Myc Phosphorylation in the bHLH-LZ Inhibits Binding to Max and DNA Supplementary Information

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1 Protein constructs

 $\rm MycWT$ bHLH-LZ MHHHHHHENLYFQGSNVKRRTHNVLERQRRNELKRSFFALRDQIPELENNEKAPKVVILK KATAYILSVQAEEQKLISEEDLLRKRREQLKHKLEQLRNS

Myc S373D/T400D bHLH-LZ MHHHHHHENLYFQGSNVKRRTHNVLERQRRNELKRDFFALRDQIPELENNEKAPKVVILK KADAYILSVQAEEQKLISEEDLLRKRREQLKHKLEQLRNS

 $\label{eq:myc} Myc \ S373E/T400E \ bHLH-LZ \\ MHHHHHHENLYFQGSNVKRRTHNVLERQRRNELKREFFALRDQIPELENNEKAPKVVILK \\ KAEAYILSVQAEEQKLISEEDLLRKRREQLKHKLEQLRNS \\$

 $\label{eq:myc} Myc \ S373D \ bHLH-LZ \\ MHHHHHHENLYFQGSNVKRRTHNVLERQRRNELKRDFFALRDQIPELENNEKAPKVVILK \\ KATAYILSVQAEEQKLISEEDLLRKRREQLKHKLEQLRNS \\$

Myc T400D bHLH-LZ MHHHHHHENLYFQGSNVKRRTHNVLERQRRNELKRSFFALRDQIPELENNEKAPKVVILK KADAYILSVQAEEQKLISEEDLLRKRREQLKHKLEQLRNS

Omomyc MHHHHHHENLYFQGSNVKRRTHNVLERQRRNELKRSFFALRDQIPELENNEKAPKVVILK KATAYILSVQAETQKLISEIDLLRKQNEQLKHKLEQLRNS

2 E-box DNA

5'-AGTTGACCACGTGGTCTGGG-3' 5'-CCCAGACCACGTGGTCAACT-3'

3 Supplementary Tables and Figures

phosphorylation site	phospho peptide seq.	exp. mass	theor. mass
pT358	R R T HNVLER	1259.624	1259.647
pS373	$\mathbf{R}_{\mathbf{S}}^{\mathbf{S}}$ FFALR	975.469	974.492
pT400	KA <mark>T</mark> AYILSVQAEEQK	1757.860	1757.883
pY402	KATA <mark>Y</mark> ILSVQAEEQK	1757.860	1757.883
pS405	ATAYIL <mark>S</mark> VQAEEQK	1629.763	1629.788
pS437	LEQLRN <mark>S</mark>	928.422	938.445

Table S1: Phosphorylation sites identified using MS with corresponding peptides including the monoisotopic experimental and theoretical masses. Corresponding MSMS spectra and Mascot (Matrix Science) match are highlighted in Figures S1-S3.



Figure S1: Output of the MSMS search results from Mascot (Matrix Science). Identified peptide sequence The table shows theoretical b and y MSMS ion masses of the corresponding peptide (center column) fragments. MS spectra and match of experimetal and theoretical fragment ions for each peptide are highlighted in red. The matched ions confirm the identity of the peptides containing phosphorylated T358 and S373.



Figure S2: MSMS search results from MASCOT for T400 and Y402 phosphorylation. See Figure S1 for further details.



Figure S3: MSMS search results from MASCOT for S3405 and S437 phosphorylation. See Figure S1 for further details.



Figure S4: NMR spectra with assignment and corresponding BMRB entry ID codes of My_{CWT} (BMRB: 27414), My_{CWT} -2P (BMRB: 27422), $My_{C_{S373E/T400E}}$ (BMRB: 27419), $My_{C_{S373D/T400D}}$ (BMRB: 27416), $My_{C_{S373D}}$ (BMRB: 27418), and My_{CT400D} (BMRB: 27421). Minor differences between the spectra of mutants and WT (main text Fig. 1) indicate no effect of the mutations on the structure of Myc bHLH-LZ.



Figure S5: Gel filtration DNA shift assay of MycWT, MycWT 2xPhos, and Max. Max (blue) homodimerizes and binds to DNA, shifting the peak of DNA to a shorter elution time relative to the DNA control (black). In contrast to Max (blue), MycWT (green) and MycWT 2xPhos (red) are not able to form stable complex with DNA.



Figure S6: MS quality control of (A) Myc_{WT} before phosphorylation and (B) Myc_{WT}-2P after the phosphorylation by PAK2 and purification on ResS cation exchange. Difference of 160 Da indicate two additional phospho groups on Myc_{WT}.



Figure S7: ITC injection heats, including fit and fit residuals for Max dissociation. Max bHLH-LZ (160 μ M) was titrated into the buffer as described in experimental procedures.



Figure S8: ITC injection heats, including fit and fit residuals for MycWT and MycWT 2xPhos at pH 7.4 and MycWT, MycWT 2xPhos, and MycS373E/T400E at pH 6.5. Data were obtained at 2μ M Max and 20μ M titrant (red) and 4μ M Max and 40μ M titrant (blue). Data at different concentrations were fitted simultaneously to obtain binding constant and thermodynamic information.

MycS373D/T400D : Max pH 6.5



Figure S9: ITC injection heats, including fit and fit residuals for MycS373D/T400D, MycT400D and Myc373D at ITC setup for expected 1:1 stoichiometry followed by MycT400D at ITC setup for 2:1 stoichiometry. Data were obtained at 2μ M Max and 20μ M titrant (red), 4μ M Max and 40μ M titrant (blue), and 8μ M Max and 80μ M titrant (magenta). Data at different concentrations were fitted simultaneously to obtain binding constant and thermodynamic information.



Figure S10: Raw circular dichroism spectra of MycWT (black), MycWT 2xPhos (green), Myc S373D/T400D (red), Myc S373E/T400E (blue), Myc T400D (magenta), and Myc T373D (cyan). Θ minimum at 205nm indicate presence of a random coil mixed with α -helix. The presence of α -helix charecterized by Θ at 222nm.