Glucan Binding Protein C of *Streptococcus mutans* Mediates both Sucrose-Independent and Sucrose-Dependent Adherence

Joshua Mieher, Matthew R. Larson, Norbert Schormann, Sangeetha Purushotham, Ren Wu, Kanagalaghatta R. Rajashankar, Hui Wu and Champion Deivanayagam

Supplemental Data Contents:

Supplemental Figure 1: Proteolytic degradation of GbpC¹¹¹⁻⁵²² displayed on SDS-PAGE gel Supplemental Figure 2: Cartoon describing the orientation of the V-region of GbpC and Agl/II Supplemental Figure 3: Scatchard and Hill plots from ITC data with 9400, 38800 and 70800 Da Dextran Supplemental Figure 4: Reverse ITC of GbpC with dextran, and ITC for interactions of Agl/II and SspB with dextran Supplemental Figure 5: Sensorgram for GbpC interaction with immobilized SRCR1 in the presence/absence of calcium Supplemental Figure 6: The effect of 5000Da Dextran on the adherence of Agl/II to immobilized SRCR1 Supplemental Figure 7: Ligplots for interactions of Ca²⁺ in native GbpC and GbpC-BGC complex structures Supplemental Figure 8: Ligplots for interactions of BGC1 and BGC2 in Ca²⁺ binding site Supplemental Figure 9a: Sensorgrams for interaction of GbpC with SRCR1 or SRCR₁₂₃, and deglycosylated SRCR₁ or SRCR₁₂₃ Supplemental Figure 9b: Sensorgram for GbpC interaction with immobilized SAG Supplemental Table 1: Affinities assessed from dextran inhibition experiments Supplemental Table 2: B-factors for residues of loop region 410-418

Supplemental Figures and Tables:



Supplemental Figure 1: SDS-PAGE gel shows the purified product within one and three months of purification. GbpC¹¹¹⁻⁵²² undergoes proteolytic degradation and results in GbpC¹²⁴⁻⁴⁷⁷, whose crystal structure has now been resolved.



Supplemental Figure 2: These cartoons describe the orientation of the distant V-region of GbpC and Agl/II as it would appear based on the orientation of the helical region that anchors it to the cell surface. In this case, it is particularly interesting to note that the ligand binding regions of GbpC and Agl/II will be in opposite spatial orientation. This gives rise to the idea that while Agl/II could be thought of as the extended grabber of SAG/SRCR, the propensity for GbpC to bind to glucans (which are predominantly on the tooth surface) would serve to bring *S. mutans* closer to the surface, promoting colonization.



Supplemental Figure 3: Scatchard and Hill plots for 9400, 38800 and 70800 Da dextran. The linearity of the Scatchard plots describes a non-cooperative interaction that occurs between GbpC and dextran. In the Hill plot, the gradual reduction in the slopes indicates the saturation of the substrate over multiple injections, thereby describing the molecular model of interaction as shown in Figure 8.



3400 D			Jobbo Da Dextrait 70800 Da Dextrait					
Tabulated results from reverse ITC experiments								
Syringe	C	ell		Ka	лн	45		
GbpC (mM)	Dextran (mM)	Dextran (Da)	N	(M)	(cal/mol)	(cal/mol/deg)		
1.30	0.025	9400	3.55	$1.62 \pm 0.109 imes 10^{-4}$	-1.495 +/- 0.05099 × 104	-32.8		
1.30	0.010	38800	16.33	$8.93 \pm 2.47 \times 10^{\text{-5}}$	-9.417 +/- 892.8 × 10³	-13.1		
1.30	0.005	70800	26	$1.02\pm0.18\times10^{\text{-}4}$	-1.02 +/- 0.06994 × 104	-16		



Α



Supplemental Figure 4: *Panel A* shows preliminary reverse titration ITC experiments performed with GbpC in the syringe and dextran in the cell as opposed to those reported in figure 4. While auto fitting the thermograms in Origin 7.0, fixed N-values from previous experiments (Figure 4) were needed to achieve convergence. The affinity (K_d), enthalpy (Δ H) and entropy (Δ S) values are very similar to the experimental values reported in figure 4, thus confirming the nature of interactions that occur between GbpC and dextran. Unfortunately, better thermograms with higher 'C' values were not obtainable given the instrument limitations. *Panel B* contains ITC experiments to determine the interaction of Agl/II (left) and SspB (right) with dextran, indicating no specific adherence.



GbpC interaction with immobilized SRCR1 in the

Supplemental Figure 5: SPR sensorgram for the interaction between GbpC and *i*SRCR₁ in the presence and absence of calcium. In the absence of calcium there is no interaction.



Supplemental Figure 6: The effect of 5000 Da Dextran on the adherence of AgI/II to immobilized SRCR₁ was evaluated in the presence of various concentrations of dextran. The result of the experiment shows that there is no change in the adherence characteristics of AgI/II to SRCR₁ in the presence of dextran, thus validating our ITC results, which demonstrates that AgI/II and SspB do not adhere to dextran. Similar values were seen for SspB (data not shown).

