

OMTM, Volume 10

Supplemental Information

Integrated Human Evaluation of the Lysophosphatidic Acid Pathway as a Novel Therapeutic Target in Atherosclerosis

Silvia Aldi, Ljubica Perisic Matic, Gregory Hamm, Daniëlle van Keulen, Dennie Tempel, Kim Holmstrøm, Agnieszka Szwajda, Boye Schnack Nielsen, Valur Emilsson, Rima Ait-Belkacem, Mariette Lengquist, Gabrielle Paulsson-Berne, Per Eriksson, Jan H.N. Lindeman, Alain J. Gool, Jonathan Stauber, Ulf Hedin, and Eva Hurt-Camejo

Supplemental Figures and Legends

Figure S1

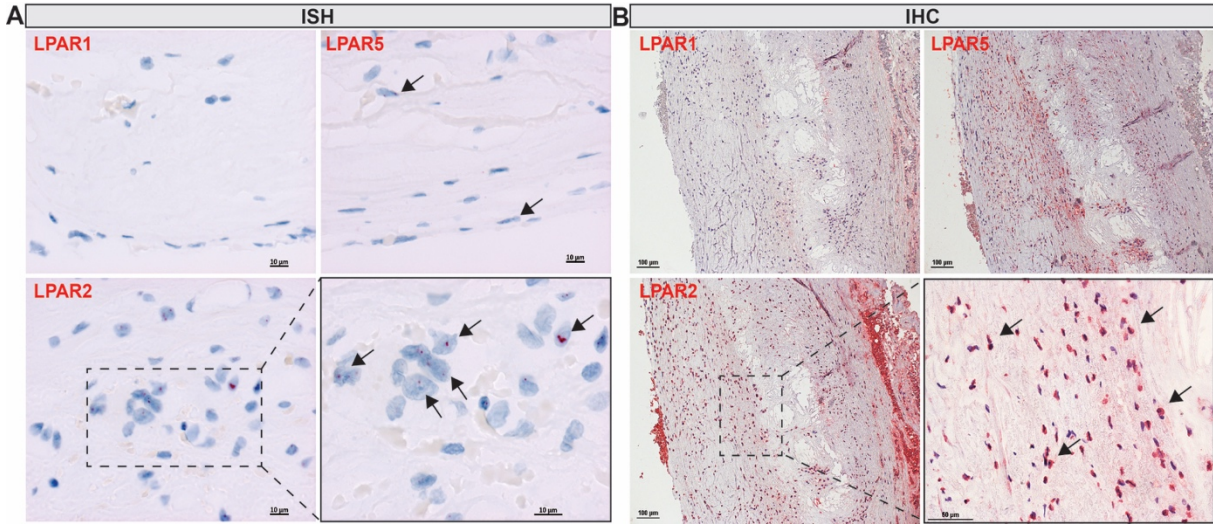


Figure S1. Localisation of LPARs in plaques. A) *In situ* hybridization detection of LPARs 1, 5 and 2 mRNA transcripts in human lesions. Arrows indicate the RNA probe signals (red). **B)** Immunohistochemistry staining of LPARs 1, 5 and 2 in plaques (red signal). Arrows in the enlarged LPAR2 image indicate positive signal in the cells within the necrotic core. Nuclei (purple) are stained with hematoxylin. Images taken with 20x, 40x and 63x objectives.

Figure S2

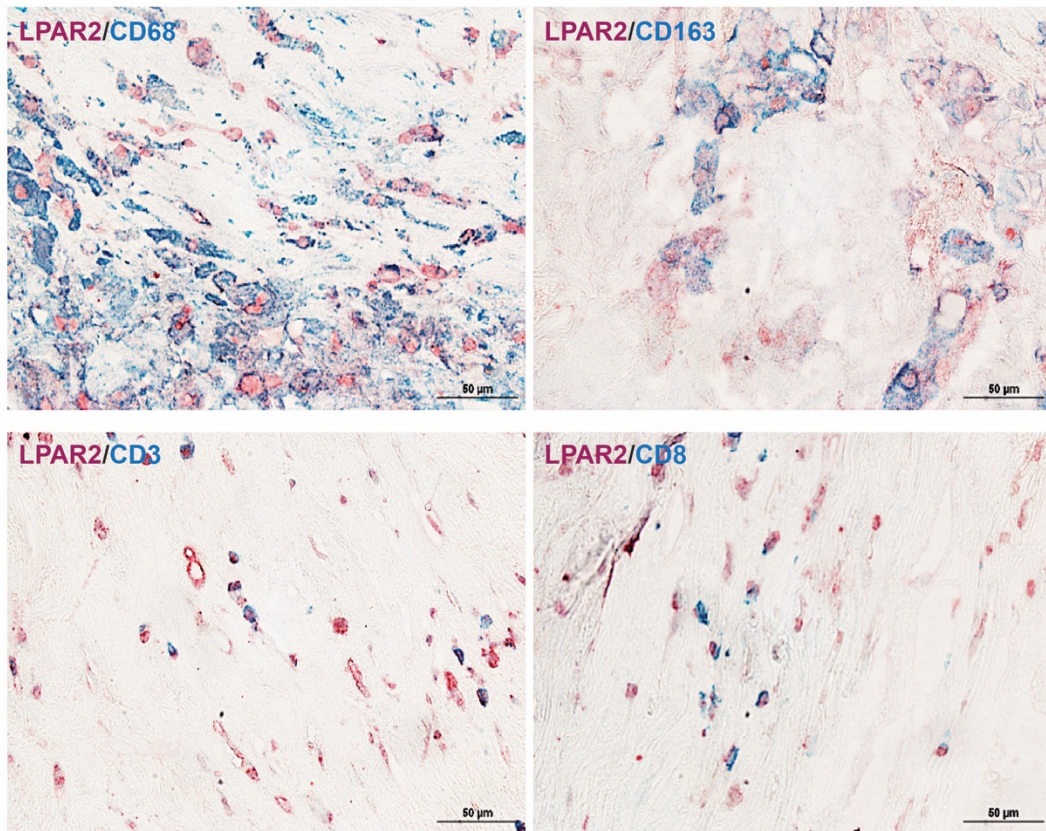


Figure S2. Immunolocalisation of LPAR2 in plaque inflammatory cells. Double immunohistochemistry stainings showing the colocalisation of LPAR2 (red signal) with macrophage cell markers (CD68 and CD163, green signal) and lymphocyte markers (CD3 and CD8, green signal). Images taken with the 40x objective.

