

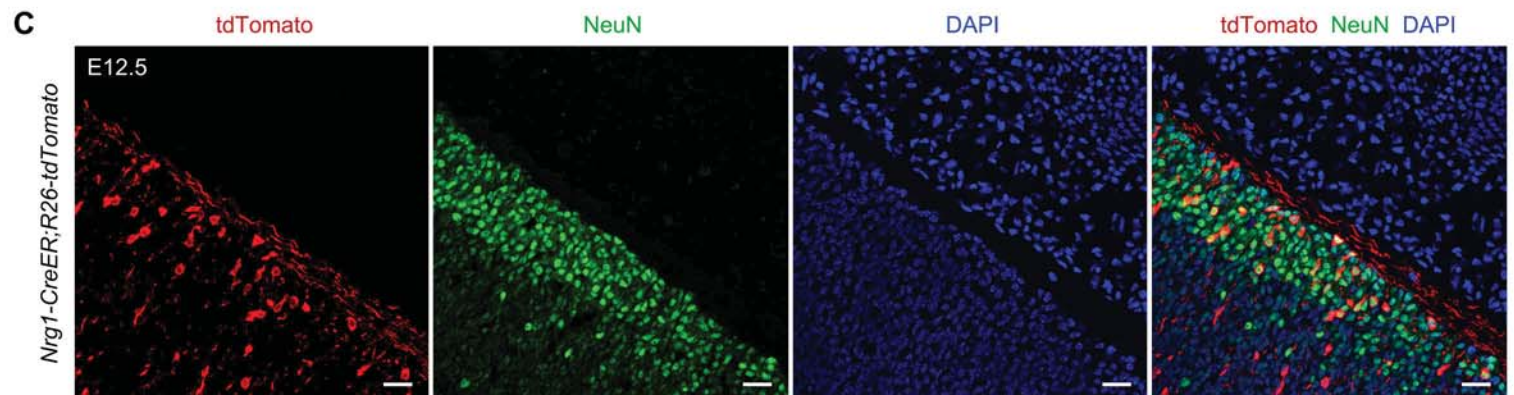
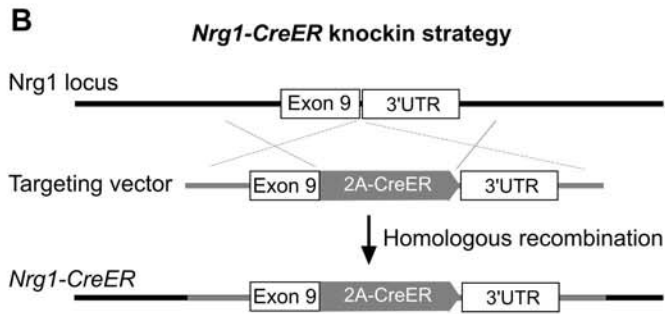
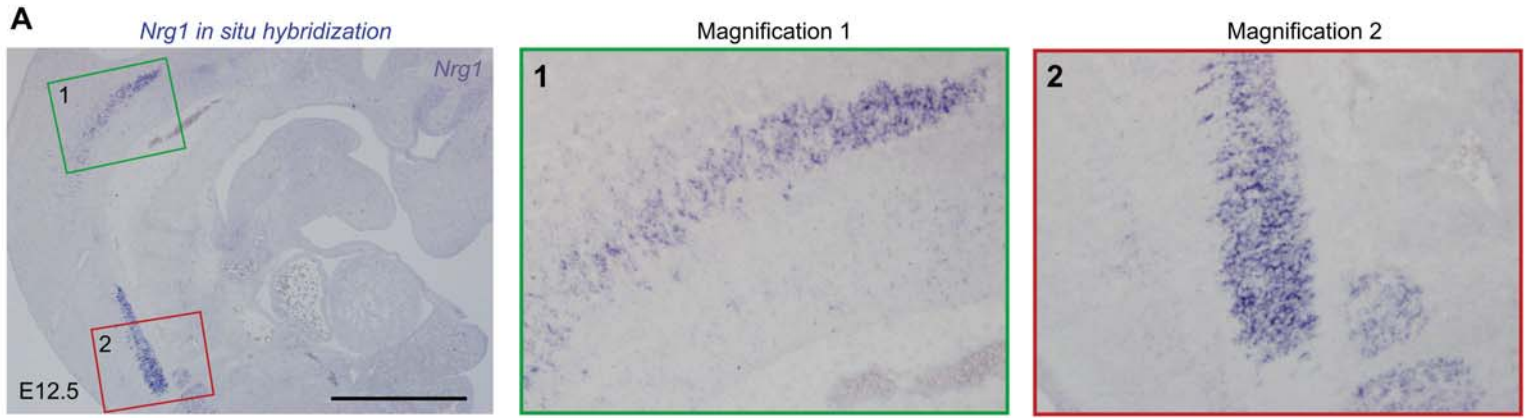
# **Supplemental Material**

