

Supplemental Material for

***In silico* genome-wide analysis reveals the potential links between core genome of *Acidithiobacillus thiooxidans* and its autotrophic lifestyle**

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SUPPLEMENTARY FIGURES

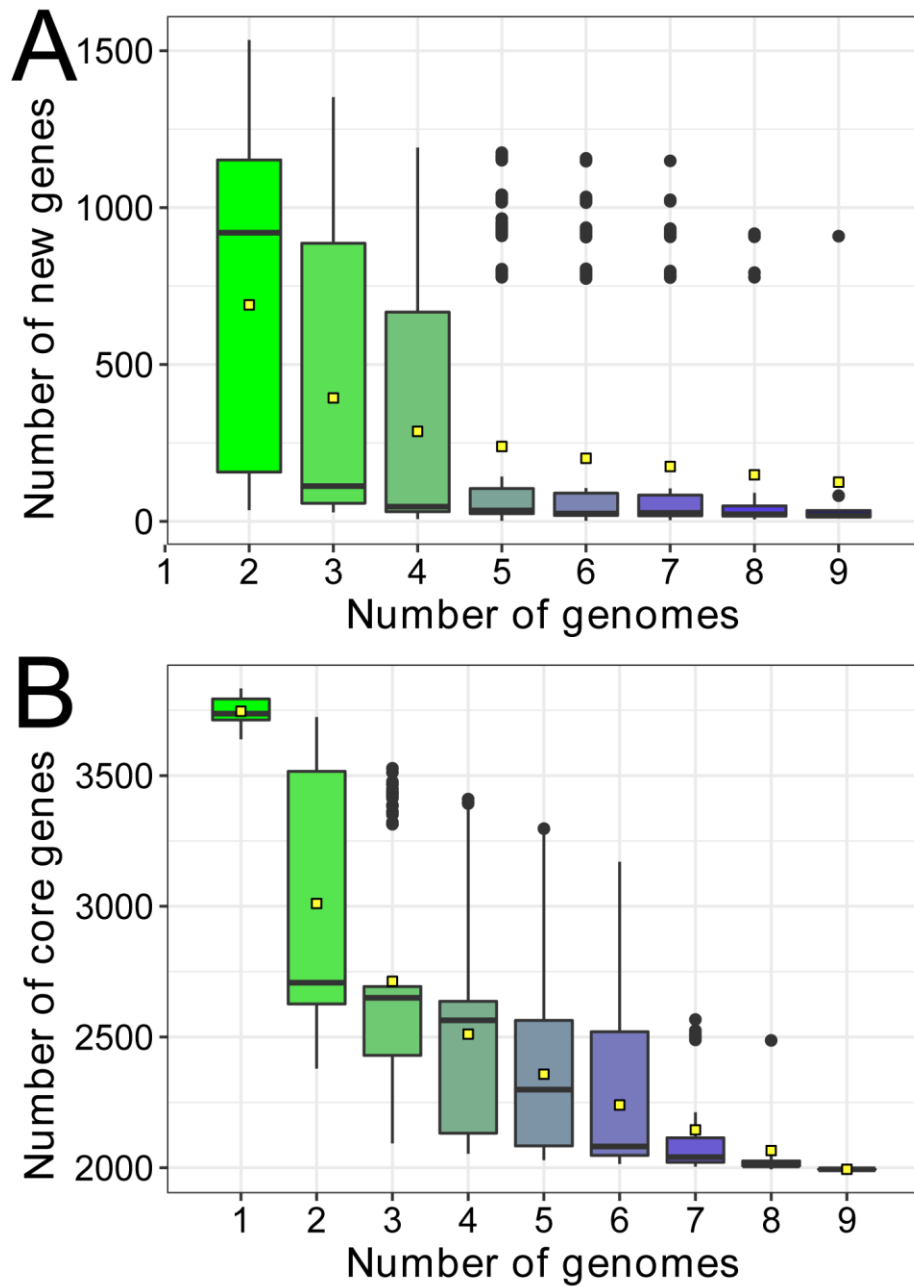


FIGURE S1 Distributions of new genes and core genes' values: averages vs. medians. Dark spots represent the distributions of new genes and core genes' values. For each N (the number of sequenced genomes), the box-plots indicating the medians (black lines) and the average (yellow rectangle) were shown.

