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1. General information

All the reactions dealing with air- or moisture- sensitive compounds were carried out in a dry reaction vessel under nitrogen protection or in the nitrogen-filled glove box. Unless otherwise noted, all reagents and solvents were purchased from commercial suppliers without further purification. THF was dried with sodium chips and indicated by benzophenone. Other anhydrous solvents were purchased from Sigma-Aldrich and transferred by syringe. Purification of products was carried out by chromatography using silica gel from ACROS (0.06-0.20mm). Thin layer chromatography was carried out using silica gel plates from Merk (GF254). [Rh(COD)Cl]₂ and other metal precursors were purchased from Heraeus.

¹H NMR, ¹³C NMR and ³¹P NMR spectra were recorded on a Bruker Avance (400 MHz) spectrometer with CDCl₃ as the solvent and tetramethylsilane (TMS) as the internal standard. Chemical shifts are reported in parts per million e (ppm, δ scale) downfield from TMS at 0.00 ppm and referenced to the CDCl₃ at 7.26 ppm for ¹H NMR or 77.0 ppm for ¹³C NMR. Data is reported as: multiplicity (s = singlet, d = doublet, t = triplet, q = quartet, m = multiplet), coupling constant in hertz (Hz) and signal area integration in natural numbers. ¹³C NMR and ³¹P NMR analyses were ran with decoupling. Enantiomeric excess values were determined with chiral columns on Agilent 7980 Series GC instrument or Agilent 1100 Series HPLC instrument. Optical rotations were measured using a 1 mL cell with a 1 dm path length on a Jasco P–2000 polarimeter at 589 nm and at 20 °C

2. Synthesis of 2-substituted quinolines



2-ethylquinoline^[1]:

To a solution of 1-methylquinoline (10 mmol) in dry THF (20 ml) at 0°C was added nbutyllithium in THF solution (2.5M) dropwise. The mixture was stirred at room temperature for 1.5h and was cooled back to 0°C. Iodomethane was added dropwise. The mixture was stirred overnight. Water was added carefully to quench the reaction and the mixture was extracted by ethyl acetate (20 ml*3). Organic phases were combined and dried with sodium sulfate. After removing solvent, the crude product was purified by flash chromatography (on silica gel, eluent: hexanes/ethyl acetate). 2methylquinoline was obtained as yellow oil with 95% yield.

Known compound. ¹H NMR (400 MHz, CDCl₃) δ 8.04 (dd, *J* = 11.8, 8.5 Hz, 1H), 7.74 (d, *J* = 8.1 Hz, 1H), 7.65 (ddd, *J* = 8.3, 6.9, 1.4 Hz, 1H), 7.45 (dd, *J* = 11.0, 4.0 Hz, 1H), 7.28 – 7.25 (d, *J* = 8.5 Hz, 1H), 2.99 (q, *J* = 7.6 Hz, 2H), 1.39 (dd, *J* = 7.6 Hz, 3H). ¹³C NMR (400 MHz, CDCl₃): δ 163.97, 147.87, 136.31, 129.30, 128.79, 127.46, 126.37, 125.63, 120.81, 32.28, 13.97.

2-isopropylquinoline^[1]

Followed the standard procedure above, using 2-ethylquinoline and iodomethane, 2isopropylquinoline was abotained as yellow oil with 76% yield.

¹H NMR (400 MHz, CDCl₃) δ 8.05 (dd, *J* = 8.2, 6.0 Hz, 2H), 7.74 (dd, *J* = 8.1, 1.1 Hz, 1H), 7.65 (ddd, *J* = 8.4, 6.9, 1.4 Hz, 1H), 7.45 (ddd, *J* = 8.1, 7.0, 1.1 Hz, 1H), 7.30 (t, *J* = 8.6 Hz, 1H), 3.26 (hept, *J* = 6.9 Hz, 1H), 1.38 (d, *J* = 6.9 Hz, 3H).

¹³C NMR (400 MHz, CDCl3): δ 167.63, 147.79, 136.36, 129.21, 129.04, 127.42, 126.95, 125.62, 119.16, 37.29, 22.52.

2-pentylquinoline^[1]

Yellow oil. ¹H NMR (400 MHz, CDCl₃) δ 8.04 (t, *J* = 8.4 Hz, 2H), 7.75 (d, *J* = 8.1 Hz, 1H), 7.66 (t, *J* = 7.7 Hz, 1H), 7.45 (t, *J* = 7.5 Hz, 1H), 7.27 (d, *J* = 8.4 Hz, 1H), 2.99 – 2.94 (m, 2H), 1.86 – 1.77 (m, 2H), 1.37 (m, 2H), 0.90 (t, *J* = 6.7 Hz, 3H). ¹³C NMR (400 MHz, CDCl3): δ 163.10, 147.97, 136.10, 129.25, 128.88, 127.45, 126.72, 125.59, 121.34, 39.35, 31.76, 29.71, 22.56, 14.00.

2-undecylquinoline^[1]

Yellow oil. ¹H NMR (400 MHz, CDCl₃) δ 7.76 (dd, *J* = 8.1, 1.4 Hz, 1H), 7.67 (ddd, *J* = 8.3, 6.9, 1.4 Hz, 1H), 7.46 (ddd, *J* = 8.0, 7.0, 1.0 Hz, 1H), 7.28 (d, *J* = 8.4 Hz, 1H), 2.96 (t, *J* = 8.0 Hz, 2H), 1.86 – 1.76 (m, 2H), 1.46 – 1.19 (m, 16H), 0.90 – 0.85 (m, 3H). ¹³C NMR (400 MHz, CDCl3): δ 163.12, 147.97, 136.09, 129.25, 128.88, 127.44, 126.72, 125.58, 121.34, 39.40, 31.68, 30.03, 29.58, 29.56, 29.52, 29.30, 22.66, 14.09.

2-(3,4-dimethoxyphenethyl)quinoline^[1]

Yellow oil. ¹H NMR (400 MHz, CDCl₃) δ 8.07 (d, *J* = 8.4, 1H), 8.03 (d, *J* = 8.4 Hz, 1H), 7.77 (dd, *J* = 8.1, 0.8 Hz, 1H), 7.69 (ddd, *J* = 8.4, 6.9, 1.4 Hz, 1H), 7.49 (d, *J* = 8.4, 6.9, 1.4 Hz, 1H), 7.22 (d, *J* = 8.4 Hz, 1H), 6.78 (d, *J* = 1.0 Hz, 1H), 6.75 (s, 1H), 3.84 (s, 1H), 3.79 (s, 1H), 3.27 (dd, *J* = 9.6, 6.2 Hz, 1H), 3.10 (dd, *J* = 9.5, 6.4 Hz, 1H). ¹³C NMR (400 MHz, CDCl3): δ 161.84, 148.83, 148.01, 147.34, 136.16, 134.17, 129.40,

128.86, 127.52, 126.60, 125.79, 121.62, 120.34, 112.01, 111.34, 55.92, 55.77, 41.23, 35.54.

