[d]-Carbon-Carbon Double Bond Engineering in Diazaphosphepines: A Pathway to Modulate the Chemical and Electronic Structures of Heteropines

Yi Ren,^a Melda Sezen,^a Fang Guo,^b Frieder Jäkle,^b Yueh-Lin Loo^a

^aDepartment of Chemical and Biological Engineering, Princeton University, NJ 08544 (USA)

^b Department of Chemistry, Rutgers University, Newark, NJ 07102 (USA)

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Materials and Experiments

General. All manipulations were carried out under a dry nitrogen atmosphere employing standard Schlenk techniques. Reagents were purchased from Sigma-Aldrich and Alfa Aesar, and were, unless otherwise noted, used as-received. NMR solvents were purchased from Cambridge Isotope Laboratories. ¹H-NMR, ¹³C{1H}-NMR, and ³¹P {1H} NMR were recorded on Bruker Avance (III) 300 and 500 MHz spectrometers. Highresolution mass spectra were obtained by Dr. Brandon Fowler of the Mass Spectroscopy Facility at Columbia University and John Eng of the Mass Spectroscopy Facility at Princeton University. We were not able to obtain high-resolution mass spectra on the other compounds as they degraded under the conditions at which they were ionized. The crystal structures of BZ-P (CCDC1427667), BTD-P (CCDC1427666), MI-PO (CCDC1427668), AN-P (CCDC1427664) and AN-PO (CCDC1427665) were obtained by Dr. Philip Jeffrey of the X-ray Crystallography Facility at Princeton University. GIXD experiments were conducted at the G1 station of the Cornell High Energy Synchrotron Source. UV-vis experiments were carried out on a UV-vis-NIR Cary 5000 spectrophotometer. The fluorescence measurements were performed using a Hitachi F-7000 fluorescence spectrophotometer. Absolute quantum yields were obtained with a pre-calibrated Quanta- φ integrating sphere attached to a Fluorolog-3 instrument. Cyclic voltammetry experiments were carried out with a Pt disk as the working electrode, a Pt wire as the counter electrode and a Ag wire as the pseudo-reference electrode. The voltammograms were recorded with ca. 10^{-3} to 10^{-4} M solutions in CH₂Cl₂ containing Bu₄N[PF₆] (0.1 M) as the supporting electrolyte. Theoretical calculations were carried out using the GAUSSIAN 03 suite of programs.^{S1}

Device Fabrication. Patterned ITO (15 Ω /sq) on glass substrates was coated with 30-nm thick poly(3,4ethylenedioxythiophene):poly-(styrenesulfonate), PEDOT:PSS (Clevios P), followed by thermal annealing at 150 °C for 10 min. The PEDOT:PSS was diluted with distilled water at a 1:1 volume ratio prior to use. The organic bulk-heterojunction layer was obtained by spin-coating solutions of P3HT and **MI-PO-C8** (1:2 wt) and P3HT and **Di-MI-PO** (1:4 wt) at 1200 rpm for 60 s atop the PEDOT:PSS layer. The total concentration of donor and acceptor in these solutions was kept constant at 20 mg/mL in CHCl₃. Aluminum (80 nm) top electrodes were thermally evaporated through a shadow mask at a pressure of 10⁻⁶ bar and an evaporation rate of 0.8 Å/s to define an active area of 0.18 cm². Devices containing P3HT and **MI-PO-C8** were annealed at 100 °C for 10 min. Devices containing P3HT and **Di-MI-PO** were annealed at 120 °C for 10 min. Current density– voltage (J-V) characteristics were acquired using a Keithley 2635 source measurement unit under AM 1.5G 100 mW/cm² illumination in a nitrogen-filled glovebox (<0.1 ppm of O₂ and H₂O).

Synthetic Procedures

General synthesis of *B*- and *O*-In

In a 1-necked 150-mL Schlenk flask, dibromo or diiodo moieties (1.0 mole eq.) was mixed with indole-2boronic acid pinacol ester (2.2 mole eq.), tris(dibenzylideneacetone)dipalladium(0) (0.05 mole eq.), tri-tertbutylphosphonium tetrafluoroborate (0.25 mole eq.), and potassium phosphate (9.0 mole eq.) in a mixture of THF and water (4:1 v/v) or toluene and water (4:1 v/v). The resulting mixture was degassed for 10 min, then refluxed under argon overnight. The reaction mixture was allowed to cool to room temperature, after which it was poured into 10 mL of water. The organic layer was extracted with chloroform. The crude product was purified by flash chromatography using dichloromethane and hexanes as eluent (from 1:9 to 4:6 by volume) to obtain *B*- and *O*-In.



BZ-In was obtained as a white solid (Yield: 64%). ¹H NMR (CDCl₃, 500 MHz, δ): 7.94 (s, 2H), 7.66 – 7.64 (m, 2H), 7.61 – 7.59 (m, 2H), 7.43 – 7.41 (m, 2H), 7.11 – 7.06 (m, 6H), 6.68 (s, 2H) ppm. ¹³C NMR (CDCl₃, 137.3, 136.8, 131.1, 131.0, 128.7, 128.6, 122.7, 120.8, 120.5, 111.4, 102.9 ppm. Due to the instability of this compound in the solution, HRMS could not be obtained by using ESI experimental conditions.



FBZ-In was obtained as a white solid (Yield: 56%). ¹H NMR (CDCl₃, 500 MHz, δ): 7.91 (s, 2H), 7.59 (d, J = 7.1 Hz, 2H), 7.12 – 7.06 (m, 6H), 6.71 (s, 2H) ppm. ¹³C NMR (CDCl₃, 125 MHz, δ): 136.7, 128.0, 126.2, 123.7, 121.4, 120.8, 111.5, 107.1 ppm. HRMS: m/z = 378.0775 ([M-2H]⁺, Calcd. 378.0780).



BTD-In was obtained as an orange solid (Yield: 67%). ¹H NMR (CDCl₃, 500 MHz, δ): 8.25 (s, 2H), 7.97 (s, 2H), 7.63 (d, J = 7.4 Hz, 2H), 7.14 – 7.09 (m, 6H), 6.77 (d, J = 2.1 Hz, 2H) ppm. ¹³C NMR (CDCl₃, 125 MHz, δ): 154.6, 137.2, 135.7, 133.7, 128.4, 123.4, 122.7, 121.2, 120.8, 111.5, 104.5 ppm. HRMS: m/z = 367.1013 ([M+H]⁺, Calcd. 367.1017).



