

Supporting Information for

Synthesis of Cyclic Chiral α -Amino Boronates by Copper-catalyzed Asymmetric Dearomative Borylation of Indoles

Lili Chen, Jun-Jian Shen, Qian Gao, and Senmiao Xu*

State Key Laboratory for Oxo Synthesis and Selective Oxidation, Suzhou Research Institute, Lanzhou Institute of Chemical Physics (LICP), Chinese Academy of Sciences,

Lanzhou 730000, China

senmiaoxu@licp.cas.cn

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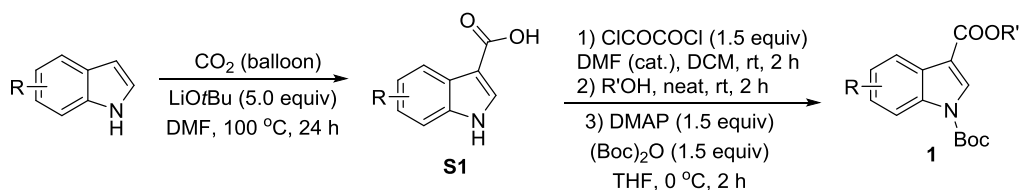
1. General

All oxygen- and moisture-sensitive manipulations were carried out under an inert atmosphere using standard Schlenk techniques or glovebox.

THF, Et₂O, CH₂Cl₂, toluene, CH₃CN, and 1,4-dioxane were purified by passing through a neutral alumina column under argon. All other chemicals and solvents were purchased and used as received.

¹H NMR, ¹³C NMR and ¹¹B NMR spectra were recorded on Zhongke-Niujin 400, Bruker 400, Bruker 600 NMR spectrometer at ambient temperature. ¹³C shifts were obtained with ¹H decoupling. ¹¹B NMR chemical shifts are externally referenced to BF₃ OEt₂ (δ 0). HPLC data were collected on a Shimadzu LC-20AT spectrometer. Optical Rotation was recorded on a Perkin Elmer 341 polarimeter. High-resolution mass spectroscopy data were obtained on Agilent 6530, Agilent 6224 TOF LC/MS spectrometer. X-ray crystallography was measured on Bruker Smart APEX II.

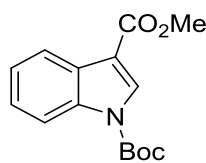
2. General procedures for the synthesis of *N*-Boc indole-3-carboxylate **1** (GPI)



The preparation of indole-3-carboxylic acid **S1** was according to literature procedures.¹ To a 100mL flame-dried two-necked flask was charged with LiOtBu (2.0 g, 25 mmol, 5.0 equiv) was added indole (5 mmol, 1.0 equiv). The reaction vessel was evacuated under high vacuum and the atmosphere was replaced with a balloon of CO₂. Then DMF (25 mL) was added and the mixture was allowed to stir for 24 h at 100 °C. Then the resulting mixture was cooled and carefully quenched by a solution of HCl (2 M, 50 mL) and extracted with EtOAc 3 times (3×30 mL). The combined organic layers were washed with water (50 mL), brine (50 mL) and dry over MgSO₄. The dried organic phase was concentrated under reduced pressure and the residue (**S1**) was used in the next step without further purification.

Steps 2 and 3 were adapted from literature procedures.^{2,3} To a 100-mL flask charged with crude **S1**, DCM (50 mL) and DMF (0.1 mL) was added (COCl)₂ (0.96 g, 7.5 mmol) slowly at 0 °C. The resulting mixture was then allowed to warm to room temperature and stir at same temperature for 2 h. After concentration, alcohol (30 mL) was introduced and the mixture was continued to stir at room temperature for additional 2 h. The alcohol was then removed and the residue was dissolved in THF (5 mL) followed by addition of DMAP (0.92 g, 7.5 mmol) and Boc₂O (1.64 g, 7.5 mmol) at 0 °C. The reaction was then allowed to warm to room temperature and stir for 2 h. After concentration, the residue was purified by column chromatography on silica gel using PE/EtOAc (20:1) as the eluent to afford corresponding product **1**.

Compound **1a**



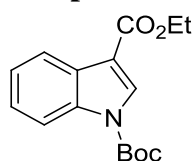
White solid, 1.32 g, 96% yield; ¹H NMR (400 MHz, CDCl₃) δ 8.26 (s, 1H), 8.19-8.15 (m, 2H), 7.39-7.32 (m, 2H), 3.94 (s, 3H), 1.69 (s, 9H); ¹³C NMR (100 MHz, CDCl₃) δ 164.6, 148.9, 135.5, 132.0, 127.4, 125.1, 123.9, 121.6, 115.1, 112.1, 85.0, 51.4, 28.0; HRMS (ESI) calcd for C₁₅H₁₈NO₄ ([M+H]⁺): 276.1236, found: 276.1232.

1. W.-J. Yoo, M. G. Capdevila, X. Du and S. Kobayashi, *Org. Lett.* 2012, **14**, 5326.

2. E. C. Linton and M. C. Kozlowski, *J. Am. Chem. Soc.* 2008, **130**, 16162.

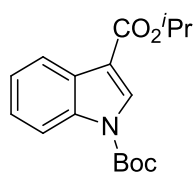
3. C. Fang, M. Li, X. Hu, W. Mo, B. Hu, N. Sun, L. Jin and Z. Shen *Adv. Syn. Catal.* 2016, **358**, 1157.

Compound 1b



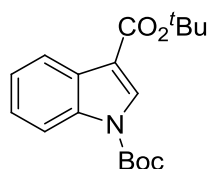
White solid, 1.39 g, 96% yield; ^1H NMR (400 MHz, CDCl_3) δ 8.27(s, 1H), 8.18-8.15 (m, 2H), 7.39-7.32(m, 2H), 4.41 (q, $J = 6.8$ Hz, 2H), 1.69 (s, 9H), 1.43 (t, $J = 7.2$ Hz, 3H); ^{13}C NMR (100 MHz, CDCl_3) δ 164.2, 149.0, 135.5, 132.0, 127.5, 125.0, 123.8, 121.7, 115.1, 112.5, 85.0, 60.3, 28.0, 14.4; HRMS (ESI) calcd for $\text{C}_{16}\text{H}_{20}\text{NO}_4$ ($[\text{M}+\text{H}]^+$): 290.1392, found: 290.1389.

Compound 1c



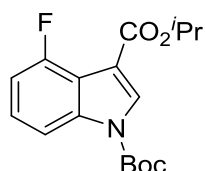
White solid, 1.48 g, 98% yield; ^1H NMR (400 MHz, CDCl_3) δ 8.26 (s, 1H), 8.18-8.16 (m, 2H), 7.38-7.32 (m, 2H), 5.33-5.27 (m, 1H), 1.69 (s, 9H), 1.41 (d, $J = 6.4$ Hz, 6H); ^{13}C NMR (100 MHz, CDCl_3) δ 163.8, 149.0, 135.5, 131.9, 127.6, 125.0, 123.8, 121.7, 115.1, 112.9, 85.0, 67.7, 28.1, 22.1; HRMS (ESI) calcd for $\text{C}_{17}\text{H}_{22}\text{NO}_4$ ($[\text{M}+\text{H}]^+$): 304.1549, found: 304.1546.

Compound 1d



White solid, 1.47 g, 93% yield; ^1H NMR (400 MHz, CDCl_3) δ 8.21 (s, 1H), 8.15-8.12 (m, 2H), 7.38-7.30 (m, 2H), 1.68 (s, 9H), 1.64 (s, 9H); ^{13}C NMR (100 MHz, CDCl_3) δ 163.7, 149.1, 135.6, 131.8, 127.6, 124.9, 123.7, 121.7, 115.1, 114.1, 84.8, 80.9, 28.4, 28.1; HRMS (ESI) calcd for $\text{C}_{18}\text{H}_{24}\text{NO}_4$ ($[\text{M}+\text{H}]^+$): 318.1705, found: 318.1701.

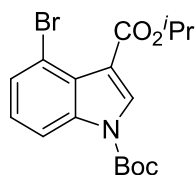
Compound 1g



White solid, 0.66 g, 41% yield; ^1H NMR (400 MHz, CDCl_3) δ 8.22 (s, 1H), 8.02 (d, $J = 8.4$ Hz, 1H), 7.33-7.29 (m, 1H), 7.05-7.00 (m, 1H), 5.32-5.23 (m, 1H), 1.69 (s, 9H), 1.39 (d, $J = 6.0$ Hz, 6H); ^{13}C NMR (100 MHz, CDCl_3) δ 162.7, 155.9 (d, $J = 252.0$ Hz), 148.7, 138.0 (d, $J = 8.8$ Hz), 132.5, 126.0 (d, $J = 7.7$ Hz), 115.5 (d, $J = 19.6$ Hz),

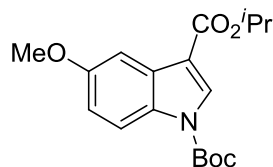
112.3 (d, $J = 3.8$ Hz), 111.2 (d, $J = 4.0$ Hz), 110.3 (d, $J = 20.8$ Hz), 85.5, 68.2, 28.0, 21.9; HRMS (ESI) calcd for $C_{17}H_{21}FNO_4$ ($[M+H]^+$): 322.1455, found: 322.1451.

Compound 1h



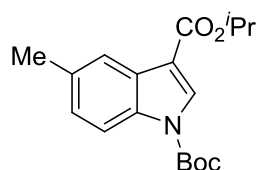
Colorless oil, 0.77 g, 40% yield; 1H NMR (400 MHz, $CDCl_3$) δ 8.22 (d, $J = 8.0$ Hz, 1H), 8.11 (s, 1H), 7.52 (d, $J = 7.6$ Hz, 1H), 7.22-7.18 (m, 1H), 5.33-5.24 (m, 1H), 1.68 (s, 9H), 1.41 (d, $J = 6.4$ Hz, 6H); ^{13}C NMR (100 MHz, $CDCl_3$) δ 163.3, 148.5, 136.7, 131.3, 128.8, 126.4, 125.9, 114.6, 114.3, 114.2, 85.4, 68.7, 28.0, 21.9; HRMS (ESI) calcd for $C_{17}H_{21}^{79}BrNO_4$ ($[M+H]^+$): 382.0654, found: 382.0650.

Compound 1i



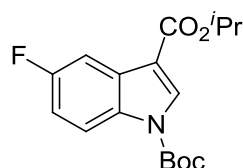
White solid, 0.69 g, 42% yield; 1H NMR (400 MHz, $CDCl_3$) δ 8.21 (s, 1H), 8.03 (d, $J = 9.2$ Hz, 1H), 7.66 (d, $J = 1.6$ Hz, 1H), 6.98-6.95 (m, 1H), 5.32-5.25 (m, 1H), 3.89 (s, 3H), 1.68 (s, 9H), 1.40 (d, $J = 6.4$ Hz, 6H); ^{13}C NMR (100 MHz, $CDCl_3$) δ 163.9, 156.7, 149.0, 132.1, 130.1, 128.7, 115.9, 114.3, 112.5, 103.7, 84.9, 67.7, 55.6, 28.1, 22.1; HRMS (ESI) calcd for $C_{18}H_{24}NO_5$ ($[M+H]^+$): 334.1654, found: 334.1650.

Compound 1j



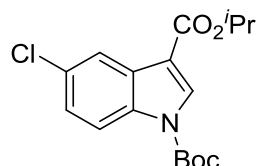
White solid, 0.68 g, 43% yield; 1H NMR (400 MHz, $CDCl_3$) δ 8.20 (s, 1H), 8.02 (d, $J = 8.4$ Hz, 1H), 7.96 (s, 1H), 7.17 (d, $J = 8.0$ Hz, 1H), 5.34-5.24 (m, 1H), 2.48 (s, 3H), 1.68 (s, 9H), 1.41 (d, $J = 6.4$ Hz, 6H); ^{13}C NMR (100 MHz, $CDCl_3$) δ 164.0, 149.1, 133.7, 133.5, 131.9, 127.8, 126.4, 121.5, 114.7, 112.5, 84.8, 67.7, 28.1, 22.1, 21.5; HRMS (ESI) calcd for $C_{18}H_{24}NO_4$ ($[M+H]^+$): 318.1705, found: 318.1703.

Compound 1k



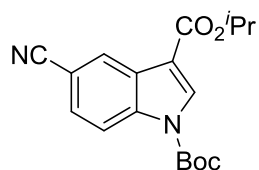
White solid, 0.69 g, 43% yield; ^1H NMR (400 MHz, CDCl_3) δ 8.27 (s, 1H), 8.13-8.10 (m, 1H), 7.82-7.79 (m, 1H), 7.11-7.06 (m, 1H), 5.33-5.26 (m, 1H), 1.69 (s, 9H), 1.41 (d, $J = 6.4$ Hz, 6H); ^{13}C NMR (100 MHz, CDCl_3) δ 162.3 (d, $J = 242.2$ Hz), 158.7, 148.8, 133.1, 131.9, 128.6, 116.2 (d, $J = 9.2$ Hz), 113.0 (d, $J = 25.2$ Hz), 112.7 (d, $J = 3.8$ Hz), 107.5 (d, $J = 25.1$ Hz), 85.3, 68.0, 28.1, 22.0; HRMS (ESI) calcd for $\text{C}_{17}\text{H}_{21}\text{FNO}_4$ ($[\text{M}+\text{H}]^+$): 322.1455, found: 322.1451.

Compound 1l



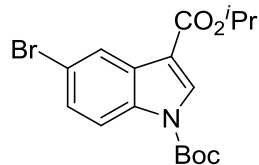
White solid, 1.23 g, 73% yield; ^1H NMR (400 MHz, CDCl_3) δ 8.24 (s, 1H), 8.13 (d, $J = 2.0$ Hz, 1H), 8.09 (d, $J = 8.8$ Hz, 1H), 7.33-7.30 (m, 1H), 5.34-5.25 (m, 1H), 1.69 (s, 9H), 1.41 (d, $J = 6.4$ Hz, 6H); ^{13}C NMR (100 MHz, CDCl_3) δ 163.4, 148.7, 133.9, 132.8, 129.7, 128.7, 125.3, 121.4, 116.2, 112.3, 85.4, 68.1, 28.0, 22.0; HRMS (ESI) calcd for $\text{C}_{17}\text{H}_{21}^{35}\text{ClNO}_4$ ($[\text{M}+\text{H}]^+$): 338.1159, found: 338.1154.

Compound 1m



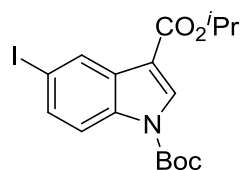
White solid, 0.66 g, 40% yield; ^1H NMR (400 MHz, CDCl_3) δ 8.50 (s, 1H), 8.33 (s, 1H), 8.29 (d, $J = 8.8$ Hz, 1H), 7.62 (d, $J = 8.4$ Hz, 1H), 5.37-5.27 (m, 1H), 1.71 (s, 9H), 1.43 (d, $J = 6.0$ Hz, 6H); ^{13}C NMR (100 MHz, CDCl_3) δ 162.9, 148.3, 137.3, 133.6, 128.1, 127.6, 126.8, 119.5, 116.1, 112.8, 107.5, 86.3, 68.5, 28.0, 22.0; HRMS (ESI) calcd for $\text{C}_{18}\text{H}_{21}\text{N}_2\text{O}_4$ ($[\text{M}+\text{H}]^+$): 329.1501, found: 329.1495.

Compound 1n



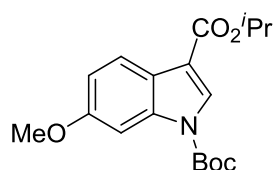
White solid, 0.92 g, 48% yield; ^1H NMR (400 MHz, CDCl_3) δ 8.30 (d, $J = 1.6$ Hz, 1H), 8.22 (s, 1H), 8.04 (d, $J = 8.8$ Hz, 1H), 7.47-7.44 (m, 1H), 5.34-5.24 (m, 1H), 1.68 (s, 9H), 1.41 (d, $J = 6.4$ Hz, 6H); ^{13}C NMR (100 MHz, CDCl_3) δ 163.3, 148.6, 134.2, 132.6, 129.2, 127.9, 124.4, 117.5, 116.5, 112.2, 85.5, 68.0, 28.0, 22.0; HRMS (ESI) calcd for $\text{C}_{17}\text{H}_{21}^{81}\text{BrNO}_4$ ($[\text{M}+\text{H}]^+$): 384.0633, found: 384.0640.

