

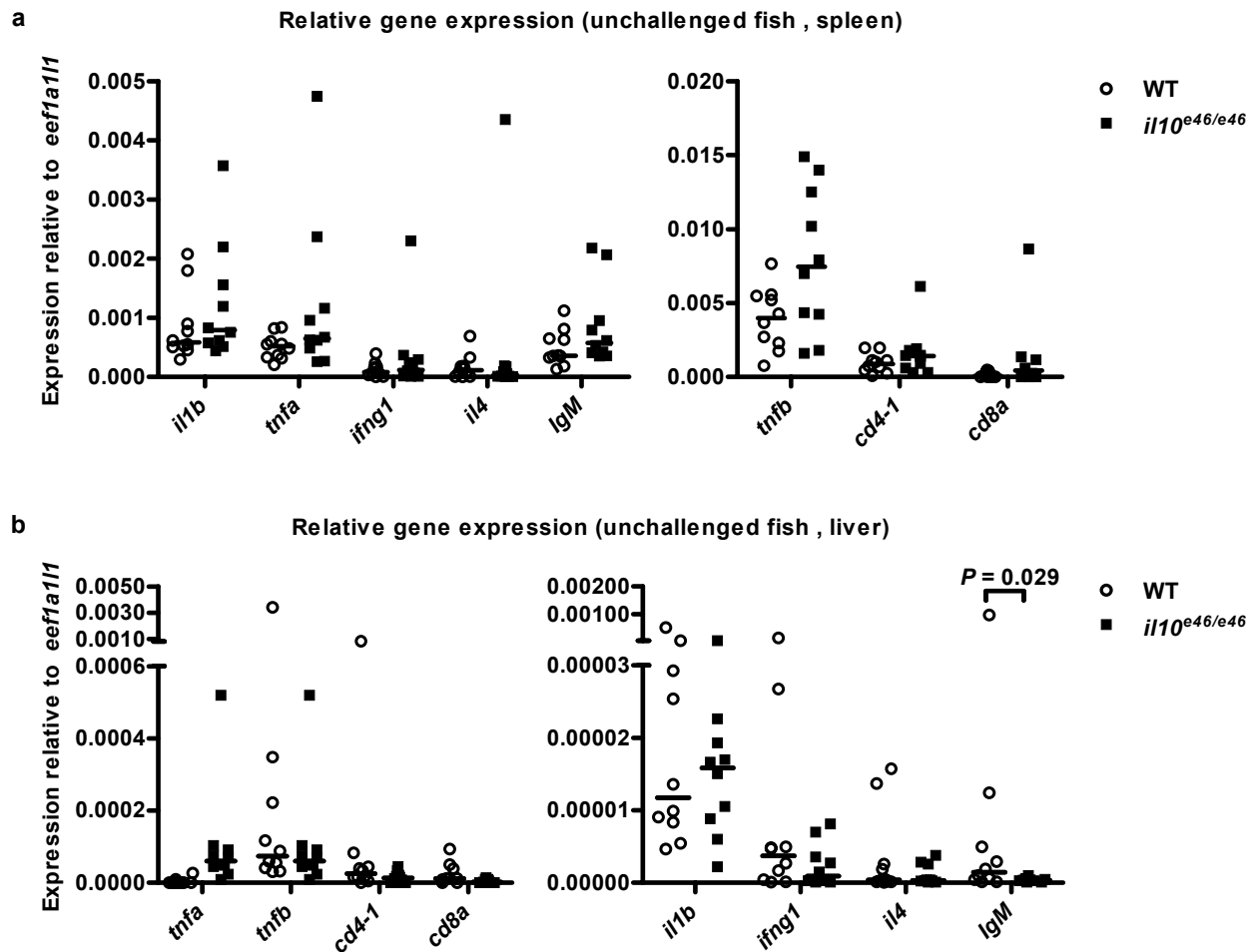
***interleukin 10* mutant zebrafish have an enhanced *interferon gamma* response and improved survival against a *Mycobacterium marinum* infection**

