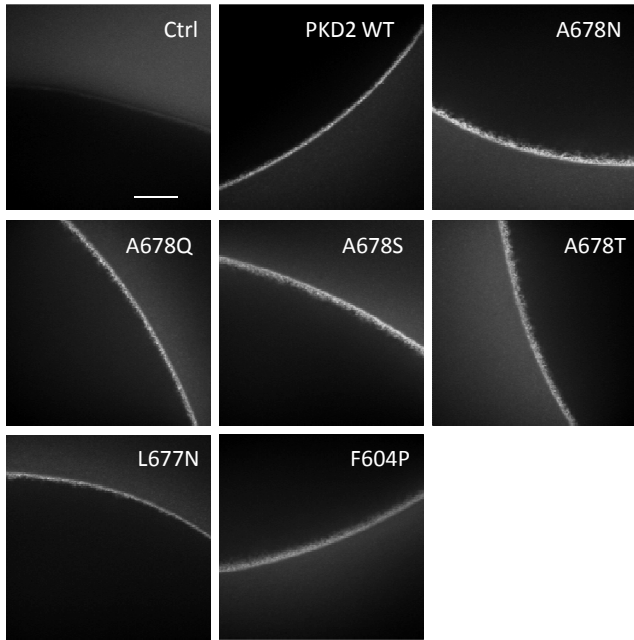
Supplementary Information

Hydrophobic pore gates regulate ion permeation in polycystic kidney disease 2 and 2L1 channels

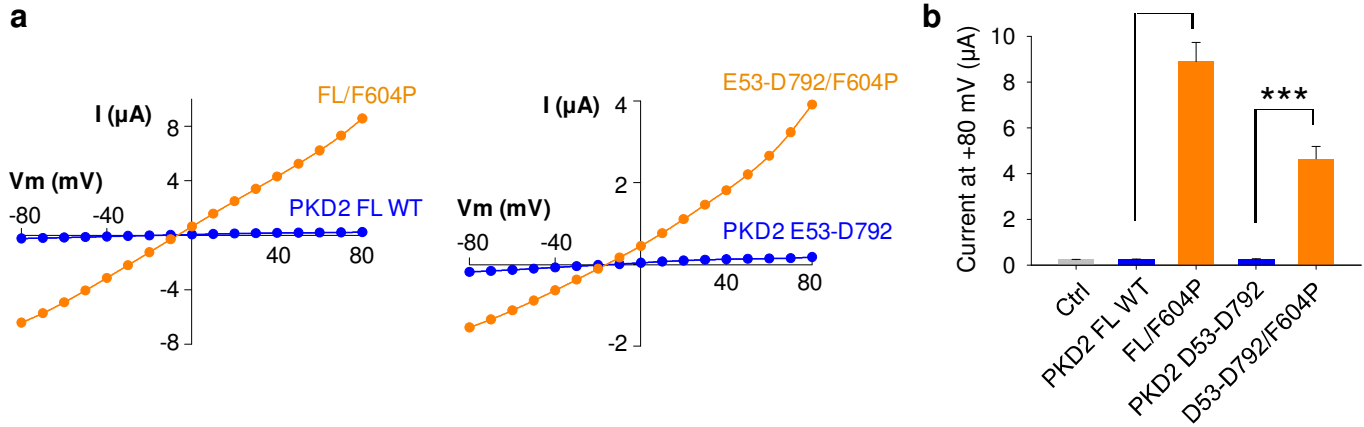
Zheng et al.



Supplementary Figure 1. Representative I-V curves for PKD2L1 A558G, A558T and A558S mutants. Currents were recorded in expressing oocytes in the presence of the Mg^{2+} -free Na-containing solution (in mM): 100 NaCl, 2 KCl, 10 HEPES at pH 7.5.



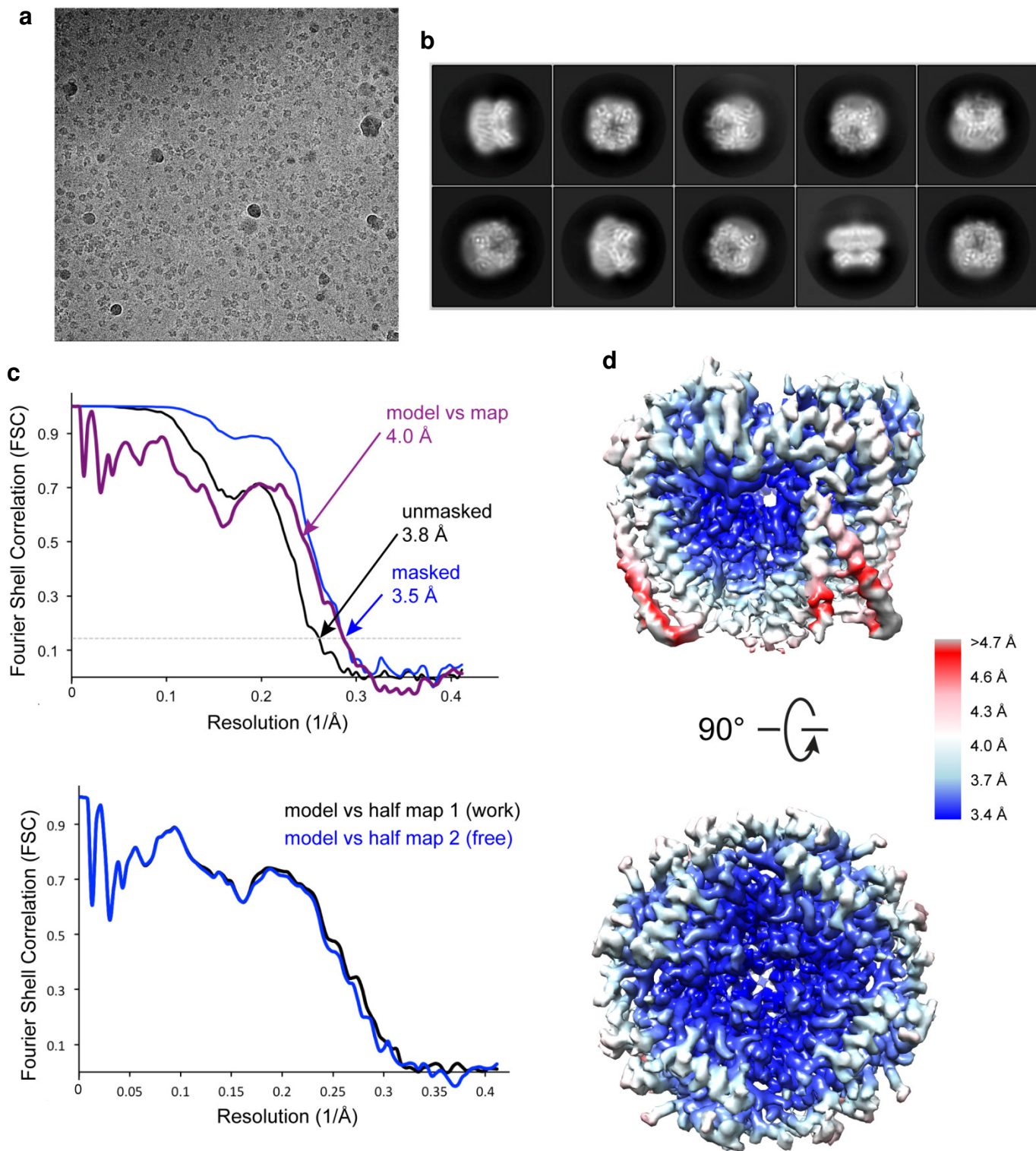
Supplementary Figure 2. Surface expression of WT or mutant PKD2 in oocytes. Left panel, whole-mount immunofluorescence using the antibody for the HA-tag showing the oocyte surface expression of the PKD2 WT or indicated mutants; Ctrl, water injected oocytes. Scale bar, 50 μ m. Right panel, averaged surface membrane expression of the PKD2 mutants normalized to that of PKD2 WT (n = 4 - 7). Data are presented as mean \pm SEM.



