

Supplementary Materials for

Bioactive polyamine production by a novel hybrid system comprising multiple indigenous gut bacterial strategies

Yusuke Kitada, Koji Muramatsu, Hirokazu Toju, Ryoko Kibe, Yoshimi Benno, Shin Kurihara, Mitsuharu Matsumoto

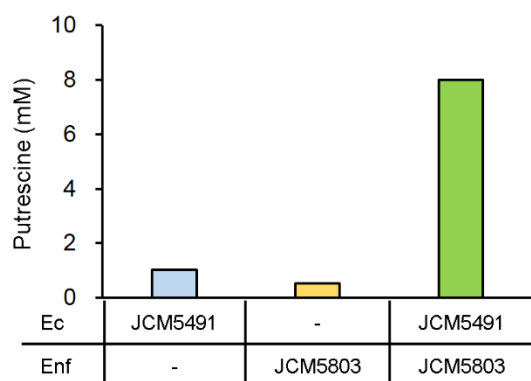
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This PDF file includes:

- fig. S1. Induction of putrescine production by coculture of *E. coli* and *En. faecalis*.
- fig. S2. Effects of extracellular pH and glucose concentration on bacterial metabolism in monocultures of *En. faecalis* and *E. coli*.
- fig. S3. Viable bacterial counts and change of extracellular pH in cocultures of putrescine-deficient *E. coli* (SK930), wild-type *En. faecalis* (V583), and each *Bifidobacterium* sp.
- fig. S4. Viable bacterial counts in feces of gnotobiotic mice.
- fig. S5. Extracellular putrescine concentration and total viable bacterial counts in human feces incubated at different pH values ($n = 5$).
- fig. S6. Effect of pH on extracellular agmatine concentration in monocultures of *E. coli*, *Ci. youngae*, and *F. varium*.
- fig. S7. Symbiont-symbiont co-occurrence networks of key genes in the putrescine production pathway using previously described human microbiome data from U.S. metropolitan areas.
- fig. S8. Symbiont-symbiont co-occurrence network patterns of key genes in the putrescine production pathway using previously described human microbiome data from Venezuela.
- fig. S9. Symbiont-symbiont co-occurrence network patterns of key genes in the putrescine production pathway using previously described human microbiome data from Malawi.
- fig. S10. Mechanistic model of a novel pathway for putrescine production from arginine through agmatine via the collaboration of three different bacterial species.

- fig. S11. Outline of gnotobiotic mouse experiments.
- table S1. List of species expressing homologs of the enzymes AdiA and AdiC, as determined by in silico analyses of 126 bacterial strains present in the human gut.
- table S2. Detection of RNA sequences of AdiA, AdiC, AguA, and AguD by metatranscriptomic analysis of human feces.
- table S3. List of bacteria used for screening of polyamine producing bacteria.
- table S4. Strains, plasmids, and oligonucleotides used in this study.

A



B

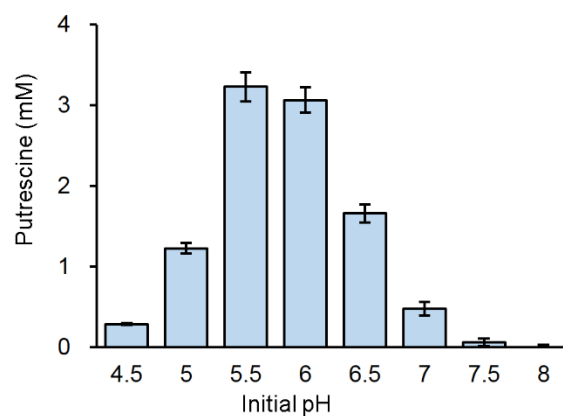


Fig. S1. Induction of putrescine production by coculture of *E. coli* and *En. faecalis*.

(A) Extracellular putrescine concentration in monocultures of *E. coli* JCM5491 and *En. faecalis* JCM 5803^T, and a coculture of both strains. Error bars represent standard errors (SEs). Ec: *E. coli*, Enf: *En. faecalis*. (B) Effect of pH on extracellular putrescine concentration in cocultures of *E. coli* and *En. faecalis*.

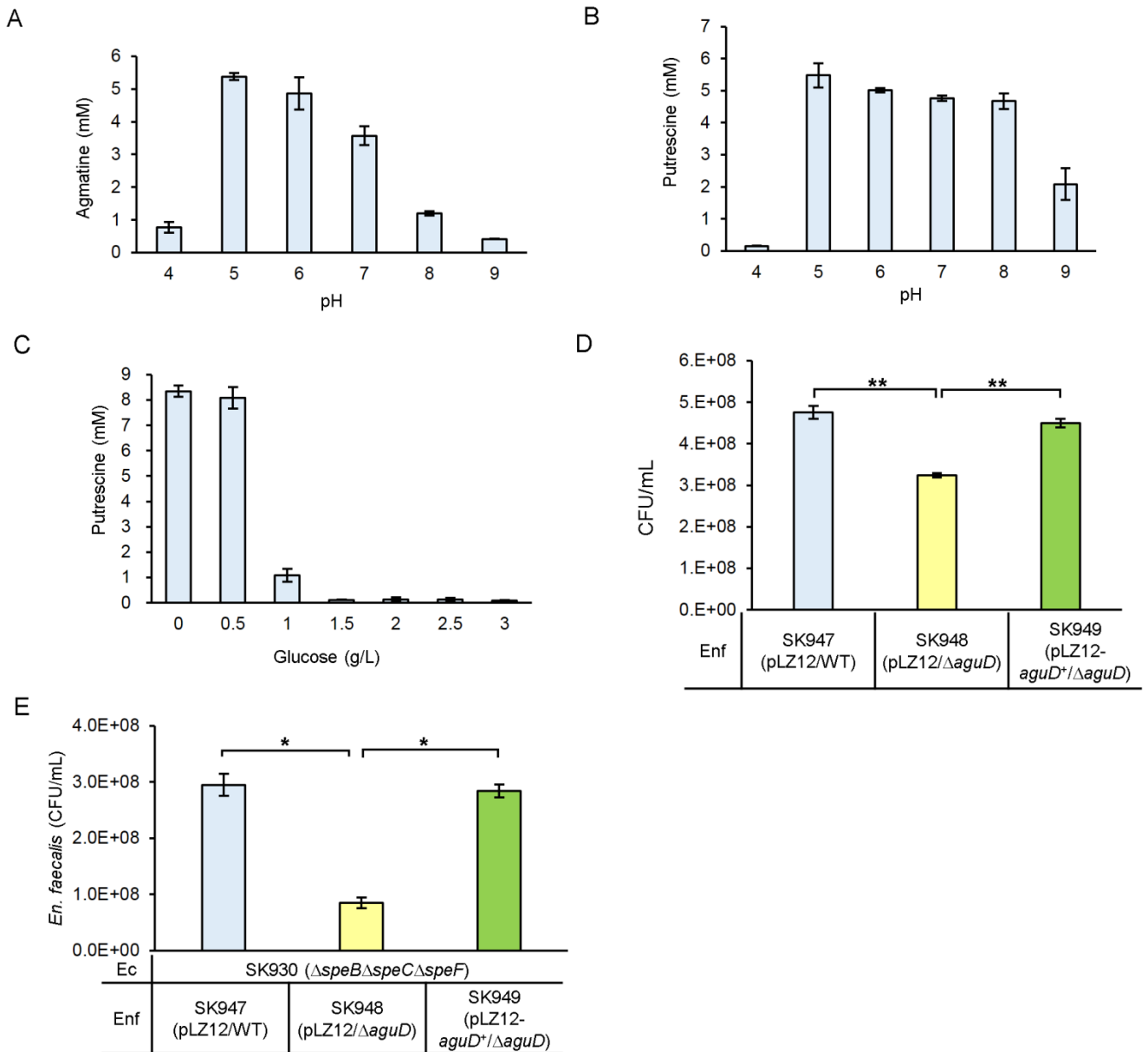


Fig. S2. Effects of extracellular pH and glucose concentration on bacterial metabolism in monocultures of *En. faecalis* and *E. coli*