Figure S3

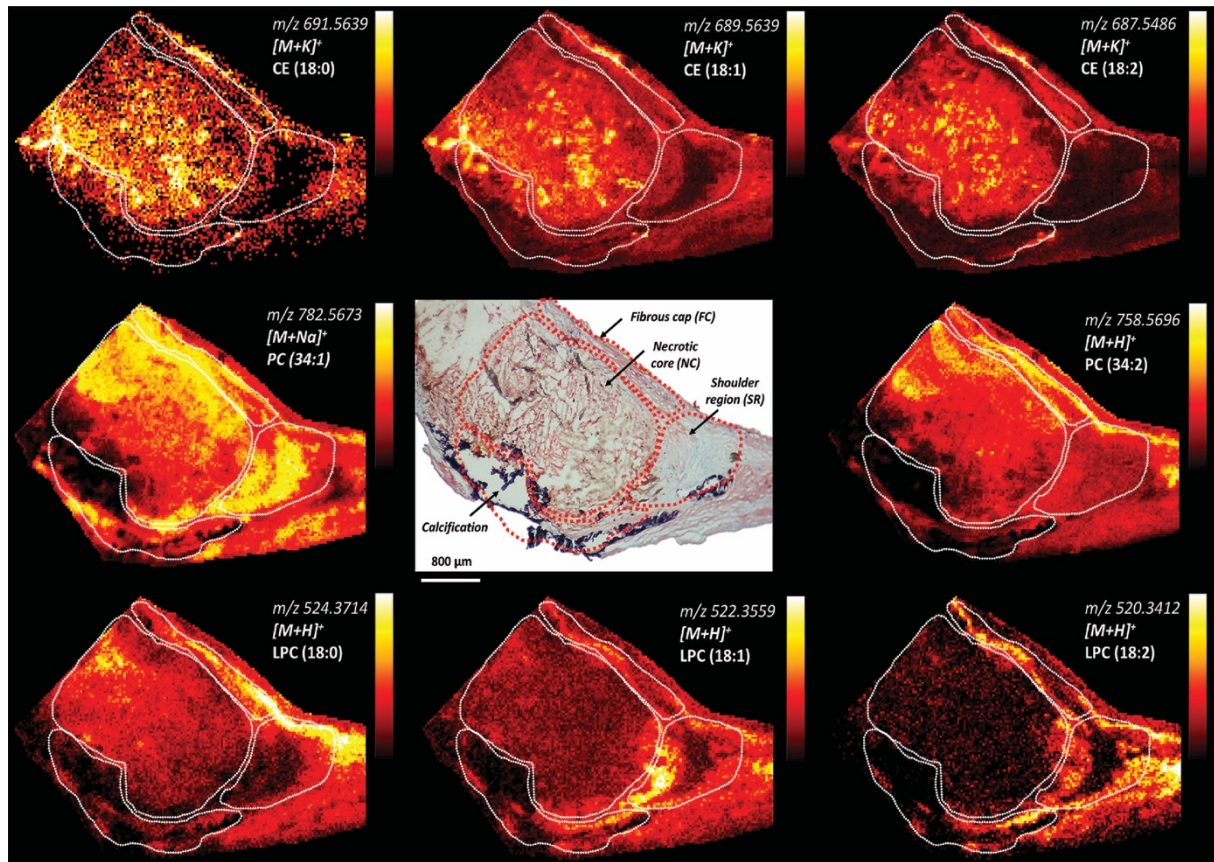


Figure S3. Molecular histology of the human atherosclerotic plaque based on its lipid fingerprint. Molecular distribution of the main lipid species from different classes (CE: Cholesteryl ester, PC: Phosphatidylcholine and LPC: lysophosphatidylcholine) detected in human plaque tissue by mass spectrometry (MS) imaging using MALDI-FTICR in positive detection mode at 30 μm of spatial resolution. Identification of molecular species was performed by accurate MS match with database (<1 ppm) and MS/MS measurement. The different lipid classes are reported on the figure. Relative intensity scale (volcano intensity scale, 0-100%) is indicated on the side of each image. Histological regions of interest were identified by Oil-red-O staining (central picture, arrows).

Figure S4

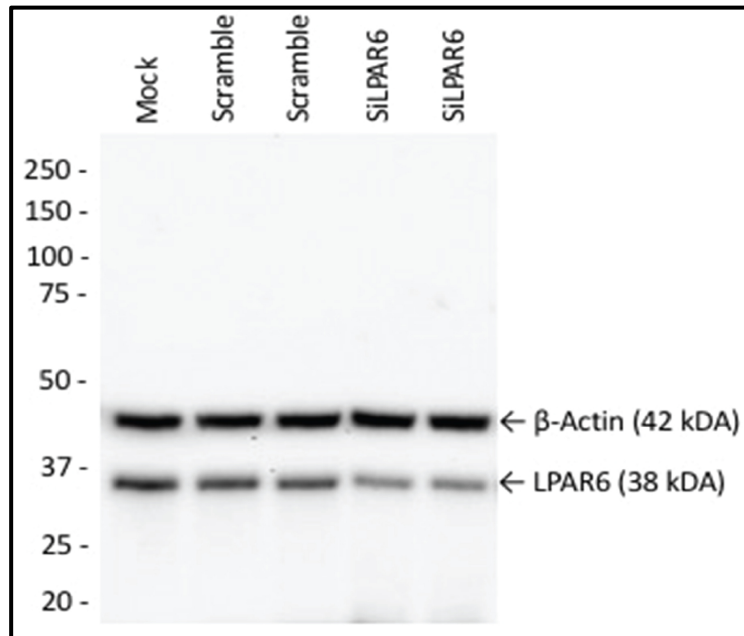


Figure S4. Western blot analysis of LPAR6 protein levels in HUVEC lysates. LPAR6 mRNA silencing resulted in repression of the protein levels compared to non-targeting scramble oligos and mock control. Protein levels of beta-actin were used as loading control. Ladder marks indicated on the left.