Calcium coordination bonds and distances										
	Gbp	C-native s	tructure		GbpC-BGC complex structure					
ID	Atom	Atom	Residue	Distance	ID	Atom	Atom	Residue	Distance	
478 CA	CA	0	888 HOH	2.49	501 CA	CA	O3	601 BGC	2.33	
478 CA	CA	0	347 SER	2.36	501 CA	CA	OE1	360 GLU	2.52	
478 CA	CA	OE1	360 GLU	2.48	501 CA	CA	02	601 BGC	2.38	
478 CA	CA	0	524 HOH	2.38	501 CA	CA	OG	347 SER	2.32	
478 CA	CA	OG	347 SER	2.45	501 CA	CA	0	347 SER	2.31	
478 CA	CA	0	1014 HOH	2.47	501 CA	CA	OD1	349 ASN	2.31	
478 CA	CA	OD1	349 ASN	2.31	501 CA	CA	04	601 BGC	2.37	
478 CA	CA	0	893 HOH	2.57	501 CA	CA	0	1 HOH	2.36	



Supplemental Figure 7: The table above and the two Ligplots below detail the interaction of the calcium atoms in the GbpC-native and the GbpC-BGC complex structures. Water molecules in the native structure are displaced to allow for the binding of the glucose molecule.

Table of hydrogen bonding distance with β -D-glucose									
ID	Atom	Atom	Residue	Distance					
601 BGC	01	0	449 THR	3.04					
601 BGC	01	OE2	360 GLU	3.37					
601 BGC	02	OE1	360 GLU	2.49					
601 BGC	02	OE2	360 GLU	2.36					
601 BGC	02	CA	501 CA	2.38					
601 BGC	02	0	1 HOH	3.31					
601 BGC	O5	0	90 HOH	3.12					
601 BGC	O4	OG	347 SER	2.25					
601 BGC	O4	OD1	349 ASN	2.99					
601 BGC	O4	OD1	408 ASP	3.17					
601 BGC	O4	CA	501 CA	2.37					
601 BGC	04	OD2	408 ASP	2.45					
601 BGC	O3	0	451 TRP	2.91					
601 BGC	O3	Ν	451 TRP	3.26					
601 BGC	O3	CA	501 CA	2.33					
601 BGC	O3	0	1 HOH	2.65					
601 BGC	O6	0	90 HOH	2.60					
601 BGC	06	ND2	349 ASN	3.27					
601 BGC	O6	OD2	408 ASP	2.59					
602 BGC	02	0	414 GLY	2.78					
602 BGC	O5	0	451 TRP	3.03					
602 BGC	04	0	17 HOH	2.61					
602 BGC	04	0	82 HOH	2.24					
602 BGC	O3	0	107 HOH	2.92					
602 BGC	O6	0	346 SER	2.28					
602 BGC	06	0	135 HOH	2.39					



Supplemental Figure 8: The table above and the Ligplots below detail the interaction of the glucose molecules BGC1 and BGC2 in close proximity to the Ca²⁺ binding site.



Supplemental Figure 9a: Panel shows SPR sensorgrams for the interaction between GbpC and SRCR, whose results are summarized in Table 2.



Supplemental Figure 9b: Panel shows SPR sensorgram for the interaction between GbpC and SAG, whose results are summarized in Table 2.

Affinities assessed from dextran inhibition experiments.									
Dextran (µM)	GbpC (µM)	K _A (1/M)	К _D (М)	Chi ²					
	4	4.43E+06	4.79E-06	1.48					
0	2	5.61E+06	1.82E-07	6.01					
	1	1.00E+07	1.03E-07	1.79					
	4	5.38E+06	1.92E-07	4.96					
40	2	7.11E+06	1.42E-07	3.02					
	1	1.24E+07	8.43E-08	1.15					
	4	3.32E+06	1.44E-06	2.17					
80	2	4.90E+06	2.05E-07	2.14					
	1	9.91E+06	1.04E-07	0.65					
	4	4.03E+06	A (1/M) K _D (M) 43E+06 4.79E-06 51E+06 1.82E-07 00E+07 1.03E-07 38E+06 1.92E-07 11E+06 1.42E-07 24E+07 8.43E-08 32E+06 1.44E-06 90E+06 2.05E-07 91E+06 1.04E-07 03E+06 2.55E-07 17E+06 6.86E-06 58E+06 2.80E-07	3.12					
160	2	5.17E+06	6.86E-06	1.60					
	1	3.58E+06	2.80E-07	0.50					

Supplemental Table 1: Table describes the observed affinities in the dextran inhibition experiment, in which the adherence of GbpC to immobilized SRCR was analyzed. While we observe that the amount of protein bound on the chip surface decreases in a dose dependent manner, their affinities remain unchanged in the presence of dextran. This indicates the possibility of steric hindrances brought by dextran that would preclude the adherence of GbpC to SRCR.

B-factor (Å ²) for residues loop region 410-418									Average	
	V410	N411	A412	D413	G414	T415	P416	R417	A418	
Native	26.7	34.5	34.5	41.3	37.5	40.2	35.0	35.3	17.2	33.6
BGC-complex	32.7	45.7	49.6	56.2	48.4	44.0	39.6	45.6	27.5	43.3
GbpC Native (for the whole protein)										18.2
Gbpc + BGC (for the whole protein)									20.7	

Supplemental Table 2: The table describes the observed B-factor values for the loop region 410-418 of GbpC in both the native and the complex structure. The B-factor values are higher for these loop regions compared to the average values for the native and the complex structure. This indicates that this region is highly flexible, and could shift to accommodate the ligand.