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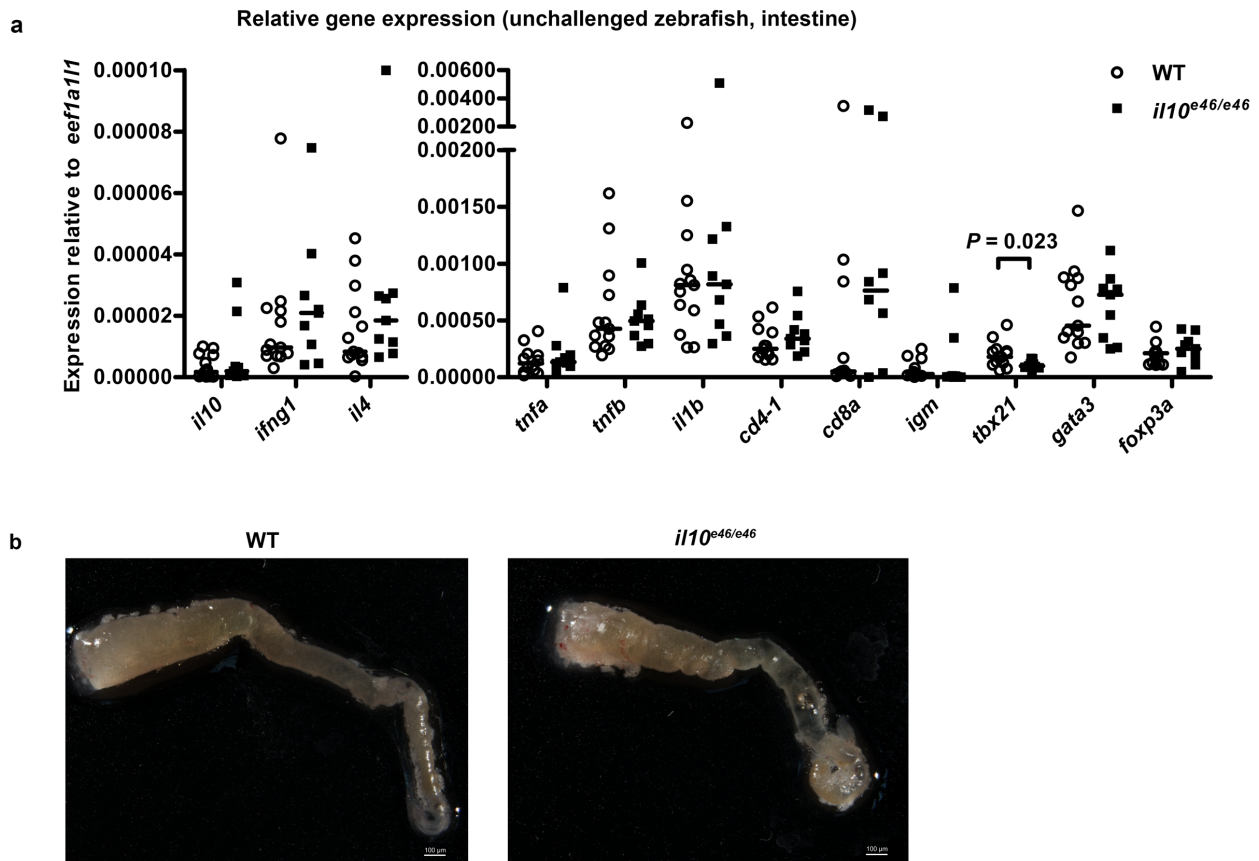
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## Supplementary Figures