Supplemental Tables and Legends

Table S1

Cell type markers	Gene Symbol	Pearson r	p-value	Significance level
Smooth muscle cells				
Myosin heavy chain 11	<i>MYH11</i>	0.2759	0.0018	**
Smoothelin	<i>SMTN</i>	0.2057	0.0209	*
Alpha smooth muscle actin	<i>ACTA2</i>	0.3752	< 0.0001	****
Myocardin	<i>MYOCD</i>	0.3399	< 0.0001	****
Transgelin	<i>TAGLN</i>	0.4809	< 0.0001	****
Endothelial cells				
von Willebrand factor	<i>VWF</i>	0.01506	0.8671	ns
PECAM-1 (CD31)	<i>PECAMI</i>	0.5245	< 0.0001	****
Dendritic cells				
ITGAX (CD11c)	<i>ITGAX</i>	-0.1212	0.1746	ns
LY75 (CD205)	<i>LY75</i>	0.04661	0.6028	ns
CD80	<i>CD80</i>	-0.1459	0.1018	ns
T Lymphocytes				
CD11b	<i>ITGAM</i>	-0.1982	0.0261	*
ITGAL	<i>ITGAL</i>	-0.1949	0.0287	*
CD27	<i>CD27</i>	-0.07268	0.4187	ns
CD28	<i>CD28</i>	0.2105	0.018	*
CD3 delta	<i>CD3D</i>	-0.02751	0.7598	ns
CD4	<i>CD4</i>	0.1938	0.0297	*
CD8A	<i>CD8A</i>	-0.07039	0.4335	ns
PTPRC (CD45RA)	<i>PTPRC</i>	0.3796	< 0.0001	****
CD69	<i>CD69</i>	-0.06686	0.4552	ns
ITGAE	<i>ITGAE</i>	0.08025	0.3698	ns
FABP4	<i>FABP4</i>	0.01908	0.8314	ns
Macrophages				
CD83	<i>CD83</i>	-0.02925	0.7451	ns
CD86	<i>CD86</i>	0.3196	0.0003	***
CD163	<i>CD163</i>	0.2817	0.0014	**
TNFRSF9	<i>TNFRSF9</i>	-0.1674	0.0611	ns
CD40	<i>CD40</i>	-0.1155	0.1977	ns
CD36	<i>CD36</i>	0.1858	0.0365	*
Inflammation/Apoptosis				
Calcification markers				
IL-1beta	<i>IL1B</i>	0.05357	0.5513	ns
NFkB	<i>NFKB1</i>	0.3644	< 0.0001	****
TNF-alpha	<i>TNFA</i>	-0.2183	0.0141	*
MCP-1	<i>CCL2</i>	0.3241	0.0002	***
Caspase-3	<i>CASP3</i>	0.1682	0.0598	ns

Caspase-7	<i>CASP7</i>	0.08284	0.3564	ns
Caspase-9	<i>CASP9</i>	0.1407	0.116	ns
BCL2	<i>BCL2</i>	0.1983	0.026	*
RANTES	<i>CCL5</i>	-0.2318	0.009	**
BMP4	<i>BMP4</i>		< 0.0001	****
Extracellular matrix/degradation				
MMP9	<i>MMP9</i>	0.03487	0.6982	ns
TIMP1	<i>TIMP1</i>	0.3178	0.0003	***
Sulfatase 1	<i>SULF1</i>	0.6295	< 0.0001	****
Sulfatase 2	<i>SULF2</i>	0.1564	0.0803	ns
Growth factors				
TGFB1	<i>TGFB1</i>	0.3565	< 0.0001	****
TGFA	<i>TGFA</i>	-0.1275	0.155	ns
IGF1	<i>IGF1</i>	0.4746	< 0.0001	****
PDGFA	<i>PDGFA</i>	-0.3214	0.0002	***
PDGFB	<i>PDGFB</i>	-0.2063	0.0205	*
PDGFC	<i>PDGFC</i>	0.3403	< 0.0001	****
PDGFD	<i>PDGFD</i>	0.4509	< 0.0001	****
Chemokines and receptors				
CCR2	<i>CCR2</i>	0.425	< 0.0001	****
CCR5	<i>CCR5</i>	0.0959	0.2835	ns
Interleukin 10	<i>IL10</i>	-0.2681	0.0024	**
Interferon gamma	<i>INFG</i>	-0.2756	0.0018	**
IL2	<i>IL2</i>	-0.1016	0.2578	ns
IL6	<i>IL6</i>	0.01629	0.8557	ns
IL4	<i>IL4</i>	-0.2411	0.0065	**
IL5	<i>IL5</i>	-0.01216	0.8925	ns
IL9	<i>IL9</i>	-0.172	0.0542	ns

Table S1. Expression correlation analyses between PPAP2B and genes of interest in plaques. Pearson correlation analyses were calculated from n=127 human plaque microarrays, p-values are corrected for multiple comparisons according to the Bonferroni method. Correlation considered weak if $r < 0.3$ moderate if $0.3 < r < 0.5$ and strong if $r > 0.5$.