(A) Extracellular agmatine concentration in monocultures of wild-type *E. coli* (MG1655) in LB-RGC at different pH. (B) Extracellular putrescine concentrations in monocultures of wild-type *En. faecalis* (V583) cultured in medium at different pH values containing 5 mM agmatine. (C) Extracellular putrescine concentration in a monoculture of wild-type *En. faecalis* (V583) in medium containing 5 mM agmatine and different concentrations of glucose. (D) Viable cell count of wild-type *En. faecalis* (SK947), *aguD*-deleted *En. faecalis* (SK948), or *aguD*-complemented *En. faecalis* (SK949) monocultured in medium containing 5 mM agmatine at pH 6. (E) Viable cell counts of *En. faecalis* when putrescine-deficient *E. coli* (SK930) was cocultured with wild-type *En. faecalis* (SK947), *aguD*-deleted *En. faecalis* (SK948), or *aguD*-complemented *En. faecalis* (SK949) in LB-RGC medium. Mono- and cocultures of these bacteria were conducted under anaerobic conditions at 37°C for 24 h. Error bars represent SEs. Student's *t*-tests; **p* < 0.05, ***p* < 0.01. Enf: *En. faecalis*

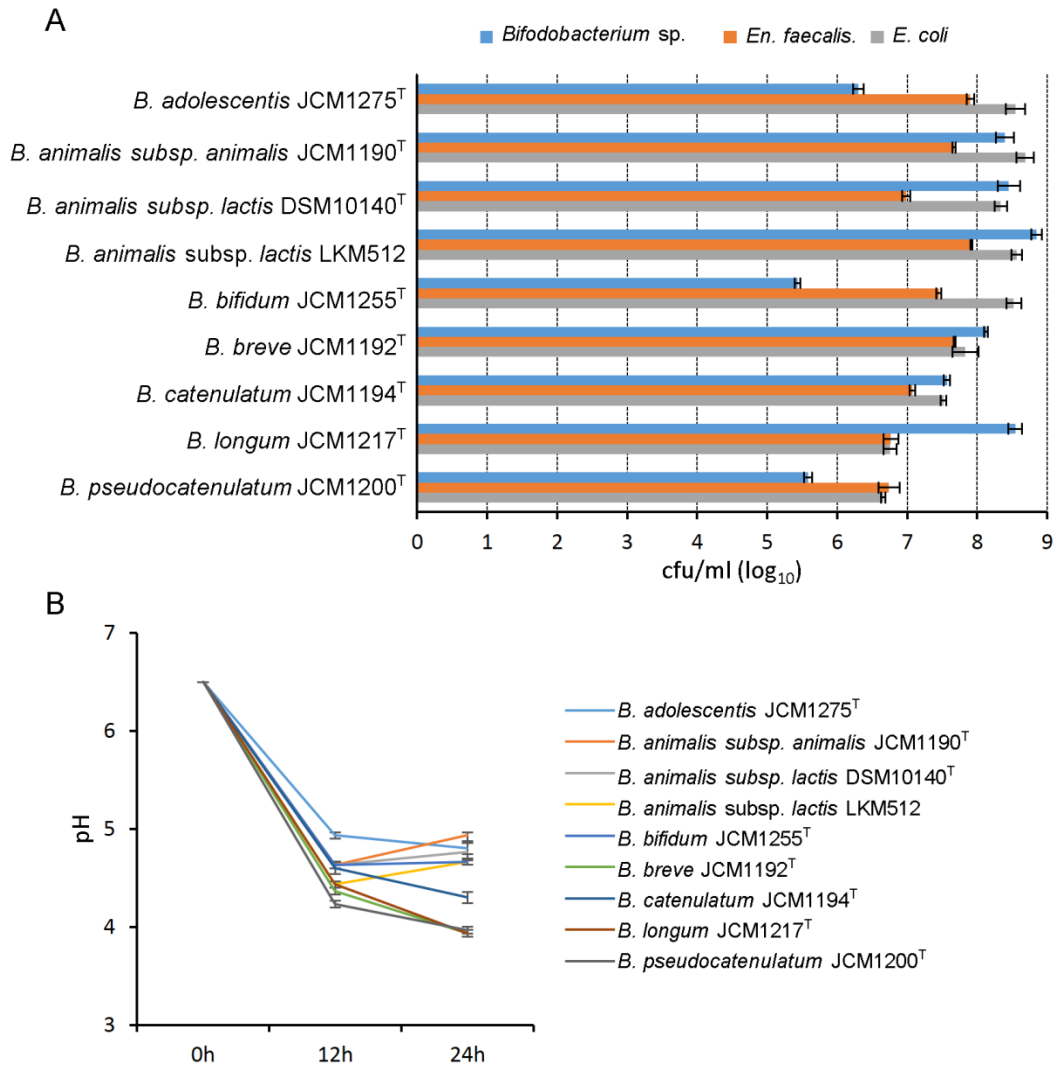


Fig. S3. Viable bacterial counts and change of extracellular pH in cocultures of putrescine-deficient *E. coli* (SK930), wild-type *En. faecalis* (V583), and each *Bifidobacterium* sp.

(A) Viable bacterial counts. Coculture of these bacteria was conducted under anaerobic conditions at 37 °C for 24 h in LB medium containing 5 mM L-arginine, 1.5 g/L D-glucose, 5 g/L galacto-oligosaccharide, 2 mM MgSO₄, 60 mM NH₄Cl, and 0.5 g/L L-cysteine-hydrochloride (pH 6.5). (B) Change of extracellular pH.

The cultures of all strains reached pH 5.5, at which point, maximal putrescine production occurred within 12 hours. However, there was no relationship between the viable count of *Bifidobacterium* sp. and pH. Interestingly, we found that the extracellular putrescine concentration positively correlated with the number of both *E. coli* and *En. faecalis*. Specifically, in the culture with *B. animalis* subsp. *lactis* LKM512 or *B. adolescentis* JCM1275, which yielded higher putrescine production, larger numbers of *E. coli* (10^{8.5} cfu/ml or more) and *En. faecalis* (10^{7.9} cfu/ml or more) were detected, whereas in the culture with *B. longum* JCM1217 or *B. pseudocatenulatum* JCM1200, which induced only slight putrescine production, 10-times lower number of *E. coli* and *En. faecalis* (less than 10^{6.8} cfu/ml) were detected.

