

## **Additional file 1**

	<b>Sex</b>	<b>WT (n=)</b>	<b>Tau<sup>-/-</sup> (n=)</b>
7 months old	<i>Male</i>	6	7
	<i>Female</i>	6	4
12 months old	<i>Male</i>	10	8
	<i>Female</i>	2	4
15 months old	<i>Male</i>	5	7
	<i>Female</i>	3	6

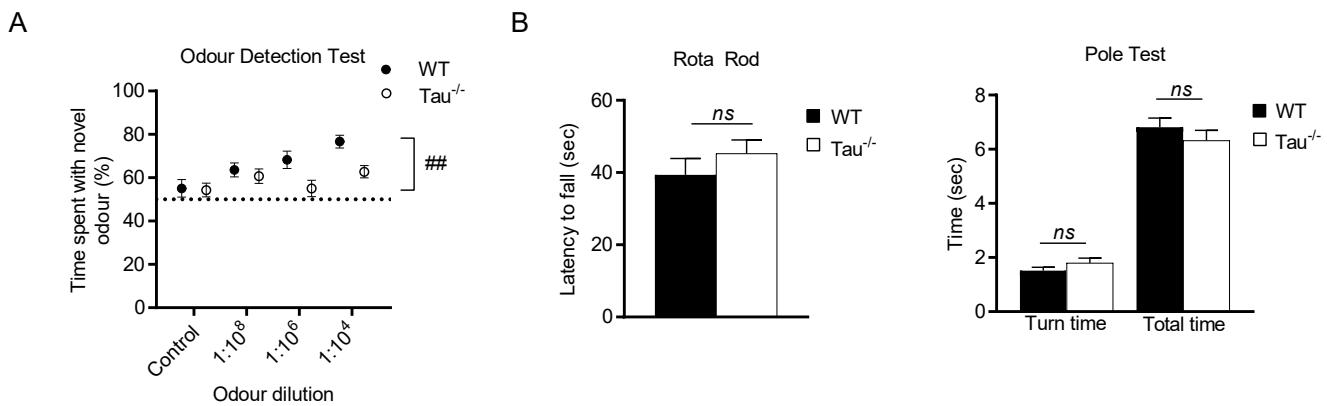
**Table S1.** Animal number breakdown based on genotype and sex.

<b>Experimental Group</b>	<b>Analysis</b>	<b>Parameter 1 Factor: genotype</b>	<b>Parameter 2 Factor: odour concentration</b>	<b>Parameter 3: interaction</b>
7 mo Sv129B/6	2-way RM ANOVA	$F_{1,57}=11$ , p=0.003	$F_{3,57}=5.3$ , p=0.003	$F_{1,57}=2.2$ , p=0.01
12 mo Sv129B/6	2-way RM ANOVA	$F_{1,99}=7.9$ , p=0.008	$F_{3,99}=6.3$ , p<0.001	$F_{3,99}=1.7$ , p=0.2
15 mo Sv129B/6	2-way RM ANOVA	$F_{1,66}=0.39$ p=0.54	$F_{3,66}=0.55$ , p=0.65	$F_{3,66}=1.01$ , p=0.4