Table S2

Cell type markers	LPAR1				LPAR2		
	Gene Symbol	Pearson r	p-value	Significance level	Pearson r	p-value	Significance level
Smooth muscle cells							
Myosin heavy chain 11	<i>MYH11</i>	0.6706	< 0.0001	****	-0.4431	< 0.0001	****
Smoothelin	<i>SMTN</i>	0.4657	< 0.0001	****	-0.4536	< 0.0001	****
Alpha smooth muscle actin	<i>ACTA2</i>	0.6343	< 0.0001	****	-0.4332	< 0.0001	****
Myocardin	<i>MYOCD</i>	0.6585	< 0.0001	****	-0.5676	< 0.0001	****
Transgelin	<i>TAGLN</i>	0.5662	< 0.0001	****	-0.5853	< 0.0001	****
Endothelial cells							
von Willebrand factor	<i>VWF</i>	-0.3296	0.0002	***	0.4895	< 0.0001	****
PECAM-1 (CD31)	<i>PECAMI</i>	-0.1619	0.0701	ns	0.2939	0.0008	***
Dendritic cells							
ITGAX (CD11c)	<i>ITGAX</i>	-0.5713	< 0.0001	****	0.5664	< 0.0001	****
LY75 (CD205)	<i>LY75</i>	-0.002261	0.9799	ns	0.4875	< 0.0001	****
CD80	<i>CD80</i>	-0.3983	< 0.0001	****	0.4718	< 0.0001	****
T Lymphocytes							
CD11b	<i>ITGAM</i>	-0.5282	< 0.0001	****	0.4891	< 0.0001	****
ITGAL	<i>ITGAL</i>	-0.4416	< 0.0001	****	0.6066	< 0.0001	****
CD27	<i>CD27</i>	-0.419	< 0.0001	****	0.3069	0.0005	***
CD28	<i>CD28</i>	-0.2708	0.0022	**	0.4252	< 0.0001	****
CD3 delta	<i>CD3D</i>	-0.3652	< 0.0001	****	0.1419	0.113	ns
CD4	<i>CD4</i>	-0.5254	< 0.0001	****	0.08309	0.355	ns
CD8A	<i>CD8A</i>	-0.3322	0.0001	***	0.2653	0.0027	**
PTPRC (CD45RA)	<i>PTPRC</i>	-0.2482	0.0049	**	0.3816	< 0.0001	****
CD69	<i>CD69</i>	-0.07406	0.408	ns	0.4384	< 0.0001	****
ITGAE	<i>ITGAE</i>	-0.1859	0.0364	*	0.5556	< 0.0001	****
FABP4	<i>FABP4</i>	-0.3392	< 0.0001	****	0.4484	< 0.0001	****
Macrophages							
CD83	<i>CD83</i>	-0.4417	< 0.0001	****	0.465	< 0.0001	****
CD86	<i>CD86</i>	-0.4989	< 0.0001	****	0.5162	< 0.0001	****
RANK	<i>TNFRSF11A</i>	-0.4012	< 0.0001	****	0.6283	< 0.0001	****
CD163	<i>CD163</i>	-0.4111	< 0.0001	****	0.5445	< 0.0001	****
TNFRSF9	<i>TNFRSF9</i>	-0.621	< 0.0001	****	0.3327	0.0001	***
CD40	<i>CD40</i>	-0.5126	< 0.0001	****	0.4241	< 0.0001	****
CD36	<i>CD36</i>	-0.3921	< 0.0001	****	0.5142	< 0.0001	****
Inflammation/Apoptosis Calcification markers							
IL-1beta	<i>IL1B</i>	-0.2719	0.0021	**	0.242	0.0063	**
NFkB	<i>NFKB1</i>	-0.1226	0.1714	ns	0.1519	0.0896	ns
TNF-alpha	<i>TNFA</i>	-0.484	< 0.0001	****	0.3666	< 0.0001	****
MCP-1	<i>CCL2</i>	-0.1699	0.0571	ns	0.01676	0.8522	ns

Caspase-3	<i>CASP3</i>	-0.05183	0.5644	ns	0.5511	< 0.0001	****
Caspase-7	<i>CASP7</i>	-0.005042	0.9553	ns	0.3474	< 0.0001	****
Caspase-9	<i>CASP9</i>	-0.02516	0.7797	ns	0.2577	0.0036	**
BCL2	<i>BCL2</i>	0.4797	< 0.0001	****	-0.4165	< 0.0001	****
RANTES	<i>CCL5</i>	-0.4499	< 0.0001	****	0.44	< 0.0001	****
BMP4	<i>BMP4</i>	0.04733	0.5987	ns	0.4274	< 0.0001	****
Extracellular matrix/degradation							
MMP9	<i>MMP9</i>	-0.4874	< 0.0001	****	0.4547	< 0.0001	****
TIMP1	<i>TIMP1</i>	-0.3947	< 0.0001	****	0.334	0.0001	***
Sulfatase 1	<i>SULF1</i>	-0.325	0.0002	***	0.5368	< 0.0001	****
Sulfatase 2	<i>SULF2</i>	0.3379	0.0001	***	-0.3382	0.0001	***
Growth factors							
TGFB1	<i>TGFB1</i>	-0.2904	0.001	***	-0.4719	< 0.0001	****
TGFA	<i>TGFA</i>	-0.3053	0.0005	***	0.5867	< 0.0001	****
IGF1	<i>IGF1</i>	-0.1903	0.0328	*	0.4513	< 0.0001	****
PDGFA	<i>PDGFA</i>	0.5479	< 0.0001	****	0.02336	0.7951	ns
PDGFB	<i>PDGFB</i>	-0.408	< 0.0001	****	0.1198	0.1813	ns
PDGFC	<i>PDGFC</i>	0.5948	< 0.0001	****	-0.1977	0.0265	*
PDGFD	<i>PDGFD</i>	0.6034	< 0.0001	****	-0.4602	< 0.0001	****
Chemokines and receptors							
CCR2	<i>CCR2</i>	-0.07643	0.3931	ns	0.3701	< 0.0001	****
CCR5	<i>CCR5</i>	-0.4306	< 0.0001	****	0.4636	< 0.0001	****
Interleukin 10	<i>IL10</i>	-0.3689	< 0.0001	****	0.5818	< 0.0001	****
Interferon gamma	<i>INFG</i>	-0.3075	0.0005	***	-0.03486	0.6984	ns
IL2	<i>IL2</i>	-0.1837	0.0395	*	-0.2099	0.0183	*
IL6	<i>IL6</i>	-0.03277	0.7145	ns	0.2466	0.0052	**
IL4	<i>IL4</i>	-0.1667	0.0621	ns	-0.2958	0.0008	***
IL5	<i>IL5</i>	-0.007118	0.9369	ns	-0.4802	< 0.0001	****
IL9	<i>IL9</i>	-0.1642	0.0662	ns	-0.206	0.0206	*