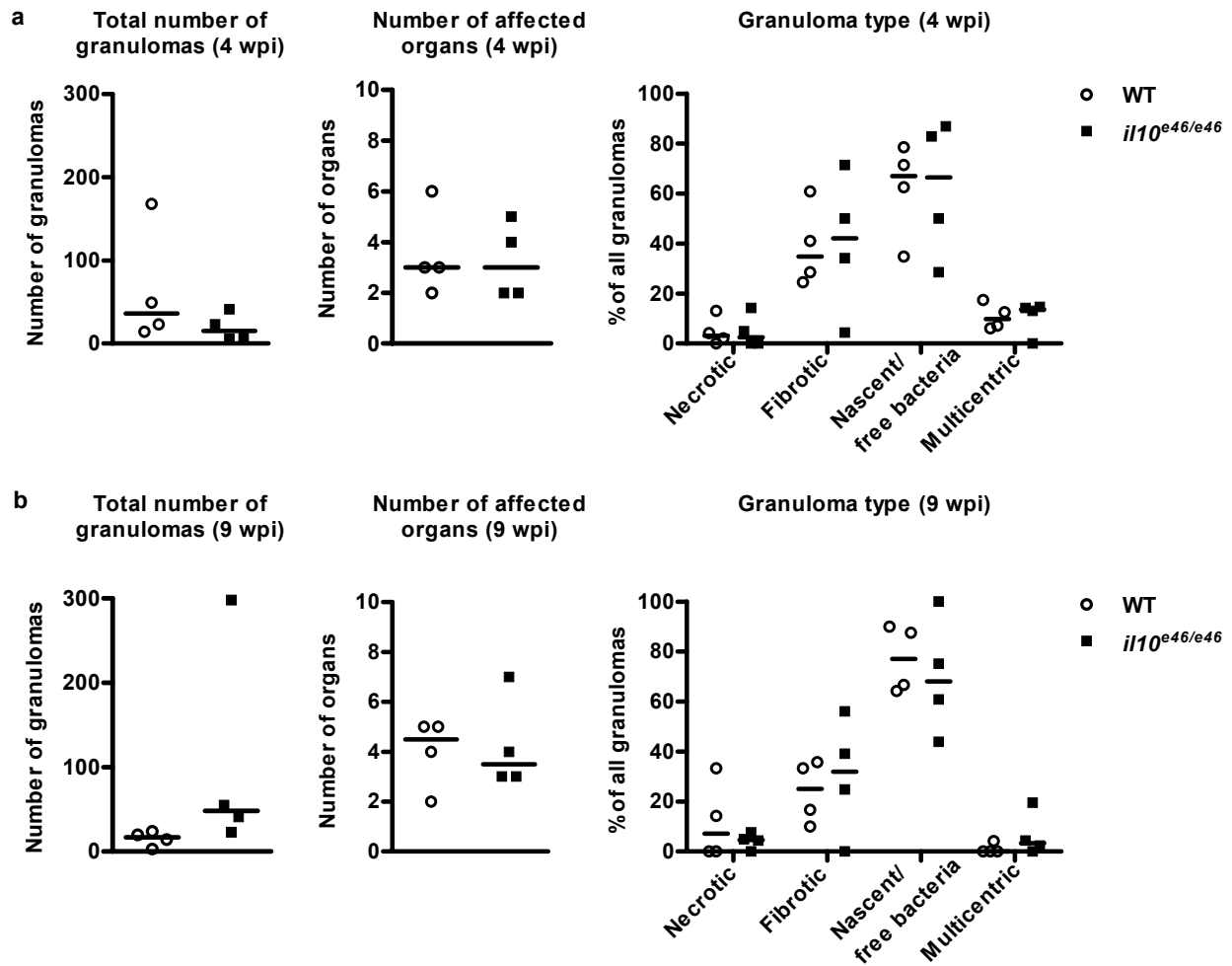
## Supplementary Figure S1

Unchallenged *il10<sup>e46/e46</sup>* zebrafish have similar expression profiles of lymphocyte markers and cytokines as WT control fish in the spleen and liver (**a - b**). The relative expression of selected proinflammatory cytokines (*il1b*, *tnfa*, *tnfb*), Th cell cytokines (*ifng1* and *il4*), T cell markers (*cd4* and *cd8*) as well as a B cell marker (*IgM*) in *il10<sup>e46/e46</sup>* mutant zebrafish and WT controls in spleen and liver from the same individuals ( $n = 10$  in both groups) were quantified with qPCR and are represented as scatter dot plot and median. Note the different scales of the y axes and the divided y axis in panel b. Gene expressions were normalized to the expression of *eef1a111*. A two-tailed Mann-Whitney test was used for the statistical comparison of differences between *il10<sup>e46/e46</sup>* zebrafish and WT controls.