3. Synthesis of 3-substituted isoquinolines

3-substituted isoquinolines were synthesized according to literature with modification:^[2]



A round-bottom flask was charged with benzylamine (10 mmol) and glyoxal 1,1-dimethyl acetal (1.1 eq., 60% soluition in water) was added dropwise. The mixture was stirred at

room temperature overnight and two layers were separated. After removal of the aqueous layer, trace water was removed by azeotropic distillation with toluene. The crude imine was used for next step without purification.

To the solution of this imine in dry THF was added Grignard reagent dropwise at 0°C. The mixture was then stirred at room temperature. Once TLC showed the completion of the reaction (usually overnight), saturated aqueous NH₄Cl was added at 0°C. The mixture was extracted with ether and the organic layer was dried with anhydrous sodium sulfate. After removal of solvent, the crude product was obtained as yellow or orange oil. After ¹H NMR shows the majority of the desired amine in the crude product, it is used for next step without purification.

A round-bottom flask was filled with nitrogen and charged with chlorosulfuric acid (5ml). The mixture was cooled at 0°C and the crude amine was added dropwise. The mixture was heated at 100°C for 15 min and then cooled to 0°C. The resulting mixture was poured on ice and the neutralized carefully with 40% NaOH solution. After the mixture turned to alkaline, ether was added to extract. The organic phase was dried with sodium sulfate and the solvent was removed under reduced pressure. The residue was purified by flash chromatography (on silica gel, eluent: hexane/ethyl acetate). 3-alkylisoquinoline was obtained as yellow oils.

3-ethylisoquinoline

Yellow oil. 43% yield for 3 steps. ¹H NMR (400 MHz, CDCl3): δ 9.06 (s, 1H), 7.75 (d, *J* = 8.1 Hz, 1H), 7.58 (d, *J* = 8.2 Hz, 1H), 7.47 (t, *J* = 6.8 Hz, 1H), 7.35 (t, *J* = 6.8 Hz, 1H), 7.31 (s, 1H), 2.84 (q, *J* = 7.5 Hz, 2H), 1.27 (t, *J* = 7.5 Hz, H). ¹³C NMR (400 MHz, CDCl3): δ 155.92, 150.95, 135.55, 129.08, 126.37, 126.02, 125.16, 125.02, 115.98, 30.06, 13.02. *m/z* (ESI–MS) 158.04 [M + H]⁺.

3-butylisoquinoline

Yellow oil. 38% yield for 3 steps. ¹H NMR (400 MHz, CDCl3): δ 9.19 (s, 1H), 7.90 (d, *J* = 8.1 Hz, 1H), 7.72 (d, *J* = 8.2 Hz, 1H), 7.62 (t, *J* = 7.5 Hz, 1H), 7.50 (t, *J* = 7.5 Hz, 1H), 7.45 (s, 1H), 2.94 (d, *J* = 8.1 Hz, 2H), 1.80 (m, 2H), 1.43 (m, 2H), 0.96 (t, *J* = 7.3 Hz, 3H).

¹³C NMR (400 MHz, CDCl3): δ 155.88, 152.04, 136.54, 130.14, 127.46, 127.07, 126.22, 126.06, 117.87, 37.84, 32.14, 22.50, 13.97.

m/z (ESI–MS) 186.06 [M + H]⁺.

3,6-dimethylisoquinoline

White solid. 42% yield for 3 steps. ¹H NMR (400 MHz, CDCl3): δ 9.10 (s, 1H), 7.81 (d, *J* = 8.4 Hz, 1H), 7.48 (s, 1H), 7.38 (s, 1H), 7.34 (dd, *J* = 8.4, 1.4 Hz, 1H), 2.38 (s, 3H), 2.52 (s, 3H).

¹³C NMR (400 MHz, CDCl3): δ 151.67, 151.51, 140.56, 136.92, 128.56, 127.28, 125.37, 124.80, 117.93, 24.18, 22.05.

MP: 76-78°C

m/*z* (ESI–MS) 158.14 [M + H]⁺.

6-chloro-3-methylisoquinoline

Yellow oil. 56% yield for 3 steps. ¹H NMR (400 MHz, CDCl3): δ 9.14 (s, 1H), 7.85 (d, *J* = 8.7 Hz, 1H), 7.69 (d, *J* = 1.8 Hz, 1H), 7.45 (dd, *J* = 8.7, 2.0 Hz, 1H), 7.38 (s, 1H), 2.69 (s, 3H).

¹³C NMR (400 MHz, CDCl3): δ 152.99, 151.72, 137.26, 129.15, 127.41, 125.04, 124.80, 117.53, 24.25.

m/*z* (ESI–MS) 178.14 [M + H]⁺.

6-fluoro-3-methylisoquinoline

Yellow oil. 34% yield for 3 steps. ¹H NMR (400 MHz, CDCl3): δ 9.12 (s, 1H), 7.91 (dd, *J* = 8.9, 5.6 Hz, 1H), 7.41 (s, 1H), 7.30 (dd, *J* = 10.1, 2.0 Hz, 1H), 7.25 (dd, *J* = 8.7, 2.5 Hz, 1H), 2.68 (s, 3H).

¹³C NMR (400 MHz, CDCl3): δ 163.28 (d, *J* = 251.9 Hz), 152.66, 151.52, 137.94 (d, *J* = 10.5 Hz), 130.43 (d, *J* = 9.9 Hz), 124.04, 118.01 (d, *J* = 5.3 Hz), 116.81 (d, *J* = 25.8 Hz), 109.14 (d, *J* = 20.9 Hz). *m*/z (ESI–MS) 162.04 [M + H]⁺.

6-trifluoromethyl-3-methylisoquinoline

Yellow oil. 27% yield for 3 steps. ¹H NMR (400 MHz, CDCl3): δ 9.26 (s, 1H), 8.05 (s, 1H), 8.03 (s, 2H), 7.68 (dd, *J* = 8.4, 1.6 Hz, 1H), 7.56 (s, 1H), 2.74 (s, 3H). ¹³C NMR (400 MHz, CDCl3): δ 153.40, 151.92, 144.38, 135.56, 131.90 (q, *J* = 128.4 Hz), 128.69, 127.47, 123.79 (q, *J* = 18.1 Hz), 121.96 (q, *J* = 12.1 Hz), 118.98, 24.20. *m/z* (ESI–MS) 211.97 [M + H]⁺.