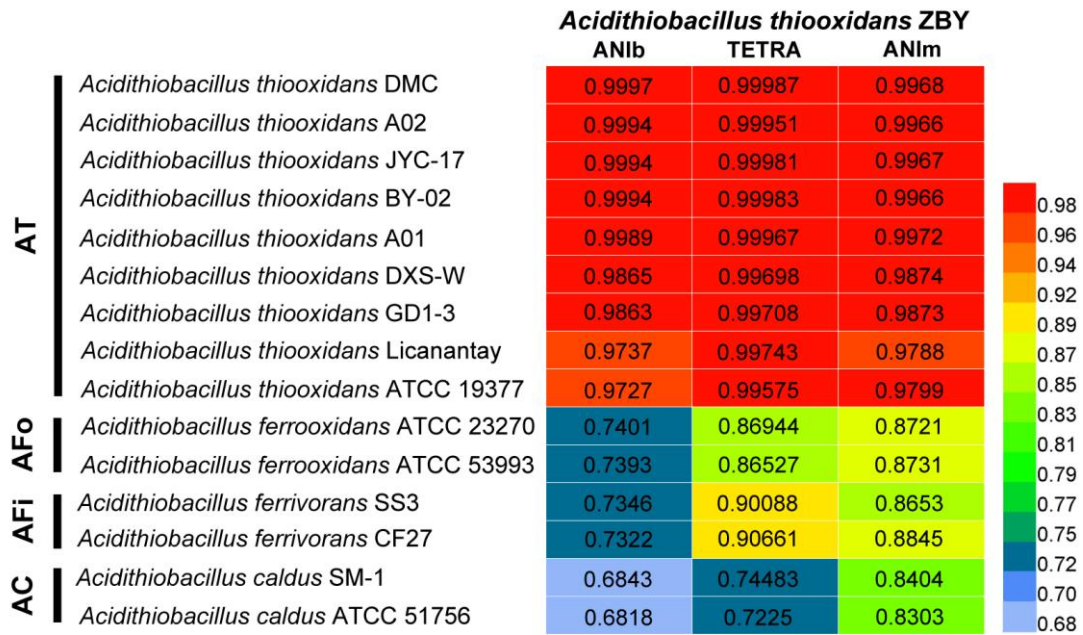


FIGURE S2 Genome-based phylogenetic indicators of *Acidithiobacillus* spp. The average nucleotide identity (ANI) based on BLAST (ANiB) and MUMmer (ANIm), as well as tetranucleotide frequency correlation coefficient (TETRA) were calculated using the software JSpecies v1.2.1 with default parameters. Values above the thresholds of $\geq 95\%$ (ANI) and ≥ 0.99 (TETRA) indicate that strains belong to the same species. Abbreviation: AT, *Acidithiobacillus thiooxidans*; AFo: *Acidithiobacillus ferrooxidans*; AFi: *Acidithiobacillus ferrivorans*; AC: *Acidithiobacillus caldus*.