Compound 1o



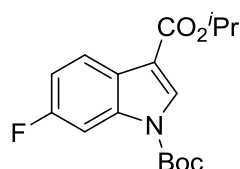
White solid, 0.90 g, 42% yield; ^1H NMR (400 MHz, CDCl_3) δ 8.52 (d, $J = 1.6$ Hz, 1H), 8.18 (s, 1H), 7.93 (d, $J = 8.8$ Hz, 1H), 7.65-7.63 (m, 1H), 5.33-5.24 (m, 1H), 1.68 (s, 9H), 1.41 (d, $J = 6.4$ Hz, 6H); ^{13}C NMR (100 MHz, CDCl_3) δ 163.3, 148.6, 134.8, 133.6, 132.2, 130.6, 129.7, 116.9, 112.0, 88.4, 85.5, 68.1, 28.0, 22.0; HRMS (ESI) calcd for $\text{C}_{17}\text{H}_{20}\text{INNaO}_4$ ($[\text{M}+\text{Na}]^+$): 452.0335, found: 452.0330.

Compound 1p



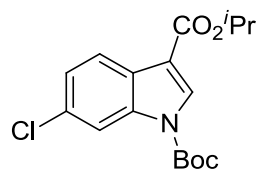
White solid, 0.55 g, 33% yield; ^1H NMR (400 MHz, CDCl_3) δ 8.13 (s, 1H), 8.00 (d, $J = 8.8$ Hz, 1H), 7.75 (s, 1H), 6.98-6.96 (m, 1H), 5.33-5.24 (m, 1H), 3.88 (s, 3H), 1.69 (s, 9H), 1.40 (d, $J = 6.4$ Hz, 6H); ^{13}C NMR (100 MHz, CDCl_3) δ 163.9, 158.1, 149.1, 136.6, 130.6, 122.2, 121.3, 113.3, 113.0, 99.0, 84.8, 67.7, 55.6, 28.1, 22.1; HRMS (ESI) calcd for $\text{C}_{18}\text{H}_{23}\text{NNaO}_5$ ($[\text{M}+\text{Na}]^+$): 356.1474, found: 356.1472.

Compound 1q



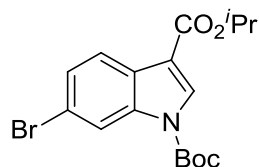
White solid, 0.85 g, 53% yield; ^1H NMR (400 MHz, CDCl_3) δ 8.21 (s, 1H), 8.10-8.07 (m, 1H), 7.90-7.88 (m, 1H), 7.11-7.06 (m, 1H), 5.34-5.24 (m, 1H), 1.69 (s, 9H), 1.40 (d, $J = 6.4$ Hz, 6H); ^{13}C NMR (100 MHz, CDCl_3) δ 163.6, 161.1 (d, $J = 240.0$ Hz), 148.7, 135.7 (d, $J = 12.9$ Hz), 131.9 (d, $J = 3.2$ Hz), 123.9, 122.6 (d, $J = 9.7$ Hz), 112.8, 112.2 (d, $J = 23.8$ Hz), 102.5 (d, $J = 28.5$ Hz), 85.4, 67.9, 28.0, 22.0; HRMS (ESI) calcd for $\text{C}_{17}\text{H}_{21}\text{FNO}_4$ ($[\text{M}+\text{H}]^+$): 322.1455, found: 322.1450.

Compound 1r



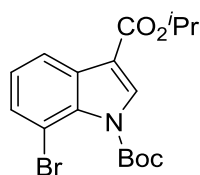
White solid, 0.62 g, 37% yield; ^1H NMR (400 MHz, CDCl_3) δ 8.21 (s, 2H), 8.06 (d, $J = 8.4$ Hz, 1H), 7.32-7.29 (m, 1H), 5.33-5.24 (m, 1H), 1.69 (s, 9H), 1.40 (d, $J = 6.4$ Hz, 6H); ^{13}C NMR (100 MHz, CDCl_3) δ 163.4, 148.6, 135.8, 132.1, 131.0, 126.1, 124.4, 122.5, 115.4, 112.8, 85.5, 67.9, 28.0, 22.0; HRMS (ESI) calcd for $\text{C}_{17}\text{H}_{21}^{35}\text{ClNO}_4$ ($[\text{M}+\text{H}]^+$): 338.1159, found: 338.1153.

Compound 1s



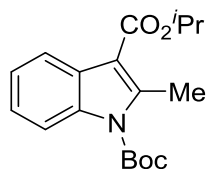
White solid, 1.24 g, 65% yield; ^1H NMR (400 MHz, CDCl_3) δ 8.38 (s, 1H), 8.19 (s, 1H), 8.01 (d, $J = 8.4$ Hz, 1H), 7.45 (d, $J = 8.4$ Hz, 1H), 5.32-5.26 (m, 1H), 1.69 (s, 9H), 1.41 (d, $J = 6.0$ Hz, 6H); ^{13}C NMR (100 MHz, CDCl_3) δ 163.4, 148.6, 136.1, 132.0, 127.1, 126.4, 122.8, 118.8, 118.3, 112.8, 85.6, 68.0, 28.0, 22.0; HRMS (ESI) calcd for $\text{C}_{17}\text{H}_{20}^{81}\text{BrNNaO}_4$ ($[\text{M}+\text{Na}]^+$): 406.0453, found: 406.0449.

Compound 1t



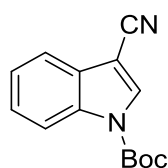
White solid, 0.99 g, 52% yield; ^1H NMR (400 MHz, CDCl_3) δ 8.18-8.16(m, 2H), 7.57 (d, $J = 7.6$ Hz, 1H), 7.22-7.19 (m, 1H), 5.32-5.26 (m, 1H), 1.68 (s, 9H), 1.41 (d, $J = 6.0$ Hz, 6H); ^{13}C NMR (100 MHz, CDCl_3) δ 163.3, 147.8, 135.0, 133.8, 130.9, 130.3, 125.0, 121.0, 112.3, 107.4, 85.7, 67.9, 27.8, 22.0; HRMS (ESI) calcd for $\text{C}_{17}\text{H}_{21}^{79}\text{BrNO}_4$ ($[\text{M}+\text{H}]^+$): 382.0654, found: 382.0651.

Compound 1u



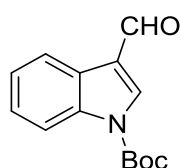
White solid, 0.16 g, 10% yield; ^1H NMR (400 MHz, CDCl_3) δ 8.11-8.06 (m, 2H), 7.29-7.27 (m, 2H), 5.35-5.29 (m, 1H), 2.98 (s, 3H), 1.70 (s, 9H), 1.43 (d, $J = 6.0$ Hz, 6H); ^{13}C NMR (100 MHz, CDCl_3) δ 165.0, 150.0, 145.6, 135.4, 127.2, 124.0, 123.5, 121.3, 114.8, 110.6, 84.9, 67.5, 28.1, 22.2, 15.0; HRMS (ESI) calcd for $\text{C}_{18}\text{H}_{24}\text{NO}_4$ ($[\text{M}+\text{H}]^+$): 318.1705, found: 318.1704.

Compound 1v



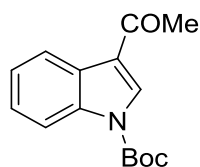
White solid, 1.20 g, 99% yield; ^1H NMR (400 MHz, CDCl_3) δ 8.17 (d, $J = 8.4$ Hz, 1H), 8.10 (s, 1H), 7.70 (d, $J = 7.6$ Hz, 1H), 7.46-7.35 (m, 2H), 1.70 (s, 9H); ^{13}C NMR (100 MHz, CDCl_3) δ 148.1, 134.2, 133.2, 128.0, 126.2, 124.2, 119.7, 115.6, 114.1, 92.2, 85.9, 27.9; HRMS (ESI) calcd for $\text{C}_{14}\text{H}_{14}\text{N}_2\text{NaO}_2$ ($[\text{M}+\text{Na}]^+$): 265.0947, found: 265.0949.

Compound 1w



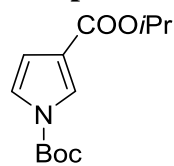
White solid, 1.04 g, 85% yield; ^1H NMR (400 MHz, CDCl_3) δ 10.10 (s, 1H), 8.29 (d, $J = 7.6$ Hz, 1H), 8.24 (s, 1H), 8.15 (d, $J = 8.0$ Hz, 1H), 7.44-7.36 (m, 2H), 1.71 (s, 9H); ^{13}C NMR (100 MHz, CDCl_3) δ 185.8, 156.6, 148.8, 136.5, 135.9, 126.1, 124.6, 122.1, 121.5, 115.1, 85.6, 28.1; HRMS (ESI) calcd for $\text{C}_{14}\text{H}_{16}\text{NO}_3$ ($[\text{M}+\text{H}]^+$): 246.1125, found: 246.1127.

Compound 1x



White solid, 1.17 g, 90% yield; ^1H NMR (400 MHz, CDCl_3) δ 8.38-8.36 (m, 1H), 8.23 (s, 1H), 8.11 (d, $J = 7.2$ Hz, 1H), 7.40-7.33 (m, 2H), 2.57 (s, 3H), 1.72 (s, 9H); ^{13}C NMR (100 MHz, CDCl_3) δ 193.9, 149.1, 135.5, 132.4, 127.3, 125.4, 124.3, 122.7, 120.6, 114.9, 85.4, 28.1, 27.7; HRMS (ESI) calcd for $\text{C}_{15}\text{H}_{17}\text{NNaO}_3$ ($[\text{M}+\text{Na}]^+$): 282.1101, found: 282.1105.

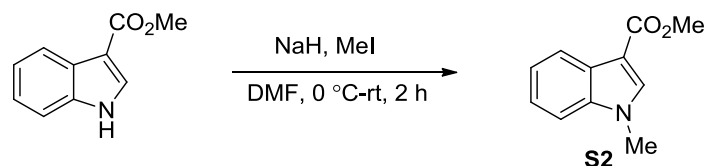
Compound 1y



Colorless oil, 0.77 g, 60% yield; ^1H NMR (400 MHz, CDCl_3) δ 7.80 (s, 1H), 7.20-7.19 (m, 1H), 6.60-6.59 (m, 1H), 5.23-5.13 (m, 1H), 1.61 (s, 9H), 1.32 (d, $J = 6.0$ Hz, 6H); ^{13}C NMR (100 MHz, CDCl_3) δ 163.6, 148.1, 124.5, 120.5, 111.9, 84.8, 81.0,

67.4, 27.8, 21.9; HRMS (ESI) calcd for C₁₃H₂₀NO₄ ([M+H]⁺): 254.1387, found: 254.1390.

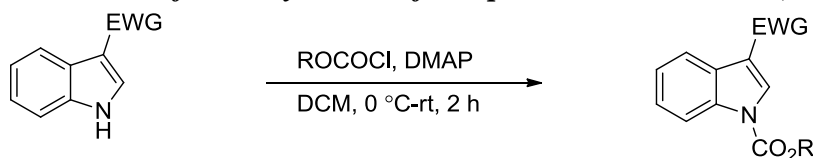
3. Synthesis of Compound S2



The preparation of compound **S2** was adapted from the literature procedures.⁴ To a 50-mL flask charged with methyl indole-3-carboxylate (1.75 g, 10 mmol) and DMF (10 mL) was added NaH (0.80 g, 60% in mineral oil, 20 mmol) in 5 portions at 0 °C. The resulting mixture was allowed to stir at same temperature for 0.5 h. Methyl iodide (1.24 mL, 20 mmol) was then introduced slowly at 0 °C. The reaction was then warmed to room temperature and continued to stir for 2 h. Water (10 mL) was then added slowly at 0 °C to quench the reaction. The mixture was then extracted with EtOAc 3 times (3 X 30 mL). The combined organic phase was then washed with water 3 times (3 X 20 mL). After removal of the solvent, the residue was purified by column chromatography on silica gel using PE/EtOAc (20:1) as the eluent to afford *N*-Methyl indole **S2** as white solid (1.88 g, 99% yield).

¹H NMR (400 MHz, CDCl₃) δ 8.18-8.16 (m, 1H), 7.75 (s, 1H), 7.33-7.27 (m, 3H), 3.90 (s, 3H), 3.79 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 165.4, 137.1, 135.1, 126.5, 122.7, 121.8, 121.5, 109.7, 106.7, 50.9, 33.4; HRMS (ESI) calcd for C₁₁H₁₂NO₂ ([M+H]⁺): 190.0868, found: 190.0861.

4. General Procedures for the synthesis of compounds 1e and S3-S6 (GP2)

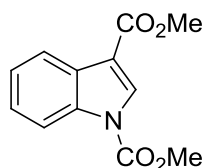


The synthesis of compounds **1e** and **S3-S6** was adapted from literature procedures.⁵ To a 100-mL flask charged with 3-substituted indole (10 mmol, carboxylic esters were prepared according to the GP1) and DCM (20 mL) was introduced DMAP (1.8 g, 15 mmol, 1.5 equiv) followed by slow addition of chloro methylformate or CbzCl (15 mmol, 1.5 equiv) at 0 °C. The resulting mixture was allowed to warm to room temperature and stir for 2 h. The reaction was then diluted by 1 M HCl (20 mL) and extracted with DCM 3 times (3 X 20 mL). The combined organic phase was dried over Na₂SO₄. After removal of the solvent, the residue was purified by column chromatography on silica gel using PE/EtOAc (10:1) as the eluent to afford corresponding 1,3-disubstituted indole.

4. M. Kitano, A. Kojima, K. Nakano, A. Miyagishi, T. Noguchi and N. Ohashi, *Chem. Pharm. Bull.* 1999, **47**, 1538.

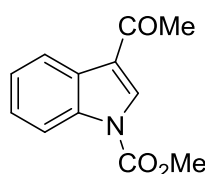
5. C. N. Rao, D. Lentz and H.-U. Reissig, *Angew. Chem., Int. Ed.* 2015, **54**, 2750.

Compound 1e



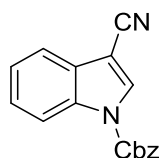
White solid, 2.05 g, 88% yield; ^1H NMR (400 MHz, CDCl_3) δ 8.27 (s, 1H), 8.18-8.15 (m, 2H), 7.41-7.34 (m, 2H), 4.07 (s, 3H), 3.93 (s, 3H); ^{13}C NMR (100 MHz, CDCl_3) δ 164.4, 150.8, 135.4, 131.5, 127.3, 125.3, 124.2, 121.6, 115.0, 112.9, 54.3, 51.5; HRMS (ESI) calcd for $\text{C}_{12}\text{H}_{12}\text{NO}_4$ ($[\text{M}+\text{H}]^+$): 234.0766, found: 234.0763.

Compound S3



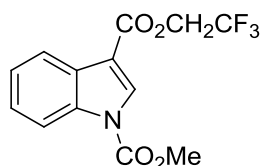
White solid, 1.98 g, 91% yield; ^1H NMR (400 MHz, CDCl_3) δ 8.38 (d, $J = 6.8$ Hz, 1H), 8.24 (s, 1H), 8.16 (d, $J = 7.6$ Hz, 1H), 7.43-7.36 (m, 2H), 4.12 (s, 3H), 2.57 (s, 3H); ^{13}C NMR (100 MHz, CDCl_3) δ 193.8, 151.0, 135.5, 131.9, 127.2, 125.8, 124.7, 122.7, 121.3, 114.8, 54.5, 27.7; HRMS (ESI) calcd for $\text{C}_{12}\text{H}_{12}\text{NO}_3$ ($[\text{M}+\text{H}]^+$): 218.0817, found: 218.0811.

Compound S4



White solid, 2.23 g, 81% yield; ^1H NMR (400 MHz, CDCl_3) δ 8.21 (d, $J = 8.0$ Hz, 1H), 8.14 (s, 1H), 7.71 (d, $J = 8.0$ Hz, 1H), 7.50-7.37 (m, 7H), 5.49 (s, 2H); ^{13}C NMR (100 MHz, CDCl_3) δ 149.5, 134.3, 134.0, 132.8, 129.2, 128.9, 128.8, 127.9, 126.6, 124.6, 119.9, 115.6, 113.8, 93.3, 69.9; HRMS (ESI) calcd for $\text{C}_{17}\text{H}_{13}\text{N}_2\text{O}_2$ ($[\text{M}+\text{H}]^+$): 277.0977, found: 277.0974.