## **Online Figures I-XII**

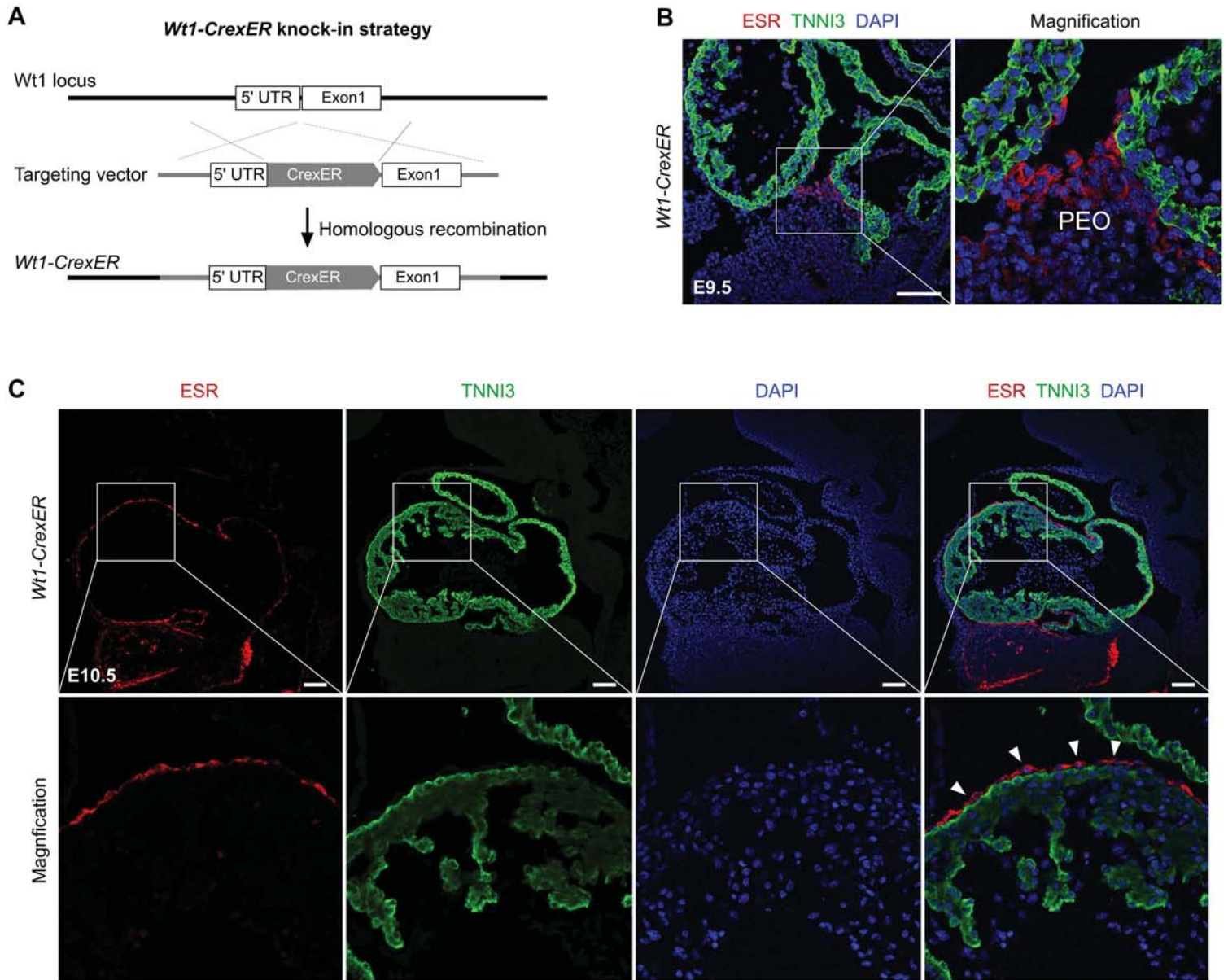
### **Online Table I**

#### **Genetic Targeting of Organ-Specific Blood Vessels**

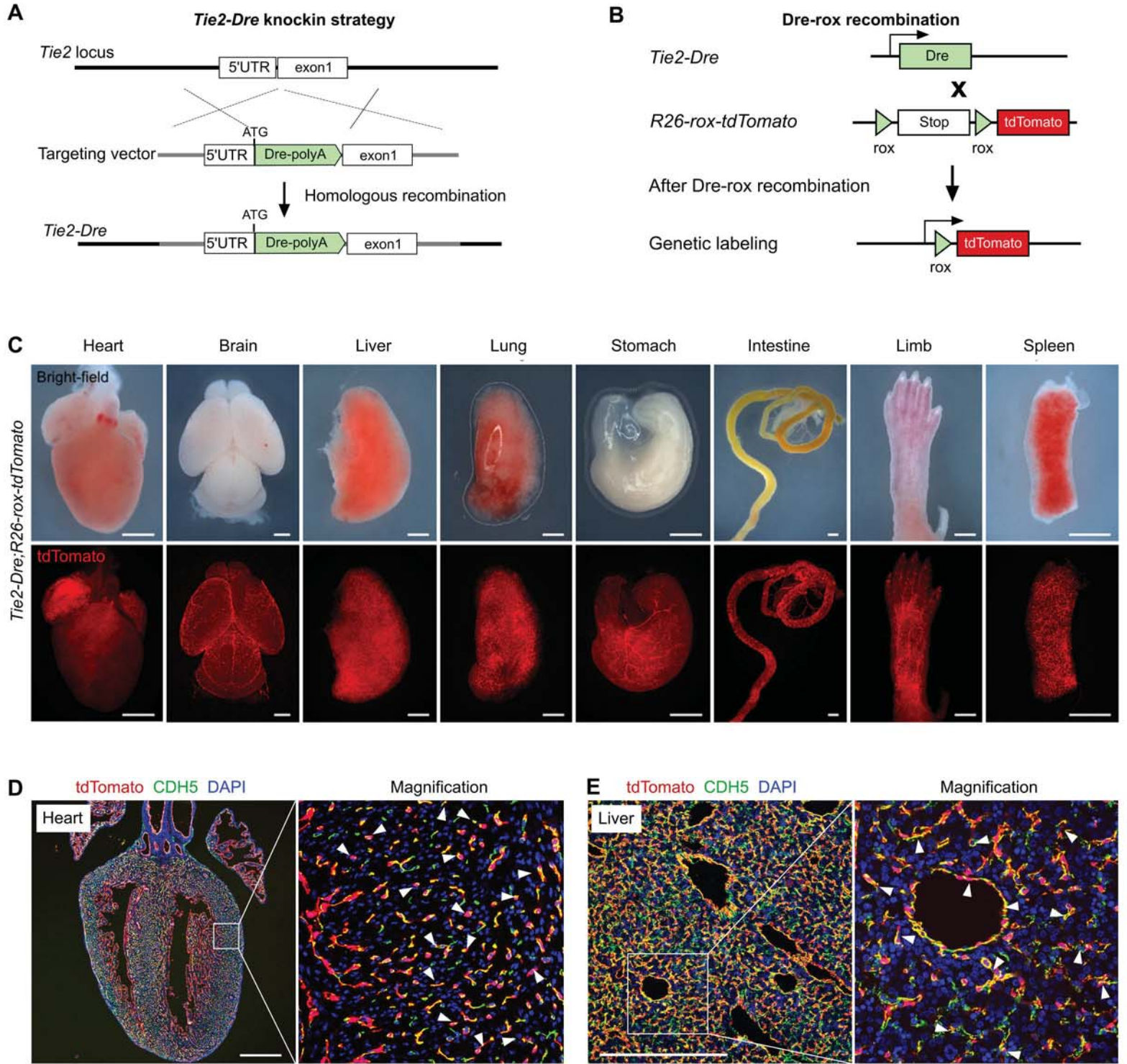
Wenjuan Pu,<sup>1,2</sup> Lingjuan He,<sup>1,2</sup> Ximeng Han,<sup>3</sup> Xueying Tian,<sup>1,2</sup> Yan Li,<sup>1,2</sup> Hui Zhang,<sup>1,3</sup> Qiaozhen Liu,<sup>1,2</sup> Xiuzhen Huang,<sup>1,2</sup> Libo Zhang,<sup>1,2</sup> Qing-Dong Wang,<sup>4</sup> Zhenyang Yu,<sup>5</sup> Xiao Yang,<sup>5</sup> Nicola Smart,<sup>6</sup> Bin Zhou<sup>1,2,3,7</sup>