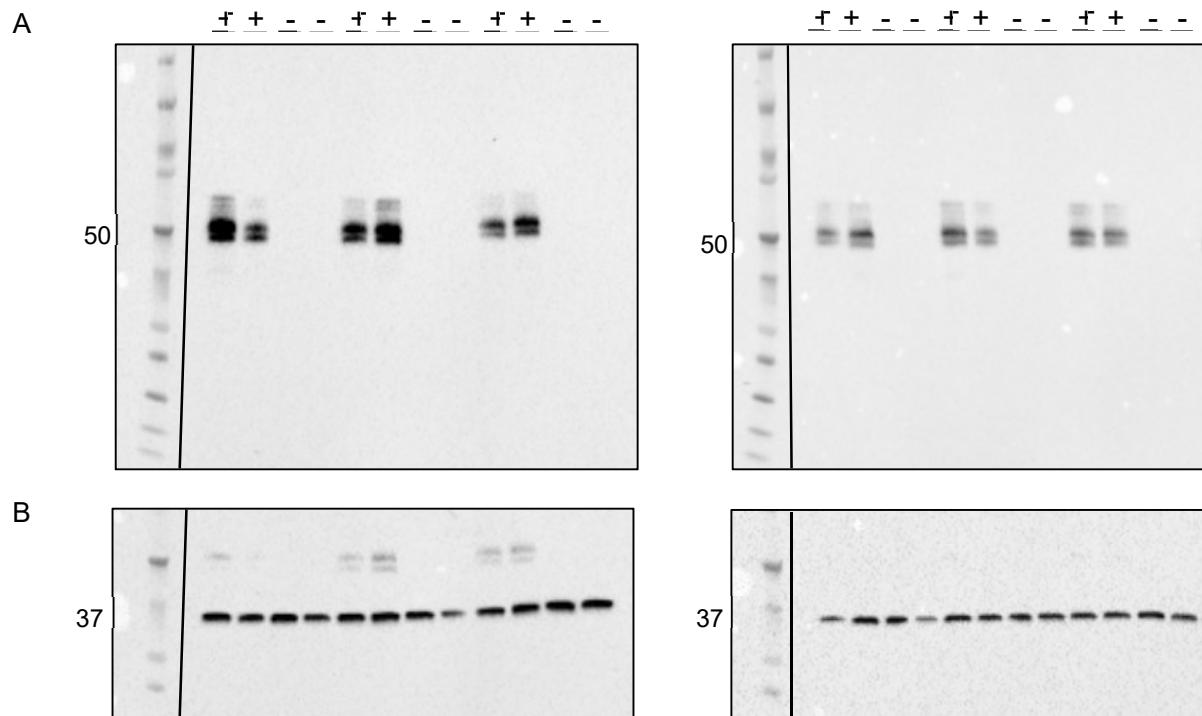
**Table S2.** ODT ANOVA factors. Normality test: Shapiro-Wilk

<b>Experimental Group</b>	<b>Odour concentration</b>	<b>Hypothetical mean</b>	<b>Actual mean</b>	<b>SEM</b>	<b>n</b>	<b>p</b>
7 mo WT	0	50	49.46	3.67	11	0.88
	1:10 <sup>8</sup>	50	62.13	6.743	12	0.10
	1:10 <sup>6</sup>	50	68.39	3.672	10	0.0007
	1:10 <sup>4</sup>	50	72.89	3.613	10	0.0001
7 mo tau <sup>-/-</sup>	0	50	45.04	2.53	10	0.08
	1:10 <sup>8</sup>	50	58.91	3.343	10	0.03
	1:10 <sup>6</sup>	50	46.79	5.355	11	0.56
	1:10 <sup>4</sup>	50	56.67	6.396	11	0.32
15 mo WT	0	50	51.40	2.785	8	0.63
	1:10 <sup>8</sup>	50	51.81	3.96	8	0.66
	1:10 <sup>6</sup>	50	54.76	4.06	7	0.29
	1:10 <sup>4</sup>	50	59.56	5.41	5	0.15
15 mo tau <sup>-/-</sup>	0	50	48.78	5.27	12	0.82
	1:10 <sup>8</sup>	50	58.73	4.41	10	0.08
	1:10 <sup>6</sup>	50	52.15	4.27	12	0.63
	1:10 <sup>4</sup>	50	49.79	4.18	11	0.96