8-chloro-3-methylisoquinoline

Yellow oil. 41% yield for 3 steps. ¹H NMR (400 MHz, CDCl₃) δ 9.56 (s, 1H), 7.64 (dd, *J* = 5.5, 3.7 Hz, 1H), 7.54 (s, 1H), 7.53 (d, *J* = 2.3 Hz, 1H), 7.47 (s, 1H), 2.72 (s, 3H). ¹³C NMR (400 MHz, CDCl3): δ 152.79, 148.94, 137.89, 132.52, 130.19, 126.52, 125.14, 124.01, 118.15, 24.08.

m/*z* (ESI–MS) 178.16 [M + H]⁺.

4. General procedure for preparation of quinoline/isoquinoline hydrochlorides

Quinolines or isoquinolines (1mmol) was dissolved in 2 ml dry THF, 1 ml hydrogen chloride solution (2M) in ether (2 eq.) was added dropwse. White precipitate was form. The resulting mixture was filtered and the solid was washed with dry THF (2 ml*2). The white solid was collected and dried under vacuum. Quinolimium/isoquinolinium chloride was obtained as white solid.

5. General procedure for asymmetric hydrogenation of quinolium/isoquinolinium chlorides

In the nitrogen-filled glovebox, solution of [Rh(COD)CI]₂ (4.9 mg, 0.01 mmol) and ligand (2.1 eq.) in 5.0 ml anhydrous solvent was stirred at room temperature for 30 min. A specified volume of the resulting solution (0.5 ml, 1% Rh catalyst) was transferred) by syringe to a Score-Break ampule charged with substrate solution (0.2 mmol in 0.5 ml). The ampule was placed into an autoclave, which was then charged with 40 atm H₂. The autoclave was stirred at desired temperature for the indicated period of time. After release of H₂, the resulting mixture was concentrated under vacuum. Saturated potassium carbonate solution and dichloromethane was added and the mixture was stirred for 30 min. The organic layer was dried with anhydrous sodium sulfate. After removal of solvent, the crude product was analysed by ¹H NMR to determine the conversion. The enantiomeric excess was determined by GC or HPLC analysis of the crude product or its corresponding trifluoroacetamides. The absolute configurations were assigned according to literature^[3,4,5] and their analogues.

6. Counterion effects in asymmetric hydrogenation of isoquinolines

When tetrabutylammonium chloride (TBAC) was added, no significant changes in conversion and enantioselectivity was observed (Table S1, entry 2 vs 1). This suggests the spectator role of tetrabutylammonium cation. The introduction of bromide anion from TBAB does not influence this catalytic reaction (Table S1, entry 3 vs 2). The presence of lodide anion decreases the conversion, but the it shows trace influence on the enantioselectivity (Table S1, entry 4 vs 2). Fluoride anion, however, inhibits this catalytic reaction dramatically: no product was observed after adding TBAF (Table S1, entry 5). To gain plausible explanation of these observations, further study will be needed in the future.

Table S1, Optimization of condition. ^a



Entry	additive	conversion ^b	eec
1	none	99%	99%
2	TBAC (1.0 eq.)	98%	99%
3	TBAB (1.0 eq.)	99%	99%
4	TBAI (1.0 eq.)	90%	98%
5	TBAF (1.0 eq.)	0	-

^aReaction condition: 1a (0.1 mmol) in 0.6 ml solvent, 1/[Rh(COD)Cl]2/**L1** ratio=100/0.5/1, 40 atm H₂, 25 °C, 48 h; ^bconversion was determined by 1H NMR analysis, no by product was observed; ^cee was determined by GC with a chiral stationary phase.

7. Result of deuterium labeling experiments

Following standard hydrogenation procedure, deuterium labeling experiments were conducted with specific modification.









For clarity, combined spectrum for the original 2a and the two deuterated 2a were shown as below:



8. Characterization data of chiral THQs and THIQs

3-methyl-1,2,3,4-tetrahydroisoquinoline (2a)

Yellow oil. ¹H NMR (400 MHz, CDCl3): δ 7.11 (m, 2H), 7.06 (m, 1H), 7.01 (t, 1H), 4.11 (d, *J* =16.0 Hz, 1H), 4.02 (d, *J* =16.0 Hz, 1H), 3.02 (dqd, *J* = 10.3, 6.3, 4.0 Hz, 1H), 2.78

(dd, *J* = 16.4, 3.8 Hz, 1H), 2.51 (dd, *J* = 16.3, 10.7 Hz, 1H), 1.73 (br, 1H), 1.24 (d, *J* = 6.3 Hz, 3H).

¹³C NMR (400 MHz, CDCl3): δ 135.39, 134.92, 129.11, 126.01, 126.00, 125.70, 49.25, 48.59, 37.26, 22.50.

[α]²²_D +103.6 (*c* 0.5, CHCl₃).

0 - 30

35

40

m/z (ESI–MS) 148.22 [M + H]⁺.

Supelco gama Dex 225 column (30 m ×0.25 mm × 0.25 μ m), He 1.0 mL/min, column 100 °C, t₁ =46.3 min, t₂ =47.7 min.



45

50

55

min

Signal 1: FID1 B, Back Signal

Peak RetTime Type Width Area Height Area [min] [pA*s] [pA] 2 # [min] ----| 1 47.642 BB 0.4541 1052.56445 32.00426 1.000e2 Totals : 1052.56445 32.00426 _____ _____

*** End of Report ***

3-ethyl-1,2,3,4-tetrahydroisoquinoline (2b)

Yellow oil. ¹H NMR (400 MHz, CDCl3): δ 7.12 (m, 3H), 7.02 (m, 1H), 4.09 (d, *J* = 12.0 Hz, 1H), 4.03 (d, *J* = 12.0 Hz, 2H), 2.79 (tdd, *J* = 10.4, 6.8, 3.8 Hz, 1H), 2.50 (dd, *J* = 17.0, 11.5 Hz, 1H), 1.74 (br, 1H), 1.64-1.47 (m, 2H), 1.02 (t, 3H).

¹³C NMR (400 MHz, CDCl3): δ 135.76, 134.92, 129.46, 126.01, 125.99, 125.65, 55.25, 48.62, 35.11, 29.61, 10.39.

[α]²²_D +97.3 (*c* 0.5, CHCl₃).

m/*z* (ESI–MS) 162.09 [M + H]⁺.

Supelco gama Dex 225 column (30 m ×0.25 mm × 0.25 μ m), He 1.0 mL/min, column 100 °C, t₁ =83.2 min, t₂ =85.3 min.