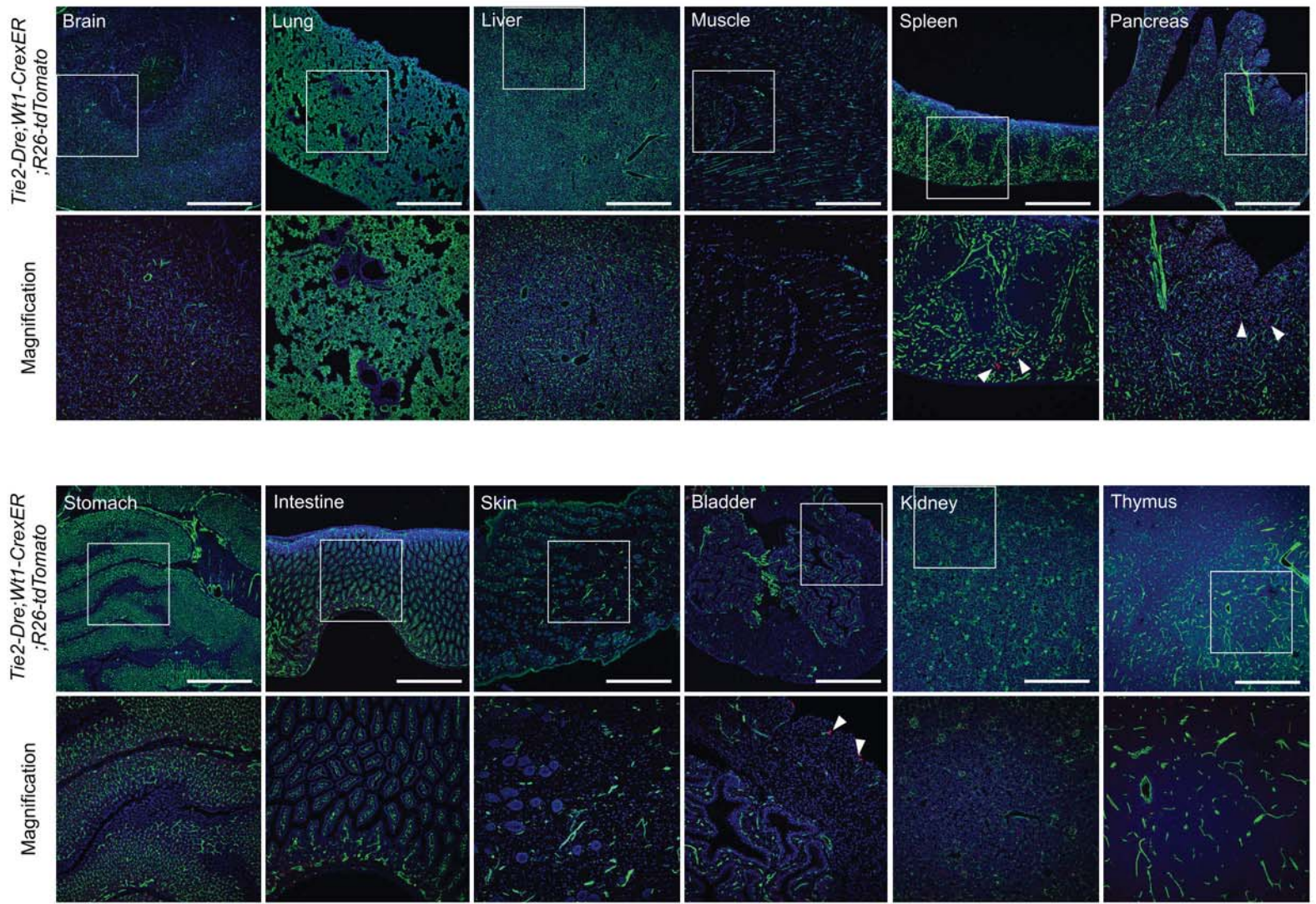
**Online Figure I. Generation and characterization of *Nrg1-CreER* line.** **A**, *In situ* hybridization of *Nrg1* on E12.5 wild-type embryonic sections. **B**, Schematic figure showing generation of *Nrg1-CreER* allele by homologous recombination. **C**, Immunostaining for tdTomato and neuron marker NeuN on E12.5 *Nrg1-CreER;R26-tdTomato* embryonic sections. Tamoxifen was induced at E9.5. Scale bars, 100  $\mu$ m. Each figure is representative of 5 individual samples.



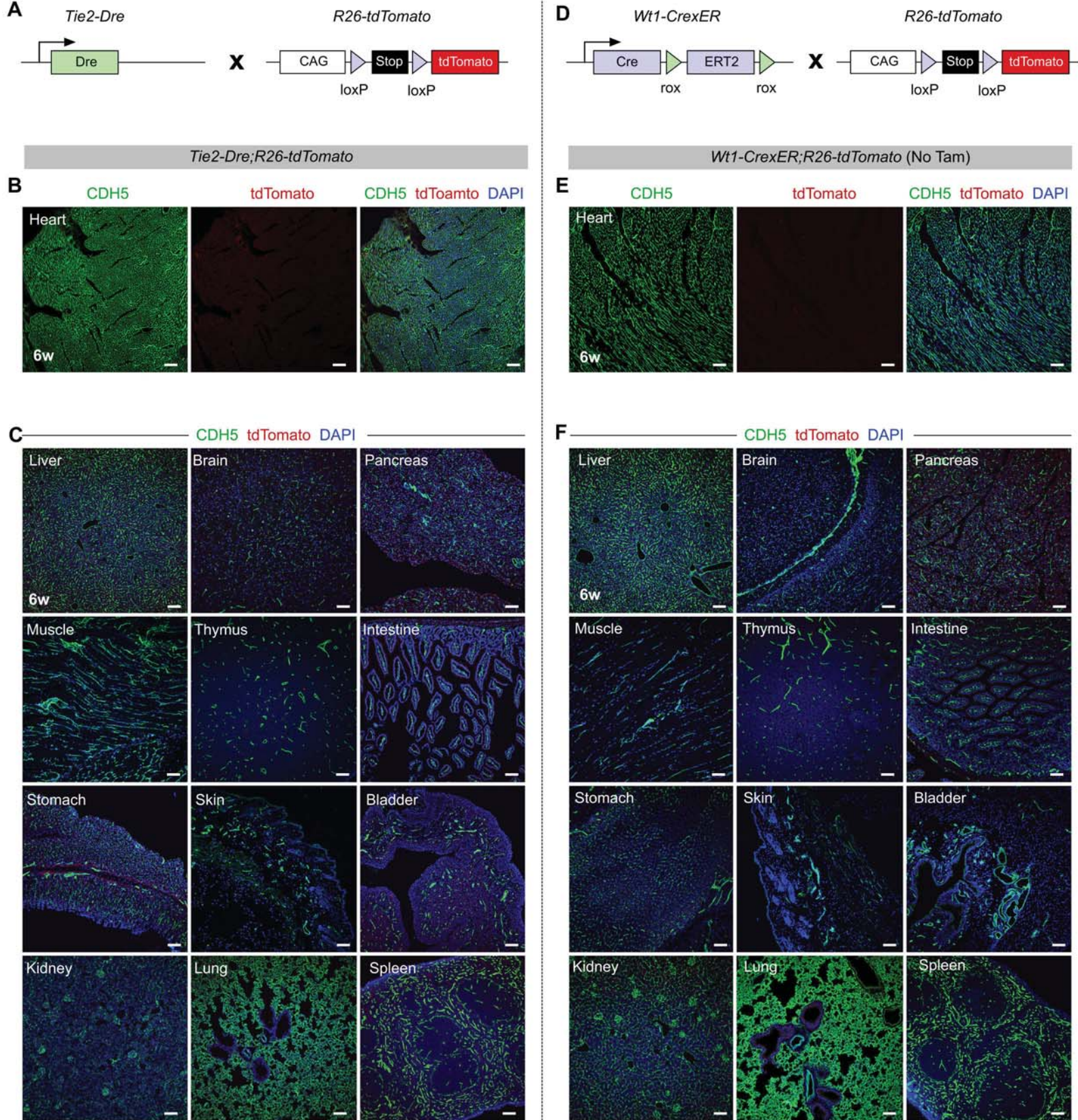
**Online Figure II. Generation and characterization of *Wt1-CrexER*.** **A**, Schematic figure showing knock-in strategy for *Wt1-CrexER* allele. **B**, Immunostaining for ESR and TNNI3 on E9.5 *Wt1-CrexER* section shows ESR<sup>+</sup> cells in proepicardium (PEO). **C**, Immunostaining for ESR and TNNI3 on E10.5 *Wt1-CrexER* embryonic section shows ESR<sup>+</sup> cells in epicardium (arrowheads). Scale bars, 100  $\mu$ m. Each image is representative of 5 individual samples.