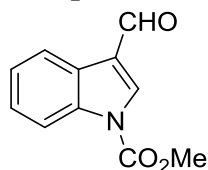
Compound S5



White solid, 2.80 g, 93% yield; ^1H NMR (400 MHz, CDCl_3) δ 8.37 (s, 1H), 8.21 (d, $J = 7.6$ Hz, 1H), 8.14 (d, $J = 7.6$ Hz, 1H), 7.45-7.38 (m, 2H), 4.76-4.70 (m, 2H), 4.11 (s, 3H); ^{13}C NMR (100 MHz, CDCl_3) δ 162.0, 150.7, 135.5, 132.6, 127.0, 125.8, 124.6,

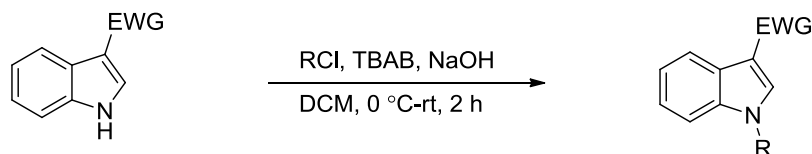
123.1 (q, $J = 275.7$ Hz), 121.5, 115.2, 111.3, 60.1 (q, $J = 36.4$ Hz), 54.6; HRMS (ESI) calcd for $C_{13}H_{11}F_3NO_4$ ($[M+H]^+$): 302.0640, found: 302.0636.

Compound S6



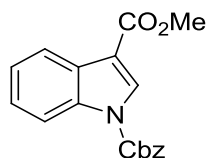
White solid, 1.87 g, 92% yield; 1H NMR (400 MHz, $CDCl_3$) δ 10.08 (s, 1H), 8.28 (d, $J = 7.6$ Hz, 1H), 8.22 (s, 1H), 8.16 (d, $J = 8.0$ Hz, 1H), 7.45-7.36 (m, 2H), 4.11 (s, 3H); ^{13}C NMR (100 MHz, $CDCl_3$) δ 185.7, 150.6, 136.0, 135.8, 126.3, 125.9, 124.8, 122.1, 115.0, 54.6 (two aromatic carbon signals overlap); HRMS (ESI) calcd for $C_{11}H_9NNaO_3$ ($[M+Na]^+$): 226.0480, found: 226.0479.

5. General Procedures for the synthesis of compounds 1f and S7-S10 (GP3)



The synthesis of compounds **1f** and **S7-S10** was adapted from the literature procedures.⁶ To a 100-mL flask charged with 3-substituted indole (10 mmol, carboxylic esters were prepared according to the GP1), DCM (10 mL), TBAB (0.32 g, 1.0 mmol), and NaOH (0.80 g, 20 mmol) was added corresponding CbzCl (15 mmol, 1.5 equiv), acyl chloride (15 mmol, 1.5 equiv) or TsCl (15 mmol, 1.5 equiv) slowly at 0 °C. The resulting mixture was allowed to warm to room temperature and stir for 3 h. The reaction was then diluted by 1 M HCl (20 mL) and extracted with DCM 3 times (3 X 10 mL). The combined organic phase was dried over Na_2SO_4 . After removal of the solvent, the residue was purified by column chromatography on silica gel using PE/EtOAc (10:1) as the eluent to afford corresponding 1,3-disubstituted indole.

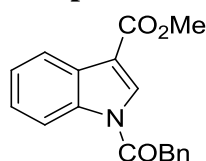
Compound 1f



White solid, 2.93 g, 95% yield; 1H NMR (400 MHz, $CDCl_3$) δ 8.28 (s, 1H), 8.20-8.14 (m, 2H), 7.49-7.32 (m, 7H), 5.46 (s, 2H), 3.91 (s, 3H); ^{13}C NMR (100 MHz, $CDCl_3$) δ 164.4, 150.2, 135.5, 134.4, 131.5, 129.0, 128.8, 128.7, 127.4, 125.4, 124.2, 121.7, 115.1, 113.0, 69.4, 51.5; HRMS (ESI) calcd for $C_{18}H_{16}NO_4$ ($[M+H]^+$): 310.1079, found: 310.1078.

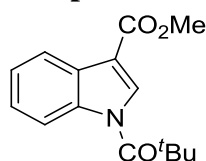
6. E. Reimann *Pharmazie* 2000, **55**, 907-912.

Compound S7



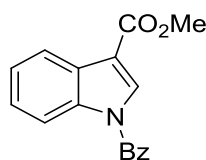
White solid, 2.46 g, 84% yield; $^1\text{H NMR}$ (400 MHz, CDCl_3) δ 8.47 (d, $J = 8.4$ Hz, 1H), 8.21 (s, 1H), 8.13 (d, $J = 7.2$ Hz, 1H), 7.42 - 7.32 (m, 7H), 4.28 (s, 2H), 3.94 (s, 3H); $^{13}\text{C NMR}$ (100 MHz, CDCl_3) δ 169.5, 164.3, 136.1, 132.5, 130.7, 129.1, 129.0, 127.7, 127.2, 126.1, 124.9, 121.5, 116.6, 114.0, 51.6, 42.6; HRMS (ESI) calcd for $\text{C}_{18}\text{H}_{16}\text{NO}_3$ ($[\text{M}+\text{H}]^+$): 294.1130, found: 294.1126.

Compound S8



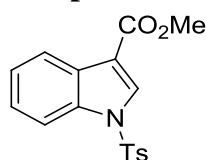
White solid, 1.86 g, 72% yield; $^1\text{H NMR}$ (400 MHz, CDCl_3) δ 8.48 (d, $J = 8.0$ Hz, 1H), 8.42 (s, 1H), 8.16 (d, $J = 7.6$ Hz, 1H), 7.43-7.36 (m, 2H), 3.96 (s, 3H), 1.55 (s, 9H); $^{13}\text{C NMR}$ (100 MHz, CDCl_3) δ 177.2, 164.6, 137.2, 131.5, 126.4, 125.9, 124.6, 121.2, 117.1, 112.9, 51.6, 41.6, 28.7; HRMS (ESI) calcd for $\text{C}_{15}\text{H}_{18}\text{NO}_3$ ($[\text{M}+\text{H}]^+$): 260.1287, found: 260.1281.

Compound S9



White solid, 2.18 g, 78% yield; $^1\text{H NMR}$ (400 MHz, CDCl_3) δ 8.39 (d, $J = 8.0$ Hz, 1H), 8.20 (d, $J = 7.2$ Hz, 1H), 7.99 (s, 1H), 7.76 (d, $J = 7.6$ Hz, 2H), 7.68-7.65 (m, 1H), 7.59-7.55 (m, 2H), 7.46-7.41 (m, 2H), 3.91 (s, 3H); $^{13}\text{C NMR}$ (100 MHz, CDCl_3) δ 168.6, 164.4, 136.3, 133.4, 133.3, 132.7, 129.4, 128.9, 127.6, 125.7, 125.0, 121.6, 116.2, 113.2, 51.6; HRMS (ESI) calcd for $\text{C}_{17}\text{H}_{13}\text{NNaO}_3$ ($[\text{M}+\text{Na}]^+$): 302.0793, found: 302.0787.

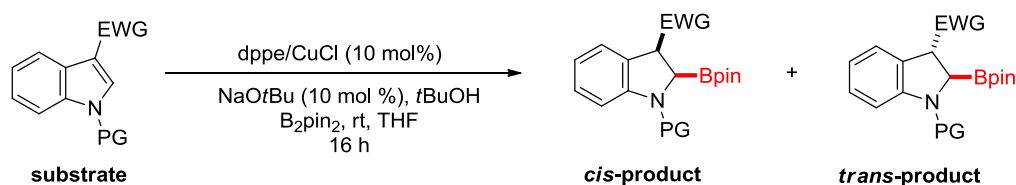
Compound S10



White solid, 2.30 g, 70% yield; $^1\text{H NMR}$ (400 MHz, CDCl_3) δ 8.28 (s, 1H), 8.13 (d, $J = 7.6$ Hz, 1H), 7.97 (d, $J = 7.6$ Hz, 1H), 7.82 (d, $J = 7.2$ Hz, 2H), 7.38-7.31 (m, 2H), 7.24 (d, $J = 7.6$ Hz, 2H), 3.92 (s, 3H), 2.33 (s, 3H); $^{13}\text{C NMR}$ (100 MHz, CDCl_3) δ 164.0, 145.7, 134.7, 134.5, 132.0, 130.1, 127.7, 127.0, 125.3, 124.3, 122.0, 113.4,

113.2, 51.5, 21.5; HRMS (ESI) calcd for C₁₇H₁₆NO₄S ([M+H]⁺): 330.0800, found: 330.0797.

6. Table S1. Initial Survey of Copper-catalyzed Dearomative Borylation Using Achiral ligand (dppe).

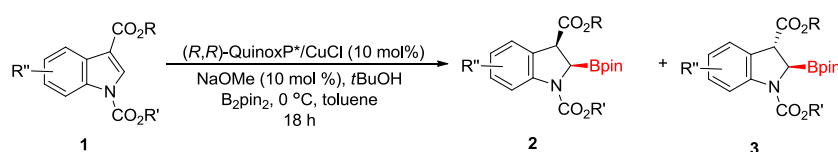


To entry	substrate	PG, EWG	results
1	S2	Me, CO ₂ Me	no reaction
2	S3	CO ₂ Me, COMe	yes, major <i>trans</i> -product.
3	S4	Cbz, CN	no reaction
4	S5	CO ₂ Me, CO ₂ CH ₂ CF ₃	yes, major <i>trans</i> -product
5	S6	CO ₂ Me, CHO	messy
6	S7	COBn, CO ₂ Me	trace
7	S8	COtBu, CO ₂ Me	trace
8	S9	Bz, CO ₂ Me	trace
9	S10	Ts, CO ₂ Me	no reaction
10	1a	<i>Boc</i> , CO ₂ Me	<i>yes, major cis</i> -product
11	1e	CO ₂ Me, CO ₂ Me	yes, major <i>trans</i> -product
12	1f	Cbz, CO ₂ Me	yes, major <i>trans</i> -product

Conclusions: 1) When indole substrate bearing electron-withdrawing group such as acyl, formyl, or alkoxy carbonyl in its 3-position was used, alkoxy carbonyl substitution in nitrogen was responsible for the reaction to take place; 2) In the presence of achiral ligand dppe, only substrate **1a** could give *cis*-product preferentially; 3) *Trans*-product was not stable towards isolation by chromatography on either silica gel or alumina whereas *cis*-one was isolable.

Therefore, we chose indole **1a** as model substrate to further evaluate reactivity and stereoselectivity in the presence of chiral catalyst.

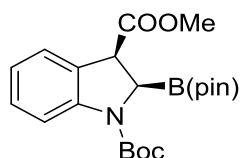
7. General procedures for the catalytic asymmetric dearomative borylation of indole 1



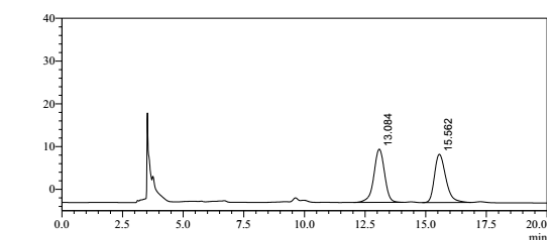
In a nitrogen-filled glovebox, to a 25-mL flame-dried Schlenk tube charged with CuCl (2.0 mg, 0.02 mmol), NaOMe (1.1 mg, 0.02 mmol) and (*R,R*)-QuinoxP* (6.7 mg, 0.02 mmol) was added toluene (0.5 mL). The resulting mixture was allowed to

stir at room temperature for 0.5 h. B₂pin₂ (0.30 mmol in 0.5 mL) was then introduced and the reaction was allowed to stir at room temperature for 10 min followed by addition of indole **1** (0.20 mmol in 1 mL toluene) and *t*BuOH (37.5 μL, 0.4 mmol) at 0 °C. The resulting mixture was continued to stir at 0 °C for 18-48 hours. After removal of the solvent, the ¹H NMR of crude mixture was taken to determine the d.r. value. The residue was purified by column chromatography on silica gel using PE/EtOAc (12:1) as the eluent to affording corresponding 2-boryl indoline **2** or **3**.

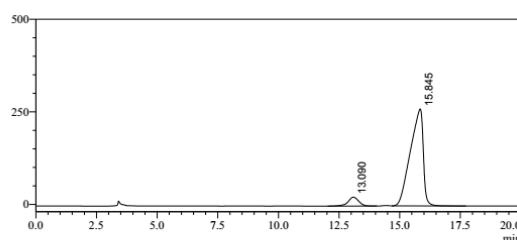
Compound 2a



Colorless oil, 70.9 mg, 88% yield, 95:5 *d.r.*, 86% *ee*, $[\alpha]_D^{25} = -57.7$ (*c* 0.40, CHCl₃). ¹H NMR (400 MHz, CDCl₃) δ 7.49 (brs, 1H), 7.27-7.17 (m, 2H), 6.93-6.90 (m, 1H), 4.44 (brs, 1H), 4.11 (d, *J* = 11.6 Hz, 1H), 3.74 (s, 3H), 1.58 (s, 9H), 1.31 (s, 6H), 1.30 (s, 6H); ¹³C NMR (100 MHz, CDCl₃) δ 172.4, 152.4, 142.1, 128.6, 128.3, 125.1, 121.8, 114.8, 83.8, 81.4, 52.4, 48.8, 47.2, 28.3, 25.3, 24.7; ¹¹B NMR (128 MHz, CDCl₃) δ 31.6; HRMS (ESI) calcd for C₂₁H₃₁BNO₆ ([M+H]⁺): 404.2244, found: 404.2245. The enantiopurity was determined by HPLC analysis (Daicel Chiralpak IE column, Hexane/*i*PrOH = 98: 2, flow rate = 1.0 mL/min., wavelength = 230 nm, *t*_R = 13.09 (minor), 15.85 (major)).

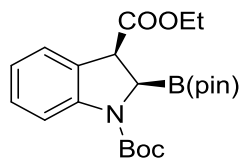


Peak ID	Ret. Time	Height	Area	Area%
1	13.084	12441	382199	51.065
2	15.562	11280	366262	48.935
Total				100.000



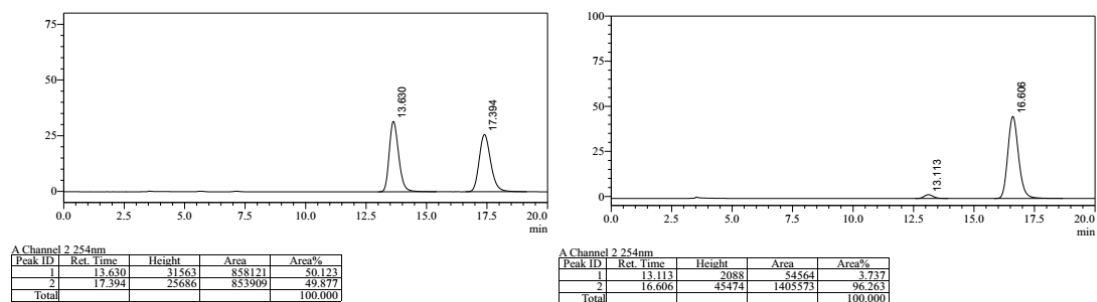
Peak ID	Ret. Time	Height	Area	Area%
1	13.090	23995	735536	7.036
2	15.845	261743	9718475	92.964
Total				100.000

Compound 2b

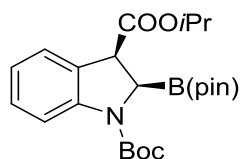


Colorless oil, 72.6 mg, 87% yield, 95:5 *d.r.*, 93% *ee* $[\alpha]_D^{25} = -66.5$ (*c* 0.2, CHCl₃). ¹H NMR (400 MHz, CDCl₃) δ 7.50 (brs, 1H), 7.28 (d, *J* = 7.2 Hz, 1H), 7.20-7.17 (m, 1H), 6.92-6.89 (m, 1H), 4.47 (d, *J* = 11.2 Hz, 1H), 4.21 (q, *J* = 7.2 Hz, *J* = 14.0 Hz, 2H), 4.11 (d, *J* = 12.0 Hz, 1H), 1.58 (s, 9H), 1.32-1.26 (m, 15H); ¹³C NMR (100 MHz, CDCl₃) δ 172.0, 152.4, 142.3, 128.5, 128.3, 124.8, 121.8, 115.1, 83.8, 81.2, 61.4, 48.6, 47.5, 28.3, 25.3, 24.8, 14.1; ¹¹B NMR (128 MHz, CDCl₃) δ 31.6; HRMS (ESI) calcd for C₂₂H₃₂BNNaO₆ ([M+Na]⁺): 440.2220, found: 440.2216. The enantiopurity was

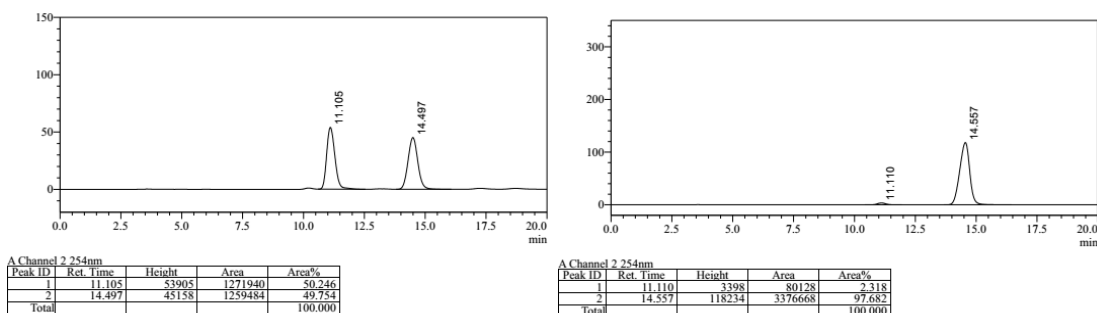
determined by HPLC analysis (Daicel Chiralpak IE column, Hexane/*i*PrOH = 98: 2, flow rate = 1.0 mL/min., wavelength = 254 nm, t_R = 13.11 (minor), 16.61 (major).