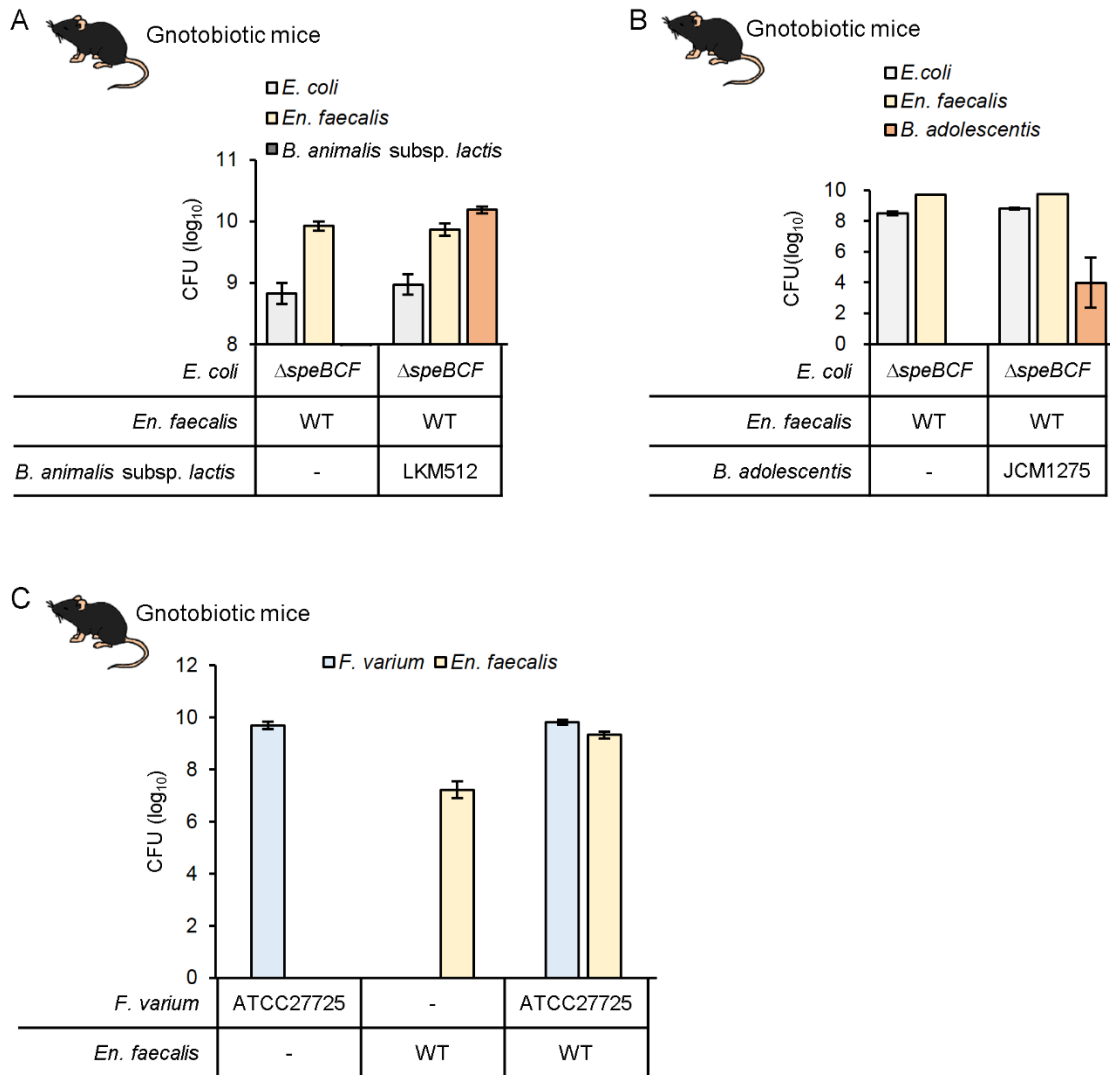


Fig. S4. Viable bacterial counts in faeces of gnotobiotic mice

(A) Viable bacterial counts in feces derived from gnotobiotic mice colonized with putrescine-deficient *E. coli* (SK930), wild-type *En. faecalis* (V583), and *B. animalis subsp. lactis* LKM512. (B) Viable bacterial counts in feces derived from gnotobiotic mice colonized with putrescine-deficient *E. coli* (SK930), wild-type *En. faecalis* (V583), and *B. adolescentis* JCM1275^T. (C) Viable bacterial counts in feces derived from gnotobiotic mice inoculated with *F. varium* ATCC 27725 and wild-type *En. faecalis* (V583). Error bars represent SEs.

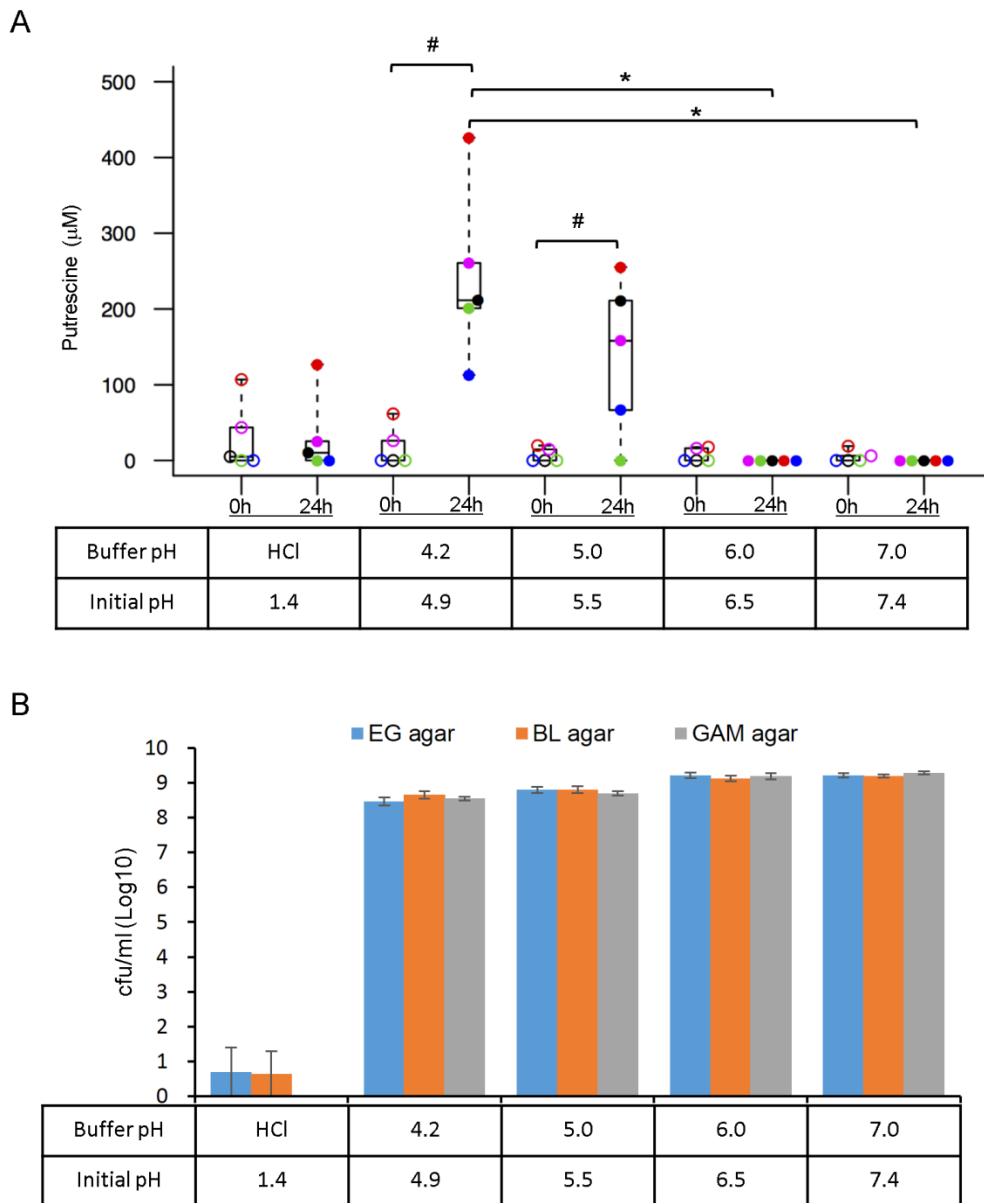


Fig. S5. Extracellular putrescine concentration and total viable bacterial counts in human feces incubated at different pH values (n = 5)

(A) Extracellular putrescine concentration. Same color plots indicate the same individual. Open and closed symbols indicate the putrescine concentration before and after incubation, respectively. (B) Total bacterial counts at 24 h were measured by cultivation on blood-liver (BL) agar plate, Eggerth-Gagnon (EG) agar plate, and GAM agar plate. Steel-Dwass test; * $p < 0.05$, Wilcoxon signed rank test; # $p < 0.05$.

Five minutes after suspension (0 h of incubation), putrescine concentration was higher at pH 1.4 (31 μM) and pH 4.9 (18 μM) than that at pH 5.5, 6.5, and 7.4 (approximately 6 μM), indicating that bacterial cells were ruptured and a small amount of putrescine was released into the suspension at pH 1.4 and pH 4.9. Furthermore, after incubation for 24 h, at pH 4.2, putrescine concentration increased to 12-times the concentration (250 μM) observed before incubation, and the increase was observed in all samples, confirming that putrescine was produced by intestinal bacteria. In contrast, putrescine concentration was not increased at pH 1.4 because almost all bacteria were dead.