CP-In was obtained as a light yellow solid (Yield: 56%). ¹H NMR (CDCl₃, 500 MHz, δ): 8.42 (s, 2H), 7.58 (d, J = 7.8 Hz, 2H), 7.24 – 7.23 (m, 2H), 7.24 – 7.23 (m, 2H), 7.17 – 7.14 (m, 2H), 7.11 – 7.07 (m, 2H), 6.61 (s, 1H), 2.98 (t, J = 7.5 Hz, 4H), 2.09 (q, J = 7.6 Hz, 2H) ppm. ¹³C NMR (CDCl₃, 125 MHz, δ): 136.5, 135.07, 128.9, 128.6, 123.0, 120.8, 123.0, 120.8, 120.5, 111.1, 103.5, 38.2, 22.3 ppm. HRMS: m/z = 299.1551 ([M+H]⁺, Calcd. 299.1548).



MI-In was obtained as a purple solid (Yield: 59%). ¹H NMR (CDCl₃, 500 MHz, δ): 10.10 (s, 2H), 7.65 (d, J = 0.6 Hz), 7.65 (d, J = 8.0 Hz, 2H), 7.42 (d, J = 8.2 Hz, 2H), 7.28 (t, J = 7.6 Hz, 2H), 7.13 (t, J = 7.5 Hz, 2H), 3.15 (s, 3H) ppm. ¹³C NMR (CDCl₃, 125 MHz, δ): 172.3, 137.8, 128.0, 125.6, 122.1, 121.9, 121.3, 119.1, 108.5, 30.0, 24.6 ppm. HRMS: m/z = 342.1220 ([M+H]⁺, Calcd. 342.1242).



AN-In was obtained as a red solid (Yield: 62%). ¹H NMR (CDCl₃, 500 MHz, δ): 8.36 (s, 2H), 7.97 (d, J = 6.9 Hz, 2H), 7.86 (d, J = 8.1 Hz, 2H), 7.70 (d, J = 7.7 Hz, 2H), 7.61 (d, J = 15.1 Hz, 2H), 7.25 (d, J = 8.0 Hz, 2H), 7.20 (d, J = 8.0 Hz, 2H), 7.16 (d, J = 14.8 Hz, 2H), 7.05 (d, J = 1.6 Hz, 2H) ppm. ¹³C NMR (CDCl₃, 125 MHz, δ): 139.3, 136.8, 132.3, 128.9(4), 128.8(9), 128.3(2), 128.2(9), 128.1, 124.6, 123.2, 121.0, 111.3, 105.1 ppm. HRMS: m/z = 383.1546 ([M+H]⁺, Calcd. 383.1548).



EHx-MI-In was obtained as a purple solid (Yield: 52%). ¹H NMR (CDCl₃, 300 MHz, δ): 10.08 (s, 2H), 7.78 – 7.71 (m, 2H), 7.65 (d, J = 8.7 Hz, 2H), 7.41 (d, J = 8.3 Hz, 2H), 7.30 – 7.25 (m, 2H), 7.16 – 7.11 (m, 2H), 3.53 (d, J = 7.1 Hz, 2H), 1.82 – 1.74 (m, 1H), 1.36 – 1.29 (m, 8H), 0.94 – 0.86 (m, 6H) ppm. ¹³C NMR (CDCl₃, 125 MHz, δ): 172.6, 137.8, 128.1, 128.0, 125.5, 121.9, 121.8, 121.2, 111.9, 108.4, 42.5, 38.8, 30.8, 28.9, 24.2, 23.3, 14.4, 10.7 ppm. HRMS: m/z = 440.2322 ([M+H]⁺, Calcd. 440.2338).

General synthesis of mono-substituted mB- and O-In

In a 1-necked 150-mL Schlenk flask, dibromo or diiodo moieties (1.0 mole eq.) was mixed with indole-2boronic acid pinacol ester (2.0 mole eq.), tris(dibenzylideneacetone)dipalladium(0) (0.05 mole eq.), tri-tertbutylphosphonium tetrafluoroborate (0.25 mole eq.), and potassium phosphate (9.0 mole eq.) in a mixture of THF and water (4:1 v/v). The resulting mixture was degassed for 10 min, then refluxed under argon overnight. The reaction mixture was allowed to cool to room temperature, after which it was poured into 10 mL of water. The organic layer was extracted with chloroform. The crude product was purified by flash chromatography using dichloromethane and hexanes as eluent (from 1:9 to 4:6 by volume).



mFBZ-In was obtained as a white solid (Yield: 52%). ¹H NMR (CDCl₃, 500 MHz, δ): 7.63 (s, 1H), 7.41 (d, J = 8.2 Hz, 1H), 7.35 – 7.30 (m, 1H), 7.25 – 7.22 (m, 1H), 5.13 (t, J = 7.5 Hz, 1H), 6.89 (d, J = 1.9 Hz, 1H) ppm. ¹³C NMR (CDCl₃, 125 MHz, δ): 147.8 (d, J = 240.7 Hz), 144.8 (d, J = 233.1 Hz), 139 (d, J = 26.08 Hz), 137.2, 129.8, 128.11, 123.9, 121.3, 121.1, 111.5, 108.7 (dt, J = 21.0, J = 3.8 Hz), 103.6 ppm. HRMS: m/z = 266.0585 ([M+H]⁺, Calcd. 266.0593).



mCP-In was obtained as a white solid (Yield: 43%). ¹H NMR (CDCl₃, 500 MHz, δ): 8.08 (s, 1H), 7.53 (d, J = 7.8 Hz), 7.29 (d, J = 8.0 Hz, 2H), 7.12 (d, J = 15.8 Hz, 2H), 7.04 (d, J = 15.6 Hz, 2H), 6.40 (s, 1H), 5.99 (s, 1H), 2.75 – 2.72 (m, 2H), 2.56 – 2.53 (m, 2H), 2.04 – 1.98 (m, 2H) ppm. ¹³C NMR (CDCl₃, 125 MHz, δ): 136.8, 136.7, 135.2, 129.2, 124.9, 122.5, 120.8, 120.1, 110.7, 33.7, 33.3, 23.5 ppm. HRMS: m/z = 184.1125 ([M+H]⁺, Calcd. 184.1126).