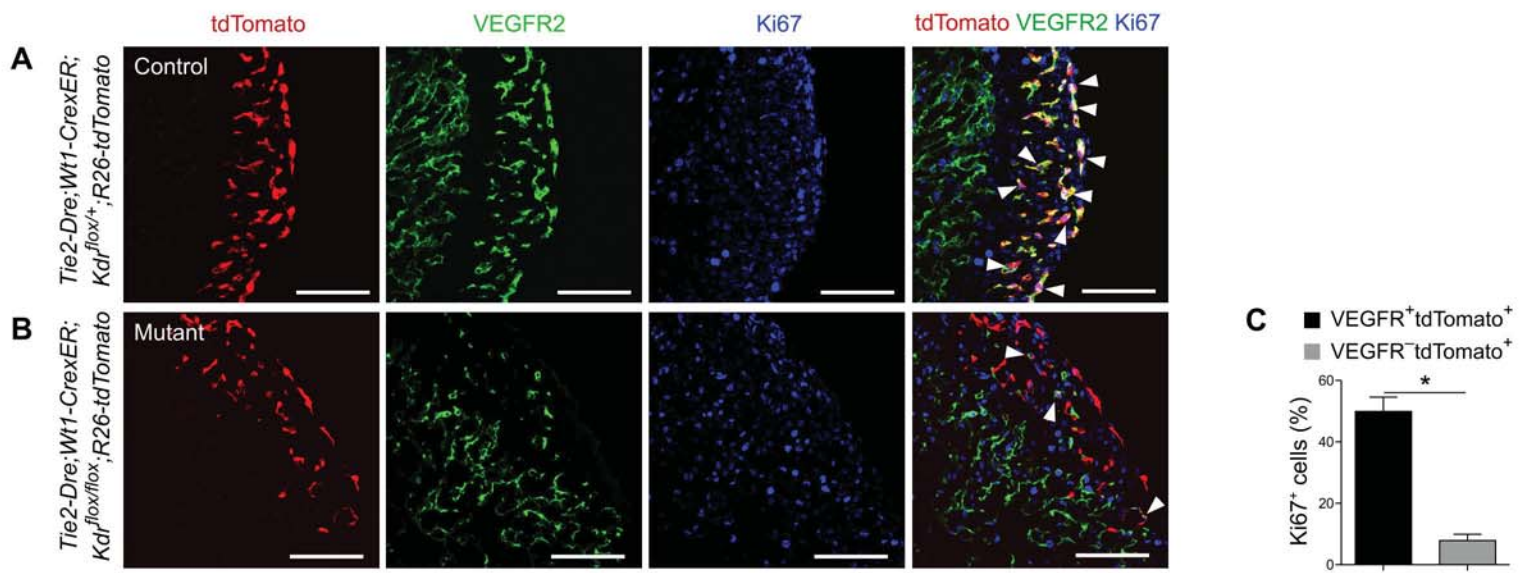
**Online Figure III. Generation and characterization of *Tie2-Dre* line.** **A**, Schematic showing strategy for knockin of *Tie2-Dre* allele by homologous recombination. **B**, Schematic showing *Tie2-Dre* mediated Dre-rox recombination on R26-rox-tdTomato reporter. **C**, Whole-mount view of organs from P0 *Tie2-Dre;R26-rox-tdTomato* mouse. **D,E**, Immunostaining for tdTomato and CDH5 on heart and liver sections. Arrowheads indicate tdTomato<sup>+</sup>CDH5<sup>+</sup> endothelial cells. Scale bars, 1 mm in C; 500  $\mu$ m in D,E. Each image is representative of 5 individual samples.



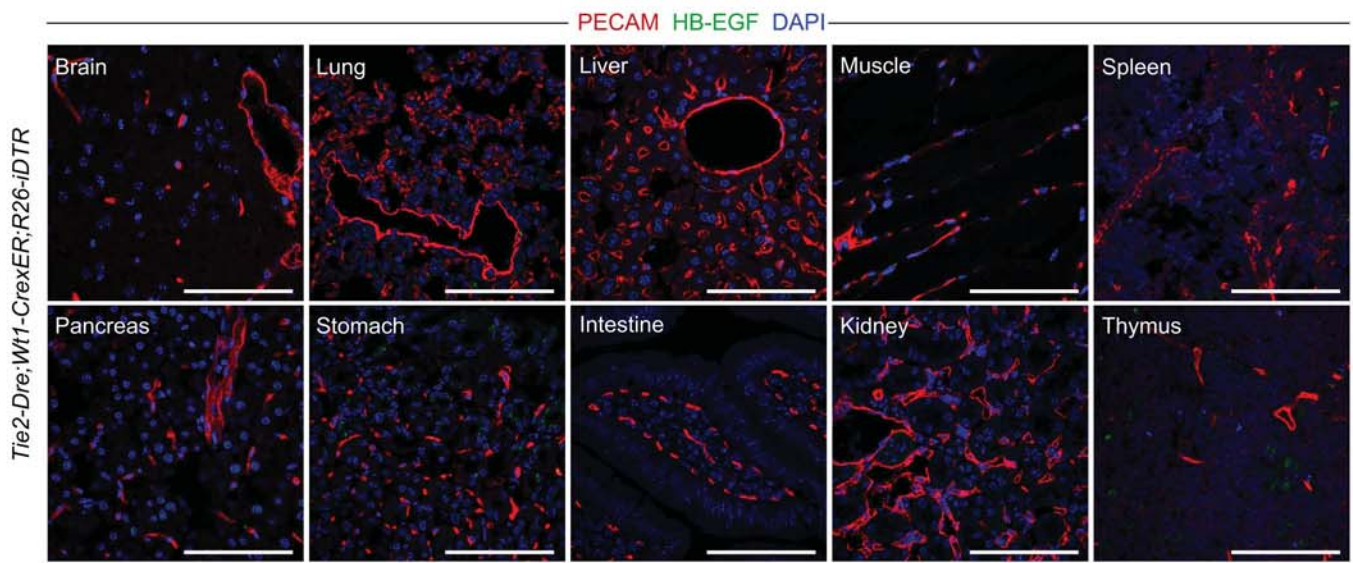
**Online Figure IV. Magnification of immunostaining images shows negligible recombination in other organs.** Immunostaining for *tdTomato* and *CDH5* on tissue sections collected from different organs of *Tie2-Dre;Wt1-CrexER;R26-tdTomato* mice. Arrowheads indicate *tdTomato*<sup>+</sup> cells. Scale bars, 1 mm. Each image is representative of 5 individual biological samples.