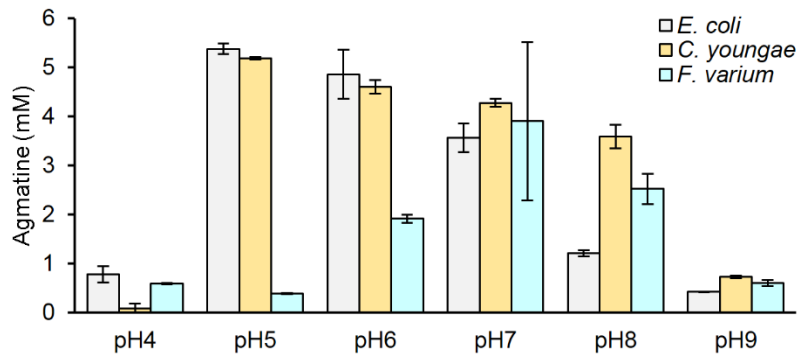
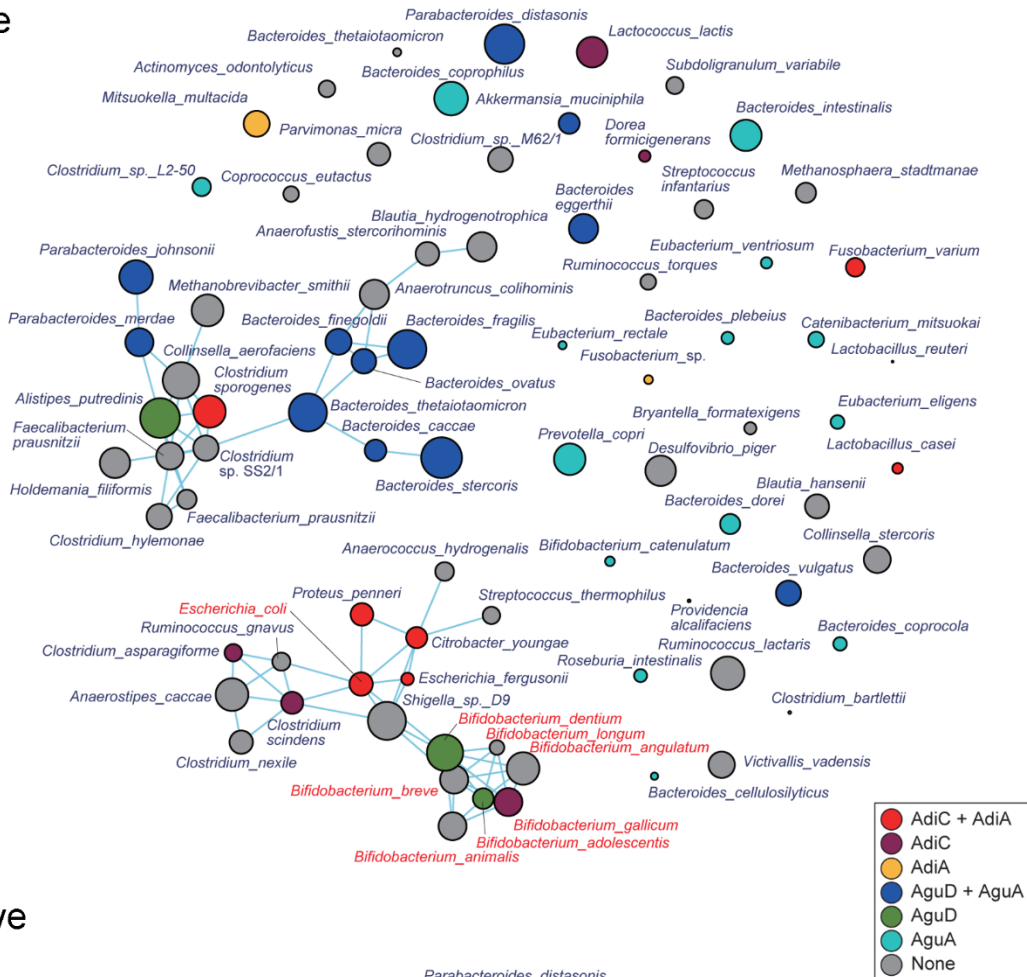


Fig. S6. Effect of pH on extracellular agmatine concentration in monocultures of *E. coli*, *C. youngae*, and *F. varium*.

Monoculture of these bacteria was conducted under anaerobic conditions at 37 °C for 24 h in LB-RGC (pH 4.0–9.0). *C. youngae* produced agmatine due to pH decrease similar to *E. coli*, indicating the presence of bacteria possessing the AdiC/AdiA pathway, which is triggered by environmental acidification in the gut, whereas *F. varium* did not show the same trend, and the highest agmatine production was observed at neutral pH (pH 7). These findings indicate that agmatine production does not depend on environmental acidification

Positive

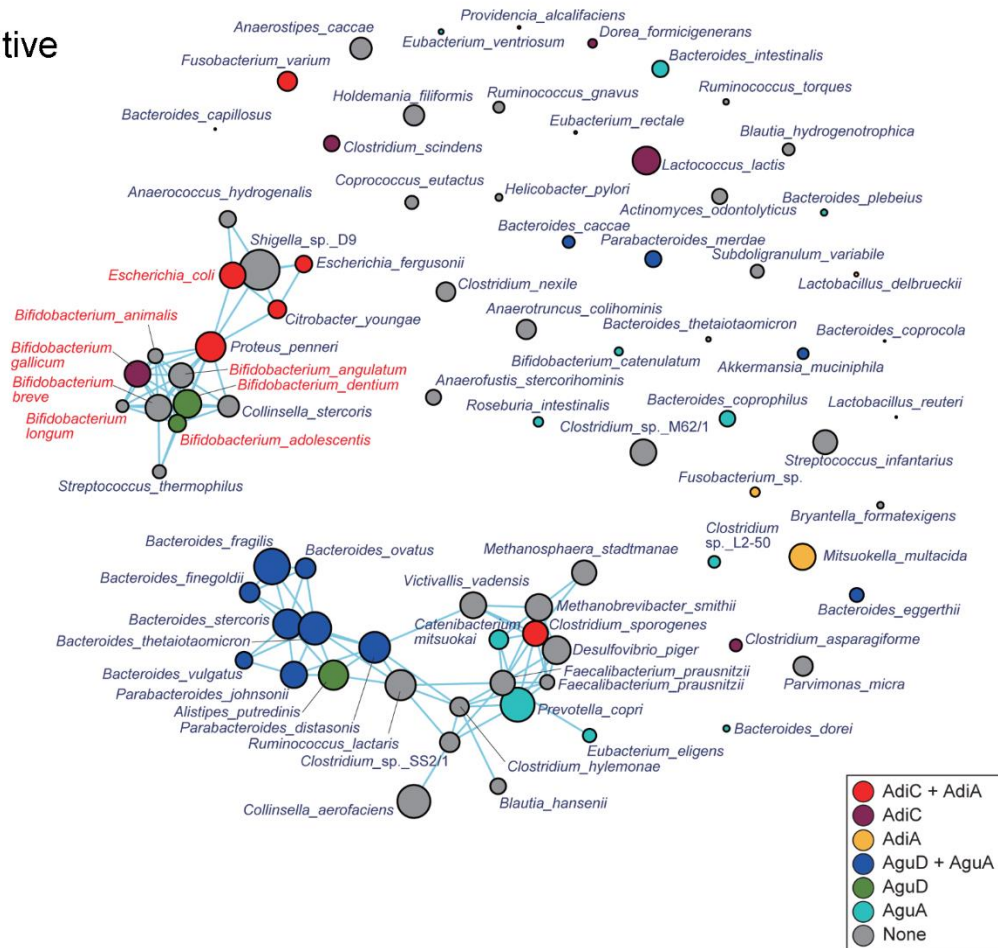


Negative



Fig. S7. Symbiont-symbiont co-occurrence networks of key genes in the putrescine production pathway using previously described human microbiome data from US metropolitan areas . Details are shown in Fig. 5C