Signal 1: FID1 B, Back Signal

Peak #	RetTime [min]	Туре	Width [min]	Area [pA*s]	Height [pA]	Area %
 1	85.317	BB	0.6529	1595.54932	33.04603	1.000e2
Total	s :			1595.54932	33.04603	

*** End of Report ***

3-butyl-1,2,3,4-tetrahydroisoquinoline (2c)

Yellow oil. ¹H NMR (400 MHz, CDCl3): δ 7.11 (m, 3H), 7.02 (m, 1H), 4.08 (d, *J* =16.0 Hz, 2H), 4.03 (d, *J* =16.0 Hz, 2H), 2.82 (comp., 2H), 2.50 (dd, *J* = 16.1, 10.4 Hz, 1H), 1.50 (br, 1H), 1.30-1.60 (m, 6H), 1.02 (t, *J* = 3.2 Hz, 3H).

¹³C NMR (400 MHz, CDCl3): δ 135.78, 134.97, 129.22, 125.99, 125.66, 57.72, 48.60, 36.60, 35.58, 28.19, 22.86, 14.06.

[α]²²_D +81.8 (*c* 0.5, CHCl₃).

m/z (ESI–MS) 190.17 [M + H]⁺.

Supelco gama Dex 225 column (30 m ×0.25 mm × 0.25 μ m), He 1.0 mL/min, column 100 °C, t₁ =278.3 min, t₂ =285.0 min.



2.4380 1928.14819 13.18125 95.81793

2012.30419 13.83672

*** End of Report ***

3,6-dimethyl-1,2,3,4-tetrahydroisoquinoline (2d)

2 284.892 MM

Totals :

Yellow oil. ¹H NMR (400 MHz, CDCl3): δ 7.00 – 6.77 (m, 3H), 4.06 (d, *J* = 15.7 Hz, 1H), 3.99 (d, *J* = 15.8 Hz, 1H), 3.05–2.94 (m, 1H), 2.73 (dd, *J* = 16.3, 3.4 Hz, 1H), 2.46 (dd, *J* = 16.0, 10.8 Hz, 1H), 2.29 (s, 3H), 1.69 (s, 1H), 1.23 (d, *J* = 6.3 Hz, 3H).

¹³C NMR (400 MHz, CDCl3): δ 135.45, 134.76, 132.38, 129.60, 126.65, 125.88, 49.28, 48.33, 37.27, 22.49, 20.98.

[α]²²_D +87.4 (c 0.5, CHCl₃).

m/*z* (ESI–MS) 162.08 [M + H]⁺.

Supelco Chiral Select 1000 column (30 m ×0.25 mm × 0.25 μ m) for its corresponding trifluoroacetamide, He 1.0 mL/min, column 120 °C, t₁ =91.4 min, t₂ =93.0 min.





Signal 1: FID1 A, Front Signal

Peak RetTime Type Width Area Height Area [pA*s] 00 # [min] [min] [pA] 91.433 MM 1.0499 1650.89038 26.20647 98.03240 1 2 93.013 MM 0.7184 33.13491 7.68718e-1 1.96760 1684.02529 26.97519 Totals :

----- *** End of Report ***

6-trifluoromethyl-3-methyl-1,2,3,4-tetrahydroisoquinoline (2e)

Yellow oil. ¹H NMR (400 MHz, CDCl3): δ 7.36 (d, *J* = 8.1 Hz, 1H), 7.32 (s, 1H), 7.12 (d, *J* = 8.0 Hz, 1H), 3.14 (d, *J* = 16.4 Hz, 1H), 3.09 (d, *J* = 16.4 Hz, 1H), 2.83 (dd, *J* = 16.5, 3.7 Hz, 1H), 2.54 (dd, *J* = 16.4, 10.6 Hz, 1H), 1.26 (d, *J* = 6.3 Hz, 3H).

¹³C NMR (400 MHz, CDCl3): δ 139.32, 135.72, 128.46 (q, J = 127.6 Hz), 126.46, 125.96 (q, J = 15.2 Hz), 122.97, 122.48 (q, J = 7.0 Hz), 49.03, 48.44, 37.13, 22.27.

 $[\alpha]^{22}_{D}$ +80.2 (*c* 0.5, CHCl₃).

m/*z* (ESI–MS) 216.01 [M + H]⁺.

Supelco gama Dex 225 column (30 m ×0.25 mm × 0.25 μ m), He 1.0 mL/min, column 100 °C, t₁ =60.8 min, t₂ =64.7 min.





#	[min]	[min]	[pA*s]	[pA]	
	63.849 BB	0.8115	897.82019	14.13314	1.000e2
Total	s :		897.82019	14.13314	

*** End of Report ***

6-chloro-3-methyl-1,2,3,4-tetrahydroisoquinoline (2f)

Yellow oil. ¹H NMR (400 MHz, CDCl3): δ 7.07 (t, 2H), 6.94 (d, *J* = 8.1 Hz, 1H), 4.05 (d, *J* = 16.0 Hz, 1H), 3.99 (d, *J* = 16.0 Hz, 1H), 2.99 (dqd, *J* = 12.6, 6.3, 4.0 Hz, 1H), 2.74 (dd, *J* = 16.5, 3.7 Hz, 1H), 2.48 (dd, *J* = 16.5, 10.6 Hz, 1H), 1.71 (br, 1H), 1.24 (d, *J* = 6.3 Hz, 3H).

¹³C NMR (400 MHz, CDCl3): δ 136.86, 133.77, 131.48, 128.83, 127.33, 125.87, 48.92, 48.09, 37.04, 29.29.

[α]²²_D +72.1 (c 0.5, CHCl₃).

m/*z* (ESI–MS) 182.15, 184.02 [M + H]⁺.

Supelco gama Dex 225 column (30 m ×0.25 mm × 0.25 μ m), He 1.0 mL/min, column 120 °C, t₁ =67.5 min, t₂ =69.4 min.







*** End of Report ***

6-fluoro-3-methyl-1,2,3,4-tetrahydroisoquinoline (2g)

Yellow oil. ¹H NMR (400 MHz, CDCl3): δ 6.96 (dd, *J* = 8.3, 5.8 Hz, 1H), 6.81 (td, *J* = 8.5, 2.6 Hz, 1H), 6.76 (dd, *J* = 9.6, 2.5 Hz, 1H), 4.05 (d, *J* = 15.6 Hz, 2H), 4.00 (d, *J* = 15.6 Hz, 2H), 2.99 (dqd, *J* = 10.3, 6.3, 4.0 Hz, 1H), 2.75 (dd, *J* = 16.5, 3.8 Hz, 1H), 2.49 (dd, *J* = 16.5, 10.7 Hz, 1H), 1.24 (d, *J* = 6.3 Hz, 3H).