**Online Figure V. *Tie2-Dre* and *Wt1-CreXER* does not recombine *R26-tdTomato* reporter. A,D, Schematic showing crossing of *Tie2-Dre* with *R26-tdTomato* or *Wt1-CreXER* with *R26-tdTomato*. B,E, Immunostaining for tdTomato and CDH5 on heart sections from adult *Tie2-Dre;R26-tdTomato* mice (left panel) or adult *Wt1-CreXER;R26-tdTomato* mice (right panel). C,F, Immunostaining for tdTomato and CDH5 on tissue sections from adult *Tie2-Dre;R26-tdTomato* mice (left panel) or adult *Wt1-CreXER;R26-tdTomato* mice (right panel). Scale bars, 100  $\mu$ m. Each image is representative of 5 individual samples.**

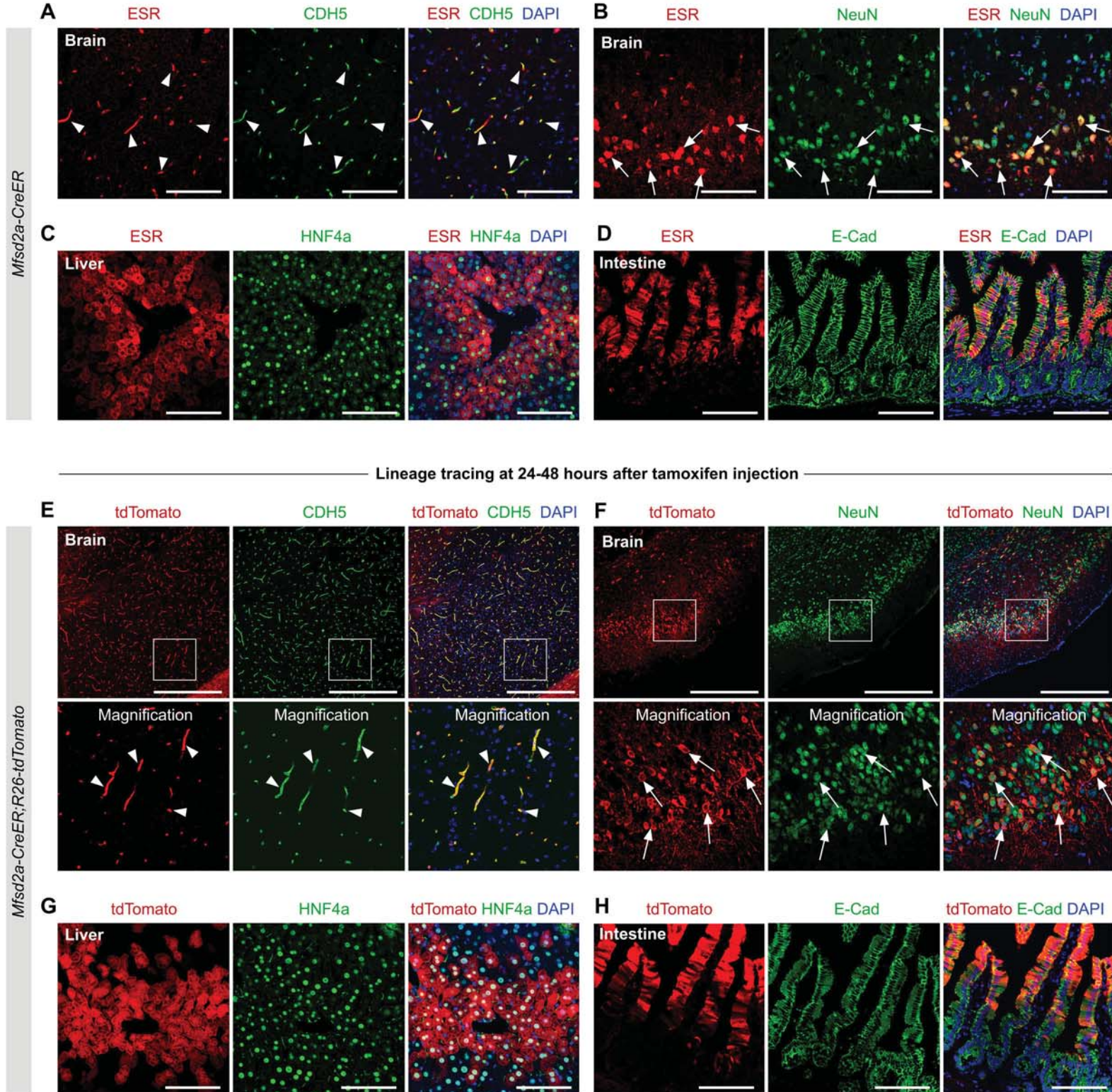


**Online Figure VI. Proliferation of CoEC-Cre labeled endothelial cells.** **A,B**, Immunostaining for tdTomato, Ki67 and VEGFR2 on E14.5 heart sections of *Tie2-Dre;Wt1-CrexER;Kdr<sup>lox/+</sup>;R26-tdTomato* (control) or *Tie2-Dre;Wt1-CrexER;Kdr<sup>lox/lox</sup>;R26-tdTomato* (mutant) embryos. Arrowheads indicate tdTomato<sup>+</sup>VEGFR2<sup>+</sup> endothelial cells in the control and mutant hearts. **C**, Quantification of the percentage of Ki67<sup>+</sup> cells in VEGFR<sup>+</sup>tdTomato<sup>+</sup> or VEGFR<sup>-</sup>tdTomato<sup>+</sup> cell populations in compact myocardium. Data are mean ± SEM.; \*P < 0.05; n = 5. Scale bars, 100 μm. Each figure is representative of 5 individual samples.