Cell type markers	Gene Symbol	LPAR3			LPAR4		
		Pearson r	p-value	Significance level	Pearson r	p-value	Significance level
Smooth muscle cells							
Myosin heavy chain 11	<i>MYH11</i>	-0.2946	0.0008	***	0.1502	0.0933	ns
Smoothelin	<i>SMTN</i>	0.4251	< 0.0001	****	-0.3744	< 0.0001	****
Alpha smooth muscle actin	<i>ACTA2</i>	-0.2514	0.0045	**	0.1165	0.1939	ns
Myocardin	<i>MYOCD</i>	-0.09252	0.3028	ns	0.03997	0.6568	ns
Transgelin	<i>TAGLN</i>	0.03132	0.7278	ns	-0.1038	0.2472	ns
Endothelial cells							
von Willebrand factor	<i>VWF</i>	-0.3106	0.0004	***	0.1736	0.0519	ns
PECAM-1 (CD31)	<i>PECAMI</i>	-0.1621	0.0698	ns	0.1127	0.2089	ns
Dendritic cells							
ITGAX (CD11c)	<i>ITGAX</i>	0.3511	< 0.0001	****	-0.07009	0.4336	ns

LY75 (CD205)	<i>LY75</i>	-0.4494	< 0.0001	****	0.06596	0.4613	ns
CD80	<i>CD80</i>	-0.2953	0.0007	***	0.1664	0.0614	ns
T Lymphocytes							
CD11b	<i>ITGAM</i>	0.4518	< 0.0001	****	-0.4083	< 0.0001	****
ITGAL	<i>ITGAL</i>	-0.1041	0.2458	ns	-0.01843	0.8377	ns
CD27	<i>CD27</i>	0.03483	0.6987	ns	-0.2123	0.017	*
CD28	<i>CD28</i>	-0.254	0.0041	**	-0.2748	0.0018	**
CD3 delta	<i>CD3D</i>	0.1412	0.1148	ns	-0.1471	0.1001	ns
CD4	<i>CD4</i>	0.2873	0.0011	**	-0.4718	< 0.0001	****
CD8A	<i>CD8A</i>	0.05853	0.515	ns	-0.09318	0.2994	ns
PTPRC (CD45RA)	<i>PTPRC</i>	-0.2953	0.0007	***	0.3897	< 0.0001	****
CD69	<i>CD69</i>	-0.4398	< 0.0001	****	0.02757	0.7583	ns
ITGAE	<i>ITGAE</i>	-0.2002	0.024	*	0.03967	0.6579	ns
FABP4	<i>FABP4</i>	-0.02759	0.7582	ns	-0.07478	0.4034	ns
Macrophages							
CD83	<i>CD83</i>	0.06016	0.5034	ns	-0.09816	0.2742	ns
CD86	<i>CD86</i>	-0.005174	0.9541	ns	-0.09719	0.279	ns
RANK	<i>TNFRSF11A</i>	0.02527	0.7788	ns	-0.1704	0.0564	ns
CD163	<i>CD163</i>	-0.268	0.0024	**	0.02635	0.7696	ns
TNFRSF9	<i>TNFRSF9</i>	0.3201	0.0003	***	-0.2192	0.0137	*
CD40	<i>CD40</i>	0.3923	< 0.0001	****	0.4597	< 0.0001	****
CD36	<i>CD36</i>	-0.1493	0.0939	ns	-0.1578	0.0765	ns
Inflammation/Apoptosis							
Calcification markers							
IL-1beta	<i>IL1B</i>	-0.01481	0.8692	ns	-0.1345	0.1333	ns
NFkB	<i>NFKB1</i>	0.007307	0.9353	ns	-0.1105	0.2179	ns
TNF-alpha	<i>TNFA</i>	0.1344	0.1334	ns	-0.1546	0.0839	ns
MCP-1	<i>CCL2</i>	-0.001336	0.9882	ns	-0.1377	0.1243	ns
Caspase-3	<i>CASP3</i>	-0.4667	< 0.0001	****	0.2803	0.0015	**
Caspase-7	<i>CASP7</i>	-0.446	< 0.0001	****	0.106	0.2374	ns
Caspase-9	<i>CASP9</i>	-0.3026	0.0006	***	0.3252	0.0002	***
BCL2	<i>BCL2</i>	0.487	< 0.0001	****	-0.1836	0.0396	*
RANTES	<i>CCL5</i>	0.07816	0.3843	ns	-0.2627	0.003	**
BMP4	<i>BMP4</i>	-0.3192	0.0003	***	0.162	0.07	ns
Extracellular matrix/degradation							
MMP9	<i>MMP9</i>	-0.004226	0.9625	ns	-0.1193	0.1834	ns
TIMP1	<i>TIMP1</i>	-0.009881	0.9126	ns	-0.2127	0.0168	*
Sulfatase 1	<i>SULF1</i>	-0.2694	0.0023	**	-0.08024	0.3718	ns
Sulfatase 2	<i>SULF2</i>	-0.5118	< 0.0001	****	0.1538	0.0856	ns
Growth factors							
TGFB1	<i>TGFB1</i>	0.4135	< 0.0001	****	-0.3427	< 0.0001	****
TGFA	<i>TGFA</i>	0.4058	< 0.0001	****	0.123	0.17	ns
IGF1	<i>IGF1</i>	-0.2969	0.0007	***	0.1296	0.148	ns
PDGFA	<i>PDGFA</i>	-0.5459	< 0.0001	****	0.3771	< 0.0001	****
PDGFB	<i>PDGFB</i>	0.2634	0.0029	**	-0.1694	0.058	ns