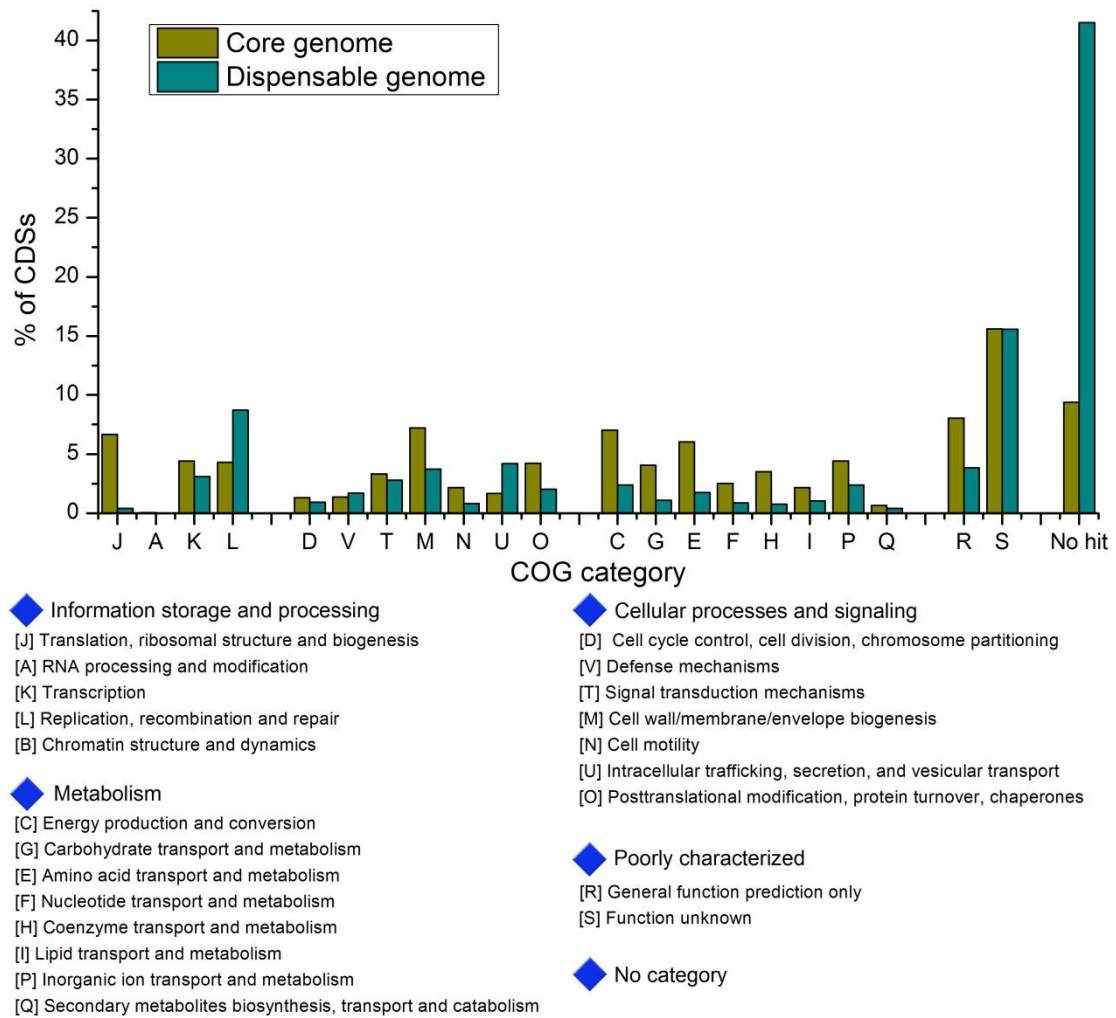


FIGURE S3 Functional assignments of core and dispensable genome via alignment against the extended COG database.