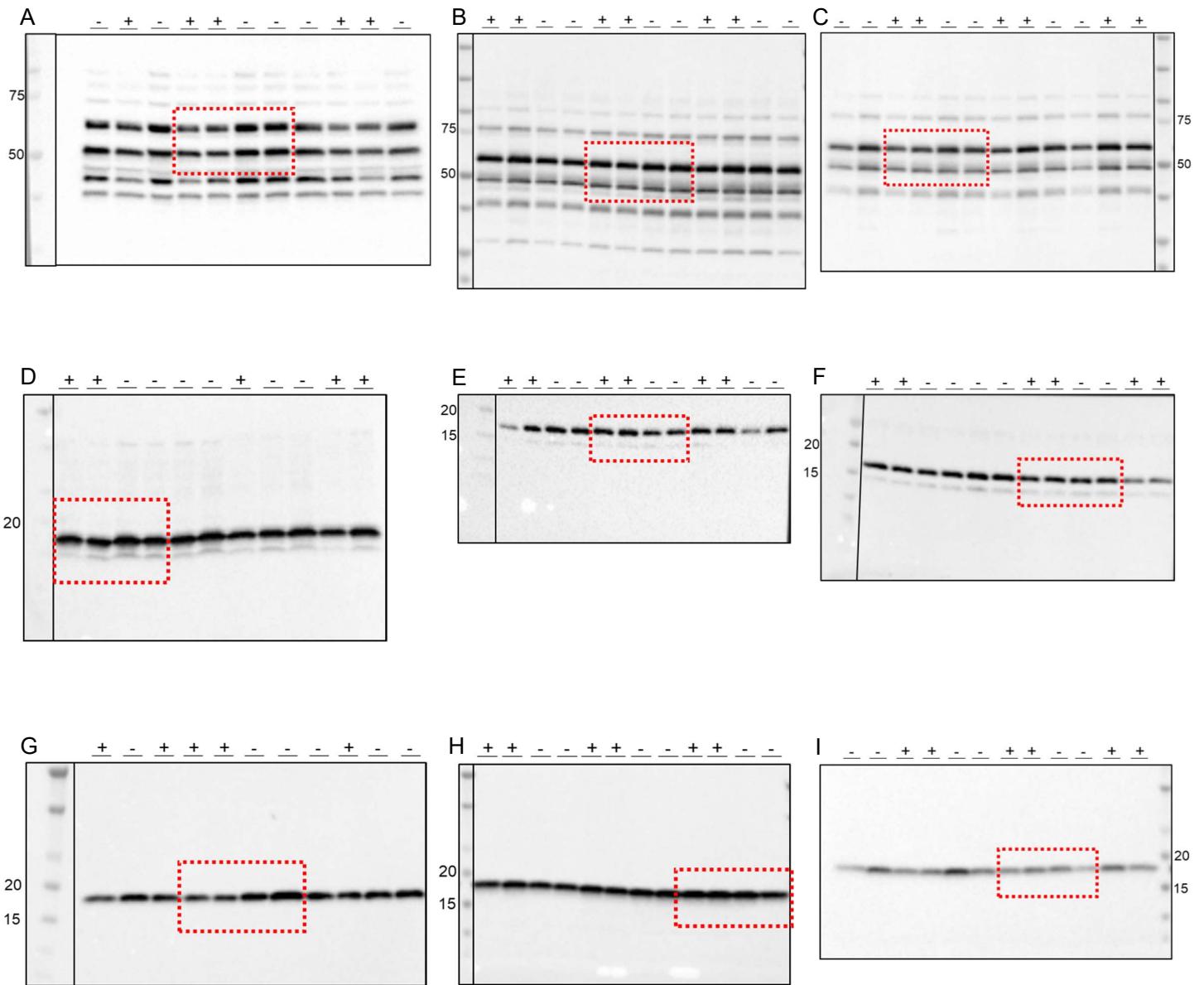
**Table S3.** ODT one sample t test, hypothetical mean = 50% (chance).