PDGFC	<i>PDGFC</i>	-0.4801	< 0.0001	****	0.269	0.0023	**
PDGFD	<i>PDGFD</i>	-0.2303	0.0095	**	0.2796	0.0015	**
Chemokines and receptors							
CCR2	<i>CCR2</i>	-0.3749	< 0.0001	****	0.05098	0.5692	ns
CCR5	<i>CCR5</i>	-0.06846	0.4444	ns	-0.2076	0.0192	*
Interleukin 10	<i>IL10</i>	-0.1088	0.2254	ns	-0.1253	0.1622	ns
Interferon gamma	<i>INFG</i>	0.1622	0.0696	ns	-0.1909	0.0323	*
IL2	<i>IL2</i>	0.3133	0.0004	***	-0.2224	0.0123	*
IL6	<i>IL6</i>	-0.2154	0.015	*	-0.05445	0.5432	ns
IL4	<i>IL4</i>	0.4896	< 0.0001	****	-0.3051	0.0005	***
IL5	<i>IL5</i>	0.404	< 0.0001	****	0.02027	0.8218	ns
IL9	<i>IL9</i>	0.4855	< 0.0001	****	-0.2934	0.0009	***

Cell type markers	Gene Symbol	LPA5			LPA6		
		Pearson r	p-value	Significance level	Pearson r	p-value	Significance level
Smooth muscle cells							
Myosin heavy chain 11	<i>MYH11</i>	0.3653	< 0.0001	****	-0.2408	0.0066	**
Smoothelin	<i>SMTN</i>	-0.3695	< 0.0001	****	-0.5444	< 0.0001	****
Alpha smooth muscle actin	<i>ACTA2</i>	-0.3167	0.0003	***	-0.2255	0.0111	*
Myocardin	<i>MYOCD</i>	-0.407	< 0.0001	****	-0.2115	0.0174	*
Transgelin	<i>TAGLN</i>	-0.422	< 0.0001	****	-0.2844	0.0013	**
Endothelial cells							
von Willebrand factor	<i>VWF</i>	0.472	< 0.0001	****	0.6393	< 0.0001	****
PECAM-1 (CD31)	<i>PECAMI</i>	0.1995	0.0251	*	0.163	0.0683	ns
Dendritic cells							
ITGAX (CD11c)	<i>ITGAX</i>	0.4244	< 0.0001	****	-0.3362	0.0001	***
LY75 (CD205)	<i>LY75</i>	0.4917	< 0.0001	****	0.4139	< 0.0001	****
CD80	<i>CD80</i>	0.5655	< 0.0001	****	0.4392	< 0.0001	****
T Lymphocytes							
CD11b	<i>ITGAM</i>	0.3943	< 0.0001	****	0.3279	0.0002	***
ITGAL	<i>ITGAL</i>	0.5303	< 0.0001	****	0.511	< 0.0001	****
CD27	<i>CD27</i>	0.2685	0.0024	**	0.3421	< 0.0001	****
CD28	<i>CD28</i>	0.6452	< 0.0001	****	0.5678	< 0.0001	****
CD3 delta	<i>CD3D</i>	0.2161	0.0151	*	0.2485	0.005	**
CD4	<i>CD4</i>	0.1155	0.1977	ns	0.05921	0.5102	ns
CD8A	<i>CD8A</i>	0.2882	0.0011	**	0.3144	0.0003	***
PTPRC (CD45RA)	<i>PTPRC</i>	0.4544	< 0.0001	****	0.6948	< 0.0001	****
CD69	<i>CD69</i>	0.5144	< 0.0001	****	0.5524	< 0.0001	****
ITGAE	<i>ITGAE</i>	0.2536	0.004	**	0.2311	0.0089	**
FABP4	<i>FABP4</i>	0.1492	0.0942	ns	0.1873	0.035	*
Macrophages							
CD83	<i>CD83</i>	0.4243	< 0.0001	****	0.2171	0.0146	*
CD86	<i>CD86</i>	0.4769	< 0.0001	****	0.5662	< 0.0001	****
RANK	<i>TNFRSF11A</i>	0.5756	< 0.0001	****	0.4303	< 0.0001	****
CD163	<i>CD163</i>	0.4787	< 0.0001	****	0.5628	< 0.0001	****