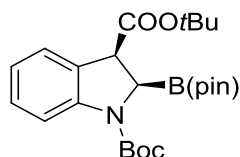
Compound 2c



Colorless oil, 75.0 mg, 87% yield, 95:5 *d.r.*, 95% *ee*, $[\alpha]_D^{25} = -66.3$ (*c* 0.15, CHCl₃). ¹H NMR (400 MHz, CDCl₃) δ 7.51 (brs, 1H), 7.29 (d, *J* = 7.6 Hz, 1H), 7.20-7.16 (m, 1H), 6.91-6.88 (m, 1H), 5.07-5.01 (m, 1H), 4.44 (d, *J* = 12.0 Hz, 1H), 4.10 (d, *J* = 12.0 Hz, 1H), 1.58 (s, 9H), 1.32-1.25 (m, 18H); ¹³C NMR (100 MHz, CDCl₃) δ 171.6, 152.4, 142.3, 128.5, 128.3, 124.7, 121.7, 115.0, 83.8, 81.2, 69.2, 48.7, 47.8, 28.4, 25.3, 24.9, 21.8, 21.7; ¹¹B NMR (128 MHz, CDCl₃) δ 31.8; HRMS (ESI) calcd for C₂₃H₃₄BNNaO₆ ([M+Na]⁺): 454.2377, found: 454.2376. The enantiopurity was determined by HPLC analysis (Daicel Chiralpak IE column, Hexane/*i*PrOH = 98:2, flow rate = 1.0 mL/min., wavelength = 254 nm, t_R = 11.11 (minor), 14.56 (major).

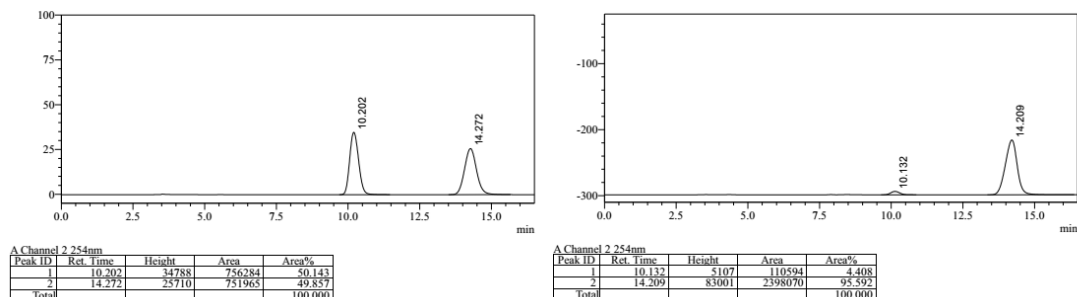


Compound 2d

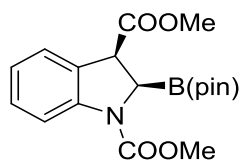


Colorless oil, 80.1 mg, 90% yield, 96:4 *d.r.*, 91% *ee*, $[\alpha]_D^{25} = -30.9$ (*c* 0.19, CHCl₃). ¹H NMR (400 MHz, CDCl₃) δ 7.51 (s, 1H), 7.30 (d, *J* = 7.2 Hz, 1H), 7.19-7.15 (m, 1H), 6.91-6.87 (m, 1H), 4.41 (d, *J* = 12.0 Hz, 1H), 4.06 (d, *J* = 12.0 Hz, 1H), 1.57 (s, 9H), 1.47 (s, 9H), 1.33 (s, 6H), 1.31 (s, 6H); ¹³C NMR (100 MHz, CDCl₃) δ 171.2, 152.2, 141.9, 128.3, 124.5, 121.5, 114.9, 83.5, 81.8, 80.8, 48.4, 28.2, 27.8, 25.2, 24.7

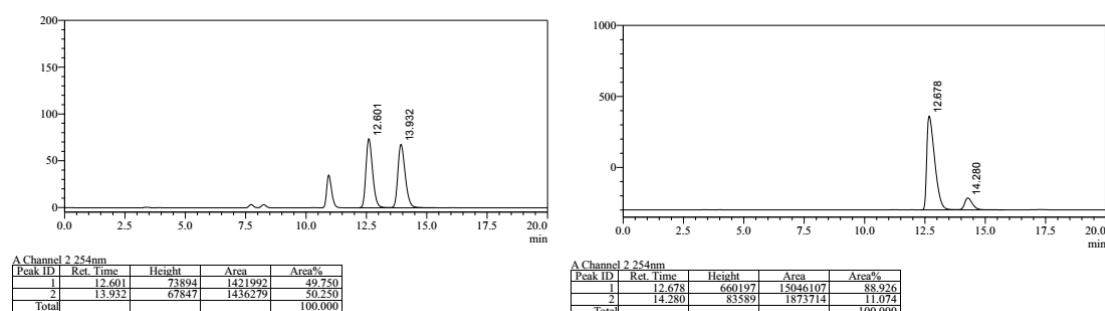
(two aromatic carbons overlap; *B*-adjacent carbon overlaps with C3-carbon); ^{11}B NMR (128 MHz, CDCl_3) δ 31.5; HRMS (ESI) calcd for $\text{C}_{24}\text{H}_{36}\text{BNNaO}_6$ ($[\text{M}+\text{Na}]^+$): 468.2535, found: 468.2531. The enantiopurity was determined by HPLC analysis (Daicel Chiralpak IE column, Hexane/*i*PrOH = 98:2, flow rate = 1.0 mL/min., wavelength = 254 nm, t_{R} = 10.13 (minor), 14.21 (major).



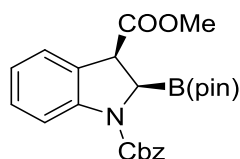
Compound 2e



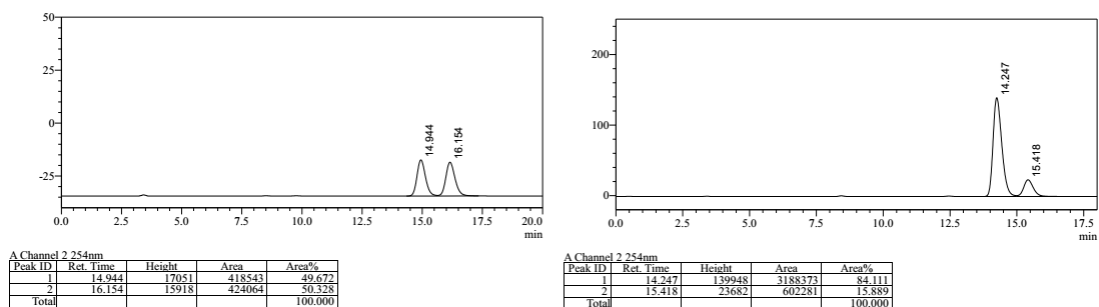
Colorless oil, 54.1 mg, 75% yield, 90:10 *d.r.*, 78% *ee*, $[\alpha]_D^{25} = -9.2$ (*c* 0.2, CHCl_3). ^1H NMR (400 MHz, CDCl_3) δ 7.93 (brs, 1H), 7.29-7.21 (m, 2H), 6.97 - 6.94 (m, 1H), 4.50 (s, 1H), 4.12 (d, $J = 11.6$ Hz, 1H), 3.81 (s, 3H), 3.75 (s, 3H), 1.31 (s, 6H), 1.30 (s, 6H). ^{13}C NMR (100 MHz, CDCl_3) δ 172.4, 153.3, 142.8, 128.8, 127.5, 124.6, 122.4, 115.1, 84.0, 60.3, 52.6, 48.4, 25.2, 24.8 (*B*-adjacent carbon overlaps with C3-carbon); ^{11}B NMR (128 MHz, CDCl_3) δ 31.9; HRMS (ESI) calcd for $\text{C}_{18}\text{H}_{24}\text{BNNaO}_6$ ($[\text{M}+\text{Na}]^+$): 384.1594, found: 384.1592. The enantiopurity was determined by HPLC analysis (Daicel Chiralpak IC column, Hexane/*i*PrOH = 90:10, flow rate = 1.0 mL/min., wavelength = 254 nm, t_{R} = 12.68 (major), 14.28 (minor).



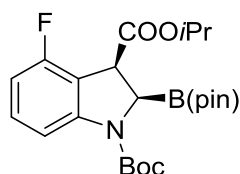
Compound 2f



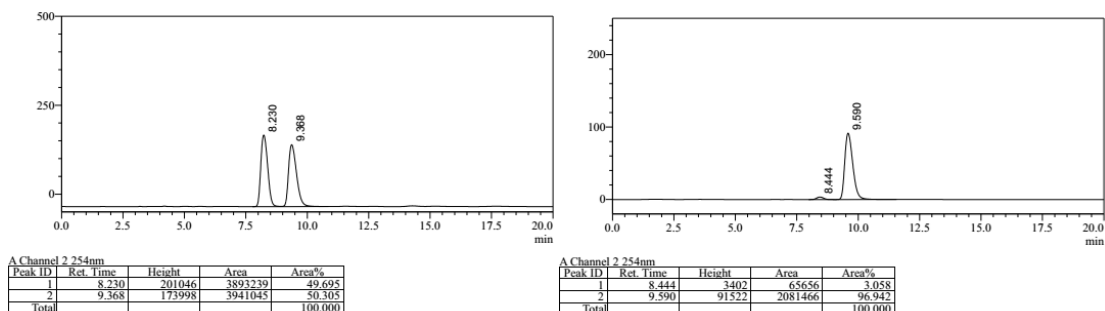
White solid, 66.4 mg, 76% yield, 95:5 *d.r.*, 68% *ee*, $[\alpha]_D^{25} = -49.2$ (*c* 0.24, CHCl₃). ¹H NMR (400 MHz, CDCl₃) δ 7.95-7.41 (m 1H), 7.41-7.28 (m, 7H), 6.94 (s, 1H), 5.39-5.07 (m, 2H), 4.50 (brs, 1H), 4.20 (brs, 1H), 3.74 (s, 3H), 1.18 (brs, 12H); ¹³C NMR (100 MHz, CDCl₃) δ 172.3, 152.8, 142.7, 136.0, 128.8, 128.4, 127.9, 127.7, 125.3, 124.5, 122.5, 115.3, 84.0, 66.9, 52.5, 48.5, 47.4, 25.1, 24.7; ¹¹B NMR (128 MHz, CDCl₃) δ 31.8; HRMS (ESI) calcd for C₂₄H₂₉BNO₆ ([M+H]⁺): 438.2088, found: 438.2090; The enantiopurity was determined by HPLC analysis (Daicel Chiralpak IC column, Hexane/*i*PrOH = 90: 10, flow rate = 1.0 mL/min., wavelength = 254 nm, *t*_R = 14.25 (major), 15.42 (minor).



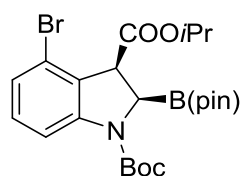
Compound 2g



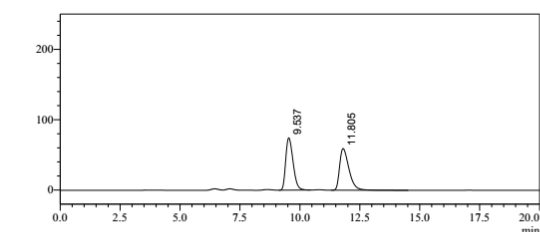
Colorless oil, 83.5 mg, 93% yield, >98:2 *d.r.*, 94% *ee*, $[\alpha]_D^{25} = -73.8$ (*c* 0.14, CHCl₃). ¹H NMR (400 MHz, CDCl₃) δ 7.53 (brs, 1H), 7.18-7.13 (m, 1H), 6.64-6.59 (m, 1H), 5.04-5.01 (m, 1H), 4.45 (d, *J* = 10.0 Hz, 1H), 4.16 (d, *J* = 12.0 Hz, 1H), 1.57 (s, 9H), 1.30-1.22 (m, 18H); ¹³C NMR (100 MHz, CDCl₃) δ 171.2, 159.1 (d, *J* = 245.6 Hz), 152.1, 145.0, 130.3 (d, *J* = 8.1 Hz), 115.7 (d, *J* = 17.9 Hz), 110.6, 108.7 (d, *J* = 20.2 Hz), 84.0, 81.7, 69.1, 50.0, 44.9, 28.3, 25.2, 24.7, 21.6, 21.4; ¹¹B NMR (128 MHz, CDCl₃) δ 31.5 ppm; HRMS (ESI) calcd for C₂₃H₃₃BFNNaO₆ ([M+Na]⁺): 472.2283, found: 472.2290. The enantiopurity was determined by HPLC analysis (Daicel Chiralpak IE column, Hexane/*i*PrOH = 98:2, flow rate = 1.0 mL/min., wavelength = 254 nm, *t*_R = 8.44 (minor), 9.59 (major).



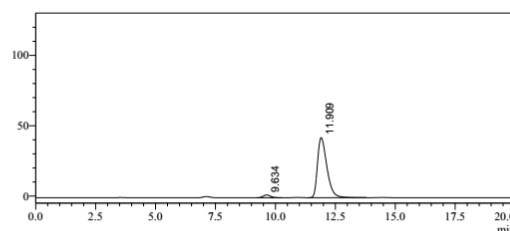
Compound 2h



Colorless oil, 65.4 mg, 64% yield, 94:6 *d.r.*, 93% *ee*, $[\alpha]_D^{25} = -17.5$ (*c* 0.16, CHCl₃). ¹H NMR (400 MHz, CDCl₃) δ 7.42 (brs, 1H), 7.07-7.06 (m, 2H), 5.01 (s, 1H), 4.27 (brs, 1H), 4.09 (brs, 1H), 1.57 (s, 9H), 1.29-1.23 (m, 18H); ¹³C NMR (100 MHz, CDCl₃) δ 170.5, 152.5, 140.0, 130.6, 130.0, 125.0, 120.2, 113.5, 84.2, 82.2, 69.0, 49.1, 49.0, 28.3, 25.4, 24.6, 21.6, 21.5; ¹¹B NMR (128 MHz, CDCl₃) δ 31.1; HRMS (ESI) calcd for C₂₃H₃₃B⁸¹BrNNaO₆ ([M+Na]⁺): 534.1462, found: 534.1464. The enantiopurity was determined by HPLC analysis (Daicel Chiralpak IE column, Hexane/*i*PrOH = 98:2, flow rate = 1.0 mL/min., wavelength = 254 nm, *t*_R = 9.63 (minor), 11.91 (major)).