¹³C NMR (400 MHz, CDCl3): δ 161.18 (d, *J* = 964.4 Hz), 137.00 (d *J* = 29.4 Hz), 130.90 (d, *J* = 11.6 Hz), 127.35 (d, *J* = 32.4 Hz), 115.34 (d, *J* = 81.2 Hz), 112.79 (d, *J* = 86.0 Hz), 48.93, 48.05, 37.32, 22.30.

¹⁹F NMR (500 MHz, CDCl3): δ -117.42.

 $[\alpha]^{22}_{D}$ +82.1 (c 0.5, CHCl₃).

m/*z* (ESI–MS) 166.24 [M + H]⁺.

Supelco gama Dex 225 column (30 m ×0.25 mm × 0.25 μ m), He 1.0 mL/min, column 100 °C, t₁ =58.0 min, t₂ =60.0 min.





Signal 1: FID1 B, Back Signal

Peak #	RetTime [min]	Туре	Width [min]	Area [pA*s]	Height [pA]	Area %	
	59.675	· BB	0.8510	1079.47339	16.33518	1.000e2	1
Total	ls :			1079.47339	16.33518		

*** End of Report ***

8-chloro-3-methyl-1,2,3,4-tetrahydroisoquinoline (2h)

Yellow oil. ¹H NMR (400 MHz, CDCl3): δ 7.16 (d, *J* = 7.8 Hz, 1H), 7.06 (t, *J* = 7.7 Hz, 1H), 6.97 (d, *J* = 7.5 Hz, 1H), 4.20 (d, *J* = 16.9 Hz, 1H), 3.94 (d, *J* = 16.9 Hz, 1H), 2.97 (dqd, *J* = 10.1, 6.3, 3.8 Hz, 1H), 2.77 (dd, *J* = 16.3, 3.5 Hz, 1H), 2.50 (dd, *J* = 16.3, 10.6 Hz, 1H), 1.67 (s, 1H), 1.24 (d, *J* = 6.3 Hz, 3H).

¹³C NMR (400 MHz, CDCl3): δ 137.42, 133.23, 132.21, 127.52, 126.84, 126.52, 48.57, 46.79, 37.36, 22.19.

[α]²²_D +117.7 (c 0.5, CHCl₃).

m/*z* (ESI–MS) 182.09, 184.01 [M + H]⁺.

Supelco gama Dex 225 column (30 m ×0.25 mm × 0.25 μ m), He 1.0 mL/min, column 100 °C, t₁ =158.2 min, t₂ =160.4 min.





*** End of Report ***

1-methyl-1,2,3,4-tetrahydroisoquinoline (2i)

Yellow oil. ¹H NMR (400 MHz, CDCl₃) δ7.22 – 7.03 (m, 4H), 4.14 (d, *J* = 6.8 Hz, 1H), 3.31 (M, 1H), 3.03 (m, 1H), 2.92 (m, 1H), 2.82 – 2.60 (m, 1H), 2.25 (br, 1H), 1.47 (d, *J* = 5.6 Hz, 3H).

¹³C NMR (400 MHz, CDCl3): δ 140.32, 134.68, 129.20, 125.99, 129.90 (overlap), 51.57, 41.71, 29.91, 22.63.

[α]²²_D -77.0 (*c* 0.5, CHCl₃).

Supelco Chiral Select 1000 column (30 m ×0.25 mm × 0.25 μ m) for its corresponding trifluoroacetamide, He 1.0 mL/min, column 120 °C, t₁ =56.8 min, t₂ =58.3 min.



*** End of Report ***

3-phenyl-1,2,3,4-tetrahydroisoquinoline (2j)

Yellow oil. ¹H NMR (400 MHz, CDCl₃) δ 7.43 (d, *J* = 7.9 Hz, 2H), 7.36 (dd, *J* = 8.0, 6.5 Hz, 2H), 7.28 (t, *J* = 7.2 Hz, 1H), 7.18-7.05 (m, 4H), 4.26 (d, *J* = 15.6 Hz, 1H), 4.16 (d, *J* = 15.6 Hz, 1H), 4.00 (t, *J* = 7.5 Hz, 1H), 2.97 (d, *J* = 7.3 Hz, 2H), 1.93 (br, 1H).

¹³C NMR (400 MHz, CDCl3): δ 144.33, 135.04, 134.92, 129.10, 128.63, 127.39, 126.55, 126.26, 126.18, 125.90, 58.61, 49.27, 37.73.

Daicel Chiralpak OD-H for its corresponding trifluoroacetamide, hexanes/i-PrOH = 98/2, Flow rate = 1.0 ml/min, UV = 220 nm, t_1 = 10.4 min, t_2 = 13.1 min.

 $[\alpha]^{22}_{D}$ +100.3 (c 0.5, CHCl₃) for (R) isomer^[7].



Signal 1: VWD1 A, Wavelength=220 nm

Peak RetTime Type Width Area Height Area 8 # [min] [min] mAU *s MAU 1 ----|-----|-----|------| 10.431 MM 0.5781 6872.61035 198.15254 12.1247 1 2 13.172 MM 0.8965 4.98101e4 925.97284 87.8753 Totals : 5.66827e4 1124.12538

2-methyl-1,2,3,4-tetrahydroquinoline (4a)

Yellow oil. ¹H NMR (400 MHz, CDCl3): δ 6.98 – 6.92 (m, 2H), 6.59 (td, *J* = 7.4, 1.1 Hz, 1H), 6.45 (dd, *J* = 5.2, 3.2 Hz, 1H), 3.67 (b, 1H), 3.43 – 3.35 (m, 1H), 2.83 (ddd, *J* = 16.9, 11.5, 5.6 Hz, 1H), 2.75 – 2.68 (m, 1H), 1.95 – 1.88 (m, 1H), 1.58 (dddd, *J* = 12.8, 11.6, 10.0, 5.4 Hz, 1H), 1.20 (d, *J* = 6.3 Hz, 3H).

¹³C NMR (400 MHz, CDCl3): δ 143.73, 128.22, 125.65, 120.09, 115.97, 112.98, 46.15, 29.12, 25.55, 21.56.

 $[\alpha]^{22}_{D}$ -44.3 (*c* 0.5, CHCl₃).

Supelco gama Dex 225 column (30 m ×0.25 mm × 0.25 μ m), He 1.0 mL/min, column 120 °C, t₁ =30.5 min, t₂ =31.3 min.