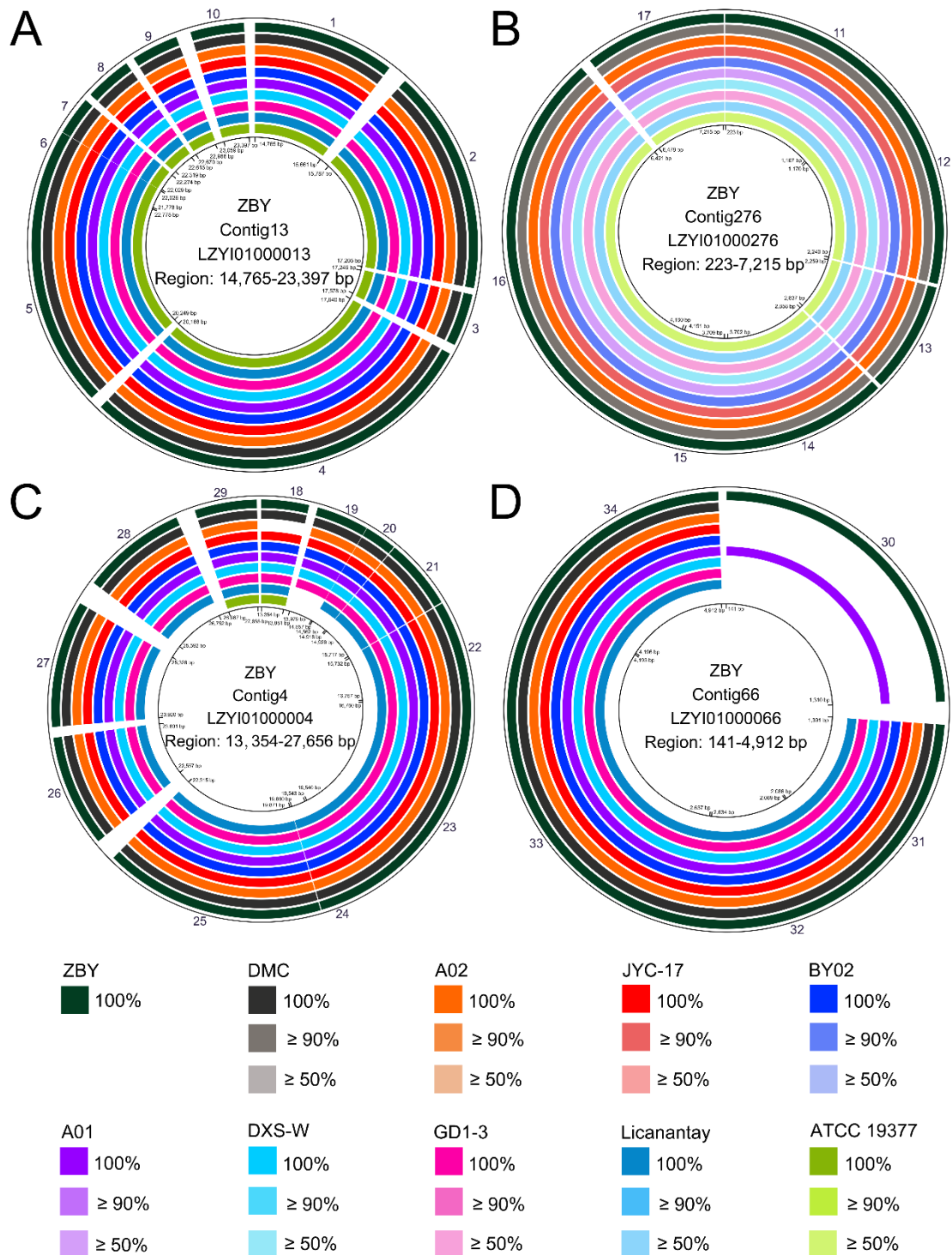


FIGURE S4 Partial genomic map of *A. thiooxidans* strains containing carboxysome-associated gene cluster (A), *coxLMS* (B), *nirBD/nasA* (C), and *narGHJI* (D). Genomic regions of *A. thiooxidans* ZBY were used as the reference. Matches to the reference genome were displayed color-coded on the rings two to ten. Color intensity indicates the level of sequence identities between query genome and reference genome (e.g. higher color intensity means the higher sequence identity). Proteins in different genomic regions of interest were numbered with 1 to 34. Abbreviation: 1, RuBisCO operon transcriptional regulator; 2, ribulose-bisphosphate carboxylase large subunit;

3, ribulose biphosphate carboxylase small subunit; 4, carboxysome shell protein CsoS2; 5, carboxysome shell carbonic anhydrase; 6, carboxysome peptide A; 7, carboxysome peptide B; 8-10, carbon dioxide-concentrating protein CcmK; 11, xanthine dehydrogenase accessory factor; 12, amidohydrolase; 13, ureidoglycolate hydrolase; 14, carbon-monoxide dehydrogenase medium subunit/xanthine dehydrogenase FAD-binding subunit; 15, carbon-monoxide dehydrogenase small subunit/xanthine dehydrogenase, iron-sulfur binding subunit; 16, carbon monoxide dehydrogenase, large subunit/xanthine dehydrogenase, molybdenum-binding subunit; 17, phosphorylase; 18, hypothetical protein; 19, response regulator NasT; 20, cobalamin biosynthesis protein CbiX; 21, uroporphyrin-III methyltransferase; 22, glycosyl transferase, family 3; 23, assimilatory nitrate reductase (NADH) catalytic subunit NasA; 24, assimilatory nitrite reductase [NAD(P)H] small subunit NirD; 25, assimilatory nitrite reductase [NAD(P)H] large subunit NirB; 26, hypothetical protein; 27, major facilitator superfamily MFS_1, nitrate/nitrite transporter; 28, nitrate transporter; 29, transcriptional regulator, Crp/Fnr family; 30, MFS transporter; 31, respiratory nitrate reductase, gamma subunit NarI; 32, respiratory nitrate reductase delta subunit NarJ; 33, respiratory nitrate reductase beta subunit NarH; 34, respiratory nitrate reductase alpha subunit NarG.

