Integrated treatment using intraperitoneal radioimmunotherapy and positron emission tomography-guided surgery with ⁶⁴Culabeled cetuximab to treat early- and late-phase peritoneal dissemination in human gastrointestinal cancer xenografts

SUPPLEMENTARY MATERIALS

Immunohistochemistry for EGFR and HER2 expression

Immunohistochemical analysis was performed with paraffin-embedded sections of HCT116-RFP spheroids cultured in 96-well nano-culture plates (SCIVAX Life Sciences), as previously described [1]. Briefly, the collected spheroids were embedded in iPGell (Genostaff, Tokyo, Japan), fixed with 10% buffered formalin (Genostaff) for 2 d at room temperature, processed for paraffin embedding, and sectioned according to standard histological procedures to obtain 6-µM sections. After deparaffinization and dehydration, the sections were treated by microwaving for antigen retrieval and 0.3% hydrogen peroxide in methanol for 30 min to quench endogenous peroxidases. To block nonspecific binding, a protein-blocking agent (G-Block, Genostaff) was applied for 10 min at room temperature. Endogenous avidin and biotin activities were blocked using the Avidin/Biotin Blocking Kit (Vector). The sections were incubated overnight at 4° C with a primary antibody against EGFR (1 µg/mL; CST) and HER2 (1 µg/mL; CST). After washing the sections with Tris-buffered saline (TBS), a biotinylated secondary antibody (E0432, Dako) was applied for 30 min at room temperature. After another TBS wash, the sections were incubated with streptavidin peroxidase reagent (426062, Nichirei) for 5 min and washed with TBS. Colour development was performed using 3,3'-diaminobenzidine tetrahydrochloride solution with H₂O₂, and sections were counterstained with hematoxylin.

Cell-binding and competitive-inhibition assays with ⁶⁴Cu-PCTA-cetuximab

Cell-binding and competitive-inhibition assays with ⁶⁴Cu-PCTA-cetuximab (synthesized in this study) were performed as previously described [2]. Briefly, for cellbinding assays, HCT116-RFP cells were serially diluted in PBS with 1% bovine serum albumin (BSA) (Sigma) and incubated with ⁶⁴Cu-PCTA-cetuximab on ice for 1 h. After washing, the radioactivity bound to the cells was measure using a γ -counter (1480 Automatic gamma counter Wizard 3; PerkinElmer). The immunoreactivity of ⁶⁴Cu-PCTA-cetuximab was determined by the Lindmo method [3]. For competitive-inhibition assays, HCT116-RFP cells were incubated with ⁶⁴Cu-PCTA-cetuximab and various concentrations of unlabelled cetuximab on ice for 1 h. After washing, the radioactivity bound to the cells was measure as described above. The dissociation constant was determined based on a competitive-inhibition assay. To compare the cell-binding affinities of ⁶⁴Cu-PCTAcetuximab and ⁶⁴Cu-PCTA-trastuzumab, 1×10^6 cells diluted in PBS with 1% BSA were incubated with each ⁶⁴Cu-labeled antibody and the radioactivity was measured in a manner similar to that described above.

Measurement of biochemical parameters

Biochemical parameters were measured at day 35, using mouse plasma prepared with blood collected from the heart. Levels of glutamate oxaloacetate transaminase (GOT), glutamate pyruvate transaminase (GPT), and alkaline phosphatase (ALP) were measured to assess liver function; urea nitrogen (BUN) and creatinine (CRE) were measured for kidney function; and amylase (AMYL) and lipase (LIP) were measured for pancreas function, using a blood biochemical analyser (Dri-Chem 7000VZ, Fuji Film).

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Supplementary Video 1: OpenPET-guided surgery with ⁶⁴Cu-PCTA-cetuximab in the case of tumours located deeply in the peritoneal cavity. Video shows OpenPET-guided surgery detected tumours located deeply in the peritoneal cavity (approximately 10 mm) and the surgeon resected the tumours while monitoring it with the OpenPET real-time imaging. The tumours could not be visually identified by eye. See Supplementary_Video_1

Supplementary Video 2: OpenPET-guided surgery with ⁶⁴Cu-PCTA-cetuximab in the case of relatively small tumours located deeply in the peritoneal cavity. Video shows OpenPET-guided surgery detected and resected 3-mm size small tumours located deeply in the peritoneal cavity using OpenPET system. See Supplementary_Video_2

Target organ	Total estimated absorbed dose (mSv/MBq)		
Adrenals	0.0138		
Brain	0.0113		
Breasts	0.0106		
Gallbladder wall	0.0148		
Lower large intestinal wall	0.0384		
Small intestine	0.0290		
Stomach wall	0.0198		
Upper large intestinal wall	0.0377		
Heart wall	0.0348		
Kidneys	0.0225		
Liver	0.0321		
Lungs	0.0119		
Muscle	0.0076		
Ovaries	0.0143		
Pancreas	0.0456		
Red marrow	0.0106		
Osteogenic cells	0.0226		
Skin	0.0096		
Spleen	0.0351		
Testes	0.0109		
Thymus	0.0119		
Thyroid	0.0112		
Urinary bladder wall	0.0178		
Uterus	0.0151		
Total body	0.0137		

Supplementary Table 1: Mean estimated human absorbed doses of ⁶⁴ Cu-PCTA-cetuximab following ip administration
extrapolated from mouse biodistribution data