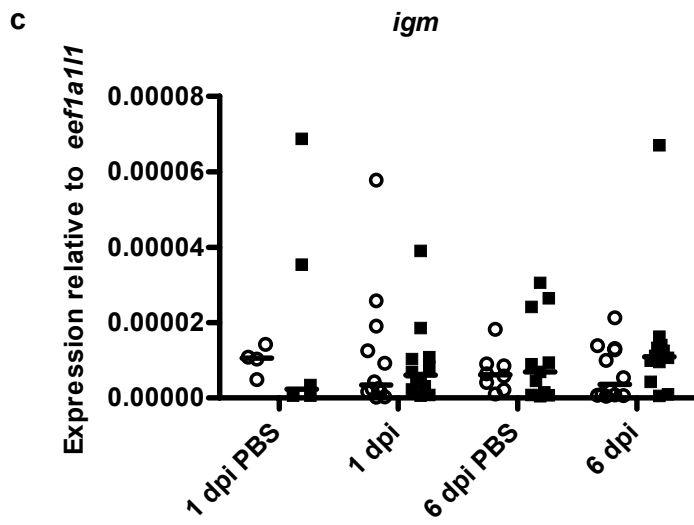
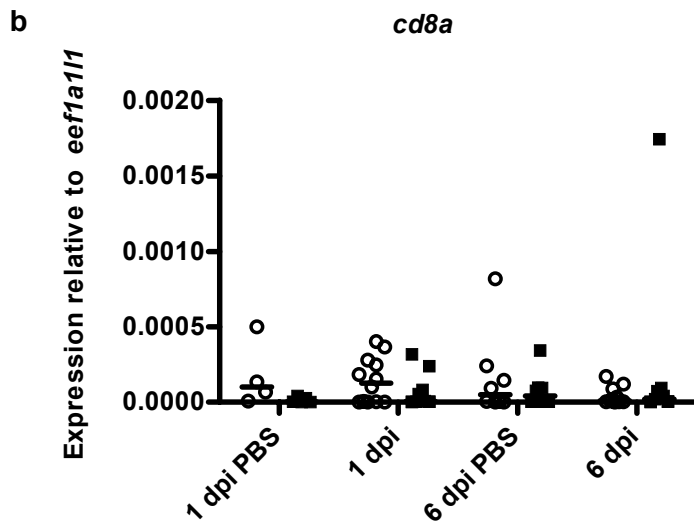
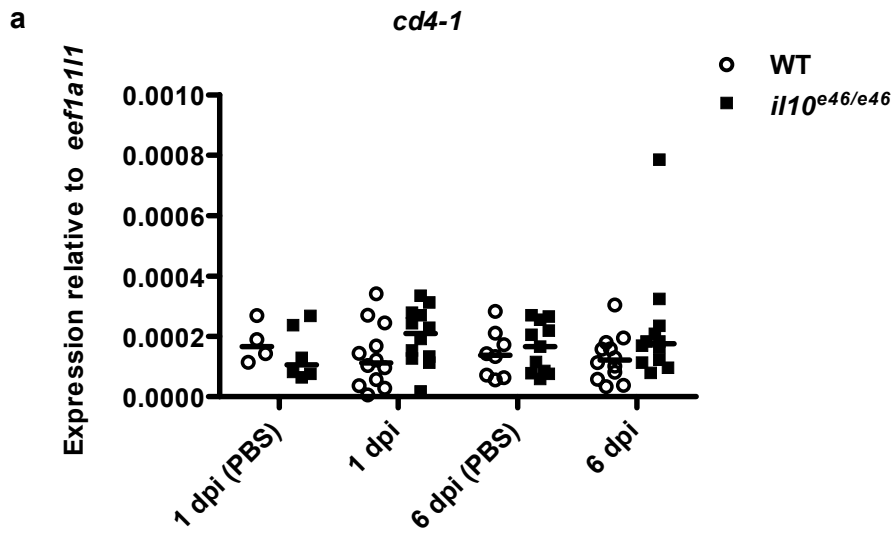
### Supplementary Figure S2