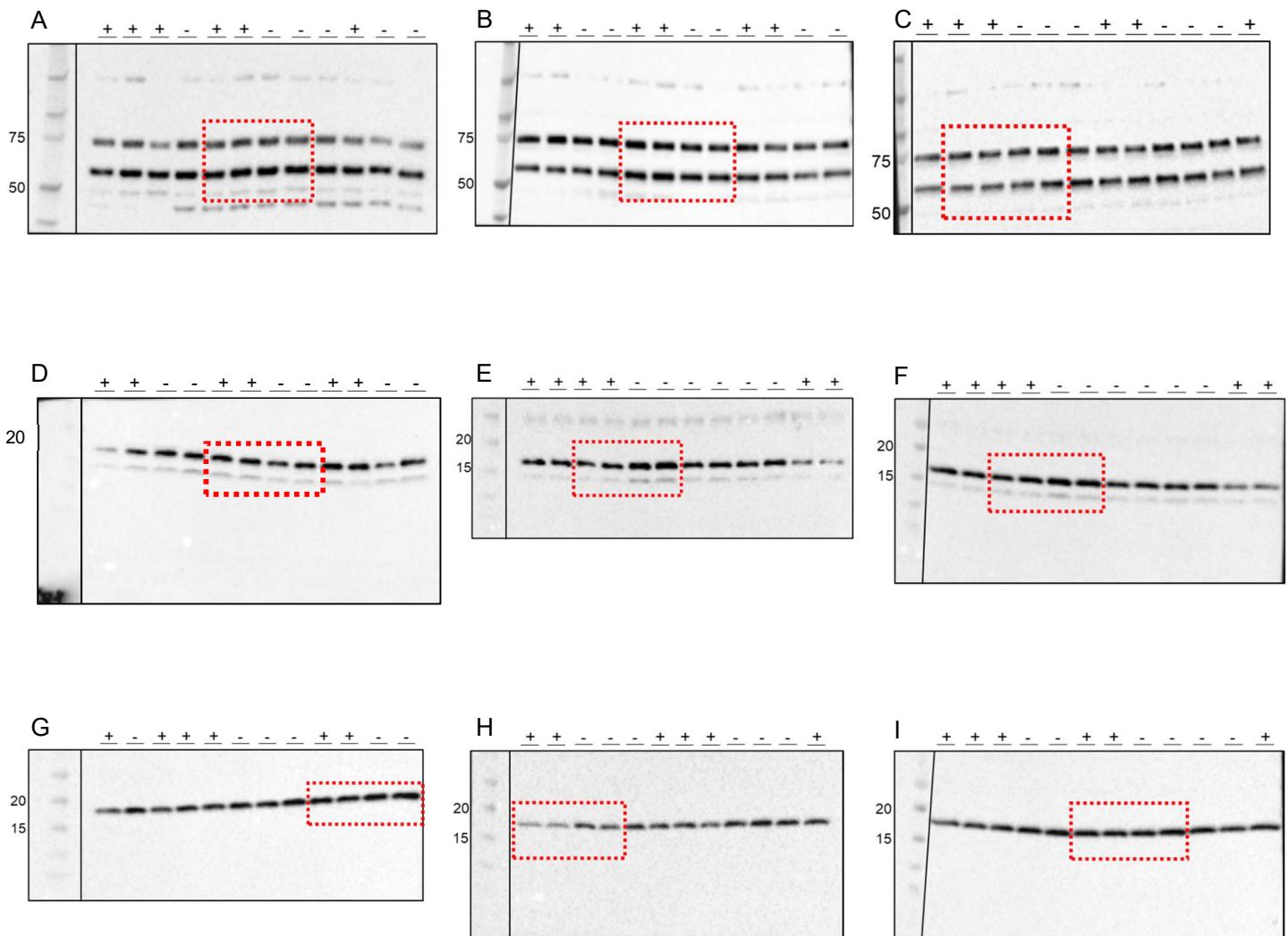
**Figure S1.** Odour detection test (A) and motor evaluation (B) (Rota Rod and Pole Test) of 12-month-old tau<sup>-/-</sup> (n=12) and littermate WT control (n=12). ODT analysed by two-way repeated measures ANOVA (one factor repetition) with Fisher LSD post-hoc comparisons. # represents significant main effect of genotype, ## p < 0.01. Motor tests analysed by unpaired two-sided t test. Ns = not significant.