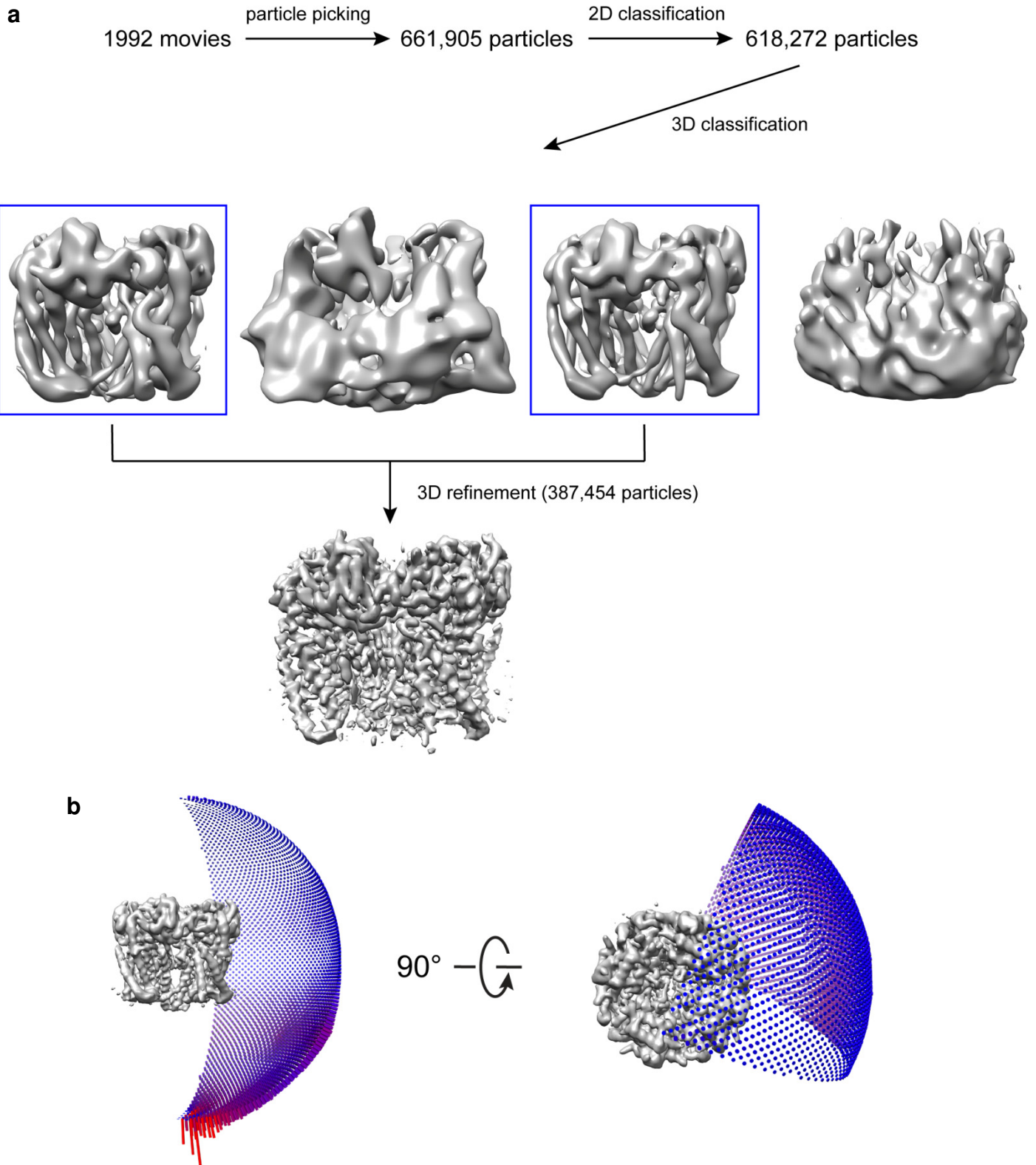
Supplementary Figure 3. Characterization of human PKD2 truncated mutant E53-D792. a

Representative I-V curves of full-length (FL) wild-type (WT) PKD2 and truncated mutant E53-D792 without or with F604P point mutation obtained under the same experimental condition as in Fig. 3a. **b**