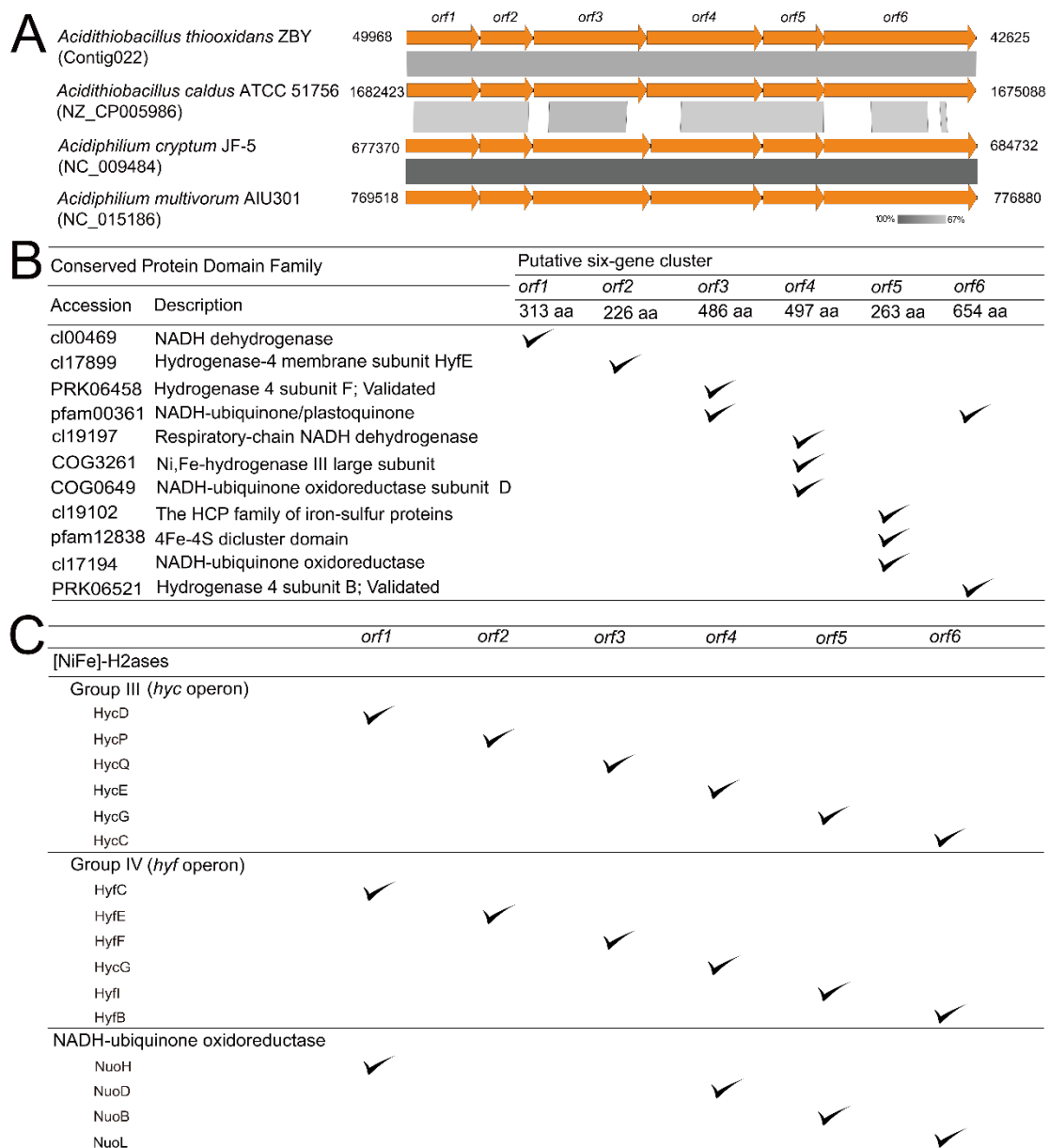


FIGURE S5 Functional assignments of putative six-gene clusters in *A. thiooxidans* strains. (A) Comparison of *A. thiooxidans* ZBY and other microorganisms containing identified or putative gene cluster; (B) Identification of conserved domains of complex enzyme encoded by six-gene cluster; (C) Comparison of amino acid sequences involved in these candidate genes to the identified [NiFe]-hydrogenases and NADH-ubiquinone oxidoreductase.

SUPPLEMENTARY TABLES

TABLE S1 List of the genomes of novel strain ZBY and other recognized *Acidithiobacillus* (*A.*) spp. used for the construction of genome-based phylogeny and the calculation of average nucleotide identity.

Organism	Accession Number	Genome size (Mb)	GC %	Reference
Strain ZBY	LZYI000000000	3.79	53.17	This study
<i>A. thiooxidans</i>				
ATCC 19377	AFOH000000000	3.02	53.20	(Valdes et al., 2011)
A01	AZMO000000000	3.82	53.10	(Yin et al., 2014)
Licanantay	JMEB000000000	3.94	52.80	(Travisany et al., 2014)
GD1-3	LWSC000000000	3.95	52.92	(Zhang et al., 2016a)
DXS-W	LWRY000000000	3.95	52.92	(Zhang et al., 2016a)
A02	LWSA000000000	3.72	53.01	(Zhang et al., 2016a)
BY-02	LWRZ000000000	3.81	53.07	(Zhang et al., 2016a)
DMC	LWSB000000000	3.85	53.06	(Zhang et al., 2016a)
TYC-17	LWSD000000000	3.83	53.06	(Zhang et al., 2016a)
<i>A. ferrooxidans</i>				
ATCC 23270	NC_011761	2.98	58.80	(Valdés et al., 2008)
ATCC 53993	NC_011206	2.89	58.90	Unpublished
<i>A. caldus</i>				
SM-1	NC_015850	3.24	60.94	(You et al., 2011)
ATCC 51756	NZ_CP005986	2.99	61.39	(Valdes et al., 2009)
<i>A. ferrivorans</i>				
SS3	NC_015942	3.21	56.60	(Liljeqvist et al., 2011)
CF27	CCCS000000000	3.43	56.40	(Talla et al., 2014)

TABLE S2 General features of bacterial strains used for comparative survey in this study.

Strain	Accession number	Genome status	Genome size (Mbp)	Number of CDS	Completeness (%)	Reference
ZBY	LZYI00000000	Draft	3.79	3,713	99.34	This study
A01	AZMO00000000	Draft	3.82	3,719	98.72	(Yin et al., 2014)
Licanantay	JMEB00000000	Draft	3.94	3,793	99.34	(Travisany et al., 2014)
DMC	LWSB00000000	Draft	3.85	3,772	99.34	(Zhang et al., 2016)
A02	LWSA00000000	Draft	3.72	3,639	99.34	(Zhang et al., 2016)
GD1-3	LWSC00000000	Draft	3.95	3,824	99.34	(Zhang et al., 2016)
BY-02	LWRZ00000000	Draft	3.81	3,683	99.03	(Zhang et al., 2016)
JYC-17	LWSD00000000	Draft	3.83	3,737	99.34	(Zhang et al., 2016)
DXS-W	LWRY00000000	Draft	3.95	3,833	99.34	(Zhang et al., 2016)

TABLE S3 The distribution of the COG functional classification in the genome of *A. thiooxidans* ZBY.