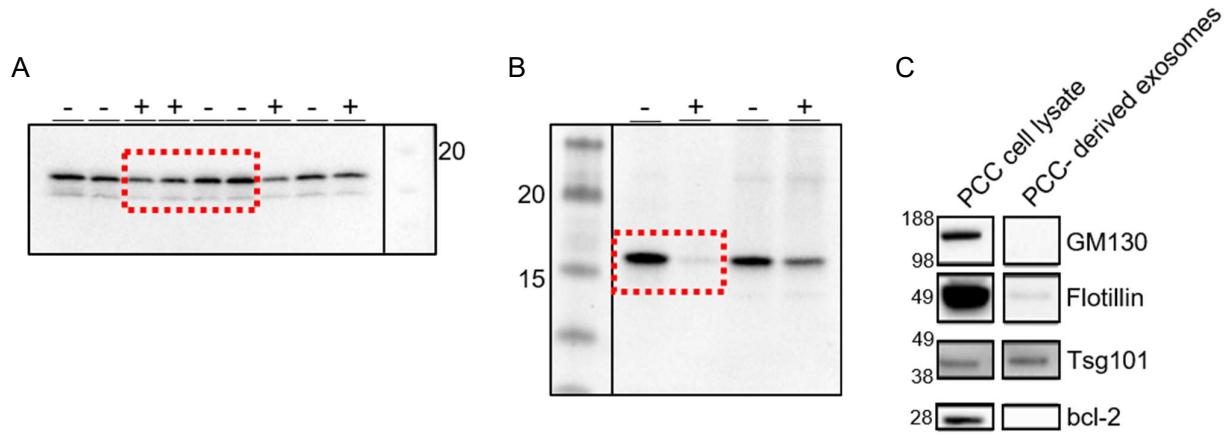
**Figure S2.** Western blot confirmation of tau ablation in 7- and 15-month old tissue used for immunoblot analysis. A) Tau antibody (Dako, catalogue number: A0024, dilution 1:10,000) confirmed tau ablation B) Protein loading in all wells confirmed by GAPDH (Cell Signaling Technology, catalogue number 2118, dilution 1:10,000).



**Figure S3.** Full Western Blots of 7 mo WT (+) and  $Tau^{-/-}$  (-) cell lysate. A, B, C: immunoblot of p62 from OB, CPU and SN respectively. D, E, F: immunoblot of LCB3 from OB, CPU and SN respectively. G, H, I: immunoblot of  $\alpha$ -synuclein from OB, CPU and SN respectively. Red box indicates the section represented in Figures.



**Figure S4.** Full Western Blots of 15 mo WT (+) and  $\text{Tau}^{-/-}$  (-) cell lysate. A, B, C: immunoblot of p62 from OB, CPU and SN respectively. D, E, F: immunoblot of LCB3 from OB, CPU and SN respectively. G, H, I: immunoblot of  $\alpha$ -synuclein from OB, CPU and SN respectively. Red box indicates the section represented in Figures.



**Figure S5.** A) Full Western Blots of primary cortical neurons for LC3B from WT (+) and  $\text{Tau}^{-/-}$  (-) cell lysate. B) Full Western Blot of primary cortical neuron-derived exosomes for  $\alpha$ -synuclein from WT (+) and  $\text{Tau}^{-/-}$  (-) cells. C) Exosome verification using exosome negative markers (GM130 & bcl-2), exosome positive marker (Tsg101) and microvesicle marker (Flotillin) in primary cortical neuron cell lysate and exosomes.