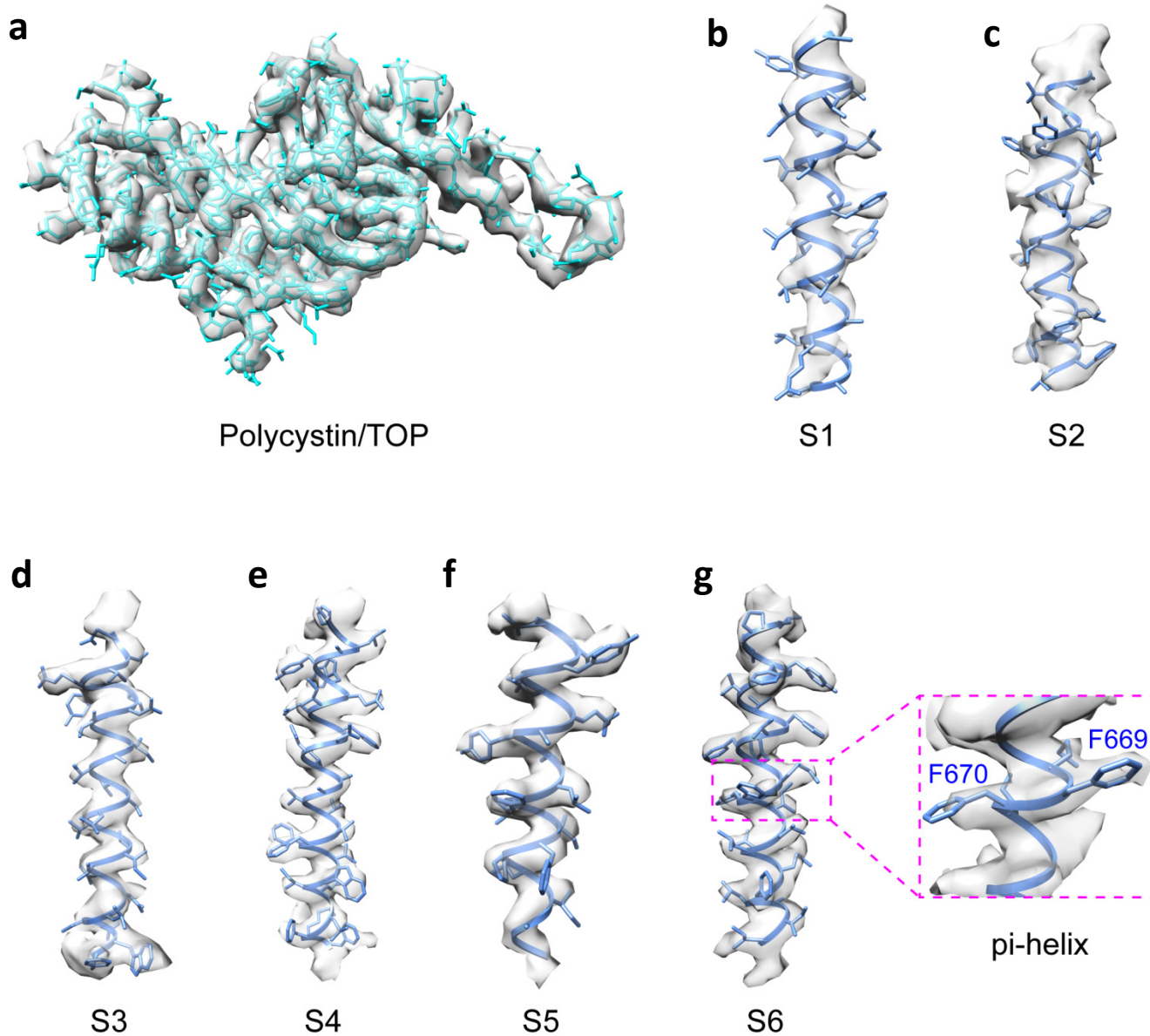
Averaged currents at +80 mV in expressing or water-injected (Ctrl) oocytes, as indicated. Data are presented as mean \pm SEM. ***P < 0.001 by Student's t-test.



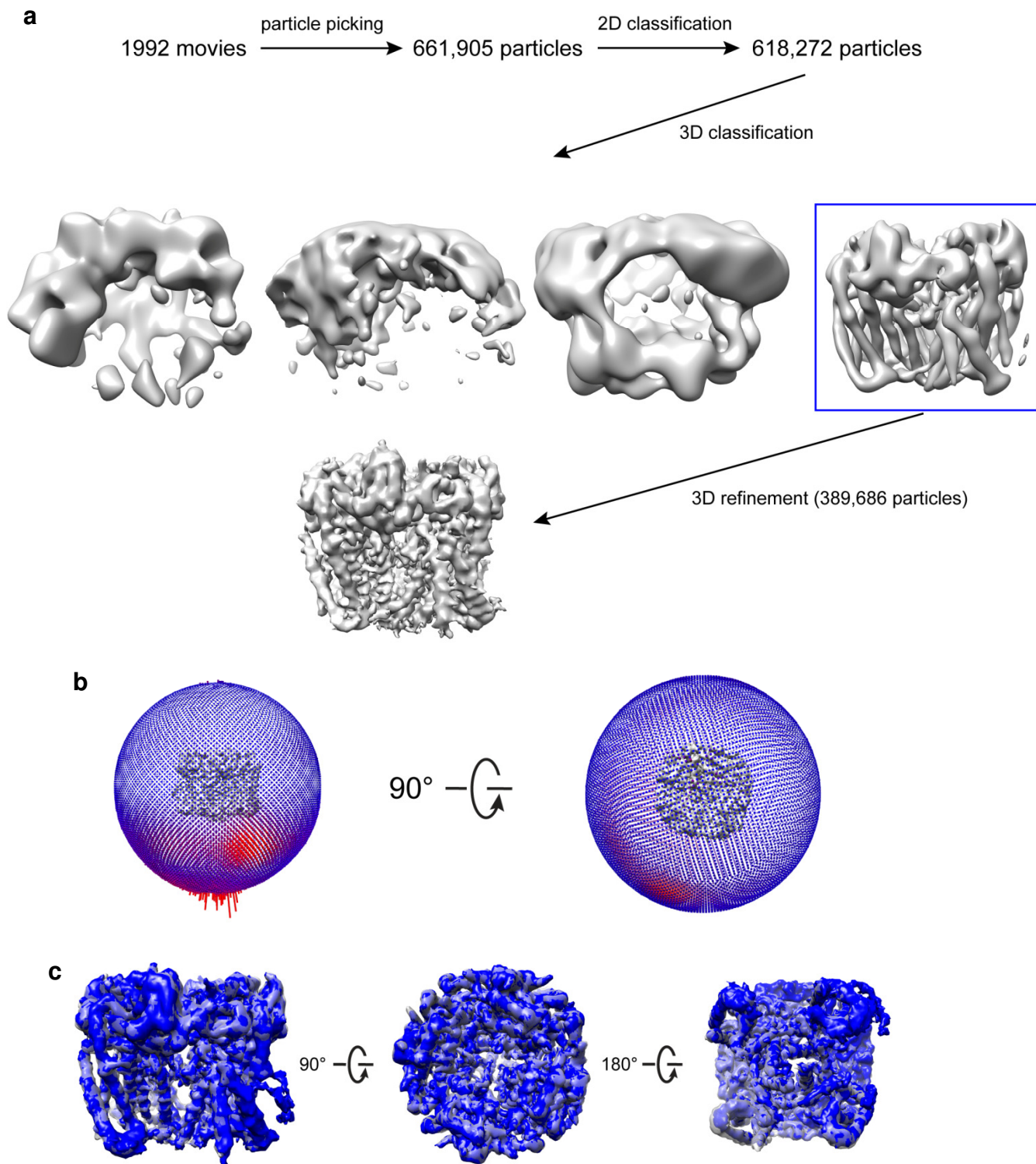
Supplementary Figure 4. Structure determination of the PKD2 F604P channel. **a** A representative micrograph of PKD2 F604P particles recorded on a TF30 Polara microscope. **b** 2D classes of PKD2 F604P calculated with RELION. **c** FSC curves for resolution estimation and model validation. (top) FSC curves reported by RELION before (unmasked, black) and after (masked, blue) postprocessing, where an automask was generated by RELION to flattening regions outside protein density prior to FSC calculation. FSC curves (model vs summed half map, purple) show overall agreement between model and map. (bottom) FSC curves are calculated for model vs half map 1 (work, black) and model versus half map 2 (free, blue). Overall agreement between these curves indicates little over-fitting of the model. **d** Local resolution determined by RELION.