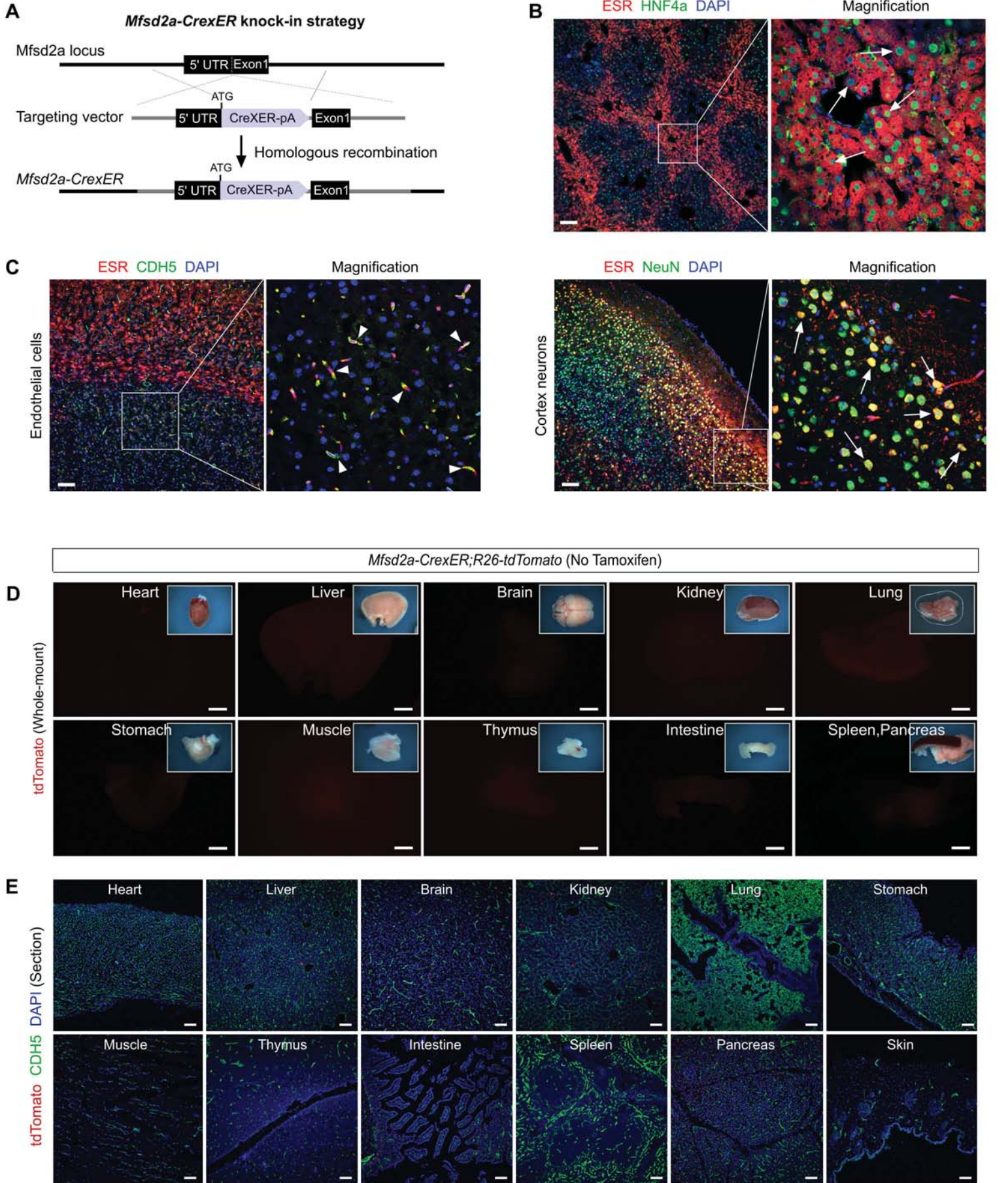


**Online Figure VII. *CoEC-Cre* minimally recombines *R26-iDTR* in endothelial cells of other organs or tissues.** Immunostaining for PECAM and HB-EGF (DTR) on tissue sections of organs collected from 6 week old *Tie2-Dre;Wt1-Cre<sup>ER</sup>;R26-iDTR* mice. Each image is representative of 5 individual samples. Scale bars, 100  $\mu$ m.