General Synthesis of *B*- and *O*-Ps: In a 1-necked 150-mL Schlenk flask, phenylphosphine dichloride (1.0 mole eq.) was added to an acetonitrile solution of *B*- or *O*-In (1.0 mole eq.) in the presence of triehtylamine (2.0 mole eq.) at 0 $^{\circ}$ C. The resulting mixture was refluxed under argon overnight. The reaction mixture was allowed to cool to room temperature. Acetonitrile was removed under vacuum. The crude product was purified by flash chromatography using dichloromethane and hexanes as eluent (from 1:9 to 9:1 by volume) to obtain **B**- and *O*-**P**.

BZ-P was obtained as a white solid (Yield: 55%). ¹H NMR (CDCl₃, 500 MHz, δ): 8.03 (d, J = 8.3 Hz, 2H), 7.61 (d, J = 7.8 Hz, 2H), 7.41 – 7.39 (m, 2H), 7.30 (t, J = 7.7 Hz, 2H), 7.19 (t, J = 15.0 Hz, 2H), 7.12 – 7.10 (m, 2H), 7.02 – 6.94 (m, 3H), 6.87 – 6.84 (m, 2H), 6.82 (s, 2H) ppm. ³¹P NMR (CDCl₃, 121 MHz, δ): 38.5 ppm. ¹³C NMR (CDCl₃, 125 MHz, δ): 143.3 (d, J = 6.7 Hz), 142.8 (d, J = 26.7 Hz), 137.1, 131.9, 130.6, 129.8 (d, J = 4.8 Hz) = 4.8 Hz

Hz), 129.3, 129.1, 128.5, 128.3 (d, J = 4.8 Hz), 123.2, 121.9, 120.9, 112.0, 109.3, ppm. HRMS: m/z = 415.1360 ([M+H]⁺, Calcd. 415.1359).

FBZ-P was obtained as a white solid (Yield: 57%). ¹H NMR (CDCl₃, 500 MHz, δ): 8.00 (d, J = 8.4 Hz, 2H), 7.62 (d, J = 7.9 Hz, 2H), 7.34 (t, J = 7.7 Hz, 2H), 7.21 – 7.20 (m, 2H), 7.17 – 7.10 (m, 3H), 7.03 – 7.00 (m, 2H), 6.92 (s, 2H) ppm. ³¹P NMR (CDCl₃, 121 MHz, δ): 38.7 ppm. ¹³C NMR (CDCl₃, 125 MHz, δ): 145.5 (dbr, J = 242.6 Hz), 142.6 (d, J = 25.5 Hz), 140.1 (d, J = 262.2 Hz), 136.7, 130.1 (d, J = 6.5 Hz), 129.2 (d, J = 4.5 Hz), 128.8 (d, J = 17.4 Hz), 128.7 (d, J = 4.5 Hz), 124.2, 122.3, 121.5, 118.0 (d, J = 10.1 Hz), 113.7, 112.0 (d, J = 19.2 Hz) ppm. HRMS: m/z = 487.0986 ([M+H]⁺, Calcd. 487.0982).

BTD-P was obtained as a green solid (Yield: 78%). ¹H NMR (CDCl₃, 500 MHz, δ): 8.04 (d, J = 8.3 Hz, 2H), 7.99 (s, 2H), 7.65 (d, J = 7.8 Hz, 2H), 7.35 (t, J = 7.7 Hz, 2H), 7.24 – 7.21 (m, 2H), 7.00 – 6.96 (m, 3H), 6.91 – 6.88 (m, 2H) ppm. ³¹P NMR (CDCl₃, 121 MHz, δ): 38.5 ppm. ¹³C NMR (CDCl₃, 125 MHz, δ): 154.2, 143.2 (d, J = 26.1 Hz), 141.5 (d, J = 6.7 Hz), 136.3, 134.7, 129.6, 129.5, 129.4 (d, J = 4.5 Hz), 128.3 (d, J = 4.7 Hz), 123.9, 122.1, 121.5, 121.2, 112.1 (d, J = 19.5 Hz), 111.4 ppm. HRMS: m/z = 473.0987 ([M+H]⁺, Calcd. 473.0984).

CP-P was obtained as a white solid (Yield: 61%). ¹H NMR (CDCl₃, 300 MHz, δ): 8.01 – 8.02 (m, 2H), 7.60 (d, J = 7.8 Hz, 2H), 7.34 – 7.27 (m, 2H), 7.19 (t, J = 7.9, 2H), 7.15 – 7.12 (m, 2H), 7.07 – 7.02 (m, 2H), 6.70 (s, 2H), 6.31 – 6.28 (m, 2H), 2.99 – 2.93 (m, 2H), 2.48 – 2,43 (m, 2H), 1.92 – 1.85 (m, 1H), 0.86 – 0.84 (m, 1H) ppm. ³¹P NMR (CDCl₃, 121 MHz, δ): 41.8 ppm. ¹³C NMR (CDCl₃, 125 MHz, δ): 142.6 (d, J = 27.7), 140.9 (d, J = 7.3 Hz), 139.2 (d, J = 7.0 Hz), 129.9, 129.9 (d, J = 5.0), 129.1 (d, J = 1.5 Hz), 128.4 (d, J = 4.5 Hz), 128.2 (d, J = 17.2 Hz), 123.3 (d, J = 1.5 Hz), 121.9 (d, J = 1.5 Hz), 121.0 (d, J = 1.3 Hz), 112.2 (d, J = 20.2 Hz), 108.8, 37.4, 22.5 ppm. HRMS: m/z = 405.1508 ([M+H]⁺, Calcd. 405.1521).