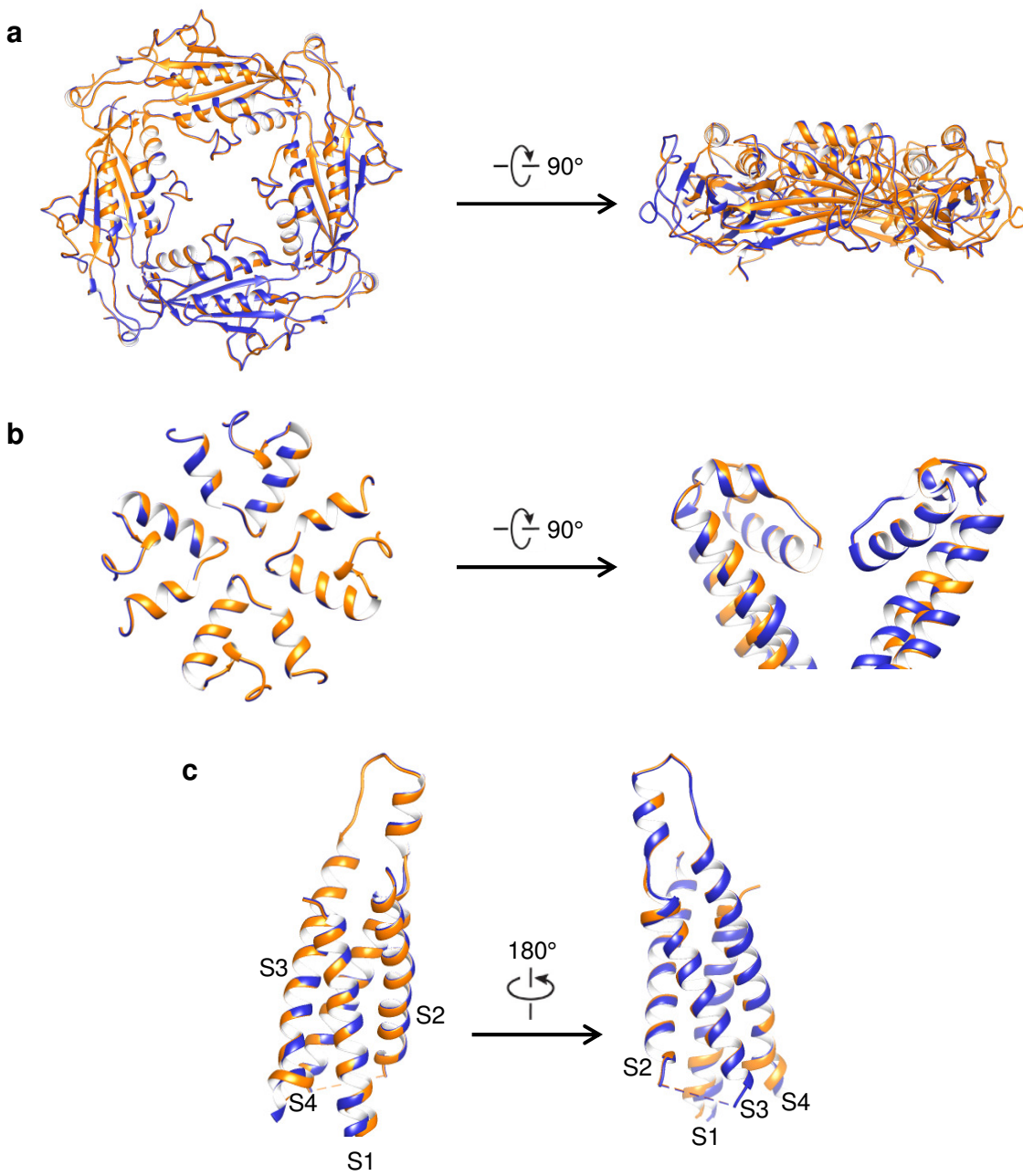
Supplementary Figure 5. Reconstruction of the PKD2 F604P channel with C4 symmetry. a Flow chart of image processing for the PKD2 F604P channel with C4 symmetry imposed. **b** Angular distribution plot of all particles projections. Cylinder heights are proportional to number of particles assigned to each set of Euler angles.



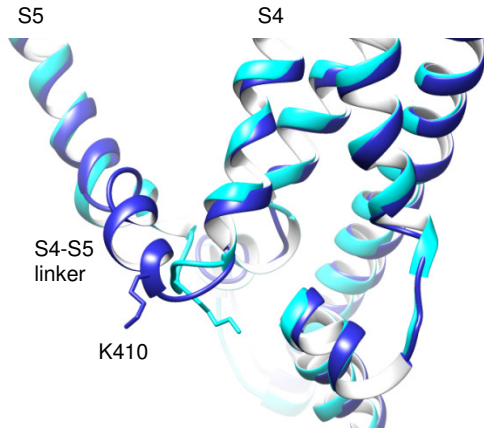
Supplementary Figure 6. EM density maps of various regions of the PKD2 F604P channel. The maps are sharpened with a b factor of -150 \AA^2 . In panel G, the pi-helix is highlighted.