TNFRSF9	<i>TNFRSF9</i>	0.2011	0.0239	*	-0.01977	0.8261	ns
CD40	<i>CD40</i>	0.4126	< 0.0001	****	0.2321	0.0089	**
CD36	<i>CD36</i>	0.3239	0.0002	***	0.3013	0.0006	***
Inflammation/Apoptosis							
Calcification markers							
IL-1beta	<i>IL1B</i>	0.4214	< 0.0001	****	0.3324	0.0001	***
NFkB	<i>NFKB1</i>	0.2796	0.0015	**	0.3435	< 0.0001	****
TNF-alpha	<i>TNFA</i>	0.3693	< 0.0001	****	0.3589	< 0.0001	****
MCP-1	<i>CCL2</i>	0.1886	0.0344	*	0.334	0.0001	***
Caspase-3	<i>CASP3</i>	0.4408	< 0.0001	****	0.5471	< 0.0001	****
Caspase-7	<i>CASP7</i>	0.4525	< 0.0001	****	0.5657	< 0.0001	****
Caspase-9	<i>CASP9</i>	0.365	< 0.0001	****	0.3399	< 0.0001	****
BCL2	<i>BCL2</i>	-0.3679	< 0.0001	****	-0.2746	0.0019	**
RANTES	<i>CCL5</i>	0.4579	< 0.0001	****	0.3982	< 0.0001	****
BMP4	<i>BMP4</i>	0.2555	0.0039	**	-0.0006809	0.994	ns
Extracellular matrix/degradation							
MMP9	<i>MMP9</i>	0.218	0.0142	*	0.1144	0.2023	ns
TIMP1	<i>TIMP1</i>	0.3771	< 0.0001	****	0.2514	0.0045	**
Sulfatase 1	<i>SULF1</i>	0.4413	< 0.0001	****	0.4089	< 0.0001	****
Sulfatase 2	<i>SULF2</i>	0.3157	0.0003	***	0.3571	< 0.0001	****
Growth factors							
TGFB1	<i>TGFB1</i>	-0.4978	< 0.0001	****	-0.2573	0.0036	**
TGFA	<i>TGFA</i>	0.5957	< 0.0001	****	0.57	< 0.0001	****
IGF1	<i>IGF1</i>	0.4343	< 0.0001	****	0.6966	< 0.0001	****
PDGFA	<i>PDGFA</i>	0.08341	0.3531	ns	0.2096	0.0185	*
PDGFB	<i>PDGFB</i>	0.1527	0.0878	ns	0.04191	0.6412	ns
PDGFC	<i>PDGFC</i>	0.06585	0.4638	ns	0.2563	0.0038	**
PDGFD	<i>PDGFD</i>	-0.2219	0.0125	*	0.1722	0.0539	ns
Chemokines and receptors							
CCR2	<i>CCR2</i>	0.5877	< 0.0001	****	0.7626	< 0.0001	****
CCR5	<i>CCR5</i>	0.4241	< 0.0001	****	0.4083	< 0.0001	****
Interleukin 10	<i>IL10</i>	0.4787	< 0.0001	****	0.3627	< 0.0001	****
Interferon gamma	<i>INFG</i>	-0.01569	0.8616	ns	0.01233	0.891	ns
IL2	<i>IL2</i>	-0.175	0.0501	ns	-0.1519	0.0895	ns
IL6	<i>IL6</i>	0.3142	0.0003	***	0.2623	0.0029	**
IL4	<i>IL4</i>	-0.1973	0.0268	*	-0.4218	< 0.0001	****
IL5	<i>IL5</i>	-0.4005	< 0.0001	****	-0.2844	0.0012	**
IL9	<i>IL9</i>	-0.2452	0.0056	**	-0.3565	< 0.0001	****

Table S2. Expression correlation analyses between LPARs and various markers in plaques.

Pearson correlation analyses were calculated from n=127 human plaque microarrays, p-values are corrected for multiple comparisons according to the Bonferroni method. Correlation considered weak if $r < 0.3$, moderate if $0.3 < r < 0.5$ and strong if $r > 0.5$.

Table S3

Correlation	Pearson r	95% confidence interval	P (two-tailed)	Significance level
PPAP2B vs. LPAR1	0,1586	-0.01608 to 0.3239	0,0749	ns
PPAP2B vs. LPAR2	-0,4009	-0.5376 to -0.2437	< 0.0001	****
PPAP2B vs. LPAR3	-0,2321	-0.3906 to -0.06033	0,0086	**
PPAP2B vs. LPAR4	-0,1145	-0.2831 to 0.06098	0,1999	ns
PPAP2B vs. LPAR5	-0,09513	-0.2650 to 0.08046	0,2874	ns
PPAP2B vs. LPAR6	0,366	0.2048 to 0.5079	< 0.0001	****

Table S3. Expression correlation analyses between PPAP2B and various LPARs in plaques. Pearson correlation analyses were calculated from n=127 human plaque microarrays, p-values are corrected for multiple comparisons according to the Bonferroni method. Correlation considered weak if $r < 0.3$ moderate if $0.3 < r < 0.5$ and strong if $r > 0.5$.

Table S4

^a Compound Name	Ion Cluster	^b <i>m/z calcd</i>	^c <i>m/z measd</i>	^d Error (ppm)	
CE (18:2)	Cholesteryl linoleate	[M+K] ⁺	687,5477	687,5486	1,3
CE (18:1)	Cholesteryl oleate	[M+K] ⁺	689,5633	689,5639	0,9
CE (18:0)	Cholesteryl stearate	[M+K] ⁺	691,5790	691,5798	1,2
PC (34:1)	Phophatydilcholine	[M+Na] ⁺	782,5670	782,5673	0,3
PC (34:2)	Phophatydilcholine	[M+H] ⁺	758,5694	758,5696	0,2
LPC (18:2)	LysoPC	[M+H] ⁺	520,3409	520,3412	0,6
LPC (18:1)	LysoPC	[M+H] ⁺	522,3554	522,3559	1,0
LPC (18:0)	LysoPC	[M+H] ⁺	524,3710	524,3714	0,8
PA (36:2)	Phosphatidic acid	[M-H] ⁻	699,4970	699,4976	0,9
LPA (18:0)	LysoPA	[M-H ₂ O-H] ⁻	419,2568	419,2572	1,0

Table S4. Analysis of lipid species in human carotid plaques.