Positive



Negative

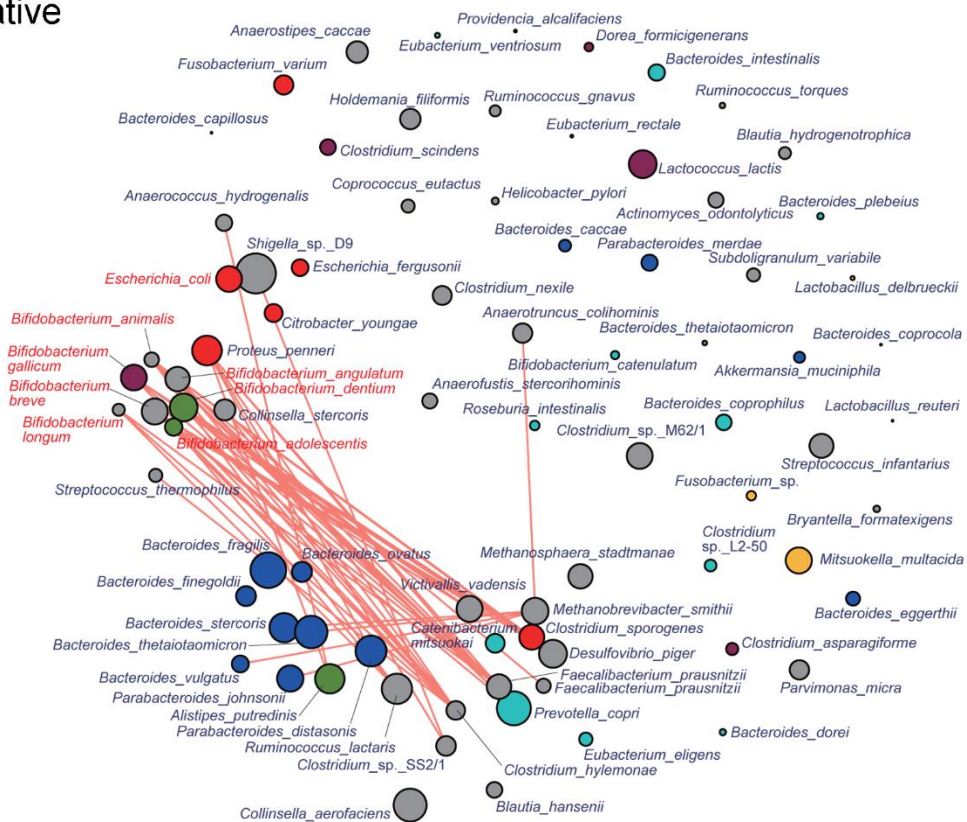
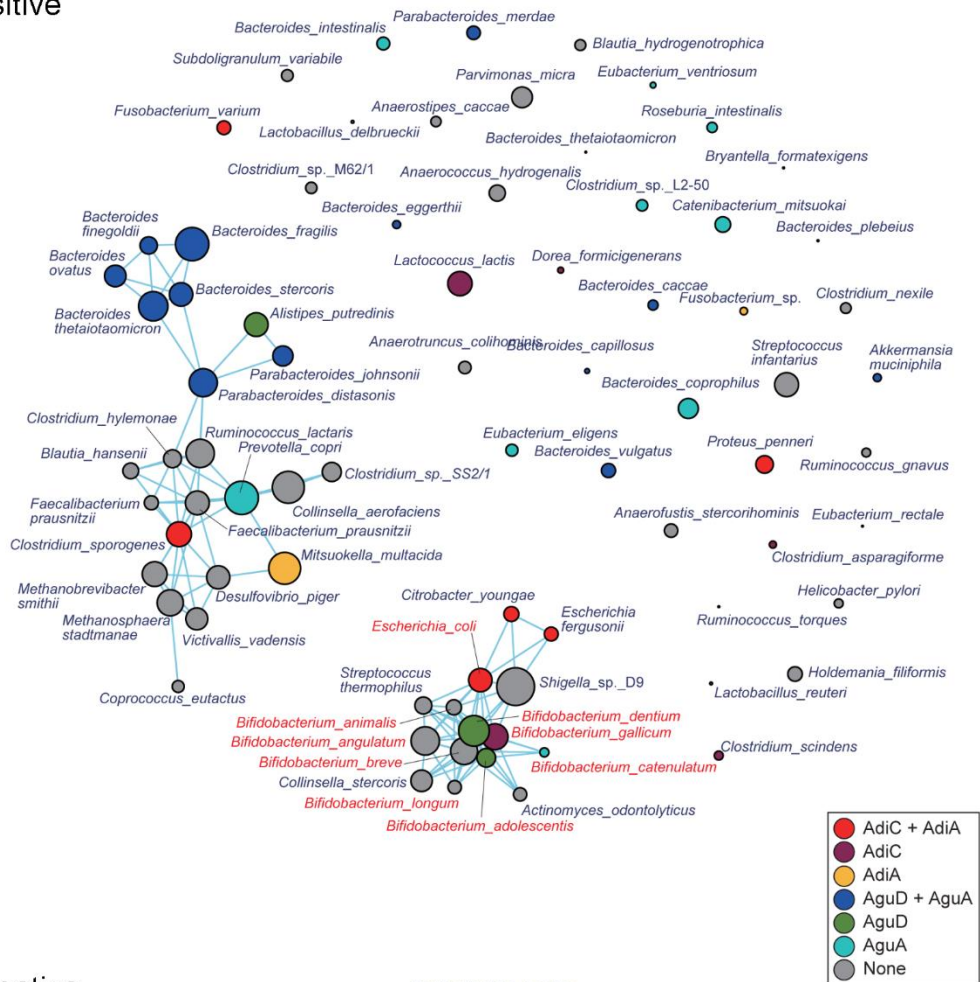


Fig. S8. Symbiont-symbiont co-occurrence network patterns of key genes in the putrescine production pathway using previously described human microbiome data from Venezuela. Details are shown in Fig. 5C

Positive



Negative



Fig. S9. Symbiont-symbiont co-occurrence network patterns of key genes in the putrescine production pathway using previously described human microbiome data from Malawi. Details are shown in Fig. 5C.