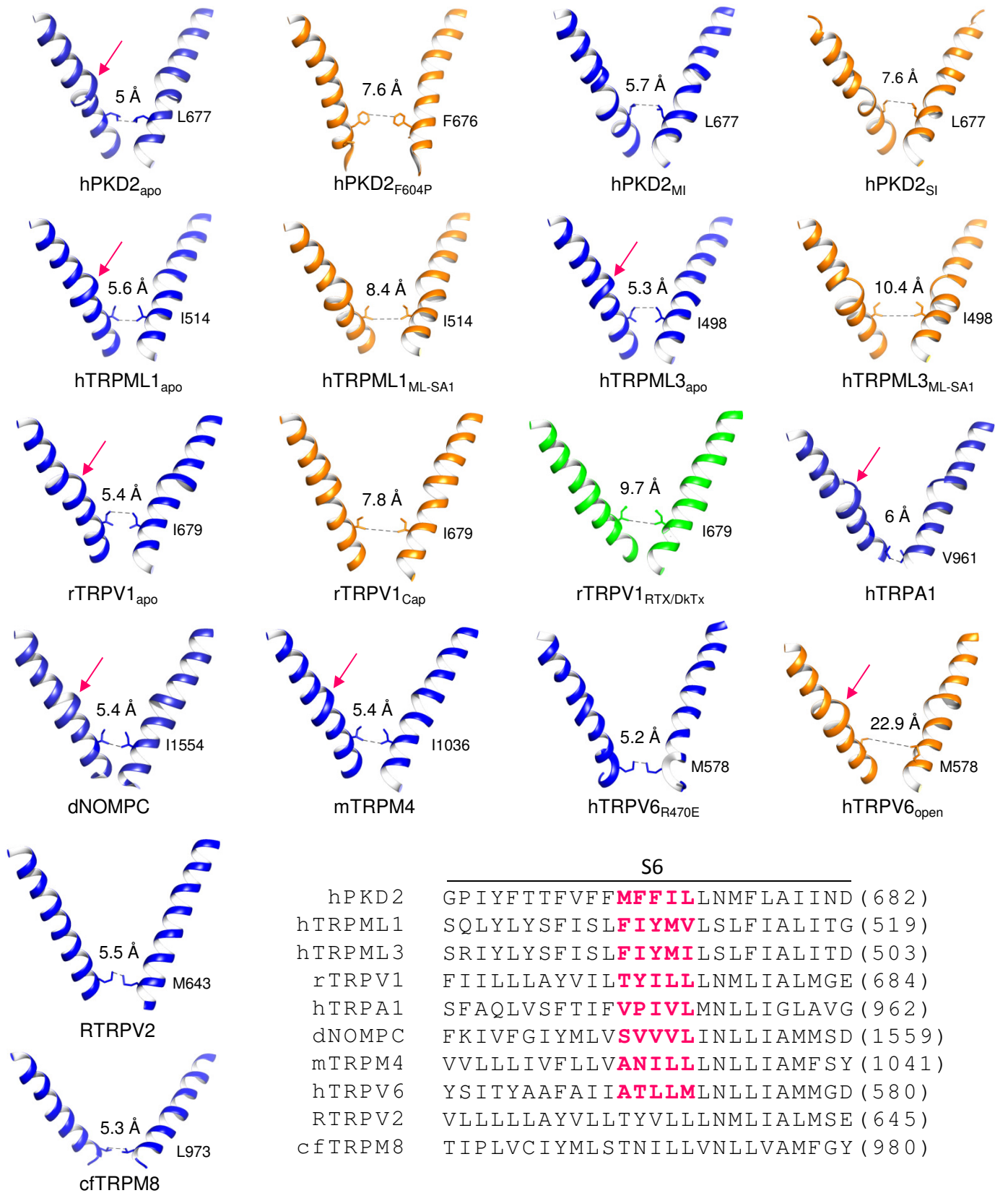
Supplementary Figure 7. Reconstruction of the PKD2 F604P channel without C4 symmetry. a Flow chart of image processing for the PKD2 F604P channel without C4 symmetry imposed. **b** Angular distribution plot of all particles projections. Cylinder heights are proportional to number of particles assigned to each set of Euler angles. **c** Comparison of the maps reconstructed with (gray) or without (blue) C4 symmetry, showing their overall agreement. Only subtle differences within the peripheral S1 helix, flexible loops, and amphipols are observed. Overall, map determined without C4 symmetry is of lower resolution as compared with the map determined with C4 symmetry. In particular, the S4-S5 linker is not well resolved in the map without imposing C4 symmetry.



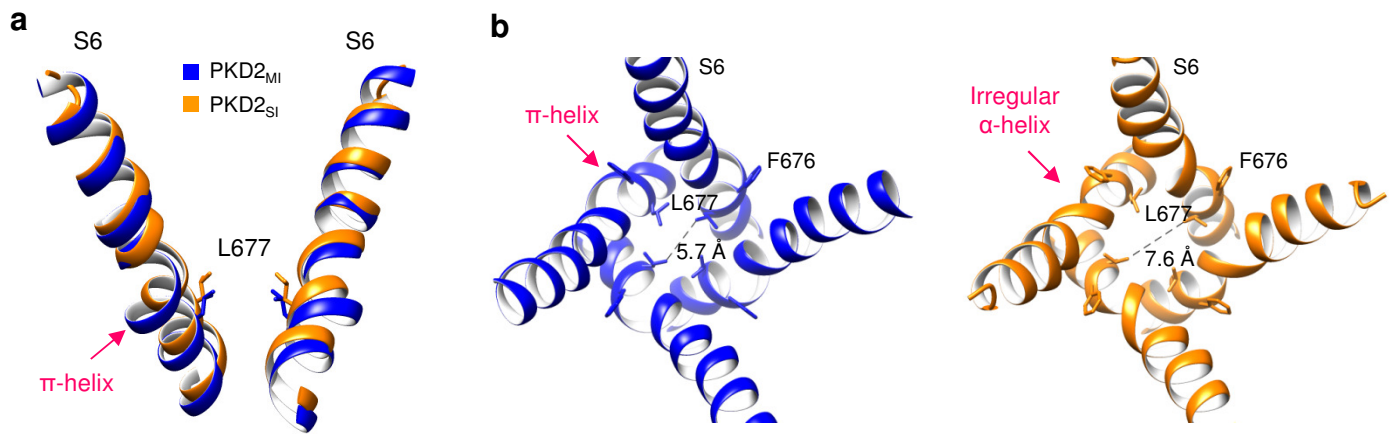
Supplementary Figure 8. Superposition of PKD2 WT and F604P mutant structures. Superposition of TOP domains (a), selectivity filters (b) and S1-S4 helices (c). PKD2 WT, blue; F604P, orange.



Supplementary Figure 9. Comparison between the TRPML1 S4-S5 linker in the closed (blue, PDB: 5WPQ) and proposed pre-open (cyan, PDB: 5WPT) states. The K410 in the linker's N-terminus, corresponding to the R581 in PKD2, is indicated.



Supplementary Figure 10. π -helix and α -helix in S6 of TRP channels. Upper and lower left panel, pore-lining S6 helices in hPKD2_{apo} (apo state, PDB: 5T4D), hPKD2_{MI} (bound with multiple Ca²⁺ ions, PDB: 5MKF), hPKD2_{SI} (bound with single Ca²⁺ ion, PDB: 5MKE), hTRPML1_{apo} (PDB: 5WJ5), hTRPML1_{ML-SA1} (bound with agonist ML-SA1, PDB: 5WJ9), hTRPML3_{apo} (PDB: 6AYE), hTRPML3_{ML-SA1} (bound with ML-SA1, PDB: 6AYF), rTRPV1_{apo} (PDB: 3J5P), rTRPV1_{Cap} (bound with capsaicin, PDB: 3J5R), rTRPV1_{RTX/DkTx} (bound with RTX/DkTx, PDB: 3J5Q), hTRPA1 (PDB: 3J9P), dNOMPC (PDB: 5VKQ), mTRPM4 (PDB: 6BCJ), hTRPV6_{R470E} (full-length human TRPV6 with the R470E mutation, PDB: 6BOA), hTRPV6_{open} (full-length human TRPV6, PDB: 6BO9), RTRPV2 (PDB: 5AN8), cfTRPM8 (PDB: 6BPQ). The gate residues revealed by the structures are shown and the pore diameters determined by the gate residues indicated. The π -helix in the middle of S6 is indicated by a red arrow. h, human; r, rat; d, *drosophila* (fruit fly); m, mouse; R, rabbit; cf, collared flycatcher. Lower right panel, amino acid sequence alignment of S6 helices from hPKD2 (Universal Protein Resource accession #: Q13563), hTRPML1 (Q9GZU1), hTRPML3 (Q8TDD5), rTRPV1 (O35433), hTRPA1 (O75762), dNOMPC (Q7KIQ2), mTRPM4 (Q7TN37), hTRPV6 (Q9H1D0), RTRPV2 (G1SNM3), cfTRPM8 (U3JD03). The last amino acid numbers are indicated and the π -helix sequences are highlighted bold and magenta.