*** End of Report ***

2-ethyl-1,2,3,4-tetrahydroquinoline (4b)

Yellow oil. ¹H NMR (400 MHz, CDCl3): δ 6.95 (t, J = 7.2 Hz, 2H), 6.59 (t, J = 7.3 Hz, 1H), 6.47 (d, J = 8.1 Hz, 1H), 3.76 (br, 1H), 3.20 – 3.12 (m, 1H), 2.81 (ddd, J = 16.4, 11.0, 5.5 Hz, 1H), 2.76 – 2.68 (m, 1H), 2.00 – 1.92 (m, 1H), 1.65 – 1.55 (m, 1H), 1.51 (dd, J = 14.3, 7.2 Hz, 2H), 0.98 (t, J = 7.5 Hz, 3H).

¹³C NMR (400 MHz, CDCl3): δ 143.72, 128.18, 125.66, 120.36, 115.84, 112.96, 52.03, 28.73, 26.57, 25.38, 8.98.

 $[\alpha]^{22}_{D}$ -74.4 (*c* 0.5, CHCl₃).

Supelco gama Dex 225 column (30 m ×0.25 mm × 0.25 μ m), He 1.0 mL/min, column 120 °C, t₁ =51.3 min, t₂ =53.7 min.



2-isopropyl-1,2,3,4-tetrahydroquinoline (4c)

Yellow oil. ¹H NMR (400 MHz, CDCl3): δ 6.95 (t, *J* = 7.7 Hz, 2H), 6.58 (t, *J* = 7.3 Hz, 1H), 6.47 (d, *J* = 8.0 Hz, 1H), 3.75 (s, 1H), 3.05 – 3.00 (m, 1H), 2.80 (ddd, *J* = 14.0, 9.9,

4.2 Hz, 1H), 2.76 – 2.68 (m, 1H), 1.91 (ddt, J = 5.9, 4.5, 3.2 Hz, 1H), 1.76 – 1.58 (m, 2iH), 0.98 (dd, J = 10.2, 6.8 Hz, 6H). δ 6.95 (t, 2H), 6.58 (t, 1H), 6.47 (d, 1H), 3.03 (m, 1H), 2.75 (m, 2H), 1.91 (m, 1H), 1.69 (m, 2H), 0.98 (dd, 6H).

¹³C NMR (400 MHz, CDCl3): δ 144.00, 128.08, 125.66, 120.40, 115.70, 112.93, 56.29, 31.51, 25.61, 23.53, 17.38 (d).

[α]²²_D -44.0 (*c* 0.5, CHCl₃).

Supelco gama Dex 225 column (30 m ×0.25 mm × 0.25 μ m), He 1.0 mL/min, column 140 °C, t₁ =27.9 min, t₂ =29.4 min.





Signal 1: FID1 A, Front Signal

Peak RetTime Type W # [min]	Width Area [min] [pA*s]	Height . [pA]	Area %
1 27.913 BB (0.1944 578.82666	45.13346 1.	 000e2
Totals :	578.82666	45.13346	
	======================================		

2-pentyl-1,2,3,4-tetrahydroquinoline (4d)

Yellow oil. ¹H NMR (400 MHz, CDCl3): δ 6.95 (t, *J* = 7.2 Hz, 2H), 6.58 (t, *J* = 7.4, 1H), 6.46 (d, *J* = 7.5 Hz, 1H), 3.75 (br, 1H), 3.22 (dtd, *J* = 9.4, 6.3, 2.9 Hz, 1H), 2.86 – 2.75 (m, 1H), 2.71 (dt, *J* = 16.3, 4.8 Hz, 1H), 1.99 – 1.91 (m, 1H), 1.64 – 1.53 (m, 1H), 1.47 (t, *J* = 6.7 Hz, 2H), 1.42 – 1.27 (m, 6H), 0.90 (t, *J* = 6.9 Hz, 3H).

¹³C NMR (400 MHz, CDCl3): δ 143.71, 128.19, 125.65, 120.36, 115.84, 112.99, 50.58, 35.66, 30.93, 27.11, 25.40, 24.36, 21.59, 12.98 (d).

[α]²²_D -78.2 (*c* 0.5, CHCl₃).

Daicel Chiralpak OD-H, hexanes/i-PrOH = 99/1, Flow rate = 1.0 ml/min, UV = 254 nm, t_1 = 6.9 min, t_2 = 8.0 min.





Signal 1: VWD1 A, Wavelength=254 nm

Peak RetTime Type Width Area Height Area # [min] [min] mAU *s [mAU 8] 6.904 VV 0.2857 1660.45337 79.54791 2.6200 1 2 8.017 VB 0.3809 6.17146e4 2262.38330 97.3800 Totals : 6.33750e4 2341.93121

2-undecyl-1,2,3,4-tetrahydroquinoline (4e0

Yellow oil. ¹H NMR (400 MHz, CDCl3): δ 6.95 (t, *J* = 7.5 Hz, 2H), 6.58 (td, *J* = 7.4, 1.1 Hz, 1H), 6.47 (dd, *J* = 9.0, 8.2 Hz, 1H), 3.74 (br, 1H), 3.27 – 3.18 (m, 1H), 2.80 (ddd, *J* = 16.4, 10.9, 5.5 Hz, 1H), 2.71 (dt, *J* = 16.3, 4.7 Hz, 1H), 1.95 (ddt, *J* = 6.9, 5.5, 4.0 Hz, 1H), 1.65 – 1.53 (m, 1H), 1.43 – 1.51 (m, 2H), 1.34 (dd, *J* = 45.4, 7.5 Hz, 18H), 0.88 (t, *J* = 7.2 Hz, 3H).

¹³C NMR (400 MHz, CDCl3): δ 143.71, 128.19, 125.65, 120.35, 115.84, 112.99, 50.59, 35.71, 30.88, 28.74, 28.59, 28.30, 27.12, 25.41, 24.70, 21.65, 13.05 (d).

Daicel Chiralpak OD-H, hexanes/i-PrOH = 98/2, Flow rate = 1.0 ml/min, UV = 254 nm, t_1 = 5.2 min, t_2 = 6.2 min.

 $[\alpha]^{22}_{D}$ -55.1 (*c* 0.5, CHCl₃).



