1	Supplementary information
2	
3	
4	
5	
6	LIM Protein Ajuba associates with the RPA complex through direct cell
7	cycle-dependent interaction with the RPA70 subunit
8	
9	Sandy Fowler, Pascal Maguin, Sampada Kalan and Diego Loayza
10	
11	
12	
13	
14	
15	
16	
17	
18	
19	
20	
21	
22	
23	

24 Supplementary Figures:

25

26 Figure S1. Ajuba-RPA70 interaction is reduced during replication stress. IP-

- 27 Western from total extracts in unsynchronized HTC75 (left) and IMR90 (right)
- cells. Cells were untreated and treated with 2mM HU for 24 hours (+2mM HU).
- 29 The * denotes a non-specific band.
- 30 **Figure S2.** Ajuba protein levels do not change during hydroxyurea treatment.

31 HTC75 cells were treated with 2mM HU for 3, 6, and 24 hours and processed for

- 32 protein extraction and Western blotting.
- 33 **Figure S3.** Flow cytometry for propidium iodide content of HTC75 cells (see
- 34 Figure 1C) and IMR90 cells (see Figure S4). Cells were untreated (UT), treated
- 35 with 2mM HU for 24 hours (24 he HU), and synchronized to S phase by double
- thymidine block and release for 2 hours (dT 2hr) or 4.5 hours (dT 4hr).
- 37 **Figure S4.** Ajuba-RPA32 interaction is reduced during replication stress but not
- in S phase cells. IMR90 cells were untreated, treated with 2mM HU for 24 hours
- 39 (24 hr 2mM HU), or synchronized to S phase by double thymidine block and
- 40 released for 4.5 hours (double thymidine S phase).
- 41 **Figure S5.** Ajuba-RPA70 nuclear co-localization is increased by Leptomycin B
- 42 treatment (10uM). A) Co-immunofluorescence of Ajuba and RPA70 in
- 43 unsynchronized HTC75 and IMR90 cells. Arrowheads point to sites of co-
- 44 localization B) Quantification of cells that exhibited >3 foci of Ajuba-RPA70 co-
- 45 localization in untreated and leptomycin B treated HTC75 cells (n=100 cells, on 3
- 46 independent experiments).

- 47 Figure S6. A) Ajuba displays significant co-localization with PCNA. Co-
- 48 immunofluorescence of Ajuba and PCNA in unsynchronized and synchronized
- 49 (double thymidine block and release) HTC75 cells. B) Ajuba displays significant
- 50 co-localization with BrdU in a subset of BrdU-positive cells. A culture of
- asynchronous HTC75 cells was pulsed with BrdU for 1 hour and processed for
- 52 BrdU and Ajuba co-immunofluorescence.
- 53 **Figure S7.** A) Ajuba does not directly interact with POT1. Binding experiment of
- 54 full-length His-Ajuba and co-translated full-length POT1 followed by
- autoradiography. B) Binding experiment between co-translated RPA70 OB folds
- 56 B-C and full-length Ajuba-His followed by autoradiography.
- 57 Figure S8. LIM domain 1 of Ajuba and OB fold A of RPA70 mediate the direct
- 58 interaction between the two proteins. Schematic of Ajuba and RPA70, arrows
- 59 pointing up represent the positions used to generate truncation mutants.
- 60 Figures S9-S13. Full-length blots and scans showed in the main figures. A
- dashed red box within each blot or scan indicates the approximate areas used to
- 62 produce the panels shown in the main figures.
- 63

64

65

Supplementary figures.

Figure S1.



Figure S2.



Figure S3.

HTC75	UT	24hr HU	dT 2hr	IMR90	UT	24hr HU	dT 4.5hr
G1	64%	58.85%	22.1%	G1	63%	55.5%	20%
S	24%	41.14%	76.7%	S	8%	44%	73%
G2/M	11%	0%	1.62%	G2/M	29%	0%	4%

Figure S4.



Figure S5.



Β.

HTC75

Lept. B



C.



Figure S6.

Α.



Β.



Α.



Β.







Figure S9.



Fig. 1A (bottom)



Fig. 1B



Fig. 1C



Fig. S2





Fig5A (top)

Fig5A (bottom)



Fig.6A (bottom)



Fig.6A (top)



Fig.6B



Fig.6C (bottom)