MI-P was obtained as a red solid (Yield: 85%). ¹H NMR (CDCl₃, 500 MHz, δ): 8.07 (d, J = 8.3 Hz, 2H), 7.83 (s, 2H), 7.74 (d, J = 7.9 Hz, 2H), 7.43 (t, J = 7.7 Hz, 2H), 7.27 (t, J = 7.5, 2H), 7.17 – 7.14 (m, 1H), 7.09 – 7.06 (m, 2H), 6.31 – 6.28 (m, 2H), 3.00 (s, 3H) ppm. ³¹P NMR (CDCl₃, 121 MHz, δ): 44.4 ppm. ¹³C NMR (CDCl₃, 125 MHz, δ): 169.6, 143.7 (d, J = 27.7 Hz), 137.4 (d, J = 7.56), 134.1 (d, J = 8.8 Hz), 130.0, 130.0, 129.2 (d, J = 5.0 Hz), 128.2 (d, J = 17.6 Hz), 126.0, 122.8 (d, J = 13.9 Hz), 117.3, 112.5, 112.4, 24.6 ppm. HRMS: m/z = 448.1215 ([M+H]⁺, Calcd. 448.1208).

AN-P was obtained as a red solid (Yield: 61%). ¹H NMR (CDCl₃, 500 MHz, δ): 8.13 (d, J = 8.2 Hz, 2H), 7.93 (d, J = 6.9 Hz, 2H), 7.75 (d, J = 8.1 Hz, 2H), 7.71 (d, J = 7.8 Hz, 2H), 7.57 – 7.48 (m, 2H), 7.36 (d, J = 7.6 Hz, 2H), 7.26 (t, J = 7.5 Hz, 2H), 7.22 (s, 2H), 6.82 – 6.76 (m, 3H), 6.71 – 6.68 (m, 2H) ppm. ³¹P NMR (CDCl₃, 121 MHz, δ): 40.30 ppm. ¹³C NMR (CDCl₃, 125 MHz, δ): 142.5 (d, J = 27.7 Hz), 138.7, 137.6 (d, J = 7.5 Hz), 137.5, 130 (d, J = 5.0 Hz), 129.7, 129.3, 129.0, 129.7, 129.3, 129.0, 128.7, 128.5, 128.3 (d, J = 3.8 Hz), 128.1, 127.8, 124.3, 123.6, 122.1, 121.2, 112.2, 112.1, 110.4 ppm. HRMS: m/z = 489.1516 ([M+H]⁺, Calcd. 489.1515).

General Synthesis of *B*- and *O*-POs: In a 1-necked 150-mL Schlenk flask, the corresponding *B*- or *O*-P (1.0 mole eq.) was mixed with H_2O_2 (5.0 mole eq.) in dichloromethane. The resulting mixture was stirred 2 days at room temperature. The reaction mixture was poured into 10 mL of water. The organic layer was extracted with chloroform. The crude product was purified by flash chromatography using dichloromethane and hexanes as eluent (from 1:9 to 9:1 by volume) to obtain the corresponding *B*- and *O*-PO.

BZ-PO was obtained as a white solid (Yield: 87%). ¹H NMR (CDCl₃, 500 MHz, δ): 8.68 (d, J = 8.4 Hz, 2H), 7.61 (dd, J = 7.8, J = 1.7 Hz, 2H), 7.40 – 7.36 (m, 2H), 7.33 (ddd, J = 8.5, J = 7.2, J = 1.4 Hz 2H), 7.28 – 7.24 (m, 4H), 7.22 – 7.21 (m, 9H), 7.16 – 7.13 (m, 9H), 7.12 – 7.08 (m, 9H), 6.82 (d, J = 3.0 Hz, 2H) ppm. ³¹P NMR (CDCl₃, 121 MHz, δ): 15.1 ppm. ¹³C NMR (CDCl₃, 125 MHz, δ): 141.5 (d, J = 3.9 Hz), 139.5 (d, J = 6.0 Hz), 133.2 (d, J = 3.1 Hz), 132.2, 130.9, 130.8, 130.7, 130.2 (d, J = 8.82 Hz), 128.5 (d, J = 15.8 Hz), 124.6, 123.1, 120.8, 115.9, 110.1 (d, J = 6.9 Hz) ppm. HRMS: m/z = 431.1307 ([M+H]⁺, Calcd. 431.1308).

FBZ-PO was obtained as a white solid (Yield: 85%). ¹H NMR (CDCl₃, 500 MHz, δ): 8.63 (d, J = 8.5 Hz), 7.62 (d, J = 7.9 Hz, 2H), 7.44 – 7.35 (m, 5H), 7.28 – 7.25 (m, 4H), 6.92 (s, 2H) ppm. ³¹P NMR (CDCl₃, 121 MHz, δ): 13.9 ppm. ¹³C NMR (CDCl₃, 125 MHz, δ): 141.2 (d, J = 3.78 Hz), 134.05 (d, J = 2.52 Hz), 131.9, 130.6 (d, J = 1.26 Hz), 129.6 (d, J = 8.82 Hz), 129.0 (d, J = 16.4 Hz), 126.8 (d, J = 5.04 Hz), 125.6, 123.5, 121.4, 115.9, 114.5 ppm. HRMS: m/z = 503.0929 ([M+H]⁺, Calcd. 503.0931).

BTD-PO was obtained as a green solid (Yield: 90%). ¹H NMR (CDCl₃, 500 MHz, δ): 8.67 (d, J = 8.2 Hz, 2H), 7.99 (s, 2H), 7.64 (d, J = 7.8 Hz, 2H), 7.40 – 7.36 (m, 4H), 7.29 – 7.26 (m, 2H), 7.19 – 7.16 (m, 1H), 7.07 – 7.05 (m, 2H), 6.98 (d, J = 2.6 Hz, 2H) ppm. ³¹P NMR (CDCl₃, 121 MHz, δ): 14.5 ppm. ¹³C NMR (CDCl₃, 125

MHz, δ): 154.2, 142.2 (d, J = 3.7 Hz), 138.0 (d, J = 5.8 Hz), 133.82 (d, J = 3.2 Hz), 133.5, 131.3, 131.3, 130.9 (d, J = 162.5 Hz), 129.9 (d, J = 8.3 Hz), 128.8, 128.6, 125.4, 123.4, 122.1, 121.2, 116.0, 112.5 (d, J = 6.9 Hz) ppm. HRMS: m/z = 489.0934 ([M+H]⁺, Calcd. 489.0933).

