## **Supplemental Material**

Aluminum fluoride-18 labeled folate enables *in vivo* detection of atherosclerotic plaque inflammation by positron emission tomography

Johanna M. U. Silvola<sup>1</sup>, Xiang-Guo Li<sup>1,2</sup>, Jenni Virta<sup>1</sup>, Päivi Marjamäki<sup>1</sup>, Heidi Liljenbäck<sup>1,3</sup>, Jarkko P. Hytönen<sup>4</sup>, Miikka Tarkia<sup>1</sup>, Virva Saunavaara<sup>5,6</sup>, Saija Hurme<sup>7</sup>, Senthil Palani<sup>1</sup>, Harri Hakovirta<sup>8</sup>, Seppo Ylä-Herttuala<sup>4,9</sup>, Pekka Saukko<sup>10</sup>, Qingshou Chen<sup>11</sup>, Philip S. Low<sup>11</sup>, Juhani Knuuti<sup>1,2,5</sup>, Antti Saraste<sup>1,5,12,13</sup> and Anne Roivainen<sup>1,3,5</sup>\*

<sup>1</sup>Turku PET Centre, University of Turku, Turku, Finland; <sup>2</sup>Turku PET Centre, Åbo Akademi University, Turku, Finland; <sup>3</sup>Turku Center for Disease Modeling, University of Turku, Turku, Finland; <sup>4</sup>A. I. Virtanen Institute for Molecular Sciences, University of Eastern Finland, Kuopio, Finland; <sup>5</sup>Turku PET Centre, Turku University Hospital, Turku, Finland; <sup>6</sup>Department of Medical Physics, Turku University Hospital, Turku, Finland; <sup>7</sup>Department of Biostatistics, University of Turku, Turku, Finland; <sup>8</sup>Department of Vascular Surgery, Turku University Hospital, Turku, Finland; <sup>9</sup>Science Service Center and Gene Therapy Unit, Kuopio University Hospital. Kuopio, Finland; <sup>10</sup>Department of Pathology and Forensic Medicine, University of Turku, Turku, Finland; <sup>11</sup>Department of Chemistry, Purdue University, West Lafayette, Indiana, USA; <sup>12</sup>Heart Center, Turku University Hospital, Turku, Finland; <sup>13</sup>Institute of Clinical Medicine, University of Turku, Turku, Finland

\*Correspondence: <u>anne.roivainen@utu.fi</u>



Figure S1. <sup>18</sup>F-FOL binds to macrophages derived from human blood monocytes. (A) Binding of <sup>18</sup>F-FOL on M1 macrophages (polarized with LPS and IFN- $\gamma$ ) and M2 macrophages (polarized with M-CSF, IL-4 and IL-10). Quantitative data are Becquerel (mean ± standard deviation, *n* = 3 triplicates). (B) Representative flow cytometric analyses of total CD68 (permeabilized cells) and surface CD206 (macrophage mannose receptor-1 [MRC-1]) from M1 and M2 macrophages. Black histograms are isotype controls and red histograms are for CD68 and CD206. MFI = mean fluorescence intensity.



**Figure S2. Uptake of <sup>18</sup>F-FOL in mice.** Uptake of <sup>18</sup>F-FOL in aortic arches of atherosclerotic LDLR<sup>-/-</sup> ApoB<sup>100/100</sup> (n = 9), healthy C57BL/6N controls (n = 6), and atherosclerotic mice with a 100-fold molar excess of folate glucosamine (FG) (n = 3), as assessed by *in vivo* PET/CT imaging. TBR = maximum target-to-background ratio, SUV<sub>max, aortic arch/SUV<sub>mean, blood</sub>. SUV<sub>max</sub> = maximum standardized uptake value determined at 60–90 min post <sup>18</sup>F-FOL injection.</sub>



**Figure S3. Mouse aorta** <sup>18</sup>**F-FOL PET.** *Ex vivo* obtained examples of <sup>18</sup>F-FOL PET images (top) and photographs of the aortas stained with Oil-Red-O (bottom) from atherosclerotic LDLR<sup>-/-</sup>ApoB<sup>100/100</sup> mouse (n = 1), atherosclerotic mouse with a 100-fold excess of folate glucosamine (blocking) (n = 1), and healthy C57BL/6N control mouse (n = 1).



Figure S4. Histology and immunohistochemistry of atherosclerotic lesions in LDLR<sup>-/-</sup>ApoB<sup>100/100</sup> mice. (A) Representative aortic root sections were stained with Movat's pentachrome (black = nuclei; yellow = collagen, reticular fibers; blue = ground substance, mucin; bright red = fibrin; red = muscle) or with anti-mouse Mac-3 (macrophages), iNOS (M1 polarized macrophages) and MRC-1 (M2 polarized macrophages) immunohistochemistry. The high-power views are of the area within the black rectangle on the left images. Mac-3, iNOS and MRC-1 positive cells appear brown in color. L = lumen; P = plaque; W = healthy vessel wall. Please note that the Movat and Mac-3 images are the same as in

Figure 3D. (B) Quantitative results of immunohistochemistry for the detection of macrophage markers (mean  $\pm$  SD).



**Figure S5. Representative HPLC chromatograms of** <sup>18</sup>**F-FOL, the reference** <sup>19</sup>**F-FOL, and precursor NOTA-folate.** (A) Radioactivity and (B) UV detection of <sup>18</sup>F-FOL (the UV peak [arrow] was low because of the high specific radioactivity of <sup>18</sup>F-FOL), (C) UV detection of <sup>19</sup>F-FOL, (D) radioactivity and (E) UV detection of <sup>18</sup>F-FOL spiked with <sup>19</sup>F-FOL, and (F) UV detection of NOTA-folate.



Figure S6. High-resolution mass spectrometry analysis of <sup>19</sup>F-FOL. The theoretical monoisotopic mass for ion  $[M+H]^+ C_{37}H_{51}AlFN_{12}O_{12}$  was 901.3544 and the observed mass was 901.3547.