Unchallenged *il10<sup>e46/e46</sup>* zebrafish have similar expression profiles of lymphocyte markers and cytokines as WT control fish in the intestine. **(a)** The relative expression of *il10*, selected proinflammatory cytokines (*il1b*, *tnfa*, *tnfb*), Th cell cytokines (*ifng1* and *il4*), T cell markers (*cd4-1* and *cd8a*), a B cell marker (*IgM*) as well as Cd4<sup>+</sup> lymphocyte transcription factors (*tbx21*, *gata3*, *foxp3a*) in *il10<sup>e46/e46</sup>* mutant zebrafish ( $n = 9$ ) and WT controls ( $n = 13$ ) in intestine were quantified with qPCR and are represented as scatter dot plot and median. Note the different scales of the y axes and the divided y axis. Gene expressions were normalized to the expression of *eef1a111*. A two-tailed Mann-Whitney test was used for the statistical comparison of differences between *il10<sup>e46/e46</sup>* zebrafish and WT controls. **(b)** The visual appearance of *il10<sup>e46/e46</sup>* mutant and WT control zebrafish intestine was studied by microscopy ( $n = 3$  in both groups). A representative image from each group is shown.



### Supplementary Figure S3

There are no differences in the quantity or quality of granulomas between *il10<sup>e46/e46</sup>* zebrafish and WT controls in histological analysis. **(a-b)** The number of *M. marinum* granulomas were counted and the affected organs and the type of granulomas were identified in *il10<sup>e46/e46</sup>* zebrafish and WT controls from Ziehl-Neelsen stained sections ( $n = 4$  in both groups at both time points) at 4 and 9 weeks post a low-dose infection (2 - 9 CFU). The granulomas were placed in one or several of the following categories: necrotic, fibrotic, multicentric, and nascent granuloma/a site of free bacteria. A two-tailed Mann-Whitney test was used for the statistical comparison of differences between *il10<sup>e46/e46</sup>* zebrafish and WT controls.



### **Supplementary Figure S4**

*il10<sup>e46/e46</sup>* mutation does not affect the expression levels of lymphocyte markers at the early stages of a low-dose mycobacterial infection. **(a - c)** The relative expression levels of T cell markers (*cd4-1* and *cd8a*) as well as the B cell marker *IgM* were measured in the abdominal organ blocks (including kidney) of *il10<sup>e46/e46</sup>* mutant fish ( $n = 4 - 12$ ) and WT controls ( $n = 5 - 12$ ) at 1 dpi and 6 days post a low-dose (2 - 9 CFU) infection and are presented as a scatter dot plot and median. Note the different scales on the y axes. Gene expressions were normalized to the expression of *eef1a111*. The data were collected from a single experiment. In panels a - c a two-tailed Mann-Whitney test was used for the statistical comparison of differences.

**Supplementary Tables S1-S3** are provided as separate files.

### **Supplementary Table S1**

All mutations found in the whole genome sequencing analysis. *P* values were calculated with Fisher's exact test,  $P < 0.05$  was considered significant.

### **Supplementary Table S2**

Stop codon or frameshift causing mutations found in the whole genome sequencing analysis. Mutations with allele fraction of  $\geq 25\%$  in either of the groups are shown here. *P* values were calculated with Fisher's exact test,  $P < 0.05$  was considered significant.