CP-PO was obtained as a white solid (Yield: 80%). ¹H NMR (CDCl₃, 500 MHz, δ): 8.70 (d, J = 8.3 Hz, 2H), 7.6 (d, J = 7.8 Hz, 2H), 6.71 (d, J = 2.5, 2H), 7.38 – 7.32 (m, 3H), 7.28 – 7.25 (m, 2H), 7.18 – 7.14 (m, 2H), 6.71 (d, J = 2.5 Hz, 2H), 6.69 – 6.64 (m, 2H), 3.05 – 2.99 (m, 2H), 2.56 – 2.51 (m, 2H), 1.98 – 1.91 (m, 1H), 0.83 – 0.78 (m, 1H) ppm. ³¹P NMR (CDCl₃, 121 MHz, δ): 15.8 ppm. ¹³C NMR (CDCl₃, 125 MHz, δ): 141.5 (d, J = 4.8 Hz), 136.5 (d, J = 5.9 Hz), 133.11 (d, J = 3.3 Hz), 132.3 (d, J = 158.4 Hz), 130.4 (d, J = 8.5 Hz), 129.6 (d, J = 11.9 Hz), 129.3, 128.8 (d, J = 15.6 Hz), 125.0, 123.4, 120.9, 116.4, 109.6 (d, J = 7.1 Hz), 37.8, 22.2 ppm. HRMS: m/z = 421.1463 ([M+H]⁺, Calcd. 421.1470).

MI-PO was obtained as a red solid (Yield: 87%). ¹H NMR (CDCl₃, 500 MHz, δ): 8.73 (d, J = 8.5 Hz, 2H), 7.96 (d, J 2.5 Hz, 2H), 7.76 (d, J = 7.8 Hz, 2H), 7.51 – 7.45 (m, 2H), 7.47 – 7.37 (m, 2H), 7.35 (t, J = 7.5 Hz, 2H), 7.19 (td, J = 7.7, J = 4.2 Hz, 2H), 6.69 – 6.61 (m, 2H), 3.02 (s, 3H) ppm. ³¹P NMR (CDCl₃, 121 MHz, δ): 15.0 ppm. ¹³C NMR (CDCl₃, 125 MHz, δ): 169.0, 142.7 (d, J = 4.3 Hz), 134.1 (d, J = 3.2 Hz), 131.5, 130.6 (d, J = 8.3 Hz), 130.2 (d, J = 4.9 Hz), 129.7 (d, J = 12.2 Hz), 129.5 (d, J = 15.7 Hz), 127.7, 124.3 122.7, 121.4, 117.8 (d, J = 6.3 Hz), 116.6, 24.7 ppm. HRMS: m/z = 464.1154 ([M+H]⁺, Calcd. 464.1158).

AN-PO was obtained as a red solid (Yield: 88%). ¹H NMR (CDCl₃, 500 MHz, δ): 8.80 (d, J = 8.80 Hz, 2H), 7.93 (d, J = 7.0 Hz, 2H), 7.78 (d, J = 8.10, 2H), 7.71 (d, J = 7.8 Hz, 2H), 7.59 – 7.53 (m, 2H), 7.39 (t, J = 8.3 Hz, 2H), 7.32 (t, J = 7.5 Hz, 2H), 7.21 (d, J = 2.7 Hz, 2H), 7.12 – 7.02 (m, 3H), 6.95 – 6.91 (m, 2H), ppm. ³¹P NMR (CDCl₃, 121 MHz, δ): 15.7 ppm. ¹³C NMR (CDCl₃, 126 MHz, δ): 141.3 (d, J = 5.04 Hz), 138.1, 134.0 (d, J = 6.3 Hz), 133.1 (d, J = 2.5 Hz), 132.2, 130.9, 130.7 (d, J = 8.8 Hz), 129.9 (d, J = 12.6 Hz), 129.0, 128,7 (d, J = 2.5 Hz), 128.6, 128.5, 128.2 (d, J = 10.8), 125.0, 124.5, 123.3, 121.0, 116.1, 110.6 (d, J = 7.5 Hz) ppm. HRMS: m/z = 505.1466 ([M+H]⁺, Calcd. 505.1464).

General Synthesis of *B*- and *O*-PO-Rs: In a 1-necked 150-mL Schlenk flask, the appropriate phosphorus reagent (1.0 mole eq.) was added to an acetonitrile solution of *B*- or *O*-In (1.0 mole eq.) in the presence of triehtylamine (2.0 mole eq.) at 0 $^{\circ}$ C. The resulting mixture was refluxed under argon overnight. The reaction mixture was allowed to cool to room temperature. Acetonitrile was removed under vacuum. The crude product

was purified by flash chromatography using dichloromethane and hexanes as eluent (from 1:9 to 9:1 by volume) to obtain **B-** and *O*-**P-R**s.

MI-PO-C8 was synthesized with octylphosphonic dichloride as the phosphorus reagent. The final product was obtained as an orange solid (Yield: 66%). ¹H NMR (CDCl₃, 500 MHz, δ): 8.61 (dd, J = 8.5, 0.5 Hz, 2H), 8.02 (s, J = 2 Hz, 2H), 7.69 (d, J = 7.9 Hz, 2H), 7.43 – 7.38 (m, 2H), 7.30 – 7.25 (m, 2H), 3.20 (s, 3H), 2.07 – 2.00 (m, 2H), 1.05 – 0.93 (m, 9H), 0.86 – 0.81 (m, 3H), 0.78 (t, J = 7.3 Hz, 3H) ppm. ³¹P NMR (CDCl₃, 121 MHz, δ): 28.8 ppm. ¹³C NMR (CDCl₃, 125 MHz, δ): 169.3, 142.0 (d, J = 3.8 Hz), 130.6 (d, J = 7.5 Hz), 129.0 (d, J = 5.0 Hz), 127.6, 124.0, 122.6, 120.9, 118.4 (d, J = 6.3 Hz), 116.4, 31.8, 30.97, 30.0, 29.9 (d, J = 15.7 Hz), 28.9 (d, J = 3.8 Hz), 24.9, 22.8, 20.8 (d, J = 5.0 Hz), 14.3 ppm. HRMS: m/z = 500.2095 ([M+H]⁺, Calcd. 500.2103).