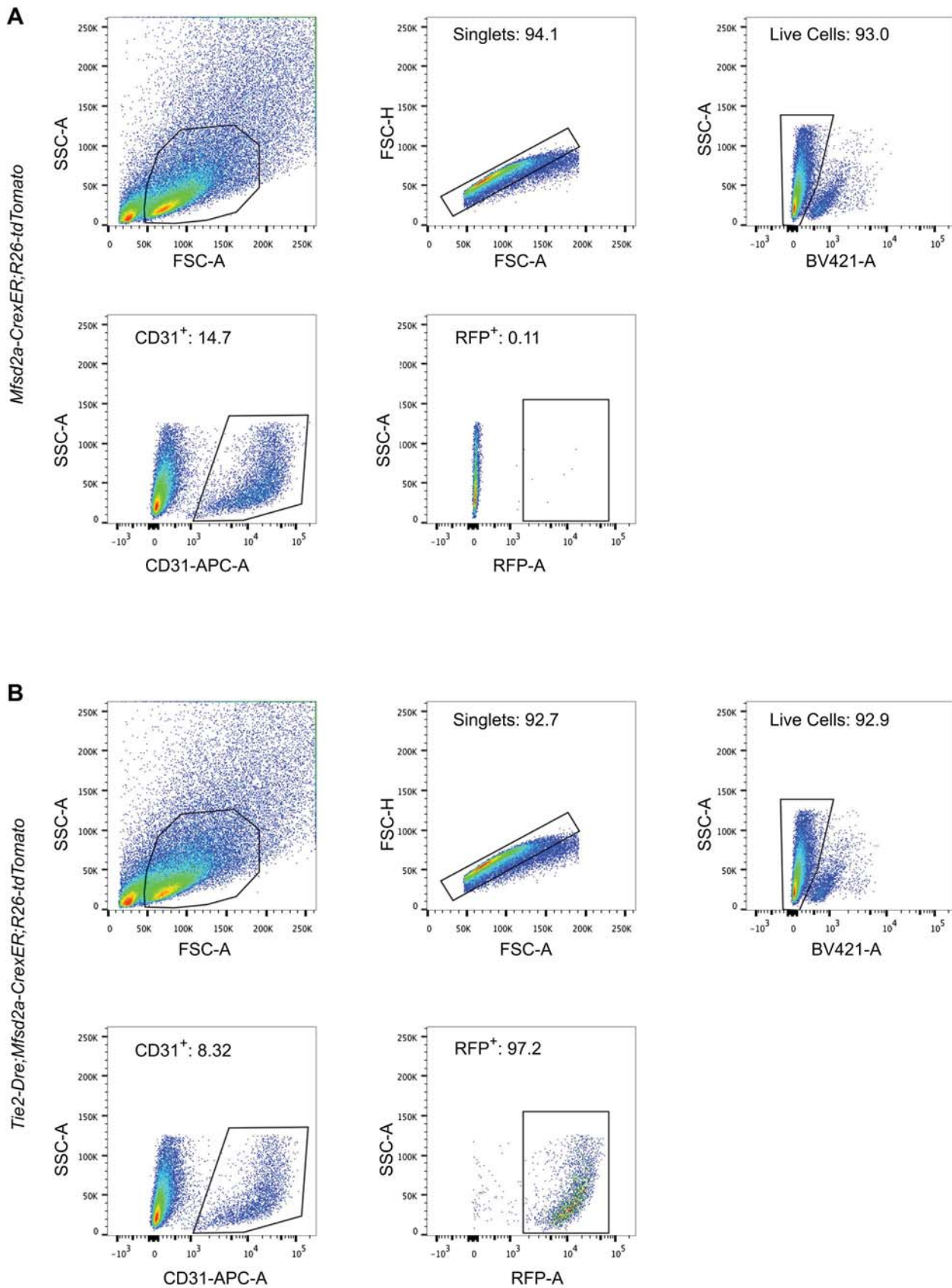




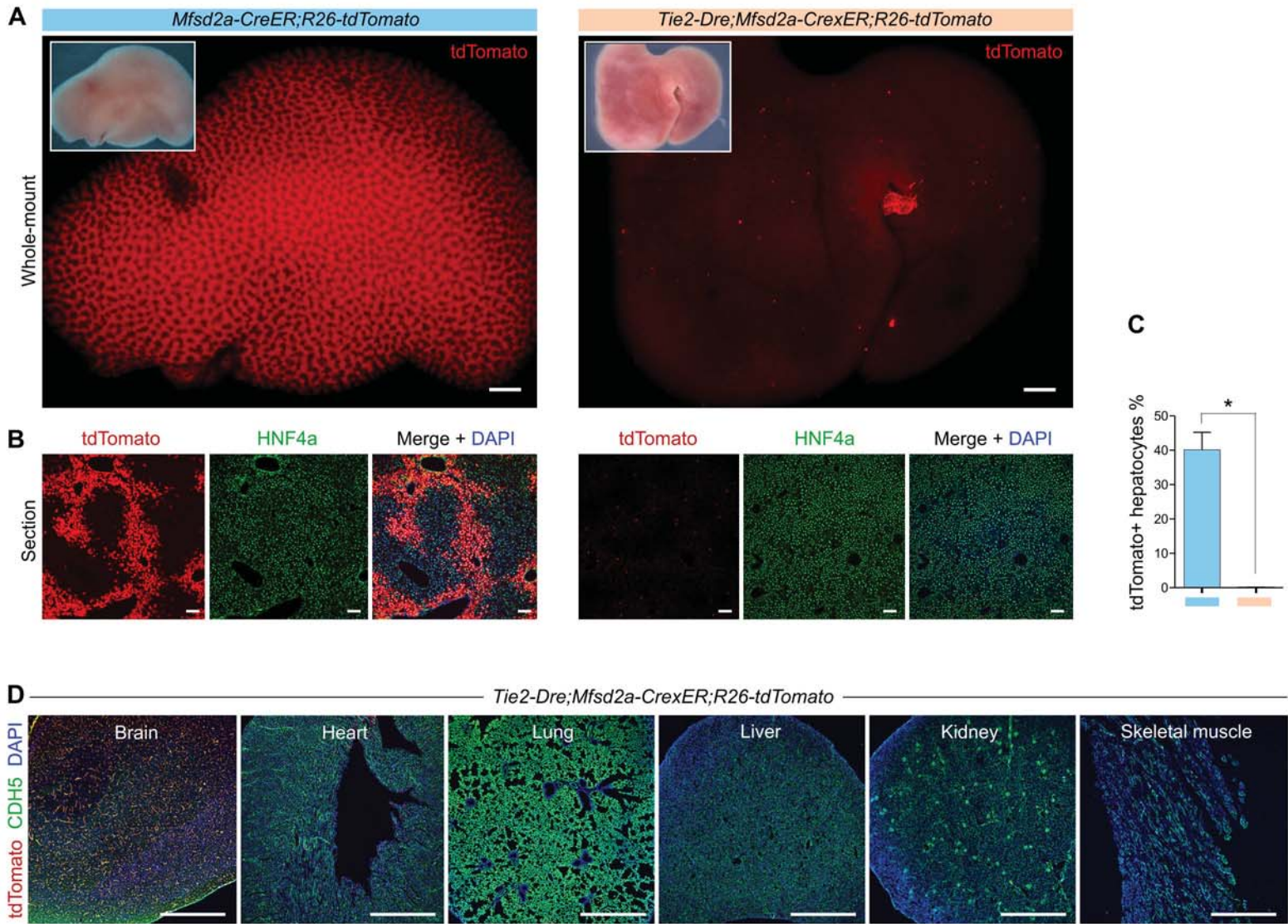
**Online Figure VIII. *Mfsd2a*<sup>+</sup> cells detected by *Mfsd2a-CreER* line.** **A**, Immunostaining for ESR (as surrogate for endogenous *Mfsd2a*) and CDH5 on *Mfsd2a-CreER* brain sections shows ESR<sup>+</sup>CDH5<sup>+</sup> BBB endothelial cells (arrowheads). **B**, Immunostaining for ESR and NeuN on *Mfsd2a-CreER* brain sections shows ESR<sup>+</sup>NeuN<sup>+</sup> neurons (arrows). **C**, Immunostaining for ESR and HNF4a on *Mfsd2a-CreER* liver sections shows ESR<sup>+</sup>HNF4a<sup>+</sup> hepatocytes. **D**, Immunostaining for ESR and E-Cad on small intestine section showed ESR<sup>+</sup>E-Cad<sup>+</sup> cells in villi. **E,F**, Immunostaining for tdTomato and CDH5 (**E**) or NeuN (**F**) on brain sections from *Mfsd2a-CreER;R26-tdTomato* mouse at 24-48 hours after tamoxifen injection. Arrowheads indicate *Mfsd2a*-expressing endothelial cells; arrows indicate *Mfsd2a*-expressing neurons. Boxed regions are magnified in the lower panels. **G**, Immunostaining for tdTomato and HNF4a on liver section shows *Mfsd2a*-expressing hepatocytes. **H**, Immunostaining for tdTomato and E-Cad on small intestine section shows *Mfsd2a*-expressing epithelial cells in villi. Scale bars, 100  $\mu$ m. Each image is representative of 5 individual samples.



