

Protocol - amplification of Oligopaints library and probe synthesis

Step 1: low cycle PCR reaction using raw library as template

PCR amplification

1 ul diluted raw library (100-200 pg/ul)
1.25 ul F primer (20 uM)
1.25 ul R primer (20 uM)
21.5 ul H₂O
50 ul total

Cycling:
1. 98°C - 3 min
2. 98°C - 5 s
3. 72 °C - 15 s
4. Repeat 2-5 30x
5. 72 °C - 2 min
6. 12 °C - hold

PCR clean-up (kit)

- elute in 30ul
- dilute to 4ng/ul

Step 2: Adding T7 sites to Oligos

- Add **TAATACGACTCACTATAGGG** to the 5' end of the reverse primer

T7 PCR (1X)

2.5 ul 4ng/ul template
4 ul 10uM F primer
4 ul 10uM R primer
20 ul 5x buffer with MgCl₂ added (Invitrogen)
2 ul dNTPs (10mM)
0.5 ul taq polymerase (Invitrogen)
H₂O to 100 ul
100ul total

Cycling:
1. 94°C – 3 min
2. 94°C – 45 sec
3. 56°C – 30 sec
4. 72°C – 30 sec
5. Repeat 2-5 35X
6. 72°C – 5 min
7. 12°C - hold

PCR clean-up (kit)

- elute in 20ul

Step 3: *in vitro* transcription

T7 RNA synthesis (Hi-Scribe kit)

7 ul clean T7 PCR
2ul ATP
2ul CTP
2ul GTP
2ul UTP
2ul T7 buffer
2ul T7 pol mix
1ul RNase OUT
20 ul total
→ incubate at 37°C O/N

Step 4: conversion of RNA to RNA:DNA duplexes with reverse transcription

RT

- 20ul T7 RNA sample
 - 7.5ul F sec primer (200uM)
 - 9.6 ul dNTPs (100 mM dNTP mix, or 25 mM of each nucleotide)
 - 30 ul 5x RT buffer
 - 1.5ul RNase OUT
 - 2ul Maxima RT-H
 - 79.4ul H2O
 - 150ul total
- incubate at 50°C for 2 hours
heat water bath to 95°C for next step

Step 5: RNA degradation and DNA probe purification

Alkaline Hydrolysis

- Make 1:1 mixture of 0.5M EDTA:1M NaOH (need 1:1 ratio with sample [150 ul])
- degrade RNA by adding 150 ul EDTA:NaOH mix and incubating at 95°C for 10 minutes

Oligo Clean-up (Zymo-100 DNA Clean and Concentrator Kit)

- For each sample, make a 15mL conical tube containing:
 - 600 ul Oligo binding buffer (D4060-1-40, Zymo)
 - 2400 ul 100% ethanol
 - 300ul sample
- Load onto column and centrifuge or vacuum
- wash 2X with 2mL wash buffer
- spin column dry in microcentrifuge tube (~1min at max speed)
- elute in 150ul dH2O

→ nanodrop to get concentrations and convert to pM