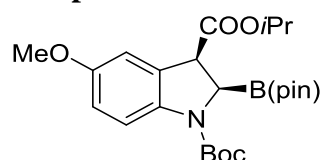


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Total				100.000

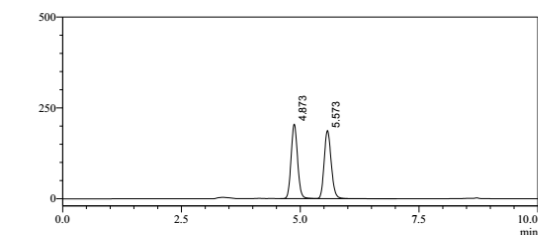


Peak ID	Ret. Time	Height	Area	Area%
1	9.634	1933	41372	3.427
2	11.909	42664	1166028	96.573
Total				100.000

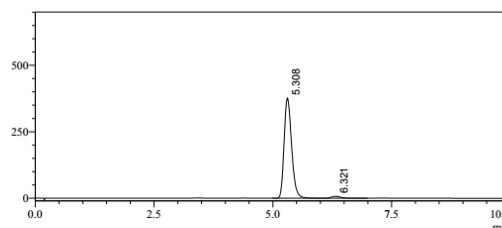
Compound 2i



Colorless oil, 77.4 mg, 84% yield, 95:5 *d.r.*, 95% *ee*, $[\alpha]_D^{25} = -46.6$ (*c* 0.38, CHCl₃). ¹H NMR (400 MHz, CDCl₃) δ 7.39 (s, 1H), 6.88 (s, 1H), 6.75-6.72 (m, 1H), 5.08-5.02 (m, 1H), 4.42 (d, *J* = 11.6 Hz, 1H), 4.10 (d, *J* = 12.0 Hz, 1H), 3.74 (s, 3H), 1.56 (s, 9H), 1.32-1.27 (m, 18H); ¹³C NMR (100 MHz, CDCl₃) δ 171.4, 155.0, 152.3, 136.0, 129.5, 115.5, 113.7, 110.7, 83.8, 81.0, 69.2, 55.6, 48.9, 47.8, 28.4, 25.3, 24.8, 21.8(2), 21.8(0); ¹¹B NMR (128 MHz, CDCl₃) δ 30.5; HRMS (ESI) calcd for C₂₄H₃₇BNO₇ ([M+H]⁺): 462.2663, found: 462.2665. The enantiopurity was determined by HPLC analysis (Daicel Chiralpak IC column, Hexane/*i*PrOH = 90:10, flow rate = 1.0 mL/min., wavelength = 254 nm, *t*_R = 5.31 (major), 6.32 (minor)).

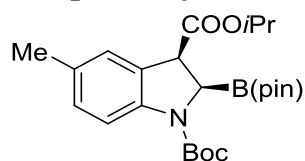


Peak ID	Ret. Time	Height	Area	Area%
1	4.873	205064	1949896	50.130
2	5.573	187695	1939776	49.870
Total				100.000

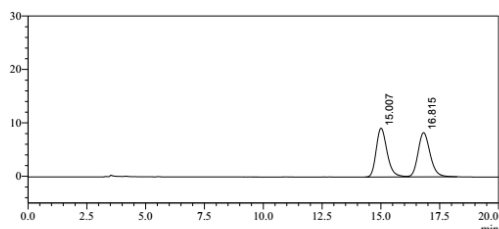


Peak ID	Ret. Time	Height	Area	Area%
1	5.308	376486	4187665	97.554
2	6.321	7279	104258	2.446
Total				100.000

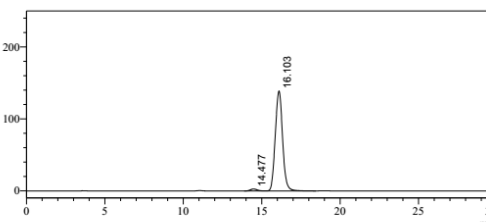
Compound 2j



Colorless oil, 81.0 mg, 91% yield, 97:3 *d.r.*, 96% *ee*, $[\alpha]_D^{25} = -39.7$ (*c* 0.08, CHCl₃). ¹H NMR (400 MHz, CDCl₃) δ 7.36 (s, 1H), 7.08 (s, 1H), 6.98 (d, *J* = 8.0 Hz, 1H), 5.07-5.01(m, 1H), 4.40 (d, *J* = 11.6 Hz, 1H), 4.09 (d, *J* = 12.0 Hz, 1H), 2.26 (s, 3H), 1.57 (s, 9H), 1.32-1.26 (m, 18H); ¹³C NMR (100 MHz, CDCl₃) δ 171.7, 152.4, 139.9, 131.1, 129.0, 128.5, 125.3, 114.7, 83.7, 81.1, 69.1, 48.7, 47.8, 28.4, 25.3, 24.9, 21.8, 21.7, 20.8; ¹¹B NMR (128 MHz, CDCl₃) δ 31.8; HRMS (ESI) calcd for C₂₄H₃₇BNO₆ ([M+H]⁺): 446.2714, found: 446.2710. The enantiopurity was determined by HPLC analysis (Daicel Chiralpak IE column, Hexane/*i*PrOH = 98:2, flow rate = 1.0 mL/min., wavelength = 254 nm, *t*_R = 14.48 (minor), 16.10 (major)).

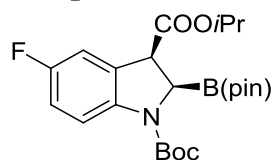


Peak ID	Ret. Time	Height	Area	Area%
1	15.007	9134	294351	49.857
2	16.815	8281	296039	50.143
Total				100.000



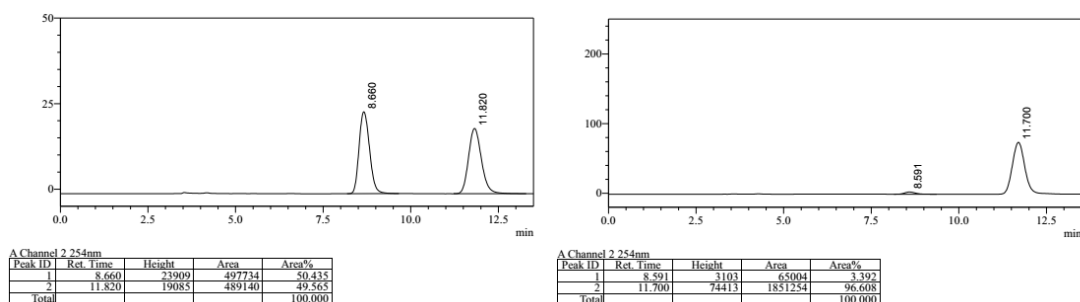
Peak ID	Ret. Time	Height	Area	Area%
1	14.477	2753	84206	1.829
2	16.103	139084	4518703	98.171
Total				100.000

Compound 2k

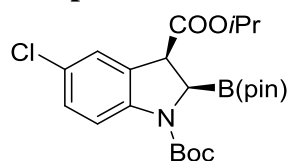


Colorless oil, 60.2 mg, 67% yield, 85:15 *d.r.*, 93% *ee*, $[\alpha]_D^{25} = -32.8$ (*c* 0.14, CHCl₃). ¹H NMR (400 MHz, CDCl₃) δ 7.41 (brs, 1H), 7.01 (d, *J* = 6.8 Hz, 1H), 6.90-6.86 (m, 1H), 5.07-5.02 (m, 1H), 4.42 (d, *J* = 11.6 Hz, 1H), 4.13 (d, *J* = 12.0 Hz, 1H), 1.57 (s, 9H), 1.32-1.27(m, 18H); ¹³C NMR (100 MHz, CDCl₃) δ 171.0, 158.2 (d, *J* = 238.1 Hz), 152.2, 138.4, 129.8, 115.5, 114.9 (d, *J* = 22.7 Hz), 112.1, 83.9, 81.4, 69.6, 49.0, 47.6, 28.3, 25.3, 24.8, 21.8; ¹¹B NMR (128 MHz, CDCl₃) δ 31.3; HRMS (ESI) calcd for C₂₃H₃₄BFNO₆ ([M+H]⁺): 450.2463, found: 450.2462. The enantiopurity was

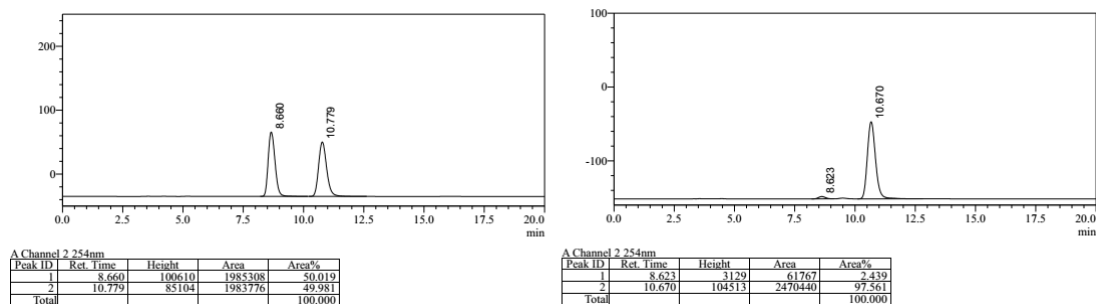
determined by HPLC analysis (Daicel Chiralpak IE column, Hexane/*i*PrOH = 98:2, flow rate = 1.0 mL/min., wavelength = 254 nm, t_R = 8.59 (minor), 11.70 (major).



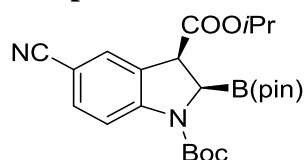
Compound 2l



Colorless oil, 77.2 mg, 83% yield, 94:6 *d.r.*, 95% *ee*, $[\alpha]_D^{25} = -25.0$ (*c* 0.20, CHCl₃). ¹H NMR (400 MHz, CDCl₃) δ 7.35 (s, 1H), 7.25 (s, 1H), 7.14 (d, *J* = 8.4 Hz, 1H), 5.08-5.02 (m, 1H), 4.41 (d, *J* = 11.6 Hz, 1H), 4.12 (d, *J* = 12.0 Hz, 1H), 1.57 (s, 9H), 1.32-1.27 (m, 18H); ¹³C NMR (100 MHz, CDCl₃) δ 171.0, 152.1, 140.8, 130.0, 128.5, 126.6, 124.9, 115.9, 83.9, 81.6, 69.6, 49.1, 47.6, 28.3, 25.3, 25.0, 21.8; ¹¹B NMR (128 MHz, CDCl₃) δ 31.4; HRMS (ESI) calcd for C₂₃H₃₃BClNO₆ ([M+Na]⁺): 488.1987, found: 488.1982. The enantiopurity was determined by HPLC analysis (Daicel Chiralpak IE column, Hexane/*i*PrOH = 98:2, flow rate = 1.0 mL/min., wavelength = 254 nm, t_R = 8.62 (minor), 10.67 (major).

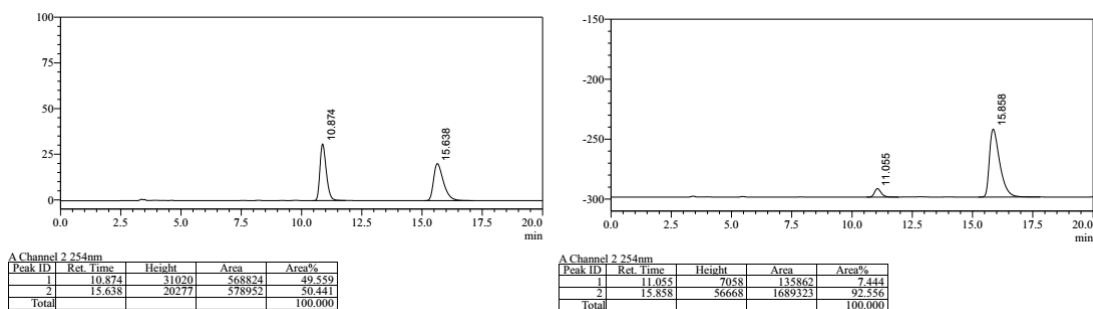


Compound 2m

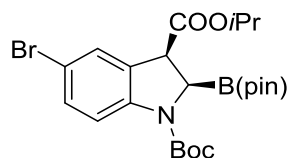


Colorless oil, 45.6 mg, 50% yield, 71:29 *d.r.*, 85% *ee*, $[\alpha]_D^{25} = -57.1$ (*c* 0.14, CHCl₃). ¹H NMR (400 MHz, CDCl₃) δ 7.84 (brs, 1H), 7.56 (s, 1H), 7.50 (d, *J* = 7.6 Hz, 1H), 5.10-5.03 (m, 1H), 4.44 (d, *J* = 12.4 Hz, 1H), 4.16 (d, *J* = 12.0 Hz, 1H), 1.58 (s, 9H), 1.32-1.27 (m, 18H); ¹³C NMR (100 MHz, CDCl₃) δ 170.5, 151.7, 145.7, 133.6, 129.4,, 128.5, 119.4, 115.2, 104.4, 84.0, 82.5, 70.0, 49.2, 47.3, 28.2, 25.2, 24.9, 21.7; ¹¹B

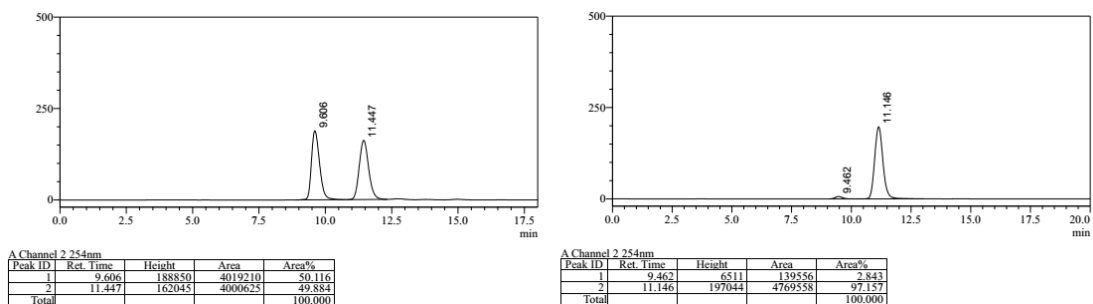
NMR (128 MHz, CDCl₃) δ 31.2 ppm; HRMS (ESI) calcd for C₂₄H₃₃BN₂NaO₆ ([M+Na]⁺): 479.2329, found: 479.2332. The enantiopurity was determined by HPLC analysis (Daicel Chiralpak IE column, Hexane/*i*PrOH = 90: 10, flow rate = 1.0 mL/min., wavelength = 254 nm, t_R = 11.06 (minor), 15.86 (major)).



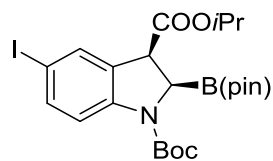
Compound 2n



Colorless oil, 89.9 mg, 88% yield, 95:5 *d.r.*, 94% *ee*, $[\alpha]_D^{25} = -20.0$ (*c* 0.76, CHCl₃). ¹H NMR (400 MHz, CDCl₃) δ 7.70-7.27 (m, 3H), 5.08-5.02 (m, 1H), 4.41 (d, *J* = 12.0 Hz, 1H), 4.11 (d, *J* = 12.0 Hz, 1H), 1.57 (s, 9H), 1.31-1.26(m, 18H); ¹³C NMR (100 MHz, CDCl₃) δ 171.0, 152.1, 141.4, 131.4, 130.5, 127.8, 116.4, 113.9, 83.9, 81.7, 69.6, 49.0, 47.5, 28.3, 25.3, 25.0, 21.8; ¹¹B NMR (128 MHz, CDCl₃) δ 31.4; HRMS (ESI) calcd for C₂₃H₃₃B⁸¹BrNNaO₆ ([M+Na]⁺): 534.1462, found: 534.1466. The enantiopurity was determined by HPLC analysis (Daicel Chiralpak IE column, Hexane/*i*PrOH = 98:2, flow rate = 1.0 mL/min., wavelength = 254 nm, t_R = 9.46 (minor), 11.15 (major)).