Signal 1: VWD1 A, Wavelength=254 nm

Peak RetTime Type Width Area Height Area # olo [min] [min] mAU *s [mAU] ----|-----|-----|-----|------| 5.243 MM 0.2518 1181.22571 78.17448 2.4676 1 2 6.241 MM 0.3466 4.66881e4 2244.74536 97.5324 4.78693e4 2322.91985 Totals :

2,6-dimethyl-1,2,3,4-tetrahydroquinoline (4f)

Yellow oil. ¹H NMR (400 MHz, CDCl3): 6.77 (d, *J* = 8.4 Hz, 2H), 6.39 (d, *J* = 8.4 Hz, 1H), 3.35 (dqd, *J* = 12.5, 6.3, 2.8 Hz, 1H), 2.80 (ddd, *J* = 17.1, 11.5, 5.8 Hz, 1H), 2.68 (ddd, *J* = 16.4, 5.3, 3.5 Hz, 1H), 2.19 (s, 3H), 1.94 – 1.86 (m, 1H), 1.56 (dddd, *J* = 12.8, 11.5, 10.0, 5.4 Hz, 1H), 1.18 (d, *J* = 6.3 Hz, 3H).

¹³C NMR (400 MHz, CDCl3): δ 142.46, 129.83, 127.23, 126.26, 121.24, 114.27, 47.35, 30.39, 26.59, 22.59, 20.40.

[α]²²_D -44.8 (*c* 0.5, CHCl₃).

Supelco gama Dex 225 column (30 m ×0.25 mm × 0.25 μ m), He 1.0 mL/min, column 120 °C, t₁ =47.8 min, t₂ =48.8 min.





Signal 1: FID1 A, Front Signal

Peak #	RetTime [min]	Туре	Width [min]	Area [pA*s]	Height [pA]	Area %		
1	47.609	 MM	0.4073	284.61505	11.64730	1.000e2	1	
Tota	ls :			284.61505	11.64730			

*** End of Report ***

6-methoxy-2-methyl-1,2,3,4-tetrahydroquinoline (4g)

Yellow oil. ¹H NMR (400 MHz, CDCl3): δ 6.58 (m, 2H), 6.44 (d, *J* = 8.3 Hz, 1H), 3.72 (s, 3H), 3.37 – 3.27 (m, 1H), 3.17 (br, 1H), 2.83 (ddd, *J* = 17.3, 11.6, 5.9 Hz, 1H), 2.70 (ddd, *J* = 16.6, 5.4, 3.2 Hz, 1H), 1.90 (ddt, *J* = 12.6, 5.9, 2.9 Hz, 1H), 1.56 (dddd, *J* = 12.8, 11.6, 10.2, 5.5 Hz, 1H), 1.19 (d, *J* = 6.3 Hz, 3H).

¹³C NMR (400 MHz, CDCl3): δ 151.92, 138.93, 122.53, 115.33, 114.72, 112.91, 55.83, 47.50, 30.34, 26.91, 22.54.

 $[\alpha]^{22}_{D}$ -34.7 (*c* 0.5, CHCl₃).

Supelco gama Dex 225 column (30 m ×0.25 mm × 0.25 μ m), He 1.0 mL/min, column 120 °C, t₁ =121.3 min, t₂ =124.5 min.





6-chloro-2-methyl-1,2,3,4-tetrahydroquinoline (4h)

Yellow oil. ¹H NMR (400 MHz, CDCl3): δ 6.93 – 6.86 (m, 2H), 6.37 (d, *J* = 8.4 Hz, 1H), 3.68 (br, 1H), 3.41 – 3.32 (m, 1H), 2.84 – 2.72 (m, 1H), 2.72 – 2.63 (m, 1H), 1.95 – 1.87 (m, 1H), 1.54 (dddd, *J* = 12.9, 11.4, 9.9, 5.4 Hz, 1H), 1.20 (t, *J* = 5.2 Hz, 3H).

¹³C NMR (400 MHz, CDCl3): δ 142.29, 127.78, 125.45, 121.56, 120.25, 113.90, 46.12, 28.67, 25.41, 21.41.

[α]²²_D -81.4 (*c* 0.5, CHCl₃).

Supelco gama Dex 225 column (30 m ×0.25 mm × 0.25 μ m), He 1.0 mL/min, column 140 °C, t₁ =55.9 min, t₂ =58.0 min.



Totals :

2377.53272 37.81902

*** End of Report ***

6-fluoro-2-methyl-1,2,3,4-tetrahydroquinoline (4i)

Yellow oil. ¹H NMR (400 MHz, CDCl3): δ 6.64 – 6.70 (m, 5.8 Hz, 2H), 6.42 – 6.35 (m, 1H), 3.57 (br, 1H), 3.34 (dqd, *J* = 12.6, 6.2, 2.7 Hz, 1H), 2.81 (ddd, *J* = 11.6, 8.7, 5.6 Hz,

1H), 2.69 (ddd, *J* = 16.6, 5.3, 3.4 Hz, 1H), 1.91 (dtd, *J* = 8.8, 5.9, 3.0 Hz, 1H), 1.55 (dddd, *J* = 12.9, 11.6, 10.1, 5.5 Hz, 1H), 1.20 (d, *J* = 6.3 Hz, 3H).

¹³C NMR (400 MHz, CDCl3): δ 154.51 (d, *J* = 938.4 Hz), 139.95, 121.44 (d, *J* = 26.4 Hz), 114.34 (d, *J* = 86.4 Hz), 113.68 (d, *J* = 30.4 Hz), 112.12 (d, *J* = 89.6 Hz), 46.29, 28.87, 25.67, 21.44.

[α]²²_D –63.84 (*c* 0.5, CHCl₃).

Supelco gama Dex 225 column (30 m ×0.25 mm × 0.25 μ m), He 1.0 mL/min, column 120 °C, t₁ =39.0 min, t₂ =40.7 min.




Signal 1: FID1 A, Front Signal

Peak #	RetTime [min]	Туре	Width [min]	Area [pA*s]	Height [pA]	Area %
1	38.985	MM	0.4684	676.17346	24.05771	97.66689
2	40.716	MM	0.4012	16.15275	6.71097e-1	2.33311
Total	ls :			692.32621	24.72881	

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6-bromo-2-methyl-1,2,3,4-tetrahydroquinoline (4j)

Yellow oil. ¹H NMR (400 MHz, CDCl3): δ 7.05 (d, *J* = 1.0 Hz, 1H), 7.02 (d, *J* = 8.4 Hz, 1H), 6.32 (d, *J* = 8.4 Hz, 1H), 3.69 (s, 1H), 3.41 – 3.31 (m, 1H), 2.84 – 2.73 (m, 1H), 2.72 – 2.64 (m, 1H), 1.90 (dtd, *J* = 8.4, 5.6, 2.8 Hz, 1H), 1.53 (dtd, *J* = 11.8, 10.7, 5.3 Hz, 2H), 1.19 (d, *J* = 6.4 Hz, 3H).