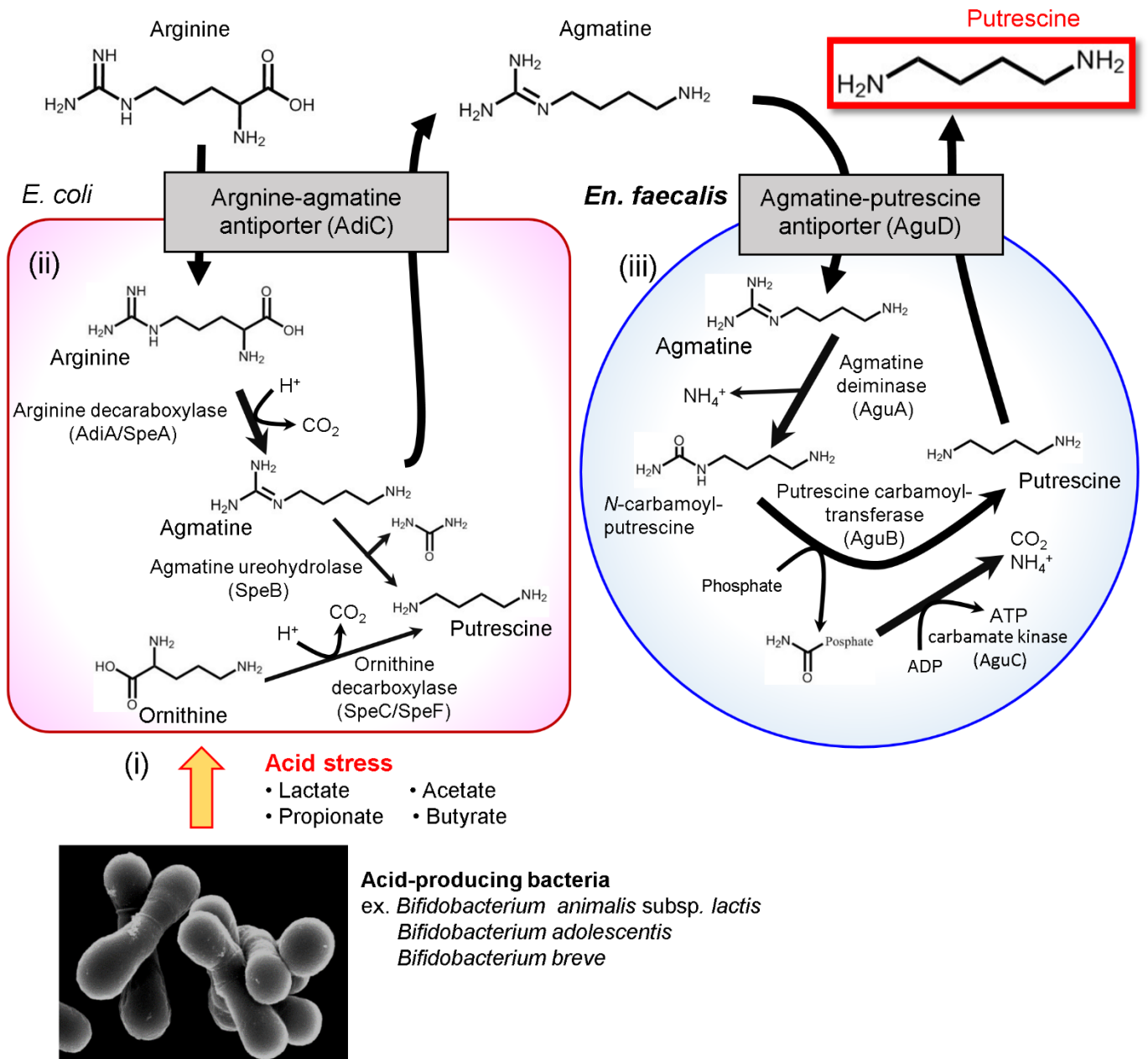


Fig. S10. Mechanistic model of a novel pathway for putrescine production from arginine through agmatine via the collaboration of three different bacterial species

(i) This pathway is triggered by environmental acidification due to acetate and lactate produced by acid-producing bacteria, represented by *Bifidobacterium* spp. (ii) In the second step, the acid-tolerance system of *E. coli* (arginine-dependent acid resistance system) is activated by acidic stress to exchange agmatine inside the cell for arginine present outside. (iii) In the third step, the energy production system of *En. faecalis* (agmatine deiminase system) is activated to exchange putrescine inside the cell for agmatine present outside (owing to the acid-tolerance system of *E. coli*). Putrescine is a byproduct of the collaboration between these different bacterial species

Abbreviations: Arg, arginine; Agm, agmatine; Put, putrescine; AdiA, arginine decarboxylase; AdiC, arginine/agmatine antiporter; AguA, agmatine deiminase; AguB, putrescine carbamoyltransferase; AguC, carbamate kinase; AguD, agmatine/putrescine antiporter.

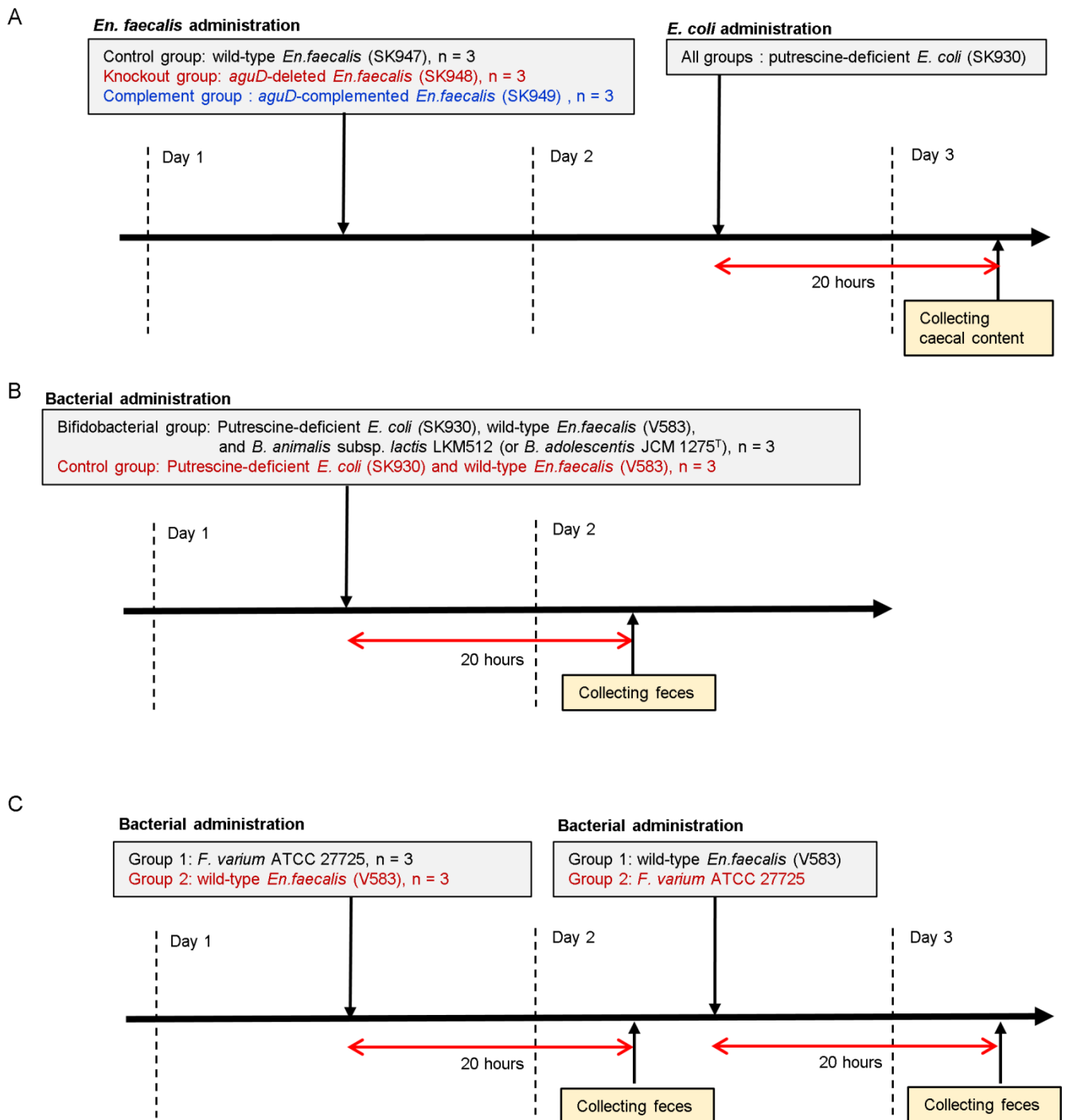


Fig. S11. Outline of gnotobiotic mouse experiments

(A) Gnotobiotic mice inoculated with putrescine-deficient *E. coli* (SK930) and *En. faecalis* gene knockout mutants or complementary transformants of the agmatine-putrescine antiporter (*aguD*). (B) Gnotobiotic mice inoculated with putrescine-deficient *E. coli* (SK930), wild-type *En. faecalis* (V583), and *B. animalis* subsp. *lactis* LKM512 (or *B. adolescentis*). (C) Gnotobiotic mice inoculated with *F. varium* ATCC27725 and wild-type *En. faecalis* (V583).