Molecular species (a) observed in human plaque by positive and negative ionization detection mode based on *m/z* value. (b) *m/z* calculated from molecular formula and (c) *m/z* measured using FTICR-MS (d) mass accuracy error unit in part per million (ppm).

Supplemental Methods

Proteomic analysis of plaques

Atherosclerotic plaques from n=18 BiKE patients (n=9 symptomatic + 9 asymptomatic; matched for male gender, age and statin medication) were analysed using LC-MS/MS as previously described¹. Briefly, protein samples were digested by trypsin and the resulting tryptic peptides were TMT-labeled and pooled. Pooled samples were cleaned by Strong Cation exchange columns (Phenomenex) and subjected to LC-MS/MS analysis. The sample pools were separated on a 4 hour gradient using an UPLC-system (Dionex UltiMate™ 3000) coupled to a Q-Exactive mass spectrometer (Thermo Fischer Scientific, San Jose, CA, USA). The fragment spectra from the mass spectrometer were matched to a database consisting of theoretical fragment spectra from all human proteins and filtered at a 1% False Discovery Rate on the peptide level to obtain protein identities (Uniprot). Quantitative information was acquired using the TMT reporter ion intensities.

Antibodies

For immunohistochemical studies, the following primary antibodies with the relative concentrations were used: PPAP2B 1:50, LPAR1 1:500 (both from Novus Biologicals, Littleton, CO, USA), LPAR2 1:250, LPAR5, LPAR6 1:400 (all from LSBio, Seattle, WA, USA), smooth muscle α -actin 1:1000 (SMA, DAKO Sweden AB, Stockholm, SE), anti-CD68 1:50 (Novocastra, Bromma, SE) and anti-CD31 1:200 (Abcam, Cambridge, UK).

Immunohistochemical (IHC) stainings

IHC reagents were from Biocare Medical (Concord, CA). Tissues were treated as previously described². In brief, tissues were fixed for 48 hours in 4% Zn-formaldehyde at room temperature and paraffin-embedded. Isotype rabbit and mouse IgG were used as negative controls. 5 μ m tissue sections were deparaffinized in Tissue Clear and rehydrated in graded ethanol. For antigen retrieval, slides were heated in DIVA buffer (pH 6.0) for 20 min with peak at 121°C. Blocking was performed with Background Sniper for 20 min and primary antibodies were diluted in Da Vinci Green solution, and incubated at room temperature for 1 hour. The single stainings were detected by a probe-polymer system for rabbit, followed by Warp Red chromogen. For double staining, a double-stain probe-polymer system containing alkaline phosphatase and horseradish peroxidase was applied, followed by detection with Warp Red and Vina Green. All slides were counterstained with Hematoxylin QS (Vector Laboratories, Burlingame, CA) and mounted in Pertex (Histolab, Gothenburg, Sweden). Images were scanned by an automated ScanScope slidescanner (Hamamatsu, Kista, Sweden) or acquired by a Nikon OPTIPHOT-2 microscope equipped with a digital camera and processed with NIS-Elements software. Magnifications are indicated in the figure legends.

In vitro experiments

Primary human umbilical vein endothelial cells (HUVECs, Lonza, The Netherlands) were cultured in EBM2 medium supplemented with bullet kit (EGM-2, Lonza) under normoxic conditions (21% O₂). Passages 5 to 7 were used throughout the study. Knockdown of LPAR6 and PPAP2B was achieved by transfection with a mix of 4 specific siRNA sequences directed against the human mRNA (SMARTpool siGENOME, GE Dharmacon, Lafayette, CO) in 70% subconfluent HUVEC cultures. Cells were incubated for 1 hour in a small volume of EGM-2 medium supplemented with DharmaFECT 1 (GE Dharmacon, Lafayette, CO) according to manufacturer's instructions. After 1-2 hours cells were supplemented with extra EGM-2 medium to complement medium volumes. As controls, HUVECs were transfected with a mix of 4 scrambled, non-targeting siRNAs (siSham Smartpool; GE Dharmacon, Lafayette, CO). 48h after siRNA transfection HUVECs were treated with 10 μ M Lysophosphatidic acid (Santa Cruz, Dallas, TX). 2 hours after LPA treatment cells were harvested and RNA was isolated with the NucleoSpin RNA kit according to the manufacturer's protocol (Macherey-Nage, Düren, Germany). Isolated RNA (500 ng) was reverse transcribed into cDNA (iScript Adv cDNA kit for RT-qPCR, Bio-Rad, Hercules, CA) and analyzed by real-time fluorescence assessment of SYBR® Green signal in the iCycler iQ Detection system (Bio-Rad). Primers were designed for the human genes of interest. mRNA levels were analyzed and corrected for the housekeeping gene beta-actin. Reported values were normalized to sham that was arbitrarily assigned an average value of 1³.

Supplemental References

1. Branca RM, Orre LM, Johansson HJ, Granholm V, Huss M, Perez-Bercoff A, Forshed J, Kall L and Lehtio J. HiRIEF LC-MS enables deep proteome coverage and unbiased proteogenomics. *Nature methods*. 2014;11:59-62.
2. Perisic L, Aldi S, Sun Y, Folkersen L, Razuvaev A, Roy J, Lengquist M, Akesson S, Wheelock CE, Maegdefessel L, Gabrielsen A, Odeberg J, Hansson GK, Paulsson-Berne G and Hedin U. Gene expression signatures, pathways and networks in carotid atherosclerosis. *Journal of internal medicine*. 2015.
3. Cheng C, Tempel D, Den Dekker WK, Haasdijk R, Chrifi I, Bos FL, Wagtmans K, van de Kamp EH, Blonden L, Biessen EA, Moll F, Pasterkamp G, Serruys PW, Schulte-Merker S and Duckers HJ. Ets2 determines the inflammatory state of endothelial cells in advanced atherosclerotic lesions. *Circ Res*. 2011;109:382-95.