¹³C NMR (400 MHz, CDCl3): δ 142.73, 130.62, 128.30, 122.08, 114.33, 107.25, 46.06, 28.60, 25.35, 21.40.

 $[\alpha]^{22}_{D}$ –66.7 (c 0.5, CHCl₃).

Supelco gama Dex 225 column (30 m ×0.25 mm × 0.25 μ m), He 1.0 mL/min, column 160 °C, t₁ =36.6 min, t₂ =37.6 min.





7-chloro-2-methyl-1,2,3,4-tetrahydroquinoline (4k)

Yellow oil. ¹H NMR (400 MHz, CDCl3): δ 6.84 (d, *J* = 8.0 Hz, 1H), 6.53 (dd, *J* = 8.0, 2.0 Hz, 1H), 6.42 (d, *J* = 2.0 Hz, 1H), 3.72 (br, 1H), 3.38 (dqd, *J* = 9.3, 6.3, 3.0 Hz, 1H), 2.81 – 2.61 (m, 1H), 1.97 – 1.82 (m, 1H), 1.54 (dddd, *J* = 12.9, 11.1, 9.8, 5.5 Hz, 1H), 1.18 (t, *J* = 10.1 Hz, 3H).

¹³C NMR (400 MHz, CDCl3): δ 144.89, 130.89, 129.11, 118.28, 115.55, 112.24, 45.96, 28.75, 25.01, 21.41.

[α]²²_D -73.3 (*c* 0.5, CHCl₃).

Supelco gama Dex 225 column (30 m ×0.25 mm × 0.25 μ m), He 1.0 mL/min, column 140 °C, t₁ =49.8 min, t₂ =50.7 min.



*** End of Report ***

8-chloro-2-methyl-1,2,3,4-tetrahydroquinoline (4I)

Yellow oil. ¹H NMR (400 MHz, CDCl3): δ 7.05 (d, *J* = 7.9 Hz, 1H), 6.86 (dd, *J* = 7.4, 0.9 Hz, 1H), 6.50 (t, *J* = 7.7 Hz, 1H), 4.25 (br, 1H), 3.46 (dqd, *J* = 9.4, 6.3, 3.1 Hz, 1H), 2.83

(ddd, *J* = 16.6, 11.2, 5.4 Hz, 1H), 2.78 – 2.70 (m, 1H), 1.98 – 1.90 (m, 1H), 1.58 (dddd, *J* = 12.9, 11.2, 9.8, 5.3 Hz, 1H), 1.27 (d, *J* = 6.3 Hz, 3H).

¹³C NMR (400 MHz, CDCl3): δ 140.74, 127.43, 126.74, 122.39, 117.84, 116.35, 47.18, 29.64, 26.74, 22.52.

[α]²²_D -68.7 (*c* 0.5, CHCl₃).

10 -

0 -

46

48

50

Supelco gama Dex 225 column (30 m ×0.25 mm × 0.25 μ m), He 1.0 mL/min, column 120 °C, t₁ =52.9 min, t₂ =54.0 min.



52

54

56

58

min

Signal 1: FID1 A, Front Signal

Peak #	RetTime Typ [min]	e Width [min]	Area [pA*s]	Height [pA]	Area %	
	54.079 BB	0.3756	1081.64124	43.63175	1.000e2	
Total	ls :		1081.64124	43.63175		

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2-(3,4-dimethoxyphenethyl)-1,2,3,4-tetrahydroquinoline (4m)

Yellow oil. ¹H NMR (400 MHz, CDCl₃) δ 6.95 (t, *J* = 7.1 Hz, 2H), 6.83–6.77 (m, 1H), 6.77–6.71 (m, 2H), 6.60 (td, *J* = 7.4, 0.9 Hz, 1H), 6.44 (dd, *J* = 8.4, 0.9 Hz, 1H), 3.87 (s, 3H), 3.85 (s, 3H), 3.77 (br, 1H), 3.30 (dtd, *J* = 9.3, 6.3, 3.0 Hz, 1H), 2.81 (ddd, *J* = 13.1, 9.0, 4.1 Hz, 1H), 2.73–2.64 (m, 2H), 2.03 – 1.95 (m, 1H), 1.81 (ddd, *J* = 10.2, 7.2, 1.2 Hz, 2H), 1.72–1.61 (m, 1H).

¹³C NMR (400 MHz, CDCl3): δ 149.02, 147.37, 144.54, 134.52, 129.27, 126.76, 121.30, 120.17, 117.05, 114.16, 111.74, 111.43, 56.00, 55.90, 51.25, 38.44, 31.86, 28.05, 26.23.
[α]²²_D -60.8 (*c* 0.5, CHCl₃).

Daicel Chiralpak AS-H, hexanes/i-PrOH = 95/5, Flow rate = 0.5 ml/min, UV = 254 nm, t_1 = 24.4 min, t_2 = 27.0 min.



Signal 1: VWD1 A, Wavelength=254 nm

Peak	RetTime	Type	Width	Area He		Heig	ght	Area
#	[min]		[min]	mAU	*s	[mAU]	00
1	24.421	BB	0.7352	2461.	57397	51.	10365	2.3987
2	26.974	BB	1.0158	1.001	59e5	1565.4	46924	97.6013

Totals :

1.02621e5 1616.57289

[1] a) B. Qian, S. Guo, J. Shao, Q. Zhu, L. Yang, C. Xia and H. Huang, *J. Am. Chem. Soc.* 2010, *132*, 3650; b) T. Wang, L. Zhuo, Z. Li, F. Chen, Z. Ding, Y. He, Q. Fan, J. Xiang, Z. Yu, and A. Chan, *J. Am. Chem. Soc.* 2011, *133*, 9878.

[2] M. Fakhfakha, X. Francka, A. Fourneta, R. Hocquemillera, B. Figadère, *Tetrah. Lett.* **2001**, *42*, 3847.

[3] K. Kido and Y. Watanabe, *Chem. Pharm. Bull.* **1987**, 35, 4964-4966.

[4] J. Mangas-Sanchez, E. Busto, V. Gotor-Fernandez and V. Gotor, *Catal. Sci. Technol.* 2012, 2, 1590.

[5] F. Gou, W. Li, X. Zhang and Y. Liang, *Adv. Synth. Catal.* **2010**, 352, 2441.

[6] J. Xie, P. Yan, Q. Zhang, K. Yuan and Q. Zhou, ACS Catal. 2012, 2, 561.

[7] D. García, B. Moreno, T. Soler, F. Foubelo, M. Yus, *Tetrahedron Lett.* 2009, 50, 4710.

9. NMR spectroscopy















































































































