Supplementary Figure 11. Conformational changes in pore-lining S6 helix of PKD2 induced by Ca²⁺. **a** Side view of superposition of S6 in the PKD2_{MI} (bound with multiple Ca²⁺ ions, PDB: 5MKF, blue) and PKD2_{SI} (bound with single Ca²⁺ ion, PDB: 5MKE, orange). The π -helix in the PKD2_{MI} structure is indicated by an arrow. The gate residue L677 is shown. **b** Top view of the pore lined by S6 in the PKD2_{MI} and PKD2_{SI} structures showing local conformational changes in the middle of S6.

Figure 1e

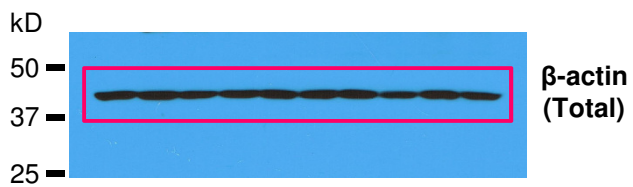
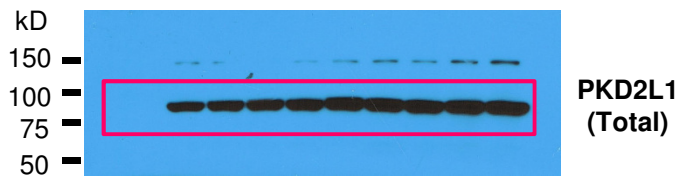
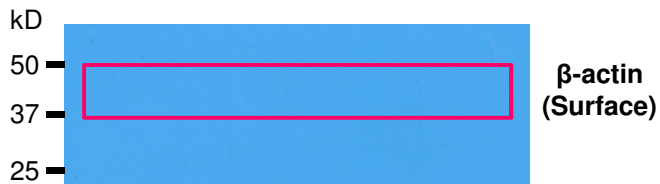
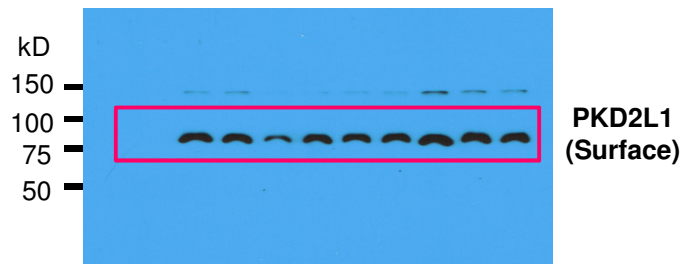
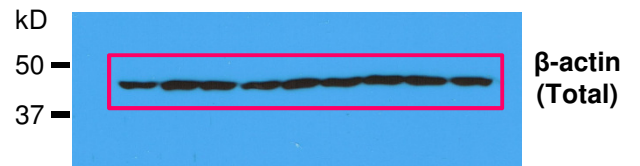
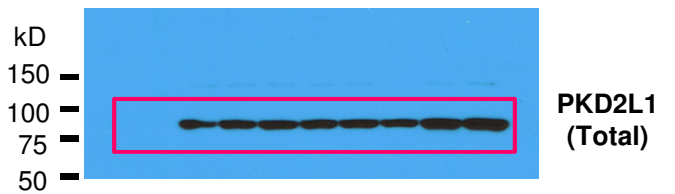
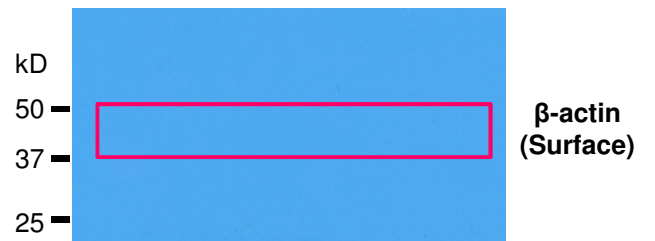
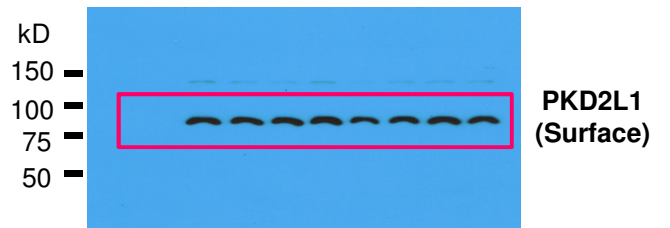


Figure 2e



Supplementary Figure 12. Uncropped images of the original scans of immunoblots.

Uncropped, full-size scans of immunoblots shown in Fig. 1e and Fig. 2e.