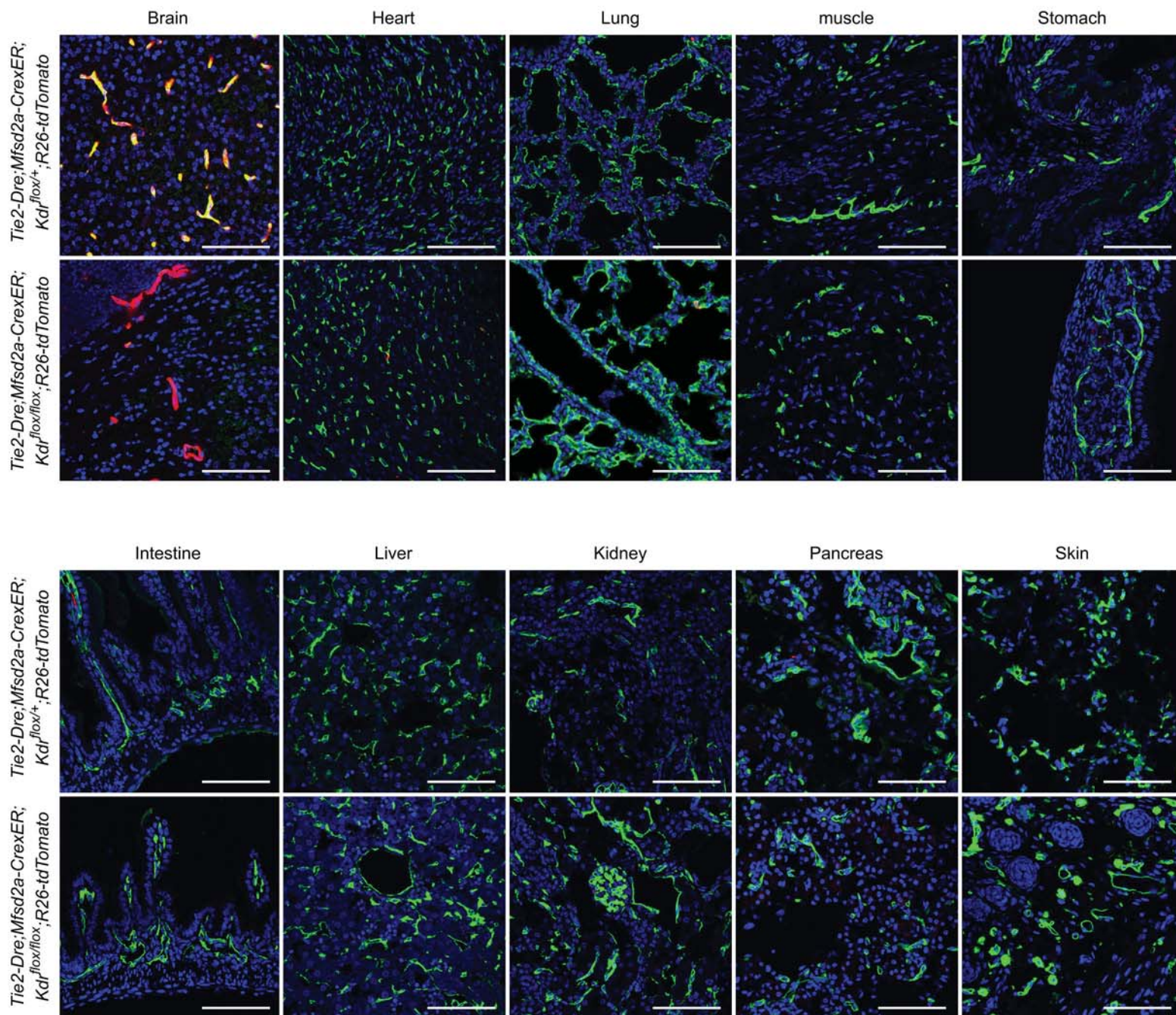
**Online Figure IX. Generation and characterization of *Mfsd2a-CrexER* mouse line.** **A**, Schematic showing knock-in strategy for *Mfsd2a-CrexER* allele by homologous recombination. **B**, Immunostaining for ESR (as surrogate for endogenous *Mfsd2a*) and HNF4a on liver sections of adult mouse. Arrows indicate ESR<sup>+</sup>HNF4a<sup>+</sup> hepatocytes. **C**, Immunostaining for ESR and CDH5 (left panel) or NeuN (right panel) on brain section. Arrowheads indicate ESR<sup>+</sup>CDH5<sup>+</sup> endothelial cells; arrows indicate ESR<sup>+</sup>NeuN<sup>+</sup> neurons. **D**, Whole-mount fluorescence views of organs collected from *Mfsd2a-CrexER;R26-tdTomato* adult mouse without tamoxifen treatment. Inserts are bright-field images of organs. **E**, Immunostaining for tdTomato and CDH5 on tissues sections from *Mfsd2a-CrexER;R26-tdTomato* adult mouse without tamoxifen treatment. Scale bars, 1 mm in D, 100  $\mu$ m in B,C,E.



**Online Figure X. Flow cytometric analysis of tdTomato<sup>+</sup> endothelial cells labeled by *BEC-Cre*.** Brain of neonatal *Mfsd2a-CrexER;R26-tdTomato* (A) and *Tie2-Dre;Mfsd2a-CrexER;R26-tdTomato* (B) mice were enzymatically dispersed for flow cytometric analysis. Single cells were gated by forward scatter. Dead cells were excluded by Violet Dye labelling. Endothelial cells were stained by CD31-APC antibody. RFP<sup>+</sup> cells were gated to analyze labeling efficiency. Each image is representative of 5 individual biological samples.



**Online Figure XI. More precise targeting by intersectional genetics.** **A,B**, Whole-mount fluorescence view of livers and immunostaining for tdTomato and HNF4a on liver sections from *Mfsd2a-CreER;R26-tdTomato* or *Tie2-Dre;Mfsd2a-CreER;R26-tdTomato* mice. For *Mfsd2a-CreER*, tamoxifen was induced at 6 weeks and samples were collected 2 days later. Inserts are bright-field images of livers. **C**, Quantification of labeled hepatocytes was shown on right panel. Data are mean  $\pm$  SEM.; \* $P < 0.05$ ;  $n = 5$ . **D**, Immunostaining for tdTomato and CDH5 on tissue sections from *Tie2-Dre;Mfsd2a-CreER;R26-tdTomato* mice. Scale bars, 1 mm in A,D; 100  $\mu$ m in B. Each image is representative of 5 individual samples.



**Online Figure XII. Expression of VEGFR2 in multiple organs.** Immunostaining for tdTomato and VEGFR2 on sections of multiple organs collected from P0 *Tie2-Dre;Mfsd2a-CrexER;Kdr<sup>flox/+</sup>;R26-tdTomato* (control) or *Tie2-Dre;Mfsd2a-CrexER;Kdr<sup>flox/flox</sup>;R26-tdTomato* (mutant) mice. VEGFR2 is not detected in the brain vessels, but remains in the vessels of other organs or tissues of mutant, in comparison with control. Scale bars, 100  $\mu$ m. Each image is representative of 5 individual samples.

Online Table I. DNA Sequence for Genotyping Primers

Name	Forward primer	Reverse primer	PCR length
<i>Nrg1-CrexER</i>	TGATTCAGCCTCTTTCAGCCG	GGATAGTTTTTACTGCCAGACCGC	868bp
<i>Nrg1-CreER</i>	TGAAGATACACCATTCTGGGC	TGCGAACCTCATCACTCGTTG	314bp
<i>CAG-Dre</i>	ACTCCTTGCCGATGTTCTCAG	TTGTCCCAAATCTGGCGGAG	415bp
<i>WT1-CrexER</i>	ATCTCCTAAAGTGACCCCGCAG	CCAGGTATGCTCAGAAAACGCC	896 bp
<i>Tie2-Dre</i>	TTCTCCTTGCCGCCAACTTG	ACTCCTTGCCGATGTTCTCAG	207bp
<i>Mfsd2a-CrexER</i>	GGACAGAAGCATTTTCCAGGTATG	TGATGATTGGTCTCGTCTGGCG	593bp
<i>Mfsd2a-CreER</i>	GGACAGAAGCATTTTCCAGGTATG	TGATGATTGGTCTCGTCTGGCG	587bp
<i>Kdr flox</i>	TGGAGAGCAAGGCGCTGCTAGC	CTTCCACTCCTGCCTACCTAG	439bp
<i>ACTB-Cre</i>	GCCTGCATTACCGGTCGATGC	CAGGGTGTTATAAGCAATCCC	481bp
<i>R26-LacZ</i>	AAAGTCGCTCTGAGTTGTTAT	GGAGCGGGAGAAATGGATAG	330bp
<i>R26-rox-LacZ</i>	TGAAATGTTACCAAGGAACT	TGACAGGAGATCCTGCCCGGCACT	716bp
<i>R26-tdTomato</i>	GGCATTAAAGCAGCGTATCC	CTGTTCTGTACGGCATGG	196bp
<i>R26-rox-tdTomato</i>	ACGGGTGTTGGGTCGTTTGTTT	TTCTTGTAATCGGGGATGTCGGCG	609bp
<i>R26-iDTR</i>	GGCTACTGCTGACTCTCAACATT	TCATGGTGGCGAATTCGAT	700bp

Note: bp, base pair.