### **Supplementary Table S3**

Stop codon or frameshift causing mutations found in the whole genome sequencing analysis that differ significantly between *il10<sup>e46/e46</sup>* and WT zebrafish. *P* values were calculated with Fisher's exact test,  $P < 0.05$  was considered significant.

## Supplementary Table S4

### qPCR primers used for gene expression analysis

Gene <sup>1</sup>	ZFIN ID	Human gene ortholog (HGNC)	Sequence 5' - 3'	Reference
<i>eef1a1l1 (ef1a)</i>	ZDB-GENE-990415-52	<i>EEF1A1</i>	F CTGGAGGCCAGCTCAAACAT R ATCAAGAAGAGTAGTACCGCTAGCATTAC	(80)
<i>il10</i>	ZDB-GENE-051111-1	<i>IL10</i>	F GCTCTGCTCACGCTTCTTC R TGGTTCCAAGTCATCGTTG	(51)
<i>tnfa</i>	ZDB-GENE-050317-1	<i>TNF</i>	F GGGCAATCAACAAGATGGAAG R GCAGCTGATGTGCAAAGACAC	(35)
<i>tnfb</i>	ZDB-GENE-050601-2	<i>TNF</i>	F GCATGTGATGAAGCCAAACG R GATTGTCCTGAAGGGTCACC	-
<i>il1b</i>	ZDB-GENE-040702-2	<i>IL1B</i>	F TGGACTTCGCAGCACAAAATG R GTTCACTTCACGCTCTTGGATG	(88)
<i>cd4-1</i>	ZDB-GENE-100922-280	<i>CD4</i>	F TAAAGCACAAAGAAAGCCATG R TACTCTGCGGGTTCCTGTTG	-
<i>cd8a</i>	ZDB-GENE-060210-2	<i>CD8A</i>	F GGAGTACCAGATCCAGTAACACAC R AACCTCCGACCAGAGATGTG	-
<i>igm</i>	ZDB-GENE-030925-46	<i>IGHM</i>	F AGATCCAATACAAAGATACTATGC R TGGTCAAATGGAATTGTGG	(21)
<i>tbx21</i>	ZDB-GENE-080104-3	<i>TBX21</i>	F GGCCTACCAGAAATGCAGACA R GGTGCGTACAGCGTGTGCATA	(27)
<i>gata3</i>	ZDB-GENE-990415-82	<i>GATA3</i>	F GGATGGCACC GGTC ACTATT R CAGCAGACAGCCTCCGTTT	(27)
<i>foxp3a</i>	ZDB-GENE-061116-2	<i>FOXP3</i>	F CAAAAGCAGAGTGCCAGTGG R CGCATAAGCAC CGATTCTGC	(27)
<i>ifng1</i>	ZDB-GENE-040629-1	<i>IFNG</i>	F CTTTCCAGGCAAGAGTGCAGA R TCAGCTCAAACAAAGCCTTTTCG	(89)
<i>il4</i>	ZDB-GENE-100204-1	<i>IL4</i>	F CCAGAGTGTGAATGGGATCC R TTTCCAGTCCCAGGTATATGC	-

<sup>1</sup> Zebrafish gene names and qPCR primer sequences are listed accompanied with the ZFIN identification codes and the names of the orthologous genes in humans. HGNC, HUGO Gene Nomenclature Committee.

## Supplementary References

88. Pressley, M. E., Phelan, P. E., 3rd, Witten, P. E., Mellon, M. T. & Kim, C. H. Pathogenesis and inflammatory response to *Edwardsiella tarda* infection in the zebrafish. *Dev. Comp. Immunol.* **29**, 501-513 (2005).
89. Vojtech, L. N., Sanders, G. E., Conway, C., Ostland, V. & Hansen, J. D. Host immune response and acute disease in a zebrafish model of *Francisella pathogenesis*. *Infect. Immun.* **77**, 914-925 (2009).