Figure 3c

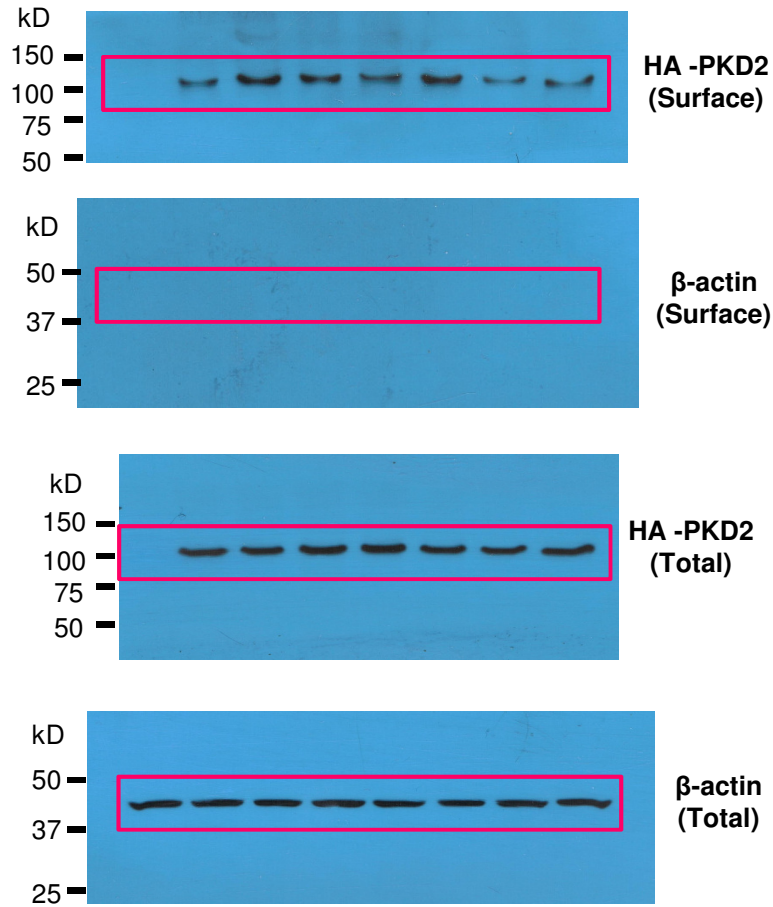
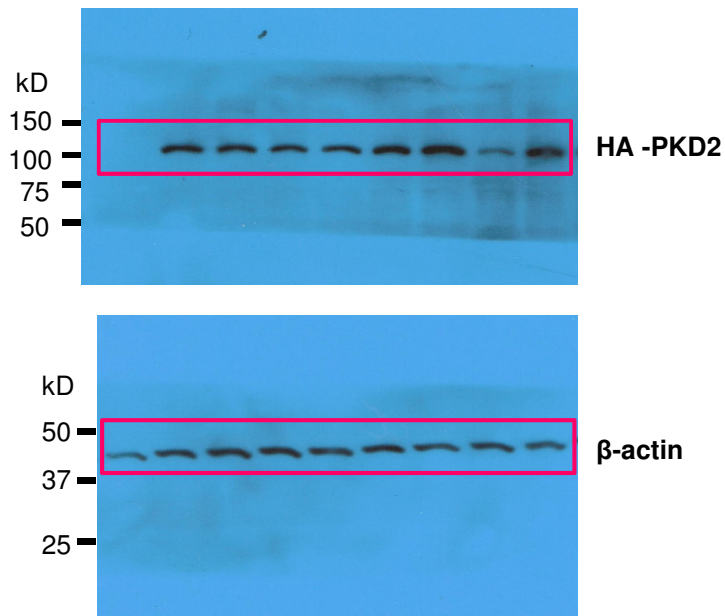


Figure 3d



Supplementary Figure 13. Uncropped images of the original scans of immunoblots.
Uncropped, full-size scans of immunoblots shown in Fig. 3c and d.

Supplementary Table 1. Cryo-EM data collection, refinement and validation statistics

PKD2 F604P (EMDB-7786) (PDB-6D1W)	
Data collection/Processing	
Voltage (kV)	300
Magnification	41,132
Defocus range (μm)	-0.6 – -2.4
Pixel size (\AA)	1.2156
Total electron dose ($\text{e}^-/\text{\AA}^2$)	80
Exposure time (s)	16
Number of images	1,992
Number of frames per image	80
Initial particle number	661,905
Final particle number	387,454
Resolution (unmasked, \AA)	3.8
Resolution (masked, \AA)	3.5
Refinement and Validation	
Number of atoms	14,452
R.M.S deviation	
Bond length (\AA)	0.008
Bond angles ($^\circ$)	0.794
Ramachandran	
Favored (%)	93.3%
Allowed (%)	6.5%
Outlier (%)	0.2%
Molprobrity score	2.20

Supplementary Table 2. Primers used in this study

Mutation	Direction	Primer (5'-3')
PKD2L1		
L552N	Forward (F)	CGTCTTCTTCGTGAACCTGAACATGTTCC
	Reverse (R)	GGAACATGTTTCAGGTTACGAAGAAGACG
L553N	F	GTCTTCTTCGTGCTCAATAACATGTTCTGTC
	R	GCCAGGAACATGTTATTGAGCACGAAGAAGAC
M555N	F	CGTGCTCCTGAACAATTTCTGGCCATCATC
	R	GATGATGGCCAGGAAATTGTTTCAGGAGCACG
F556N	F	GTCCTGAACATGAACCTGGCCATCATCAA
	R	TTGATGATGGCCAGGTTTCATGTTTCAGGAGC
L557N	F	GTCCTGAACATGTTCAATGCCATCATCAATGACAC
	R	GTGTCATTGATGATGGCATTGAACATGTTTCAGGAGC
A558N	F	GAACATGTTCTGAACATCATCAATGACAC
	R	GTGTCATTGATGATGTTTCAGGAACATGTTTC
I559N	F	GTTCTGGCCAACATCAATGACACATATTC
	R	GAATATGTGTCATTGATGTTGGCCAGGAAC
I560N	F	CCTGGCCATCAACAATGACACATATTC
	R	GAATATGTGTCATTGTTGATGGCCAGG
L557E	F	GTCCTGAACATGTTTCGAGGCCATCATCAATGACAC
	R	GTGTCATTGATGATGGCCTCGAACATGTTTCAGGAGC
L557D	F	GTCCTGAACATGTTTCGATGCCATCATCAATGACAC
	R	GTGTCATTGATGATGGCATCGAACATGTTTCAGGAGC
L557Q	F	GTCCTGAACATGTTCCAGGCCATCATCAATGACAC
	R	GTGTCATTGATGATGGCCTGGAACATGTTTCAGGAGC
L557S	F	GTCCTGAACATGTTTCAGTGCCATCATCAATGACAC
	R	GTGTCATTGATGATGGCACTGAACATGTTTCAGGAGC
L557C	F	GTCCTGAACATGTTCTGTGCCATCATCAATGACAC
	R	GTGTCATTGATGATGGCACAGAACATGTTTCAGGAGC
L557T	F	GTCCTGAACATGTTTCACGGCCATCATCAATGACAC
	R	GTGTCATTGATGATGGCCGTGAACATGTTTCAGGAGC
L557G	F	GTCCTGAACATGTTTCGGGGCCATCATCAATGACAC
	R	GTGTCATTGATGATGGCCCCGAACATGTTTCAGGAGC
L557A	F	GTCCTGAACATGTTTCGCGGCCATCATCAATGACAC
	R	GTGTCATTGATGATGGCCGCGAACATGTTTCAGGAGC
L557V	F	GTCCTGAACATGTTTCGTGGCCATCATCAATGACAC
	R	GTGTCATTGATGATGGCCACGAACATGTTTCAGGAGC
L557I	F	GTCCTGAACATGTTTCATTGCCATCATCAATGACAC
	R	GTGTCATTGATGATGGCAATGAACATGTTTCAGGAGC
L557F	F	GTCCTGAACATGTTCTTTGCCATCATCAATGACAC
	R	GTGTCATTGATGATGGCAAAGAACATGTTTCAGGAGC
L557W	F	GTCCTGAACATGTTCTGGGCCATCATCAATGACAC
	R	GTGTCATTGATGATGGCCCAGAACATGTTTCAGGAGC
A558Q	F	GAACATGTTCTGCAAATCATCAATGACAC
	R	GTGTCATTGATGATTTGCAGGAACATGTTTC
A558S	F	GAACATGTTCTGTCCATCATCAATGACAC
	R	GTGTCATTGATGATGGACAGGAACATGTTTC