COG category	Description	COGs Number	Percent (%)
A	RNA processing and modification	1	0.03
C	Energy production and conversion	177	4.77
D	Cell cycle control, cell division, chromosome partitioning	41	1.10
E	Amino acid transport and metabolism	145	3.91
F	Nucleotide transport and metabolism	61	1.64
G	Carbohydrate transport and metabolism	97	2.61
H	Coenzyme transport and metabolism	80	2.15
I	Lipid transport and metabolism	61	1.64
J	Translation, ribosomal structure, and biogenesis	139	3.74
K	Transcription	141	3.80
L	Replication, recombination, and repair	231	6.22
M	Cell wall/membrane/envelope biogenesis	207	5.58
N	Cell motility	54	1.45
O	Posttranslational modification, protein turnover, chaperones	118	3.18
P	Inorganic ion transport and metabolism	127	3.42
Q	Secondary metabolites biosynthesis, transport and catabolism	20	0.54
R	General function prediction only	196	5.28
S	Function unknown	574	15.46
T	Signal transduction mechanisms	113	3.04
U	Intracellular trafficking, secretion, and vesicular transport	104	2.80
V	Defense mechanisms	54	1.45
	No hits	972	26.18

The four most abundant functional categories are shown in bold.

TABLE S4 The fitting results of function F_c (A), F_s (B), and P_s (C) of *A. thiooxidans* pan-genome.

(a)				
ϵ_c	τ_c	Ω	R-square	Adj. R-square ¹
2,690 ± 421***	2.0 ± 0.54***	1,994 ± 118***	0.9912	0.9883
(b)				
ϵ_s	τ_s	$tg(\theta)$	R-square	Adj. R-square ¹
93,320 ± 16,520***	0.43 ± 0.04***	28 ± 3***	0.9996	0.9995
(c)				
κ	α		R-square	Adj. R-square ¹
26,798 ± 7,397***	4.9 ± 0.4***		0.9958	0.9944

The parameter results are in form of estimate ± standard error, and are labeled by '***' when $p < 0.001$.

¹ Adjust R-square = $1 - (1 - R\text{-square}) * (n - 1) / (n - k)$, where n is the number of strains, and k is the number of parameter (here, $k=3$).

TABLE S5 Classification of core genome of *A. thiooxidans* species using the online platform KAAS.

Pathway Number	Description	CDS Number
Carbohydrate metabolism		172
ko00010	Glycolysis / Gluconeogenesis	26
ko00020	Citrate cycle (TCA cycle)	14
ko00030	Pentose phosphate pathway	15
ko00040	Pentose and glucuronate interconversions	4
ko00051	Fructose and mannose metabolism	10
ko00052	Galactose metabolism	6
ko00053	Ascorbate and aldarate metabolism	2
ko00500	Starch and sucrose metabolism	10
ko00520	Amino sugar and nucleotide sugar metabolism	20
ko00620	Pyruvate metabolism	22
ko00630	Glyoxylate and dicarboxylate metabolism	21
ko00640	Propanoate metabolism	8
ko00650	Butanoate metabolism	3
ko00660	C5-Branched dibasic acid metabolism	6
ko00562	Inositol phosphate metabolism	5
Energy metabolism		133
ko00190	Oxidative phosphorylation	50
ko00195	Photosynthesis	8
ko00710	Carbon fixation in photosynthetic organisms	14
ko00720	Carbon fixation pathways in prokaryotes	13
ko00680	Methane metabolism	19
ko00910	Nitrogen metabolism	8
ko00920	Sulfur metabolism	21
Lipid metabolism		40
ko00061	Fatty acid biosynthesis	9
ko00071	Fatty acid degradation	2
ko00100	Steroid biosynthesis	1
ko00120	Primary bile acid biosynthesis	1
ko00121	Secondary bile acid biosynthesis	1
ko00561	Glycerolipid metabolism	8
ko00564	Glycerophospholipid metabolism	13
ko00565	Ether lipid metabolism	2
ko00600	Sphingolipid metabolism	1
ko00590	Arachidonic acid metabolism	1
ko01040	Biosynthesis of unsaturated fatty acids	1
Nucleotide metabolism		67
ko00230	Purine metabolism	41
ko00240	Pyrimidine metabolism	26
Amino acid metabolism		142
ko00250	Alanine, aspartate and glutamate metabolism	13