Target organ	Total estimated absorbed dose (mSv/MBq)		
Adrenals	0.0171		
Brain	0.0141		
Breasts	0.0134		
Gallbladder wall	0.0190		
Lower large intestinal wall	0.0300		
Small intestine	0.0300		
Stomach wall	0.0222		
Upper large intestinal wall	0.0299		
Heart wall	0.0554		
Kidneys	0.0328		
Liver	0.0533		
Lungs	0.0151		
Muscle	0.0091		
Ovaries	0.0164		
Pancreas	0.0145		
Red marrow	0.0129		
Osteogenic cells	0.0282		
Skin	0.0120		
Spleen	0.0393		
Testes	0.0135		
Thymus	0.0153		
Thyroid	0.0140		
Urinary bladder wall	0.0188		
Uterus	0.0165		
Total body	0.0171		

Supplementary Table 2: Mean estimated hur	an absorbed doses of	⁶⁴ Cu-PCTA-cetuximab	following iv	administration,
extrapolated from mouse biodistribution data				

	Organ*	Estimated dose in human (Sv/male)**	Tolerance dose of radiation (Sv)***
ip	Lower large intestinal wall	3.14	35–50
	Small intestine	2.37	40
	Upper large intestinal wall	3.08	35–50
	Kidneys	1.84	23
	Liver	2.62	30
	Pancreas	3.73	30
	Red marrow	0.86	2.5
iv	Lower large intestinal wall	2.46	35–50
	Small intestine	2.45	40
	Upper large intestinal wall	2.45	35–50
	Kidneys	2.69	23
	Liver	4.36	30
	Pancreas	1.19	30
	Red marrow	1.06	2.5

Supplementary Table 3: Comparison between the estimated and tolerance doses in critical organs during ⁶⁴Cu-PCTA-cetuximab therapy with ip or iv administration

*Organs normally considered as those of concern in ⁶⁴Cu-PCTA-cetuximab therapy are listed. **Estimated doses with 81.8 GBq/ adult man (73.7 kg), which were calculated from 22.2 MBq/mouse based on body weight, during ⁶⁴Cu-PCTA-cetuximab therapy. ***Reported tolerated doses of radiation in organs [4, 5]. The values were compared to the estimated doses in human organs in the text.



Supplementary Figure 1: *In vitro* characterization of ⁶⁴Cu-PCTA-cetuximab binding with HCT116-RFP cells. (A) Cellbinding assay of ⁶⁴Cu-PCTA-cetuximab with HCT116-RFP cells. The immunoreactive fraction was 96.4%. (B) Competitive-inhibition assay of ⁶⁴Cu-PCTA-cetuximab with HCT116-RFP cells. The dissociation constant was 0.097 nM.

EGFR

HER2





Supplementary Figure 2: Expression of EGFR and HER2 in HCT116-RFP spheroids. Immunohistochemistry of EGFR and HER2 with HCT116-RFP spheroids is shown (left and right panels, respectively).



Supplementary Figure 3: Distribution and safety of ip-injected ⁶⁴Cu-PCTA-cetuximab. (A, B) Distribution of ⁶⁴Cu-PCTA-cetuximab in the organs of interest in mice following ip or iv administration at 3, 6, 18, 24, and 48 h (n = 4). Values are shown as the mean \pm SD. (C) (D) Haematological toxicity in mice administered ⁶⁴Cu-PCTA-cetuximab by ip or iv injection. The numbers of WBCs, RBCs, and PLTs at day 0 (before ⁶⁴Cu-PCTA-cetuximab injection) and at 7, 14, 21, 28, and 35 days after ⁶⁴Cu-PCTA-cetuximab injection (n = 4) are shown. Values are shown as the mean \pm SD. *P < 0.05 vs day 0 in each group (1-way ANOVA with Bonferroni's post hoc test).



Supplementary Figure 4: Biochemical parameters in mice administered ⁶⁴Cu-PCTA-cetuximab by ip injection, measured at day 35. (A) Levels of GOT, GPT, and ALP as a measure of liver functions. (B) Levels of BUN and CRE for kidney functions. (C) Levels of AMYL and LIP for pancreas functions.



Supplementary Figure 5: Biochemical parameters in mice administered ⁶⁴Cu-PCTA-cetuximab by iv infection, measured at day 35. (A) Levels of GOT, GPT, and ALP as a measure of liver functions. (B) Levels of BUN and CRE for kidney functions. (C) Levels of AMYL and LIP for pancreas functions.



Supplementary Figure 6: Cell-binding affinity of ⁶⁴Cu-PCTA-cetuximab and ⁶⁴Cu-PCTA-trastuzumab for HCT116-RFP cells.



Supplementary Figure 7: Fluorescence imaging during OpenPET surgery. The images show a series of observations obtained with a fluorescence microscope equipped with the OpenPET system (upper; left and right panels), fluorescent imaging of tumours deeply located (lower, left), and fluorescent imaging of tumours identified during OpenPET surgery (lower, right). Fluorescent signals were detected from isolated tumours, but not deeply located tumours. The yellow, dotted circle highlights an isolated tumour.

Normal background



Supplementary Figure 8: OpenPET imaging with a tumour-free mouse administered ⁶⁴Cu-PCTA-cetuximab. An imaging dose of ⁶⁴Cu-PCTA-cetuximab (7.4 MBq/mouse) was administered to a mouse at 24 h before OpenPET imaging in a manner similar to that used in the OpenPET experiments represented in Figures 3, 4.