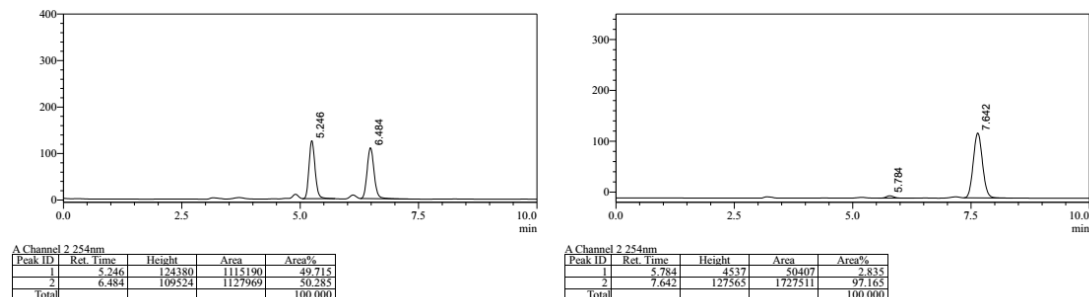


Compound 2o

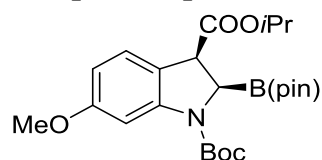


Colorless oil, 95.8 mg, 86% yield, 97:3 *d.r.*, 94% *ee*, $[\alpha]_D^{25} = -4.0$ (*c* 0.30, CHCl₃). ¹H NMR (400 MHz, CDCl₃) δ 7.58 (s, 1H), 7.46 (d, *J* = 8.0 Hz, 1H), 7.27 (brs, 1H), 5.08-5.02 (m, 1H), 4.41 (d, *J* = 12.0 Hz, 1H), 4.09 (d, *J* = 12.0 Hz, 1H), 1.56 (s, 9H),

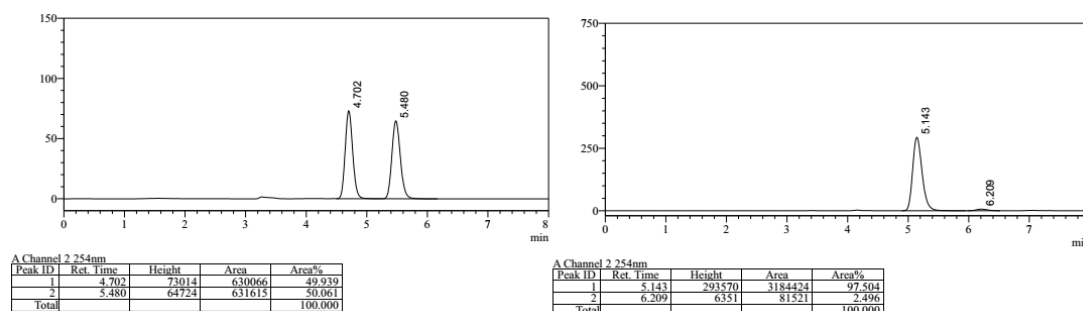
1.31-1.26 (m, 18H); ^{13}C NMR (100 MHz, CDCl_3) δ 171.0, 152.1, 141.9, 137.3, 133.6, 130.9, 117.0, 83.9, 83.6, 81.6, 69.6, 48.8, 47.2, 28.3, 25.3, 24.9, 21.8; ^{11}B NMR (128 MHz, CDCl_3) δ 30.7; HRMS (ESI) calcd for $\text{C}_{23}\text{H}_{34}\text{BINO}_6$ ($[\text{M}+\text{H}]^+$): 558.1524, found: 558.1520. The enantiopurity was determined by HPLC analysis (Daicel Chiralpak IF column, Hexane/*i*PrOH = 95:5, flow rate = 1.0 mL/min., wavelength = 254 nm, t_R = 5.78 (minor), 7.64 (major)).



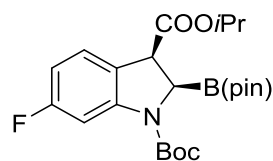
Compound 2p



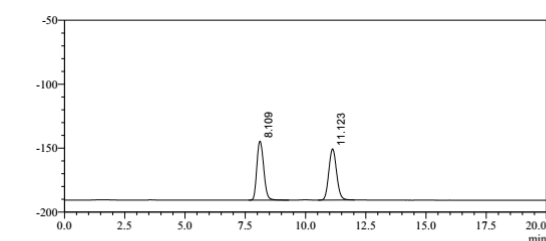
Colorless oil, 81.1 mg, 88% yield, 98:2 *d.r.*, 95% *ee*, $[\alpha]_D^{25} = -73.8$ (*c* 0.13, CHCl_3). ^1H NMR (400 MHz, CDCl_3) δ 7.45-7.14 (m, 2H), 6.47-6.44 (m, 1H), 5.07-4.98 (m, 1H), 4.37 (d, $J = 12.0$ Hz, 1H), 4.11 (d, $J = 12.0$ Hz, 1H), 3.77 (s, 3H), 1.58 (s, 9H), 1.32-1.24 (m, 18H); ^{13}C NMR (100 MHz, CDCl_3) δ 171.8, 160.4, 152.3, 143.3, 124.9, 120.3, 107.6, 101.1, 83.8, 81.2, 69.1, 55.3, 49.8, 47.1, 28.4, 25.3, 24.8, 21.8, 21.7; ^{11}B NMR (128 MHz, CDCl_3) δ 31.8; HRMS (ESI) calcd for $\text{C}_{24}\text{H}_{36}\text{BNNaO}_7$ ($[\text{M}+\text{Na}]^+$): 484.2483, found: 484.2484. The enantiopurity was determined by HPLC analysis (Daicel Chiralpak IC column, Hexane/*i*PrOH = 90: 10, flow rate = 1.0 mL/min., wavelength = 254 nm, t_R = 5.14 (major), 6.21 (minor)).



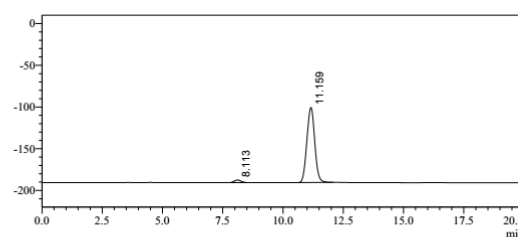
Compound 2q



Colorless oil, 72.7 mg, 81% yield, 97:3 *d.r.*, 94% *ee*, $[\alpha]_D^{25} = -73.6$ (*c* 0.38, CHCl₃). ¹H NMR (400 MHz, CDCl₃) δ 7.59-7.18 (m, 2H), 6.60-6.56 (m, 1H), 5.07-5.01 (m, 1H), 4.39 (d, *J* = 12.0 Hz, 1H), 4.13 (d, *J* = 12.0 Hz, 1H), 1.58 (s, 9H), 1.32-1.24 (m, 18H); ¹³C NMR (100 MHz, CDCl₃) δ 171.4, 163.4 (d, *J* = 241.0 Hz), 152.1, 143.6, 125.4, 123.7, 108.2 (d, *J* = 23.5 Hz), 103.1 (d, *J* = 22.6 Hz), 83.9, 81.8, 69.4, 49.6, 47.0, 28.3, 25.3, 24.8, 21.8, 21.7; ¹¹B NMR (128 MHz, CDCl₃) δ 31.1; HRMS (ESI) calcd for C₂₃H₃₃BFNNaO₆ ([M+Na]⁺): 472.2283, found: 472.2285. The enantiopurity was determined by HPLC analysis (Daicel Chiralpak IE column, Hexane/*i*PrOH = 98:2, flow rate = 1.0 mL/min., wavelength = 254 nm, *t*_R = 8.11 (minor), 11.16 (major).

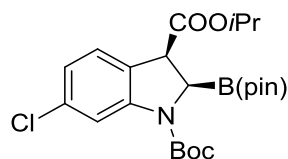


Peak ID	Ret. Time	Height	Area	Area%
1	8.109	46109	914435	50.412
2	11.123	39966	899488	49.588
Total				100.000

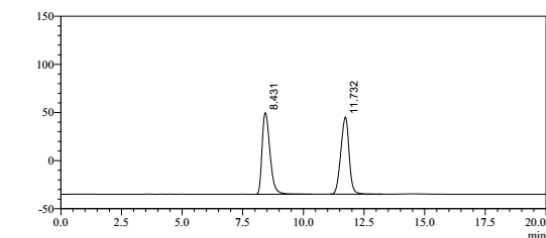


Peak ID	Ret. Time	Height	Area	Area%
1	8.113	3106	63341	3.003
2	11.159	89892	2045712	96.997
Total				100.000

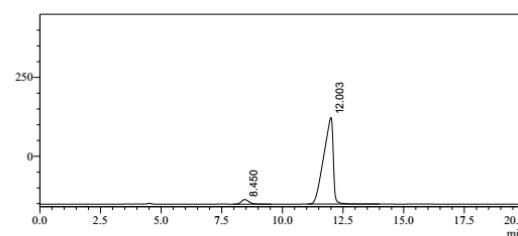
Compound 2r



Colorless oil, 78.1 mg, 84% yield, 93:7 *d.r.*, 92% *ee*, $[\alpha]_D^{25} = -67.6$ (*c* 0.21, CHCl₃). ¹H NMR (400 MHz, CDCl₃) δ 7.48 (brs, 1H), 7.19 (d, *J* = 7.6 Hz, 1H), 6.87 (d, *J* = 8.0 Hz, 1H), 5.07-5.02(m, 1H), 4.39 (d, *J* = 11.6 Hz, 1H), 4.12 (d, *J* = 12.0 Hz, 1H), 1.58 (s, 9H), 1.32-1.25 (m, 18H); ¹³C NMR (100 MHz, CDCl₃) δ 171.1, 151.9, 143.1, 134.3, 126.8, 125.4, 121.6, 115.4, 83.8, 81.6, 69.4, 49.2, 47.2, 28.2, 25.2, 24.9, 21.7; ¹¹B NMR (128 MHz, CDCl₃) δ 31.5; HRMS (ESI) calcd for C₂₃H₃₃BCINNaO₆ ([M+Na]⁺): 488.1987, found: 488.1992. The enantiopurity was determined by HPLC analysis (Daicel Chiralpak IE column, Hexane/*i*PrOH = 98:2, flow rate = 1.0 mL/min., wavelength = 254 nm, *t*_R = 8.45 (minor), 12.00 (major).

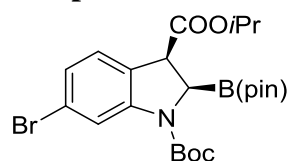


Peak ID	Ret. Time	Height	Area	Area%
1	8.431	84762	1917072	50.101
2	11.732	80202	1909363	49.899
Total				100.000

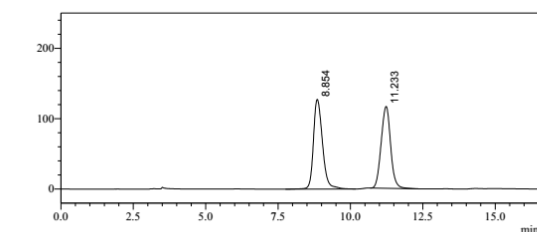


Peak ID	Ret. Time	Height	Area	Area%
1	8.450	14384	319448	4.063
2	12.003	274260	7543034	95.937
Total				100.000

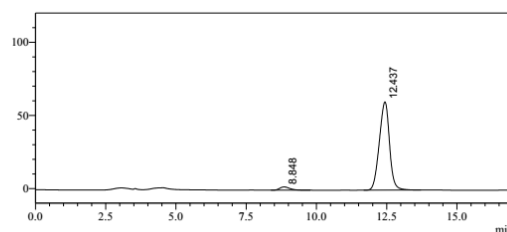
Compound 2s



White solid, 74.3 mg, 73% yield, 94:6 *d.r.*, 93% *ee*, $[\alpha]_D^{25} = -49.5$ (*c* 0.15, CHCl₃). ¹H NMR (400 MHz, CDCl₃) δ 7.64 (s, 1H), 7.14 (d, *J* = 7.6 Hz, 1H), 7.02 (d, *J* = 7.6 Hz, 1H), 5.07-5.01(m, 1H), 4.37 (d, *J* = 12.0 Hz, 1H), 4.11 (d, *J* = 12.0 Hz, 1H), 1.58 (s, 9H), 1.32-1.24 (m, 18H); ¹³C NMR (100 MHz, CDCl₃) δ 171.1, 152.0, 127.4, 125.9, 124.6, 122.4, 118.4, 83.9, 82.0, 69.5, 49.2, 47.3, 28.3, 25.3, 25.0, 21.8 (one aromatic carbon signal does not show.); ¹¹B NMR (128 MHz, CDCl₃) δ 31.8; HRMS (ESI) calcd for C₂₃H₃₃B⁷⁹BrNNaO₆ ([M+Na]⁺): 532.1482, found: 532.1478. The enantiopurity was determined by HPLC analysis (Daicel Chiralpak IE column, Hexane/*i*PrOH = 98:2, flow rate = 1.0 mL/min., wavelength = 254 nm, *t*_R = 8.85 (minor), 12.44 (major).

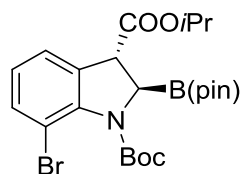


Peak ID	Ret. Time	Height	Area	Area%
1	8.854	127443	2771900	51.098
2	11.233	116293	2652743	48.902
Total				100.000

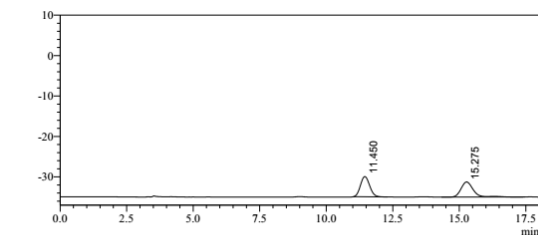


Peak ID	Ret. Time	Height	Area	Area%
1	8.848	2236	56832	3.597
2	12.437	60315	1523194	96.403
Total				100.000

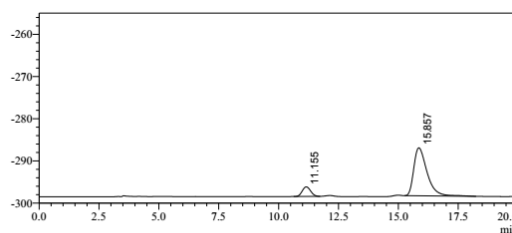
Compound 3t



Colorless oil, 52.9 mg, 52% yield, 16:84 *d.r.*, 78% *ee*, $[\alpha]_D^{25} = -50.0$ (*c* 0.16, CHCl₃). ¹H NMR (400 MHz, CDCl₃) δ 7.39 (d, *J* = 8.0 Hz, 1H), 7.23 (d, *J* = 7.2 Hz, 1H), 6.92-6.88 (m, 1H), 5.07-5.00 (m, 1H), 4.09-4.04 (m, 2H), 1.56 (s, 9H), 1.26-1.24 (m, 18H); ¹³C NMR (100 MHz, CDCl₃) δ 171.2, 155.4, 142.5, 136.5, 132.9, 125.7, 122.9, 112.5, 83.4, 82.9, 68.8, 54.3, 49.6, 28.3, 24.8, 24.7, 21.8; ¹¹B NMR (128 MHz, CDCl₃) δ 27.6 ppm; HRMS (ESI) calcd for C₂₃H₃₃B⁸¹BrNNaO₆ ([M+Na]⁺): 534.1462, found: 534.1467. The enantiopurity was determined by HPLC analysis (Daicel Chiralpak IE column, Hexane/*i*PrOH = 98:2, flow rate = 1.0 mL/min., wavelength = 254 nm, *t*_R = 11.16 (minor), 15.86 (major).

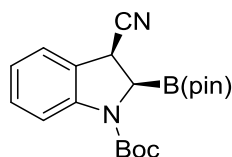


Peak ID	Ret. Time	Height	Area	Area%
1	11.450	5007	120558	50.270
2	15.275	3737	119262	49.730
Total				100.000

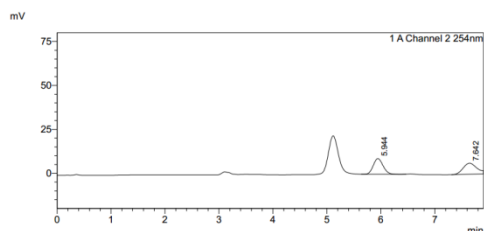


Peak ID	Ret. Time	Height	Area	Area%
1	11.155	2262	55462	11.209
2	15.857	11371	439339	88.791
Total				100.000

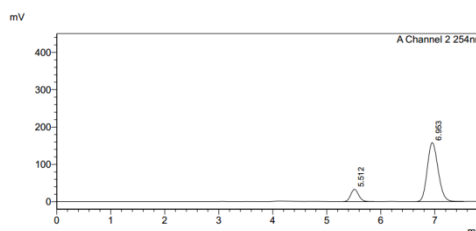
Compound 2v



Colorless oil, 62.2 mg, 84% yield, 92:8 *d.r.*, 73% *ee*, $[\alpha]_D^{25} = -35.0$ (*c* 0.37, CHCl₃). ¹H NMR (400 MHz, CDCl₃) δ 7.86-7.46 (m, 1H), 7.32 (d, *J* = 7.6 Hz, 1H), 7.28-7.25 (m, 1H), 7.02-6.98 (m, 1H), 4.50 (br s, 1H), 4.05 (br s, 1H), 1.59 (s, 9H), 1.35 (s, 12H); ¹³C NMR (100 MHz, CDCl₃) δ 152.7, 141.5, 129.8, 125.3, 122.7, 118.3, 115.0, 84.8, 82.6, 47.4, 30.8, 28.3, 25.4, 24.4; HRMS (ESI) calcd for C₂₀H₂₇BN₂NaO₄([M+Na]⁺): 393.1956, found: 393.1963. The enantiopurity was determined by HPLC analysis (Daicel Chiralpak IC column, Hexane/*i*PrOH = 90:10, flow rate = 1.0 mL/min., wavelength = 254 nm, *t*_R = 5.51 (minor), 6.95 (major)).