AN-PO-C8 was synthesized with octylphosphonic dichloride as the phosphorus reagent. The final product was obtained as a red solid (Yield: 53%). ¹H NMR (CDCl₃, 300 MHz, δ): 8.56 (d, J = 6.0 Hz, 2H), 8.36 (s, 2H), 8.12 (d, J = 7.0 Hz, 2H), 7.95 (d, J = 9.0 Hz, 2H), 7.34 – 7.29 (m, 2H), 7.25 – 7.20 (m, 2H), 7.01 (d = 2.4 Hz, 2H), 2.14 – 2.03 (m, 2H), 1.12 – 1.03 (m, 4H), 1.00 – 0.83 (m, 8H), 0.75 (t, d = 7.2 Hz, 3H) ppm. ³¹P NMR (CDCl₃, 121 MHz, δ): 29.4 ppm. ¹³C NMR (CDCl₃, 125 MHz, δ): 140.6 (d, J = 3.8 Hz), 133.0 (d, J = 6.0 Hz), 130.6 (d, J = 8.0 Hz), 129.4, 129.2, 128.7, 128.5, 128.3, 125.3, 125.0, 123.1, 120.9, 116.0, 111.0 (d, J = 6.6 Hz), 31.7, 31.2 (d, J = 109.6 Hz), 30.0, 29.6 (d, J = 16.4 Hz), 28.8 (d, J = 15.1 Hz), 22.7, 21.2 (d, J = 5.0 Hz), 14.3 ppm. HRMS: m/z = 541.2404 ([M+H]⁺, Calcd. 541.2408).

BT-PO-C8 was synthesized with octylphosphonic dichloride as the phosphorus reagent. The final product was obtained as an green solid (Yield: 71%). ¹H NMR (CDCl₃, 500 MHz, δ): 8.55 (d, J = 10.0 Hz, 2H), 8.36(s, 2H), 7.59 (d, J = 7.8 Hz, 2H), 7.33 – 7.29 (m, 2H), 7.24 – 7.21 (m, 2H), 7.01 (d, J = 2.4 Hz, 2H), 2.11 – 2.05 (m, 2H), 1.10 – 1.04 (m, 4H), 0.98 – 0.92 (m, 4H), 0.92 – 0.80 (m, 4H), 0.75 (t, d = 14.6 Hz, 3H) ppm. ³¹P NMR (CDCl₃, 121 MHz, δ): 27.6 ppm. ¹³C NMR (CDCl₃, 126 MHz, δ): 154.8, 141.4 (d, J = 3.0 Hz), 136.8 (d, J = 5.8 Hz), 129.7 (d, J = 7.6 Hz), 125.3, 123.2, 122.8, 121.1, 115.8, 112.6 (d, J = 6.6 Hz), 116.4, 32.9 (d, J = 110.4 Hz), 31.8, 29.8 (d, J = 16.1 Hz), 29.0, 28.9, 22.7, 21.4 (d, J = 4.8 Hz) ppm. HRMS: m/z = 525.1867 ([M+H]⁺, Calcd. 525.1878).

MI-POH was synthesized with phosphorus trichloride as the phosphorus reagent. The reaction was quenched with water. The final product was obtained as an orange solid (Yield: 82%). ¹H NMR (CDCl₃, 500 MHz, δ):

8.49 (d, J = 656.6 Hz, 1H), 8.42 (dd, J = 8.5, J = 1.0 Hz, 2H), 7.91 (d, J = 2.8, 2H), 7.71 (d, J = 7.9, 2H), 7.44 (ddd, J = 8.5, J = 7.2, J = 1.3 Hz, 2H), 7.31 (ddd, J = 7.9, J = 7.1, J = 1.0 Hz, 2H), 3.21 (s, 3H) ppm. ³¹P NMR (C_6D_6 , 203 MHz, δ): -1.75 (d, J = 665.7 Hz) ppm. ¹³C NMR (CDCl₃, 125 MHz, δ): 168.9, 140.7 (d, J = 4.6 Hz), 130.9 (J = 7.5 Hz), 128.3 (J = 7.5 Hz), 127.4, 122.9, 121.4, 118.1 (J = 6.3 Hz), 115.6 ppm. Due to the instability of this compound, HRMS could not be obtained by using ESI experimental conditions.

Di-MI-P was synthesized with 1,2-bis(dichlorophosphino)ethane as the phosphorus reagent. The final product was obtained as a red solid (Yield: 64%). ¹H NMR (CDCl₃, 500 MHz, δ): 7.82 (s, 4H), 7.71 (d, J = 8.4 Hz, 4H), 7.57 (d, J = 7.9 Hz, 4H), 7.36 – 7.32 (m, 4H), 7.18 – 7.15 (m, 4H), 3.52 (d, J = 7.2 Hz, 2H), 1.82 – 1.76 (m, 2H), 1.60 (t, J = 10.1, 4H), 1.37 – 1.29 (m, 16H), 0.93 (t, J = 7.4 Hz, 6H), 0.88 (t, J = 6.8 Hz, 6H) ppm. ³¹P NMR (CDCl₃, 121 MHz, δ): 50.1 ppm. ¹³C NMR (CDCl₃, 125 MHz, δ): 169.5, 142.7 (t, J = 13.8 Hz), 1433.5 (t, J = 4.1 Hz), 129.8 (t, J = 2.52 Hz), 126.1, 122.8, 122.5, 121.5, 118.0 (t, J = 1.2 Hz), 112.4 (t, J = 11.3 Hz), 42.7, 38.5, 30.8, 28.8, 24.4, 24.2, 23.3, 14.4, 10.7 ppm. HRMS: m/z = 987.3901 ([M+Na]⁺, Calcd. 987.3892).