ko00260	Glycine, serine and threonine metabolism	21
ko00270	Cysteine and methionine metabolism	25
ko00280	Valine, leucine and isoleucine degradation	4
ko00290	Valine, leucine and isoleucine biosynthesis	10
ko00300	Lysine biosynthesis	10
ko00220	Arginine biosynthesis	9
ko00330	Arginine and proline metabolism	11
ko00340	Histidine metabolism	10
ko00350	Tyrosine metabolism	4
ko00360	Phenylalanine metabolism	4
ko00380	Tryptophan metabolism	1
ko00400	Phenylalanine, tyrosine and tryptophan biosynthesis	20
Metabolism of other amino acids		36
ko00410	beta-Alanine metabolism	4
ko00430	Taurine and hypotaurine metabolism	2
ko00440	Phosphonate and phosphinate metabolism	7
ko00450	Selenocompound metabolism	3
ko00460	Cyanoamino acid metabolism	2
ko00471	D-Glutamine and D-glutamate metabolism	2
ko00473	D-Alanine metabolism	2
ko00480	Glutathione metabolism	14
Glycan biosynthesis and metabolism		41
ko00531	Glycosaminoglycan degradation	1
ko00540	Lipopolysaccharide biosynthesis	19
ko00550	Peptidoglycan biosynthesis	21
Metabolism of cofactors and vitamins		94
ko00730	Thiamine metabolism	7
ko00740	Riboflavin metabolism	5
ko00750	Vitamin B6 metabolism	4
ko00760	Nicotinate and nicotinamide metabolism	8
ko00770	Pantothenate and CoA biosynthesis	13
ko00780	Biotin metabolism	9
ko00785	Lipoic acid metabolism	3
ko00790	Folate biosynthesis	15
ko00670	One carbon pool by folate	6
ko00830	Retinol metabolism	1
ko00860	Porphyrin and chlorophyll metabolism	13
ko00130	Ubiquinone and other terpenoid-quinone biosynthesis	10
Metabolism of terpenoids and polyketides		16
ko00900	Terpenoid backbone biosynthesis	8
ko00909	Sesquiterpenoid and triterpenoid biosynthesis	2
ko00908	Zeatin biosynthesis	1
ko00523	Polyketide sugar unit biosynthesis	4
ko01055	Biosynthesis of vancomycin group antibiotics	1
Biosynthesis of other secondary metabolites		28
ko00940	Phenylpropanoid biosynthesis	1

ko00950	Isoquinoline alkaloid biosynthesis	1
ko00960	Tropane, piperidine and pyridine alkaloid biosynthesis	3
ko00965	Betalain biosynthesis	1
ko00332	Carbapenem biosynthesis	3
ko00261	Monobactam biosynthesis	4
ko00521	Streptomycin biosynthesis	8
ko00524	Neomycin, kanamycin and gentamicin biosynthesis	1
ko00525	Acarbose and validamycin biosynthesis	2
ko00401	Novobiocin biosynthesis	4
Xenobiotics biodegradation and metabolism		21
ko00627	Aminobenzoate degradation	2
ko00364	Fluorobenzoate degradation	1
ko00625	Chloroalkane and chloroalkene degradation	1
ko00361	Chlorocyclohexane and chlorobenzene degradation	1
ko00623	Toluene degradation	1
ko00633	Nitrotoluene degradation	2
ko00643	Styrene degradation	1
ko00791	Atrazine degradation	2
ko00626	Naphthalene degradation	1
ko00980	Metabolism of xenobiotics by cytochrome P450	3
ko00982	Drug metabolism - cytochrome P450	3
ko00983	Drug metabolism - other enzymes	3

TABLE S6 The predicted enzymes involved in amino sugar and nucleotide sugar metabolism in all *A. thiooxidans* genomes.