Peak ID	Ret. Time	Height	Area	Area%
1	5.944	8872	109485	49.926
2	7.642	6240	109609	50.074
Total				100.000



Peak ID	Ret. Time	Height	Area	Area%
1	5.512	32845	341806	13.408
2	6.953	157893	2207447	86.592
Total				100.000

8. Method to determine diastereoselective ratio (*d.r.*) by NMR (reaction of 1c at room temperature as an example)

To determine the *d.r.* value (**2c**/**3c**), H^{a-c} and H^{a'-c'} were chosen as diagnostic signals. Apparently, H^a and H^{a'} overlap at δ 5.07 ppm. In order to assign H^b, H^c, H^{b'}, and H^{c'}, a deuterium experiment was conducted. The reaction was carried out with MeOD instead of normal alcohol. Therefore by combination of ¹H NMR spectra from normal reaction we could elucidate the assignments of these diagnostic protons as shown in Figures S1 and S2. The disappearance of signal at δ 4.48 ppm upon use of MeOD indicates that H^c (or H^{c'}) is C3-proton in **2c** (or **3c**) (Figures S1 and S2, right spectra). Another information could be obtained is that H^b and H^{b'} overlap at δ 4.10 ppm (Figure S1). Lastly, the integration difference (0.12) of signal at δ 4.10 ppm minus that of signal at δ 4.48 ppm is twice as much as difference (0.06) between signals at δ 5.07

ppm and δ 4.48 ppm, which indicates that H^b and H^c overlap at δ 4.10 ppm. Therefore the chemical of 4.48 ppm belongs to H^c . With that, d.r. value could be obtained from singals at δ 5.07 ppm (H^a and $H^{a'}$) and δ 4.48 ppm (H^c). A total integration of H^a and $H^{a'}$ is denoted $I^{a+a'}$. The integration of H^c is denoted I^c . Therefore we could deduce the I^a by subtraction I^c from $I^{a+a'}$. Thus the d.r. could be calculated as $100 \cdot I^c / I^{a+a'} : 100 \cdot I^a / I^{a+a'}$. The d.r. values of the rest substrate scope could be calculated by analogue.

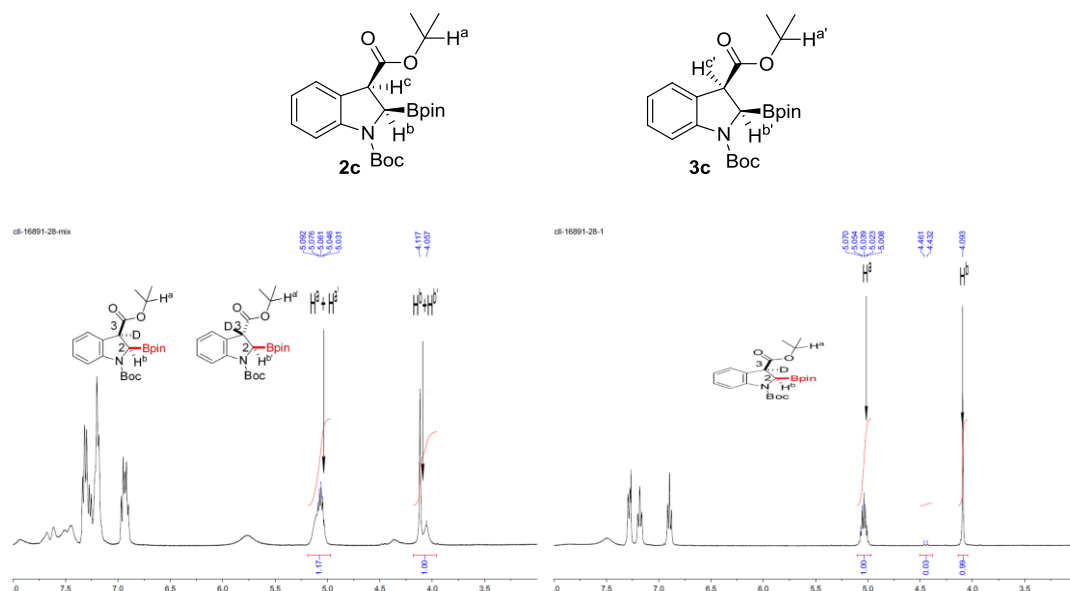


Figure S1. Partial ^1H NMR spectra of deuterium experiment. Left: ^1H NMR of crude reaction mixture; right: pure $C3\text{-D}$ *cis* product.

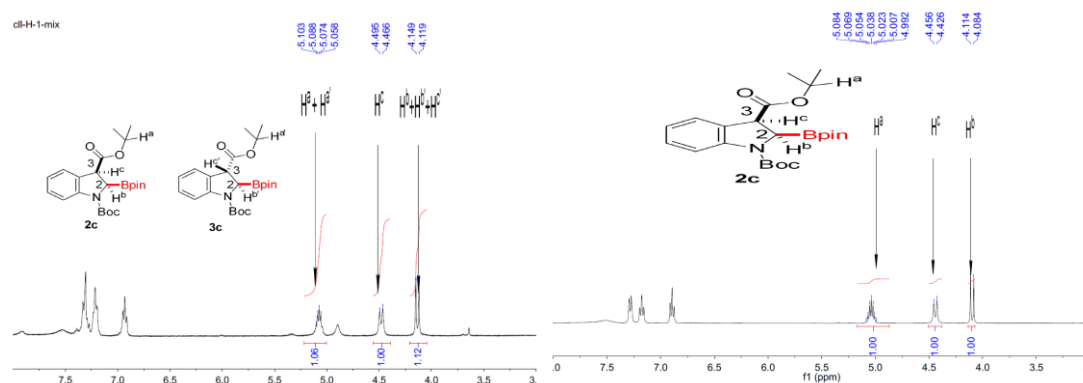
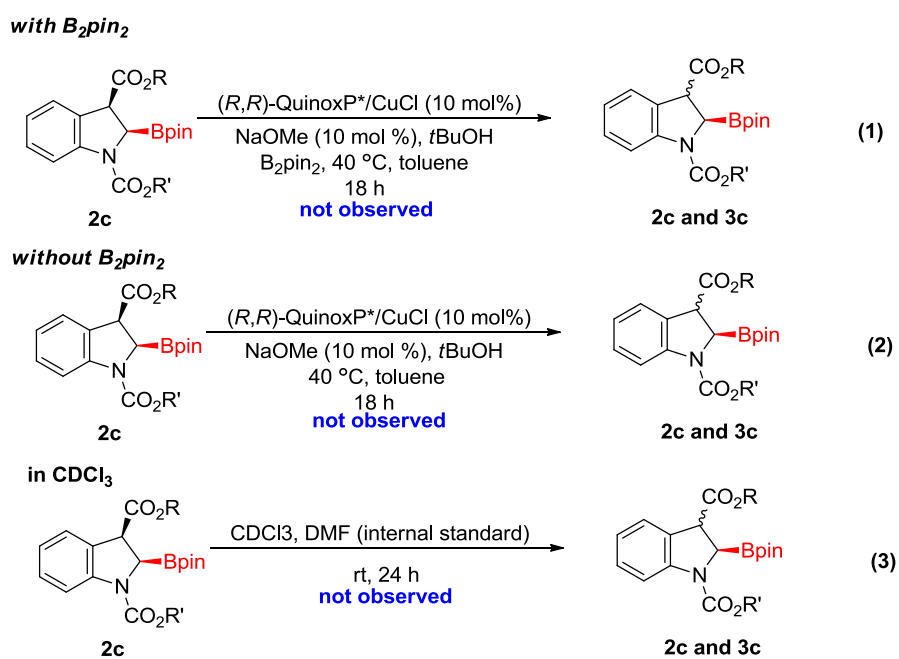


Figure S2. Partial ^1H NMR spectra of crude mixture and **2c**. Left: crude ^1H NMR of crude mixture; right: pure *cis* product **2c**.

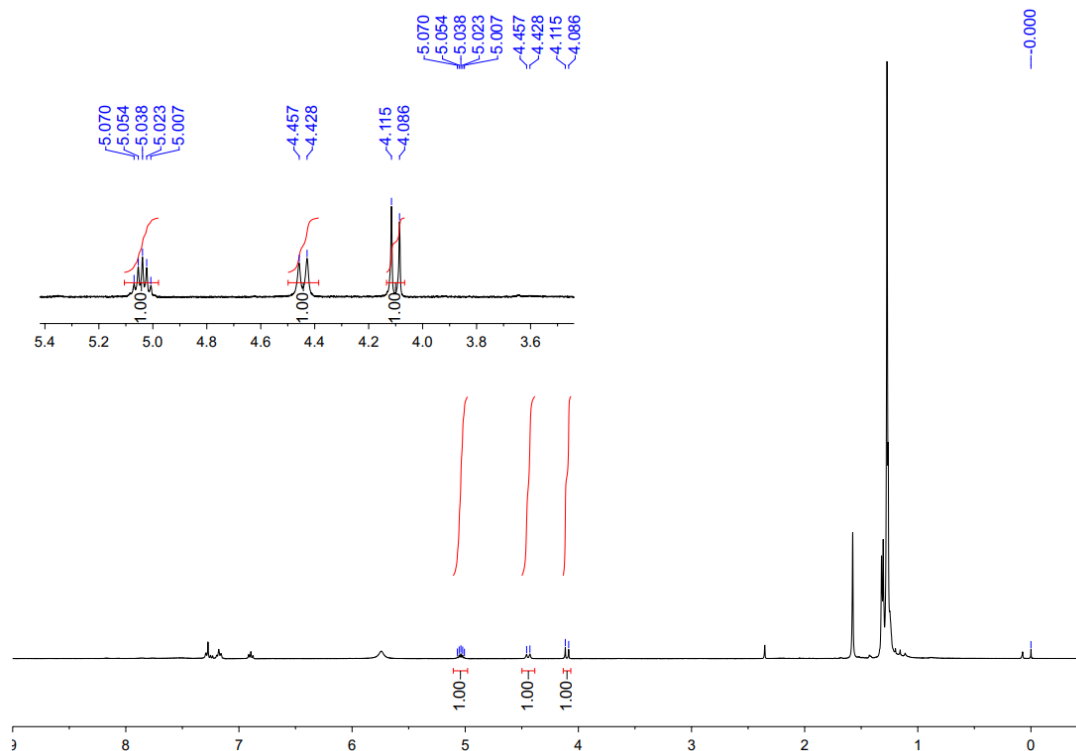
9. Control experiments for isomerization test and NMR spectrum

In order to know possibility of isomerization of product, three control experiments were conducted: 1) **2c** under reaction conditions at 40°C for 18 h; 2) **2c** under reaction conditions without B_2pin_2 at 40°C for 18 h; 3) **2c** in CDCl_3 for 24 hours. The results of these experiments show that no isomerization was observed under these

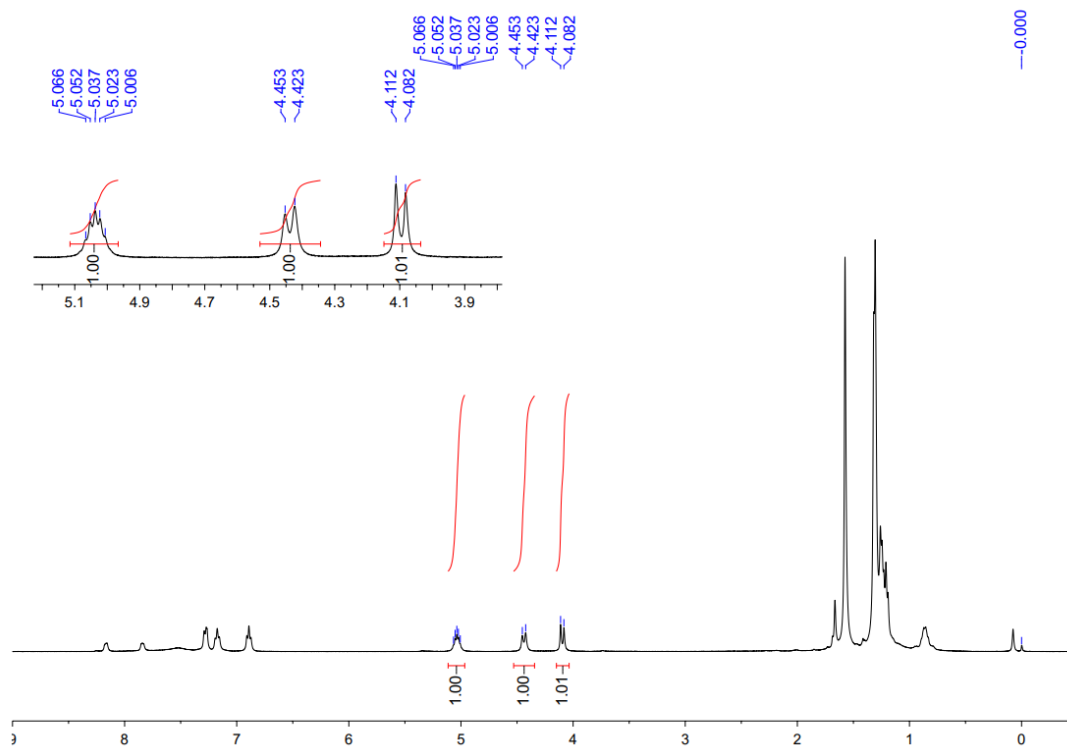
conditions, demonstrating relative stability of stereochemistry of **2c**. Additionally, the ee value of each control experiment was not changed.



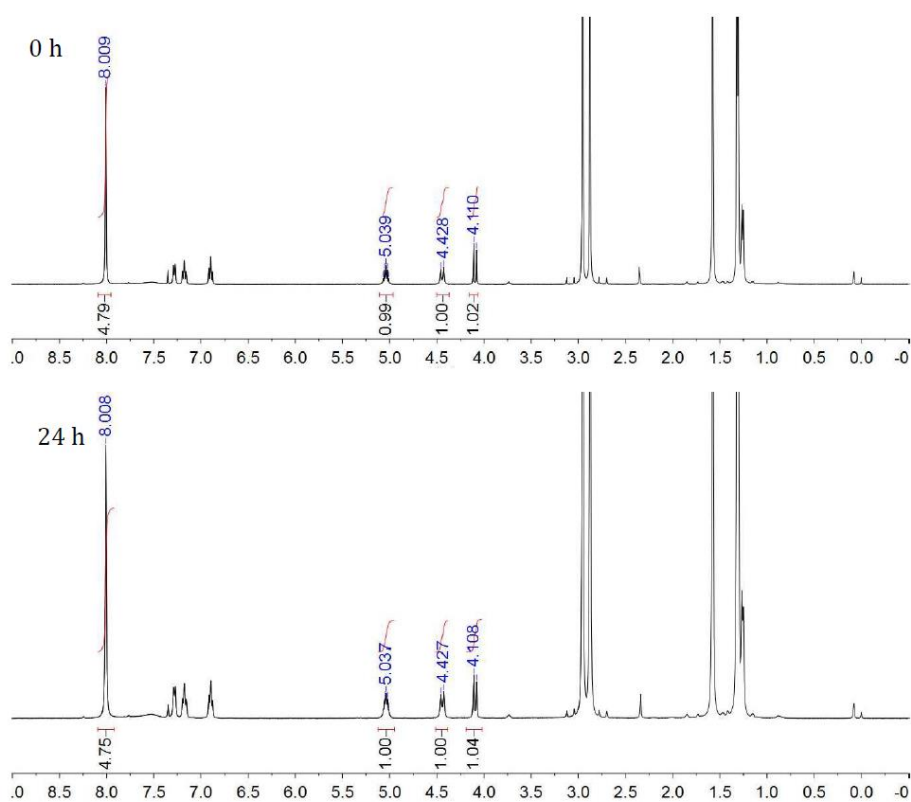
NMR of control experiment (1)



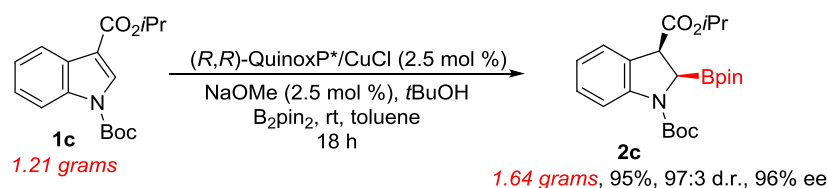
NMR of control experiment (2)



NMR of control experiment (3)