Di-MI-PO was synthesized by oxidation of **Di-MI-P** with H_2O_2 (5.0 mole eq.). The final product was obtained as a red solid (Yield: 61%). ¹H NMR (CDCl₃, 500 MHz, δ): 8.34 (d, J = 8.5 Hz, 4H), 7.99 (s, 4H), 7.58 (d, J = 7.8 Hz, 4H), 7.32 – 7.30 (m, 4H), 7.24 – 7.21 (m, 4H), 3.58 (d, J = 7.2 Hz, 2H), 1.85 – 1.81 (m, 2H), 1.73 (d, J = 3.5, 4H), 1.41 – 1.34 (m, 16H), 0.97 (t, J = 10.0 Hz, 6H), 0.92 (t, J = 10.0 Hz, 6H) ppm. ³¹P NMR (CDCl₃, 121 MHz, δ): 23.5 ppm. ¹³C NMR (CDCl₃, 125 MHz, δ): 168.6, 141.5, 130.4, 128.7, 127.9, 124.2, 119.8, 119.5, 116.1, 42.9, 38.6, 30.8, 30.0, 28.9, 24.2, 23.4, 14.4, 10.8 ppm. HRMS: m/z = 1019.3783 ([M+Na]⁺, Calcd. 1019.3791).



Figure S1. (a) The crystal structure of BZ-P and (b) its molecular stacking.



Figure S2. (a, b) The crystal structure of AN-PO and (c, d) its molecular stacking.



Figure S3. (a, b) The crystal structure of MI-PO and (c) its molecular stacking.



Figure S4. The molecular stacking of BZ-P.



Figure S5. (a) The crystal structure of AN-P and (b) its molecular stacking.



Figure S6. UV-vis absorption and emission spectra of *B*- and *O*-P derivatives (black), as well as of *B*- and *O*-P derivatives (red) in hexane (ca. 10^{-5} M).



Figure S7. Comparison of UV-vis absorption and emission spectra of FBZ and CP derivatives.

The absence of its second indole leg in **CP-In** results in a substantial blue shift in the absorption spectrum of **mCP-In**; this blue shift is more substantial than that observed when comparing the absorption spectra of **FBZ-In** with **mFBz-In**. This observation further supports our hypothesis that derivatives with aromatic substitutions exhibit strong electronic confinement.



Figure S8. UV-vis absorption and emission spectra of (a) **MI-P** (a) and (b) **BTD-P** in different solvents (Hex: hexane, Et₂O: diethylether, THF: tetrahydrofuran, CHCl₃: chloroform, DCM: dichloromethane, Ace: acetone, DMSO: dimethyl sulfoxide, MeOH: methanol. Absorption data are represented by solid lines while emission data are represented by dotted lines. (c) Solvatochromic properties, as quantified by the Stokes shift as function of solvent polarity.



Figure S9. UV-vis absorption and emission spectra of (a) **CP-P** and (b) **AN-P** in different solvents (black: hexane, red: dichloromethane, blue: methanol). Absorption data are provided in solid lines while emission data are provided in dotted lines.



Figure S10. UV-vis absorption and emission spectra of (a) **BZ-P**, (b) **FBZ-P**, (c) **BTD-P**, (d) **CP-P**, (e) **MI-P**, (f) **AN-P** in solution and in the solid state. Absorption data are in solid lines while emission data are in dotted lines.



Figure S11. Optimized structure of **BZ-P** and **CP-P** at the S_0 and S_1 ' states at theory level of B3LYP/6-31+g(d) level.



Figure S12. Cyclic voltammograms of *B*- and *O*-PO species: species (a) BZ-PO, (b) FBZ-PO, (c) BTD-PO, (d) CP-PO, (e) MI-PO and (f) AN-PO in DCM.



Figure S13. UV-vis absorption and emission spectra of (a) BTD-PO-C8 and (b) AN-PO-C8. Absorption data are in solid lines while emission data are in dotted lines.



Figure S14. UV-vis absorption and emission spectra of (a) **MI-POH**, (b) **MI-PO-C8** and (c) **Di-MI-PO**. Absorption data are in solid lines while emission data are in dotted lines. Solution spectra were acquired in XX.



Figure S15. UV-vis absorption and emission spectra of (black) **MI-PO**, (red) **MI-PO-C8** and (blue) **Di-MI-PO** in toluene. Absorption data are in solid lines while emission data are in dotted lines.

Compound	λ_{abs}^{a} [nm]	$\lambda_{\rm em}^{\ \ b}$ [nm]	ϕ^{c}	$E_{\rm red}^{d}$	$E_{\rm ox}^{\ \ d}$
BZ-PO	289	390	6.5%	n.d. ^e	1.14 ^f
FBZ-PO	289	397	17.0%	n.d. ^e	1.17 ^{<i>f</i>}
BTD-PO	402	474	21.6%	-1.62^{f}	n.d. ^e
СР-РО	364	421	30.0%	n.d. ^e	0.61 ^g
МІ-РО	485	538	68.4% 72.5% ^h 43.0% ⁱ	-1.23 ^{<i>f</i>} /-1.94 ^{<i>f</i>}	1.00 ^{<i>f</i>}
AN-PO	495	582	18.7%	-1.71 ^{<i>f</i>} /-2.03 ^{<i>g</i>}	$0.72^{f}/1.00^{f}$
BTD-PO-C8	398	474	27.9%	N/A	N/A
AN-PO-C8	495	582	14.8%	N/A	N/A
MI-PO-C8	485	538	43.5% 59.4% ^h 30.2% ⁱ	N/A	N/A
MI-POH	486	539	27.6%	N/A	N/A
Di-MI-P	493	552	35.2%	N/A	N/A
Di-MI-PO	512	545	$ \begin{array}{r} 19.1\% \\ 44.3\%^{h} \\ 1.2\%^{i} \end{array} $	N/A	N/A

Table S1: Photophysical and redox data of diazaphosphepine oxide species.

^{*a*} absorption maximum measured in hexane. ^{*b*} emission maximum in hexane. ^{*c*} Fluorescence QY determined by a calibrated integrating sphere system in CH₂Cl₂. ^{*d*} vs. Fc^{0/+}, 0.1 M Bu₄N[PF₆] in CH₂Cl₂. ^{*e*} not detected in solvent range. ^{*f*} quasi-reversible or reversible ($E_{red}(E_{ox}) = 1/2(E_{pc}+E_{pa})$). ^{*g*} irreversible ($E_{red}(E_{ox}) = E_{pc}(E_{pa})$). ^{*h*} in toluene. ^{*I*} in DMF.