Gene	Description	KO Number	Enzyme Number
<i>nagZ</i>	beta-N-acetylhexosaminidase	K01207	3.2.1.52
<i>amgK</i>	anomeric MurNAc/GlcNAc kinase	K07102	2.7.1.-
<i>murU</i>	MurNAc alpha-1-phosphate uridylyltransferase	K00992	2.7.7.-
<i>glmU</i>	bifunctional UDP-N-acetylglucosamine pyrophosphorylase / Glucosamine-1-phosphate N-acetyltransferase	K04042	2.7.7.23 / 2.3.1.157
<i>murA</i>	UDP-N-acetylglucosamine 1-carboxyvinyltransferase	K00790	2.5.1.7
<i>murB</i>	UDP-N-acetylmuramate dehydrogenase	K00075	1.3.1.98
<i>glmM</i>	phosphoglucosamine mutase	K03431	5.4.2.10
<i>scrK</i>	fructokinase	K00847	2.7.1.4
<i>glk</i>	glucokinase	K00845	2.7.1.2
<i>pgm</i>	phosphoglucomutase	K01835	5.4.2.2
<i>galU</i>	UTP--glucose-1-phosphate uridylyltransferase	K00963	2.7.7.9
<i>ugd</i>	UDPglucose 6-dehydrogenase	K00012	1.1.1.22
<i>galE</i>	UDP-glucose 4-epimerase	K01784	5.1.3.2
	UDP-glucuronate 4-epimerase	K08679	5.1.3.6
<i>manB</i>	phosphomannomutase	K01840	5.4.2.8
<i>rfbA</i>	mannose-1-phosphate guanylyltransferase / mannose-6-phosphate isomerase	K16011	2.7.7.13 / 5.3.1.8
<i>gmd</i>	GDPmannose 4,6-dehydratase	K01711	4.2.1.47
<i>rmd</i>	GDP-4-dehydro-6-deoxy-D-mannose reductase	K15856	1.1.1.281
<i>rfbI</i>	CDP-4-dehydro-6-deoxyglucose reductase, E3	K00523	1.17.1.1
<i>glgC</i>	glucose-1-phosphate adenylyltransferase	K00975	2.7.7.27

TABLE S7 Genes involved in sulfur oxidation and electron transfer in core genome of *A. thiooxidans* species.

Gene	Description	KO Number	Enzyme Number
<i>sqr</i>	sulfide quinone oxidoreductase	K17218	1.8.5.4
Tetrathionate hydrolase operon			
<i>tetH</i>	tetrathionate hydrolase		
<i>doxD</i>	thiosulfate:quinone oxidoreductase subunit DoxD	K16936	1.8.5.2
Sulfur-oxidizing protein			
<i>soxA</i>	sulfur-oxidizing protein SoxA	K17222	
<i>soxX</i>	sulfur-oxidizing protein SoxX	K17223	
<i>soxY</i>	sulfur-oxidizing protein SoxY	K17226	
<i>soxZ</i>	sulfur-oxidizing protein SoxZ	K17227	
<i>soxB</i>	sulfur-oxidizing protein SoxB	K17224	
Heterodisulfide reductase			
<i>hdrA</i>	heterodisulfide reductase subunit A		
<i>hdrB</i>	heterodisulfide reductase subunit B		
<i>hdrC</i>	heterodisulfide reductase subunit C		
<i>tst</i>	thiosulfate sulfurtransferase		2.8.1.1
<i>sat</i>	sulfate adenylyltransferase		2.7.7.4
<i>bd</i> ubiquinol oxidases			1.10.3.14
<i>cydA</i>	cytochrome <i>d</i> ubiquinol oxidase subunit I	K00425	
<i>cydB</i>	cytochrome <i>d</i> ubiquinol oxidase subunit II	K00426	
<i>bo₃</i> ubiquinol oxidases			1.10.3.10
<i>cyoA</i>	cytochrome <i>o</i> ubiquinol oxidase, subunit II	K02297	
<i>cyoB</i>	cytochrome <i>o</i> ubiquinol oxidase subunit I	K02298	
<i>cyoC</i>	cytochrome <i>o</i> ubiquinol oxidase subunit III	K02299	
<i>cyoD</i>	cytochrome <i>o</i> ubiquinol oxidase subunit IV	K02300	
NADH-ubiquinone oxidoreductase (complex I)			1.6.5.3
<i>nuoA</i>	NADH-ubiquinone oxidoreductase, A subunit	K00330	
<i>nuoB</i>	NADH-ubiquinone oxidoreductase, B subunit	K00331	
<i>nuoC</i>	NADH-ubiquinone oxidoreductase, C subunit	K00332	
<i>nuoD</i>	NADH-ubiquinone oxidoreductase, D subunit	K00333	
<i>nuoE</i>	NADH-ubiquinone oxidoreductase, E subunit	K00334	
<i>nuoF</i>	NADH-ubiquinone oxidoreductase, F subunit	K00335	
<i>nuoG</i>	NADH-ubiquinone oxidoreductase, G subunit	K00336	
<i>nuoH</i>	NADH-ubiquinone oxidoreductase, H subunit	K00337	
<i>nuoI</i>	NADH-ubiquinone oxidoreductase, I subunit	K00338	
<i>nuoJ</i>	NADH-ubiquinone oxidoreductase, J subunit	K00339	
<i>nuoK</i>	NADH-ubiquinone oxidoreductase, K subunit	K00340	
<i>nuoL</i>	NADH-ubiquinone oxidoreductase, L subunit	K00341	
<i>nuoM</i>	NADH-ubiquinone oxidoreductase, M subunit	K00342	
<i>nuoN</i>	NADH-ubiquinone oxidoreductase, N subunit	K00343	

SUPPLEMENTARY REFERENCES

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