Table S1. List of species expressing homologs of the enzymes AdIA and AdIC, as determined by in silico analyses of 126 bacterial strains present in the human gut

	AdiC		AdIA		SpeA		AguA		
	Bit	Accession ID	Bit	Accession ID	Bit	Accession ID	Bit	Accession ID	
<i>Actinomyces odontolyticus</i> ATCC 17982									
<i>Bifidobacterium adolescentis</i> ATCC 15703						111	gb EDN82401.1		
<i>Bifidobacterium adolescentis</i> L 2-37									
<i>Bifidobacterium animalis</i> DSM 20089									
<i>Bifidobacterium animalis</i> subsp. lactis AD011									
<i>Bifidobacterium animalis</i> subsp. lactis HN019									
<i>Bifidobacterium breve</i> DSM 20213									
<i>Bifidobacterium catenulatum</i> DSM 16992							189	ref WP_033513568.1	
<i>Bifidobacterium celeratum</i>						107	ref WP_003839068.1		
<i>Bifidobacterium gallicum</i> DSM 20093	105	gb KF59536.1							
<i>Bifidobacterium longum</i> DJ010A									
<i>Bifidobacterium longum</i> NCC2705									
<i>Bifidobacterium pseudocatenulatum</i> DSM 20438									
<i>Collinsella aerofaciens</i> ATCC 25986									
<i>Collinsella intestinalis</i> DSM 13280									
<i>Collinsella stercoris</i> DSM 13279						103	ref WP_004329135.1		
<i>Alistipes putredinis</i> DSM 17216									
<i>Bacteroides caecae</i> ATCC 43185					545	ref NP_076979213.1	168	gb EDM22213.1	
<i>Bacteroides capillus</i> ATCC 23799					545	ref NP_007786434.1	108	gb EDM21916.1	
<i>Bacteroides cellulosilyticus</i> DSM 14838					531	ref NP_007710943.1	102	ref WP_007209680.1	
<i>Bacteroides coprococcus</i> DSM 17136					529	gb EEDV02459.1	176	gb EEF87524.1	
<i>Bacteroides coprohilus</i> DSM 18228					531	gb EEF76458.1	155	gb EEDU99579.1	
<i>Bacteroides dorei</i> DSM 17855					538	gb EEB25237.1	132	gb EEF74901.1	
<i>Bacteroides equorhi</i> DSM 20697					538	gb EEB25237.1	129	gb EEB24739.1	
<i>Bacteroides finlayi</i> DSM 17565					541	ref WP_004291990.1	103	gb EEC53335.1	
<i>Bacteroides fragilis</i> 3 1 12					545	ref NP_007756434.1	107	gb EEX43976.1	
<i>Bacteroides fragilis</i> NCTC 9343					537	emb CAH05973.1	106	emb CAH06244.1	
<i>Bacteroides fragilis</i> YCH46					537	ref NP_087522.1	107	ref NP_087522.1	
<i>Bacteroides intestinalis</i> DSM 17393					533	gb EEDV06321.1	159	ref YP_099665.1	
<i>Bacteroides ovatus</i> ATCC 8483					547	gb EDO12412.1	104	gb EDO14173.1	
<i>Bacteroides plebeius</i> DSM 17135					530	gb EEDV97001.1	162	gb EEX45314.1	
<i>Bacteroides</i> sp. 11 6									
<i>Bacteroides</i> sp. D1									
<i>Bacteroides</i> sp. D2									
<i>Bacteroides stercoris</i> ATCC 43183					541	gb EDS14810.1	105	gb EDS16535.1	
<i>Bacteroides thetaioamicron</i> 3731									
<i>Bacteroides thetaioamicron</i> 7330									
<i>Bacteroides thetaioamicron</i> VPI-5482					550	ref NP_812306.1	108	ref NP_811486.1	
<i>Bacteroides uniformis</i> ATCC 8492					527	gb EDO5269.1	147	ref NP_809789.1	
<i>Bacteroides vulgatus</i> ATCC 8482					538	gb ABR39026.1	100	gb ABR41504.1	
<i>Bacteroides</i> WH2									
<i>Bacteroides xylosoxydans</i> XB1A					549	emb CBK67757.1	106	emb CBK66780.1	
<i>Parabacteroides distasonis</i> ATCC 8503							108	ref NP_005857294.1	
<i>Parabacteroides johnsonii</i> DSM 18315					538	gb EEC95655.1	111	gb EEC97972.1	
<i>Parabacteroides merdae</i> ATCC 43184					541	gb EDN84739.1	112	gb EDN84739.1	
<i>Prevotella copri</i> DSM 18205					519	gb EFB33967.1	112	gb EDN84739.1	
<i>Methanobrevibacter smithii</i> ATCC 35061									
<i>Methanobrevibacter smithii</i> DSM 2374									
<i>Methanobrevibacter smithii</i> DSM 2375									
<i>Methanosphaera stadtmanae</i> DSM 3091									
<i>Anaerococcus hydrogenalis</i> DSM 7454									
<i>Anaerofustis stercorihominis</i> DSM 17244									
<i>Anaerostipes caccæ</i> DSM 14662									
<i>Anaerotruncus colibiformis</i> DSM 17241									
<i>Blautia hansenii</i> DSM 20583									
<i>Blautia hydroxynitrohalica</i> DSM 10507									
<i>Butyrivibrio formatorgens</i> DSM 14469									
<i>Butyrivibrio crossotus</i> DSM 2876								301	gb EFF68065.1
<i>Catenibacterium mitsuokai</i> DSM 15897								338	gb EFF93927.1
<i>Clostridium asparaniforme</i> DSM 15981	105	gb EEG57719.1							
<i>Clostridium bartlettii</i> DSM 16795									
<i>Clostridium botulinum</i> ATCC BAA-113									
<i>Clostridium hiranonis</i> DSM 13275									
<i>Clostridium tylenoneae</i> DSM 15053									
<i>Clostridium leptum</i> DSM 753									
<i>Clostridium methyloxylosum</i> DSM 5476									
<i>Clostridium nexile</i> DSM 1787									
<i>Clostridium ramosum</i> DSM 1402									
<i>Clostridium scindens</i> ATCC 35704	104	gb EDS08131.1							
<i>Clostridium</i> sp. L2-50								534	gb EDO66807.1
<i>Clostridium</i> sp. M621									
<i>Clostridium</i> sp. SS21									
<i>Clostridium sporiforme</i> DSM 1552									
<i>Clostridium sporogenes</i> ATCC 15579	204	gb EDU38352.1	132	gb EDU39429.1			152	gb EDU37786.1	
<i>Clostridium symbiosum</i>									
<i>Coccoloccus comes</i> ATCC 27768									
<i>Coccoloccus eutactus</i> ATCC 27769									
<i>Dorea formicigenerans</i> ATCC 27755	104	ref WP_005330286.1							
<i>Dorea longicatena</i> DSM 13814									
<i>Eubacterium bifforme</i> DSM 3989									
<i>Eubacterium dolium</i> DSM 3991									
<i>Eubacterium eligens</i> ATCC 27750								334	gb ACR72415.1
<i>Eubacterium hallii</i> DSM 3353									
<i>Eubacterium rectale</i> ATCC 33656								316	gb ACR77067.1
<i>Eubacterium rectale</i> DSM 17629								320	emb CBK99636.1
<i>Eubacterium ventriosum</i> ATCC 27560								315	emb CRL40832.1
<i>Faecalibacterium prausnitzii</i> A2-165									
<i>Faecalibacterium prausnitzii</i> M212									
<i>Hoblermanii</i> DSM 12042									
<i>Lactobacillus casei</i> ATCC 334	111	gb BAN74254.1	279	ref NP_011674560.1					
<i>Lactobacillus delbrueckii</i> subsp. <i>bulgaricus</i> ATCC 11842				308	emb CAI98549.1				
<i>Lactobacillus reuteri</i> DSM 20018	119	gb ABQ83632.1							
<i>Lactococcus lactis</i> subsp. <i>cremoris</i> MG1363	102	emb CAL98875.1							
<i>Lactococcus lactis</i> subsp. <i>cremoris</i>	102	emb CAB93581.2							
<i>Lactococcus lactis</i> subsp. <i>lactis</i> 11403	101	ref NP_268192.1							
M23A									
<i>Mitsuokella multacida</i> DSM 20544			101	gb EEX68878.1					
<i>Parvimonas micro</i> ATCC 33270									
<i>Roseburia intestinalis</i> L1-82								305	gb EEV01176.1
<i>Ruminococcus bromii</i> L263									
<i>Ruminococcus gnavus</i> ATCC 29149									
<i>Ruminococcus lactis</i> ATCC 29176									
<i>Ruminococcus obeum</i> ATCC 29174									
<i>Ruminococcus torques</i> ATCC 27756									
<i>Streptococcus infantarius</i> subsp. <i>infantarius</i> ATCC BAA-102									
<i>Streptococcus thermophilus</i> CNRZ1086									
<i>Streptococcus thermophilus</i> LMD-9									
<i>Streptococcus thermophilus</i> LMG 18311									
<i>Subdoligranulum variabile</i> DSM 15176									
<i>Fusobacterium</i> sp. 41 13			122	gb EEO39391.1					
<i>Fusobacterium varium</i> ATCC 27725	125	gb EE895190.1	1009	gb EES866191.1					
<i>Citrobacter youngae</i> ATCC 29220	849	gb EFF05427.1	1514	gb EFF05425.1	1280	gb EFF06567.1			
<i>Desulfovibrio piger</i> ATCC 29098					432	gb EEB32660.1			
<i>Desulfovibrio piger</i> GOR1					462	WP_040369942.1			
<i>Enterobacter cancerogenus</i>	259	ref WP_006177175.1	454	ref WP_034829451.1	1292	ref NP_042321927.1			
<i>Escherichia coli</i> str. K-12 substr. MG1655	881	ref NP_416539.1	1509	ref NP_418541.2	1368	ref NP_417413.1			
<i>Escherichia ferusonii</i> ATCC 35469	295	emb CAQ89410.1	481	emb CAQ89409.1	1361	emb CAQ89371.1			
<i>Helicobacter pylori</i> HPAG1					325	gb AER85039.1			
<i>Proteus penneri</i> ATCC 35198	113	gb EEG86390.1	587	gb EEO39391.1	924	gb EEB38448.1			
<i>Providencia alcalifaciens</i> DSM 30120	122	gb EEB44926.1	1155	gb EEB44562.1	1044	gb EEB44175.1			
<i>Providencia rettgeri</i> DSM 1131	121	gb EEF55447.1	1159	gb EEF52717.1	1039	gb EEF53196.1	101	gb EEB44240.1	
<i>Providencia rustiganii</i> DSM 4541	121	gb EEF71127.1	1159	gb EEF72852.1	1043	gb EEF71688.1			
<i>Providencia stuartii</i> ATCC 25827	117	gb EDU59312.1	1170	gb EDU61801.1	1044	gb EDU60243.1			
<i>Shigella</i> sp. D9									
<i>Akkermansia muciniphila</i> ATCC BAA-835					479	gb ACD05477.1	115	gb ACD03884.1	
<i>Vivivallis vadensis</i> ATCC BAA-548							204	gb ACD03862.1	