A558C	F	GAACATGTTCCCTGTGCATCATCAATGACAC
	R	GTGTCATTGATGATGCACAGGAACATGTTC
A558T	F	GAACATGTTCCCTGACCATCATCAATGACAC
	R	GTGTCATTGATGATGGTCAGGAACATGTTC
A558G	F	GAACATGTTCCCTGGGCATCATCAATGACAC
	R	GTGTCATTGATGATGCCCAGGAACATGTTC
A558V	F	GAACATGTTCCCTGGTCATCATCAATGACAC
	R	GTGTCATTGATGATGACCAGGAACATGTTC
A558I	F	GAACATGTTCCCTGATCATCATCAATGACAC
	R	GTGTCATTGATGATGATCAGGAACATGTTC
A558L	F	GAACATGTTCCCTGCTCATCATCAATGACAC
	R	GTGTCATTGATGATGAGCAGGAACATGTTC
A558F	F	GAACATGTTCCCTGTTTCATCATCAATGACAC
	R	GTGTCATTGATGATGAACAGGAACATGTTC
A558W	F	GAACATGTTCCCTGTGGATCATCAATGACAC
	R	GTGTCATTGATGATCCACAGGAACATGTTC
L557W/A558N	F	CCTGAACATGTTCTGGAACATCATCAATGACAC
	R	GTGTCATTGATGATGTTCCAGAACATGTTTCAGG
A558W/L557N	F	CCTGAACATGTTCAACTGGATCATCAATGACAC
	R	GTGTCATTGATGATCCAGTTGAACATGTTTCAGG
K461A	F	CATCAGCTTCAACGCAACCATGACCCAG
	R	CTGGGTCATGGTTGCGTTGAAGCTGATG
PKD2		
F676N	F	CTTCATTCTTTTGAATATGAATTTGGCTATCATCAATG
	R	CATTGATGATAGCCAAATTCATATTCAAAAGAATGAAG
L677N	F	CATTCTTTTGAATATGTTTAATGCTATCATCAATGATAC
	R	GTATCATTGATGATAGCATTAAACATATTCAAAAGAATG
A678N	F	CTTTTGAATATGTTTTTGAATATCATCAATGATACTTAC
	R	GTAAGTATCATTGATGATATTCAAAACATATTCAAAAG
I679N	F	GAATATGTTTTTGGCTAACATCAATGATACTTAC
	R	GTAAGTATCATTGATGTTAGCCAAAACATATTC
A678Q	F	CTTTTGAATATGTTTTTGCAAATCATCAATGATACTTAC
	R	GTAAGTATCATTGATGATTTTGCAAACATATTCAAAAG
A678S	F	CTTTTGAATATGTTTTTGCTATCATCAATGATACTTAC
	R	GTAAGTATCATTGATGATAGACAAAACATATTCAAAAG
A678T	F	CTTTTGAATATGTTTTTGACTATCATCAATGATACTTAC
	R	GTAAGTATCATTGATGATAGTCAAAAACATATTCAAAAG
L677G	F	CATTCTTTTGAATATGTTTGGGGCTATCATCAATGATAC
	R	GTATCATTGATGATAGCCCAAACATATTCAAAAGAATG
L677A	F	CATTCTTTTGAATATGTTTGCGGCTATCATCAATGATAC
	R	GTATCATTGATGATAGCCGCAAACATATTCAAAAGAATG
L677F	F	CATTCTTTTGAATATGTTTTTGGCTATCATCAATGATAC
	R	GTATCATTGATGATAGCAAACATATTCAAAAGAATG
L677Y	F	CATTCTTTTGAATATGTTTTACGCTATCATCAATGATAC
	R	GTATCATTGATGATAGCGTAAAACATATTCAAAAGAATG
L677W	F	CATTCTTTTGAATATGTTTTGGGCTATCATCAATGATAC
	R	GTATCATTGATGATAGCCCAAACATATTCAAAAGAATG
W201A	F	GGCTGCGAGGTCTCGCGGGAACAAGACTCATG
	R	CATGAGTCTTGTTCGCGGAGACCTCGCAGCC

K688A	F	CTTACTCTGAAGTGGCATCTGACTTGGC
	R	GCCAAGTCAGATGCCACTTCAGAGTAAG
D511V	F	GTTTCTGGAATTGTCTGGTTGTTGTGATCGTTGTGC
	R	GCACAACGATCACAACAACCAGACAATTCCAGAAAC
ΔY684	F	CTATCATCAATGATACTTCTGAAGTGAAATCTGACT
	R	AGTCAGATTTCACTTCAGAAGTATCATTGATGATAG
Y684A	F	CTATCATCAATGATACTGCCTCTGAAGTGAAATCTGACT
	R	AGTCAGATTTCACTTCAGAGGCAGTATCATTGATGATAG
L677N/N68 1L	F	GTTTATTGCTATCATCCTTGATACTTACTCTG
	R	CAGAGTAAGTATCAAGGATGATAGCAATAAAC
R581A	F	CAAATTCATCAATTTTAACGCGACCATGAGCCAG
	R	CTGGCTCATGGTCGCGTTAAAATTGATGAATTTG
F676G	F	CTTCATTCTTTTGAATATGGGTTTGGCTATCATCAATG
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F676A	F	CTTCATTCTTTTGAATATGGCTTTGGCTATCATCAATG
	R	CATTGATGATAGCCAAAGCCATATTCAAAGAATGAAG
F676Y	F	CTTCATTCTTTTGAATATGTATTTGGCTATCATCAATG
	R	CATTGATGATAGCCAAATACATATTCAAAGAATGAAG
F676W	F	CTTCATTCTTTTGAATATGTGGTTGGCTATCATCAATGATAC
	R	GTATCATTGATGATAGCCAACCACATATTCAAAGAATGAAG
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	R	CATTGATGATAGCGTTACCCATATTCAAAGAATGAAG
L677N/F67 6A	F	CTTCATTCTTTTGAATATGGCTAACGCTATCATCAATG
	R	CATTGATGATAGCGTTAGCCATATTCAAAGAATGAAG
L677N/F67 6Y	F	CTTCATTCTTTTGAATATGTATAACGCTATCATCAATG
	R	CATTGATGATAGCGTTATACATATTCAAAGAATGAAG
L677N/F67 6W	F	CTTCATTCTTTTGAATATGTGGAACGCTATCATCAATG
	R	CATTGATGATAGCGTTCCACATATTCAAAGAATGAAG