

Figure S5

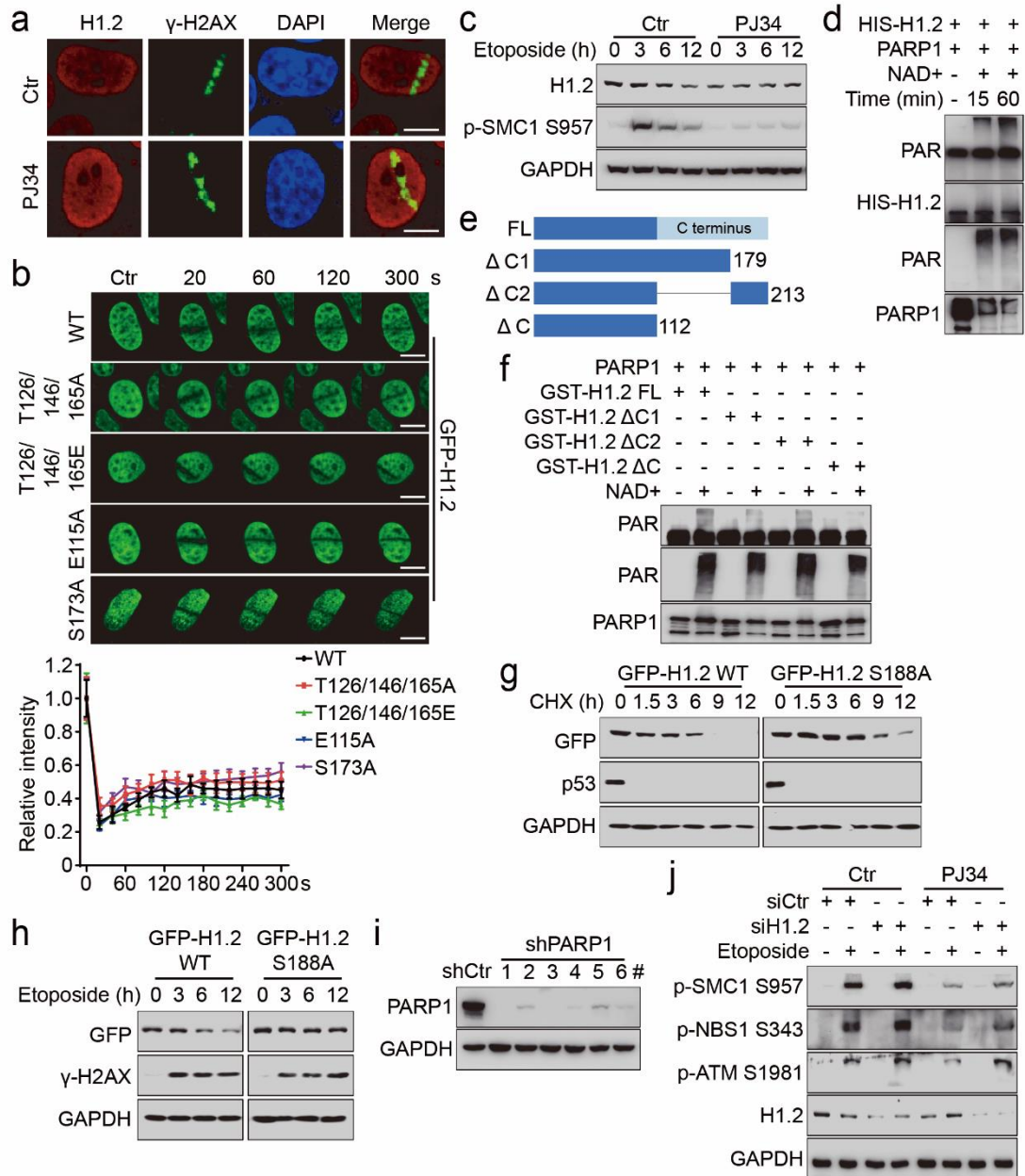


Figure S5. PARylation of linker histone H1.2 is required for its displacement and ATM activation

a HeLa cells were treated with or without 5 μ M PJ34 for 1 h, subjected to laser micro-irradiation and fixed immediately before immunofluorescence assay. Cells were stained with the indicated antibodies or DAPI. Scale bars, 10 μ m. **b** HeLa cells were transfected with the indicated WT or mutated GFP-H1.2 plasmids and subjected to laser micro-irradiation. Images were captured every 20 s for 5 min and quantifications of the IR path signal intensity are shown. The data represent the mean \pm SD. Scale bars, 10 μ m. **c** HeLa cells were treated with 20 μ M etoposide for the indicated times, with or without 5 μ M PJ34 for 1 h prior to etoposide treatment and then analyzed by immunoblotting. **d** Recombinant HIS-H1.2 was incubated with or without NAD⁺ and PARP1 as indicated. The samples were analyzed by immunoblotting after *in vitro* PARylation assay. **e** Location of the H1.2 C-terminal deletion mutants. **f** Recombinant full-length (FL), C1-deleted (Δ C1), C2-deleted (Δ C2) or C-deleted (Δ C) GST-H1.2 were assessed for *in vitro* PARylation assay. **g, h** HeLa cells were transfected with the indicated plasmids and treated with 1 μ M CHX or 20 μ M etoposide for the indicated times before analyzed by immunoblotting. **i** HeLa cells were stably transfected with shRNAs targeting PARP1 and several clones (1-6#) were selected for immunoblotting to determine the efficiency of PARP1 knockdown. **j** HeLa cells were transfected with the indicated siRNAs and treated with 40 μ M etoposide for 15 min with or without 5 μ M PJ34 for 1 h and then analyzed by immunoblotting.