Data from 126 bacterial strains was obtained from article of Yatsunen et al. (Nature 486: 222-227, 2012)



Table S3. List of bacteria used for screening of polyamine producing bacteria

Strains	Phylum
<i>Alistipes putredinis</i> JCM16772	Bacteroidetes
<i>Bacteroides caccae</i> JCM9498	Bacteroidetes
<i>Bacteroides thetaiotaomicron</i> JCM 5827 ^T	Bacteroidetes
<i>Bacteroides uniformis</i> JCM5828	Bacteroidetes
<i>Bacteroides vulgatus</i> JCM 5826 ^T	Bacteroidetes
<i>Bifidobacterium adolescentis</i> JCM 1275 ^T	Actinobacteria
<i>Bifidobacterium longum</i> JCM 1217 ^T	Actinobacteria
<i>Blautia hansenii</i> JCM14655 ^T	Firmicutes
<i>Clostridium leptum</i> DSM753	Firmicutes
<i>Enterococcus faecalis</i> JCM 5803 ^T	Firmicutes
<i>Escherichia coli</i> JCM 5491	Proteobacteria
<i>Parabacteroides johnsonii</i> JCM13406	Bacteroidetes
<i>Parabacteroides merdae</i> JCM9497	Bacteroidetes
<i>Streptococcus thermophiles</i> NBRC13957	Firmicutes

Table S4. Strains, plasmids, and oligonucleotides used in this study

Strain, plasmid, or oligonucleotide	Characteristic or sequence	Source or reference
Strains		
<i>Escherichia coli</i> K-12		
JW4076	ME9062 except Δ adiC :: FRT-kan ⁺ -FRT	Baba et al. 2006
JW5731	ME9062 except Δ adiA :: FRT-kan ⁺ -FRT	Baba et al. 2006
LKM10096	pBeloBAC11/MG1655	This study
LKM10097	pBelobac11/SK903	This study
LKM10100	pBelobac11-adiC ⁺ /SK903	This study
ME9062	<i>rrmB</i> Δ lacZ4787 <i>hsdR514</i> Δ (<i>araBAD</i>)567 Δ (<i>rhaBAD</i>)568 <i>rph-1</i>	Baba et al. 2006
MG1655	F prototrophic	C. A. Gross
SK900	MG1655 except Δ adiA :: FRT-kan ⁺ -FRT	This study
SK901	MG1655 except Δ adiA :: FRT	This study
SK902	MG1655 except Δ adiC :: FRT-kan ⁺ -FRT	This study
SK903	MG1655 except Δ adiC :: FRT	This study
SK912	pBelobac11-adiA ⁺ /SK901	This study
SK914	pBelobac11/SK901	This study
SK930	pBelobac11/MG1655 except Δ speB :: FRT Δ speC :: FRT Δ speF :: FRT	This study
<i>Enterococcus faecalis</i>		
SK932	pLT06- Δ aguD/V583	This study
SK934	V583 except Δ aguD <i>lacZ</i> ⁺ <i>cat</i> ⁺ <i>pheS</i> ⁺ <i>aguD</i> ⁺	This study
SK937	V583 except Δ aguD	This study
SK947	pLZ12/V583	This study
SK948	pLZ12/SK937	This study
SK949	pLZ12- <i>aguB</i> ⁺ <i>D</i> ⁺ /SK937	This study
V583	Clinical isolate, TIGR sequence strain; Van ^R	American Type Culture Collection
Plasmids		
harbouring genes of <i>E. coli</i> K-12 MG1655		
pBelobac11	Mini-F replicon <i>cat</i> ⁺	New England Biolabs Inc., Beverly, Massachusetts, USA
pBelobac11- <i>adiA</i> ⁺	Mini-F replicon <i>cat</i> ⁺ <i>adiA</i> ⁺	This study
pBelobac11- <i>adiC</i> ⁺	Mini-F replicon <i>cat</i> ⁺ <i>adiC</i> ⁺	This study
harbouring genes of <i>En. faecalis</i> V583		
pLT06	<i>repA</i> -ts replicon <i>pheS</i> ⁺ - <i>cat</i> ⁺ - <i>lacZ</i> ⁺	Thurlow et al. 2009
pLT06- Δ aguD	<i>repA</i> -ts replicon Δ aguD <i>pheS</i> ⁺ <i>cat</i> ⁺ <i>lacZ</i> ⁺	This study
pLZ12	pSH71 replicon <i>cat</i> ⁺	Perez-Casal et al. 1991
pLZ12- <i>aguB</i> ⁺ <i>D</i> ⁺	pSH71 replicon <i>cat</i> ⁺ <i>aguB</i> ⁺ <i>D</i> ⁺	This study
Oligonucleotides		
adiA_300bp-up_for	ccgtgccggcagcgtTCAAGTAGTCGTTTTTACAG	
adiA_term_rev	ggatgcagcccggTTTACGCTTTCACGCACATAA	
adiC_300bp-up_for	ccgtgccggcagcgtAAAAGTTGAAAAGCGAAAAC	
adiC_term_rev	ggatgcagcccggTTAATCTTTGCTTATTGGTG	
Clone_agcB+0.5K_FWD	tgaattcattagatcGTGCGGTCATAAAGGTCTGA	
Clone_agcD+COMPL	aggaagatctggatcTTAGCCATTTTCTTTCGTTCCCTT	