The photophysical properties of **Di-MI-PO** are characteristically different from those of the other MIderivatives, both in solution and in the solid state. Its fluorescence quantum yield decreases substantially with increasing solvent polarity whereas the fluorescence of **MI-PO-C8** and **MI-PO** do not (Table S1). We attribute the low quantum yield of **Di-MI-PO** in DMF, to the formation of a non-emissive symmetry-breaking charge transfer (SBCT) state in polar solvents. The presence of SBCT in dimeric systems has previously been reported.^{S2-S4} The presence of SBCT in electron acceptors have been shown to improve the V_{oc} of organic solar cells, respectively.^{S3}

Compound	NICS(0)	$NICS(-1Å)^a$	$NICS(+1Å)^{b}$	
-		ppm	ppm	
BZ-P	3.86	0.72	0.20	
FBZ-P	3.23	-0.54	0.55	
BTD-P	4.12	0.09	1.04	
CP-P	3.24	0.38	0.54	
MI-P	2.46	-0.19	0.53	
AN-P	2.66	-0.23	0.48	

Table S2: Nucleus-independent chemical shifts data at level of GIAO/B3LYP/6-31+g(d).

^{*a*} H located at the same direction of Phenyl group on P-center. ^{*b*} H located at the opposite direction of Phenyl group on P-center.

Table S3: Devices characteristics.

acceptor	blend ratio of donor and acceptor	$V_{\rm oc}[V]$	$J_{\rm sc}[{\rm mA/cm}^2]$	FF [%]	PCE [%]
Di-MI-PO	1:1 wt as-cast	0.77±0.01	0.57±0.1	26±0.5	0.11±0.02
	1:1 wt annealed a	0.65±0.01	2.04±0.1	30±0.3	0.40±0.02
	1:2 wt annealed a	0.67±0.01	2.50±0.05	30±0.2	0.50±0.01
	1:4 wt annealed a	0.71±0.02	3.77±0.7	39±3	1.04±0.2
	1:5 wt annealed ^{<i>a</i>}	0.66±0.01	2.38±0.06	29±0.4	0.46±0.02
MI-PO-C8	1:1 wt annealed ^{b}	0.61±0.004	1.93±0.1	37±0.6	0.44±0.02
	1:2 wt annealed ^{b}	0.61±0.01	3.07±0.1	38±0.5	0.71±0.02
	1:4 wt annealed b	0.55±0.06	1.22±0.02	44±0.6	0.29±0.01

^a annealed at 120C for 10 min. ^bannealed at 100C for 10 min,



Figure 16. GIXD images of Di-MI-C8 and Di-MI-PO as-cast and annealed films.



q_{xy} (A⁻¹) q_{xy} (A⁻¹) Figure 17. GIXD images of active layers comprising Di-MI-C8 and P3HT as well as Di-MI-PO and P3HT before and after thermal annealing.

P3HT

P3HT

1

2

N

0

0

P3HT

SHT

1

2

0

0







Figure S21. ¹³C NMR spectrum of FBZ-In in CDCl₃.



Figure S23. ¹³C NMR spectrum of BTD-In in CDCl₃.



Figure S24. ¹H NMR spectrum of CP-In in CDCl₃.



Figure S25. ¹³C NMR spectrum of CP-In in CDCl₃.



Figure S27. ¹³C NMR spectrum of MI-In in CDCl₃.



Figure S28. ¹H NMR spectrum of MI-In-EHx in CDCl₃.





Figure S31. ¹³C NMR spectrum of AN-In in CDCl₃.



Figure S32. ¹H NMR spectrum of mFBZ-In in CDCl₃.



Figure S33. ¹³C NMR spectrum of mFBZ-In in CDCl₃.



Figure S34. ¹H NMR spectrum of mCP-In in CDCl₃.



Figure S35. ¹³C NMR spectrum of mCP-In in CDCl₃.



Figure S37. ¹³C NMR spectrum of **BZ-P** in CDCl₃.



Figure S39. ¹³C NMR spectrum of FBZ-P in CDCl₃.

Figure S41. ¹H NMR spectrum of BTD-P in CDCl₃.

Figure S42. ¹H NMR spectrum of CP-P in CDCl₃.

Figure S43. ¹³C NMR spectrum of CP-P in CDCl₃.

Figure S45. ¹³C NMR spectrum of MI-P in CDCl₃.

Figure S46. ¹H NMR spectrum of AN-P in CDCl₃.

Figure S47. ¹³C NMR spectrum of AN-P in CDCl₃.

Figure S49. ¹³C NMR spectrum of BZ-PO in CDCl₃.

Figure S51. ¹³C NMR spectrum of FBZ-PO in CDCl₃.

Figure S52. ¹H NMR spectrum of BTD-PO in CDCl₃.

Figure S53. ¹³C NMR spectrum of BTD-PO in CDCl₃.

Figure S55. ¹³C NMR spectrum of CP-PO in CDCl₃.

Figure S56. ¹H NMR spectrum of MI-PO in CDCl₃.

Figure S57. ¹³C NMR spectrum of MI-PO in CDCl₃.

Figure S58. ¹H NMR spectrum of AN-PO in CDCl₃.

Figure S59. ¹³C NMR spectrum of AN-PO in CDCl₃.

Figure S61. ¹³C NMR spectrum of MI-PO-C8 in CDCl₃.

Figure S62. ¹H NMR spectrum of BTD-PO-C8 in CDCl₃.

Figure S63. ¹³C NMR spectrum of BTD-PO-C8 in CDCl₃.

Figure S65. ¹³C NMR spectrum of AN-PO-C8 in CDCl₃.

80 70 f1 (ppm)

Figure S67. ¹³C-NMR spectrum of MI-PO-H in CDCl₃.

Figure S68. ¹H NMR spectrum of Di-MI-P in CDCl₃.

Figure S69. ¹³C NMR spectrum of Di-MI-P in CDCl₃.

Figure S70. ¹H NMR spectrum of Di-MI-PO in CDCl₃.

Figure S71. ¹³C NMR spectrum of Di-MI-PO in CDCl₃.

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