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Life Sciences Reporting Summary

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Please do not complete any field with "not applicable" or n/a. Refer to the help text for what text to use if an item is not relevant to your study. For final submission: please carefully check your responses for accuracy; you will not be able to make changes later.

Experimental design

1. Sample size

Describe how sample size was determined.

Previous studies and pilot experiments form the basis of power calculations for the various studies. Depending on the experiment, calculations indicated that 3-12 mice per group would enable the testing of our hypotheses based on an expected 25-30% coefficient of variation and an 80% chance of detecting a 33% difference in the key specified endpoints (P = 0.05)

2. Data exclusions

Describe any data exclusions.

Pre-specified exclusion criteria were weight loss>10% of initial body weight or signs of illness or injury requiring euthanasia. According to these pre-specified criteria, the maximum number of mice removed before analysis was 3, but more typically 0-2.

3. Replication

Describe the measures taken to verify the reproducibility of the experimental findings.

All experiments were reproducible as assessed in multiple wells of cells, tissue samples or mice. We did not have cases of irreproducibility. Please note that the bar graphs depicting spectral counts (Ext. Data Fig. 2g and Ext. Data Fig. 8a) simply represent a single MS/MS experiment, and likewise the FPLC fractions (Ext. Data Fig. 2f and Ext. Data Fig. 7c) were screened using technical duplicates, but all of these data were subjected to extensive reproducibility in all the subsequent biological assays related to these data.

4. Randomization

Describe how samples/organisms/participants were allocated into experimental groups.

Mice of the same age and similar weight were randomly assigned to experimental and control groups. On occasion, we analyzed a subset of mice for a particular parameter, and the subset was chosen randomly from a full cohort.

5. Blinding

Describe whether the investigators were blinded to group allocation during data collection and/or analysis. The investigators were not blinded during cell and mouse experiment assays.

Note: all in vivo studies must report how sample size was determined and whether blinding and randomization were used.

For all figures and tables that use statistical methods, cor Methods section if additional space is needed).	ifirm that the following items are present in relevant figure legends (or in the
n/a Confirmed	
The exact sample size (n) for each experimental group/o	condition, given as a discrete number and unit of measurement (animals, litters, cultures, etc.)
A description of how samples were collected, noting sample was measured repeatedly	g whether measurements were taken from distinct samples or whether the same
A statement indicating how many times each experi	ment was replicated
The statistical test(s) used and whether they are one Only common tests should be described solely by name; de	e- or two-sided scribe more complex techniques in the Methods section.
A description of any assumptions or corrections, such	ch as an adjustment for multiple comparisons
Test values indicating whether an effect is present Provide confidence intervals or give results of significance in	tests (e.g. P values) as exact values whenever appropriate and with effect sizes noted.
A clear description of statistics including central ten	dency (e.g. median, mean) and variation (e.g. standard deviation, interquartile range)
Clearly defined error bars in <u>all</u> relevant figure capti	ons (with explicit mention of central tendency and variation)
See the web collection on st	atistics for biologists for further resources and guidance.
► Software	
Policy information about availability of computer code	
7. Software	
Describe the software used to analyze the data in this study.	Image J for density ratio measurements; Proteome Discoverer 1.4
	central to the paper but not yet described in the published literature, software must be made courage code deposition in a community repository (e.g. GitHub). <i>Nature Methods</i> guidance for er information on this topic.
► Materials and reagents	
Policy information about availability of materials	
8. Materials availability	
Indicate whether there are restrictions on availability of unique materials or if these materials are only available for distribution by a third party.	No restrictionsall mutant mice are commercially available or available from the labs indicated in the manuscript, and the methods for GeRP construction is published.
9. Antibodies	
Describe the antibodies used and how they were validated for use in the system under study (i.e. assay and species).	The source and catalogue number of all antibodies appear in Methods. Antibodies were validated by showing lack of signal after knockout or siRNA or by a single major band at the proper MW on immunoblot.
10. Eukaryotic cell lines	
a. State the source of each eukaryotic cell line used.	N/A
b. Describe the method of cell line authentication used.	N/A
 Report whether the cell lines were tested for mycoplasma contamination. 	N/A
d. If any of the cell lines used are listed in the database of commonly misidentified cell lines maintained by ICLAC, provide a scientific rationale for their use.	N/A
▶ Animals and human research participar	its

Policy information about studies involving animals; when reporting animal research, follow the ARRIVE guidelines

11. Description of research animals

6. Statistical parameters

Provide all relevant details on animals and/or animal-derived materials used in the study.

All details appear under "Mouse experiments" section in methods

Policy information about studies involving human research participan	Policy	v information	about	studies	involving	human	research	particin	oan [.]
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12.	Description of human research participants
	Describe the covariate-relevant population
	characteristics of the human research participants.

N/A	
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