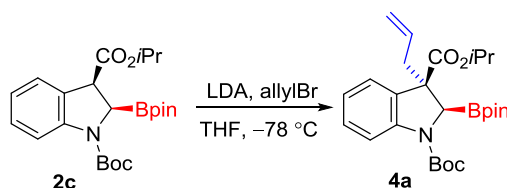


10. Gram-scale dearomative borylation of indole 2c



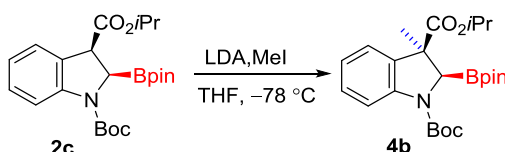
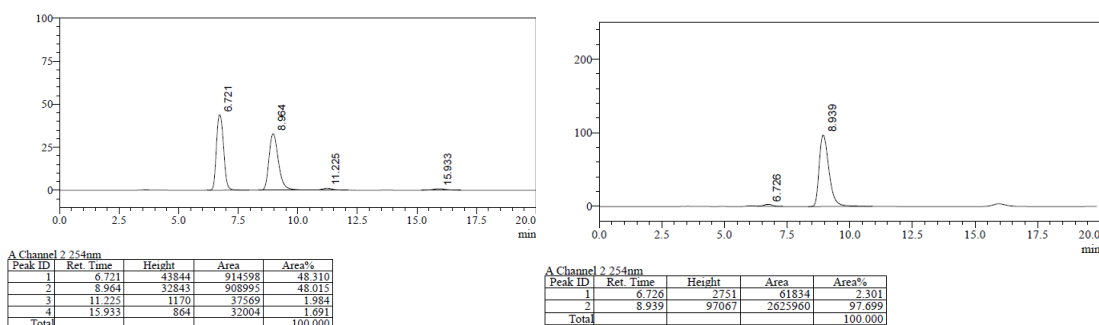
To a 100-mL flame-dried flask charged with CuCl (9.9 mg, 0.10 mmol), NaOMe (5.4 mg, 0.10 mmol) and (R,R)-QuinoxP* (33.5 mg, 0.10 mmol) was added toluene (10 mL). The resulting mixture was allowed to stir at room temperature for 0.5 h. B₂pin₂ (1.53 g, 6.0 mmol in 10 mL) was then introduced and the reaction was allowed to stir at same temperature for 10 min followed by addition of indole **1c** (1.21 g, 4.0 mmol in 20 mL toluene) and tBuOH (0.77 mL, 8.0 mmol) at same temperature. The resulting mixture was continued to stir at room temperature for 12 h. After removal of the solvent, the ¹H NMR of crude mixture was taken to determine the d.r. value (97:3). The residue was purified by column chromatography on silica gel using PE/EtOAc (12:1) as the eluent to afford corresponding 2-boryl indole **2c** as colorless oil (1.64 g, 95% yield, 97:3 d.r., 96% ee).

11. C3 functionalization of 2-borylindoline 4

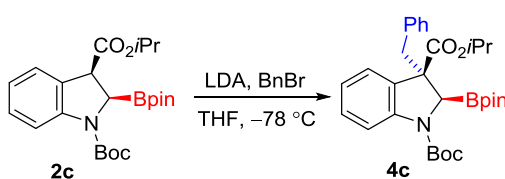
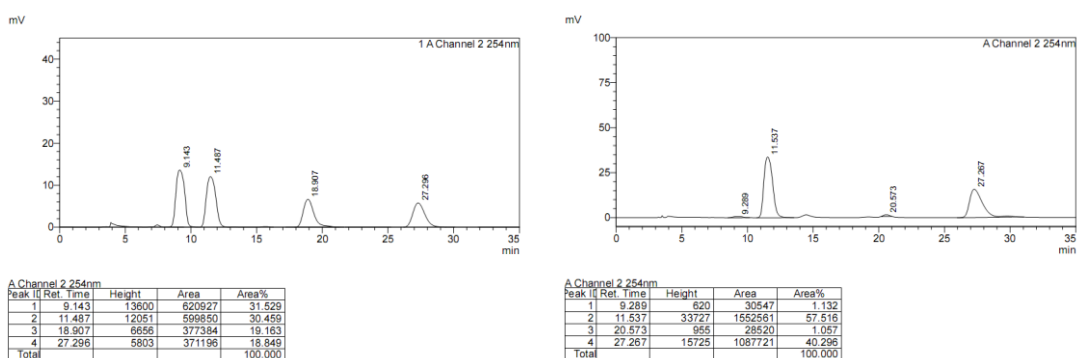


To a 50-mL flame-dried flask charged with 2-borylindoline **2c** (1.64 g, 3.80 mmol) and THF (30 mL) was added LDA (2.85 mL, 2M in hep/ethylbenzene, 5.7 mmol, 1.5 equiv) at -78 °C dropwisely. The resulting mixture was allowed at same temperature for 30 min. The allyl bromide (0.49 mL, 5.7 mmol, 1.5 equiv) was then added slowly at -78 °C. The reaction mixture was then allowed to warm to room temperature and continued to stir at same temperature for 3 h. After removal of the solvent, the residue was purified by column chromatography on silica gel using PE/EtOAc (20: 1) as the eluent to afford C3 allylated 2-borylindoline **4a** as colorless oil (1.75 g, 98%). The relative configuration was determined by 2D NMR NOSEY spectrum.

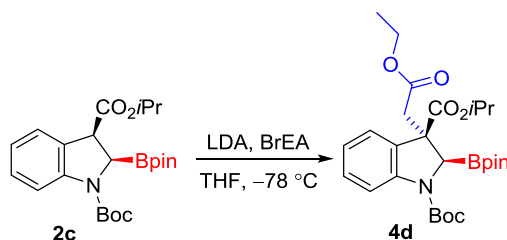
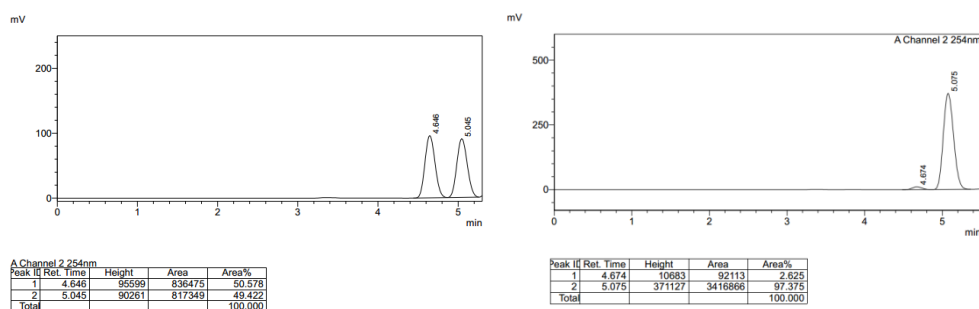
96:4 d.r., 95% ee, $[\alpha]_D^{25} = -50.0$ (c 0.16, CHCl₃). ¹H NMR (400 MHz, CDCl₃) δ 7.84-7.47 (m, 1H), 7.35-7.33 (m, 1H), 7.18 (s, 1H), 6.91 (s, 1H), 5.60 (s, 1H), 5.13-5.05 (m, 3H), 3.87 (s, 1H), 2.60-2.66 (m, 2H), 1.57 (s, 9H), 1.27 (brs, 18H); ¹³C NMR (100 MHz, CDCl₃) δ 173.0, 152.3, 142.1, 132.6, 132.0, 128.6, 124.4, 121.7, 119.2, 114.9, 83.4, 81.1, 69.5, 54.1, 45.1, 28.4, 28.0, 25.3, 24.9, 21.7, 21.6; HRMS (ESI) calcd for C₂₆H₃₈BNNaO₆ ([M+Na]⁺): 494.2690, found: 494.2689. The enantiopurity was determined by HPLC analysis (Daicel Chiralpak IE column, Hexane/iPrOH = 98:2, flow rate = 1.0 mL/min., wavelength = 254 nm, t_R = 6.73 (minor), 8.94 (major))



To a 25-mL flame-dried flask charged with 2-borylindoline **2c** (86.2 mg, 0.20 mmol) and THF (1 mL) was added LDA (150 μL , 2M in hep/ethylbenzene, 0.3 mmol, 1.5 equiv) at $-78\text{ }^\circ\text{C}$ dropwisely. The resulting mixture was allowed at same temperature for 10 min. The iodomethane (24.4 μL , 0.4 mmol, 2.0 equiv) was then added slowly at $-78\text{ }^\circ\text{C}$. The reaction mixture was then allowed to warm to room temperature and continued to stir at same temperature for 10 min. After removal of the solvent, the residue was purified by column chromatography on silica gel using PE/EtOAc (15: 1) as the eluent to afford C3 methylated 2-borylindoline **8** as colorless oil (59.6 mg, 67%). d.r. = 2:1, 96% *ee*, $[\alpha]_D^{25} = -18.0$ (*c* 0.3, CHCl_3). $^1\text{H NMR}$ (400 MHz, CDCl_3) δ 7.46 (s, 1H), 7.28-7.27 (m, 1H), 7.20-7.16 (m, 1H), 6.93-6.90 (m, 1H), 5.00-4.97 (m, 1H), 4.38 (s, 0.3H), 3.68 (s, 0.6H), 1.58 (s, 9H), 1.29-1.20 (m, 18H); $^{13}\text{C NMR}$ (100 MHz, CDCl_3) δ 174.0, 173.5, 152.6, 129.0, 128.5, 124.6, 123.7, 122.7, 121.9, 121.8, 115.3, 114.9, 83.5, 81.1, 69.3, 68.8, 57.7, 28.4, 28.0, 27.2, 25.3, 24.9, 21.7, 21.5; HRMS (ESI) calcd for $\text{C}_{24}\text{H}_{36}\text{BNNaO}_6$ ($[\text{M}+\text{Na}]^+$): 468.2528, found: 468.2542. The enantiopurity was determined by HPLC analysis (Daicel Chiralpak IE column, Hexane/*i*PrOH = 99:1, flow rate = 1.0 mL/min., wavelength = 254 nm, t_R = 9.29 (minor), 11.54 (major))

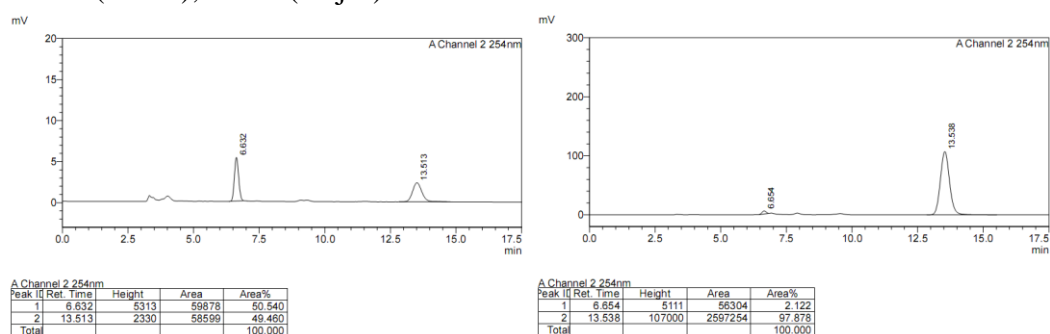


To a 25-mL flame-dried flask charged with 2-borylindoline **2c** (86.2 mg, 0.20 mmol) and THF (1 mL) was added LDA (150 μ L, 2M in hep/ethylbenzene, 0.3 mmol, 1.5 equiv) at -78 $^{\circ}$ C dropwisely. The resulting mixture was allowed at same temperature for 10 min. The benzyl bromide (35.6 μ L, 0.3 mmol, 1.5 equiv) was then added slowly at -78 $^{\circ}$ C. The reaction mixture was then allowed to warm to room temperature and continued to stir at same temperature for 10 min. After removal of the solvent, the residue was purified by column chromatography on silica gel using PE/EtOAc (15: 1) as the eluent to afford C3 benzylated 2-borylindoline **4c** as colorless oil (67.8 mg, 65%). 95% *ee*, $[\alpha]_D^{25} = -23.0$ (*c* 0.57, CHCl_3). ^1H NMR (400 MHz, CDCl_3) δ 7.68-7.34 (m, 2H), 7.19-7.09 (m, 4H), 6.94-6.74 (m, 3H), 5.09 (s, 1H), 3.97 (s, 1H), 3.27 (d, *J* = 6.8 Hz, 1H), 3.10-3.02 (m, 1H), 1.46 (s, 9H), 1.31-1.23 (m, 18H); ^{13}C NMR (100 MHz, CDCl_3) δ 173.4, 151.7, 142.9, 135.7, 131.8, 129.9, 128.7, 127.7, 126.6, 124.9, 121.4, 115.1, 83.3, 80.9, 69.7, 58.9, 53.6, 46.3, 28.3, 25.2, 24.7, 21.7; HRMS (ESI) calcd for $\text{C}_{30}\text{H}_{41}\text{BNO}_6$ ($[\text{M}+\text{H}]^+$): 522.3021, found: 522.3030. The enantiopurity was determined by HPLC analysis (Daicel Chiralpak IE column, Hexane/*i*PrOH = 99:1, flow rate = 1.0 mL/min., wavelength = 254 nm, t_R = 4.67 (minor), 5.08 (major))

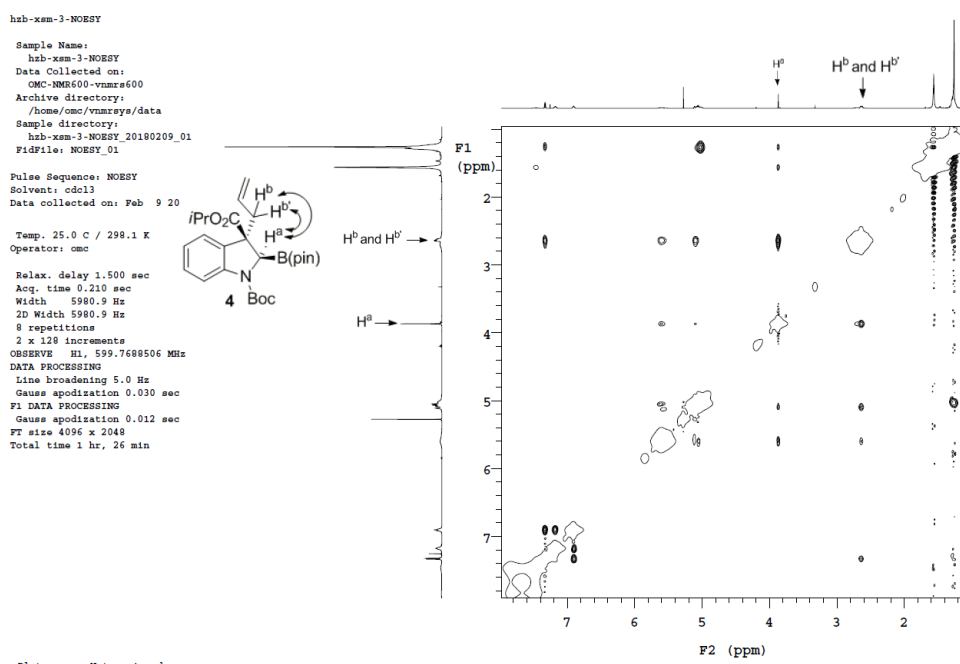


To a 25-mL flame-dried flask charged with 2-borylindoline **2c** (86.2 mg, 0.20 mmol) and THF (1 mL) was added LDA (150 μ L, 2M in hep/ethylbenzene, 0.3 mmol, 1.5 equiv) at -78 $^{\circ}$ C dropwisely. The resulting mixture was allowed at same temperature for 10 min. The Ethyl bromoacetate (35.6 μ L, 0.3 mmol, 1.5 equiv) was then added slowly at -78 $^{\circ}$ C. The reaction mixture was then allowed to warm to room temperature and continued to stir at same temperature for 10 min. After removal of the solvent, the residue was purified by column chromatography on silica gel using PE/EtOAc (12: 1) as the eluent to afford C3-alkylate 2-borylindoline **4d** as colorless oil (64.2mg, 62%). 96% *ee*, $[\alpha]_D^{25} = -32.0$ (*c* 0.32, CHCl_3). ^1H NMR (400 MHz, CDCl_3) δ 7.89-7.48 (m, 1H), 7.27-7.18 (m, 2H), 6.91-6.89 (m, 1H), 5.08-5.05 (m, 1H), 4.13 (q, *J* = 6.6 Hz, 2H), 3.97 (s, 1H), 3.06(bris, 1H), 2.83 (d, *J* = 12.8 Hz, 1H), 1.58 (s, 9H), 1.27-1.22 (m, 21H); ^{13}C NMR (100 MHz, CDCl_3) δ 171.9, 169.9, 152.2, 141.3, 131.8, 129.1, 123.9, 121.8, 115.0, 83.4, 81.4, 69.7, 60.9, 56.2, 54.7, 45.2, 28.4, 25.4,

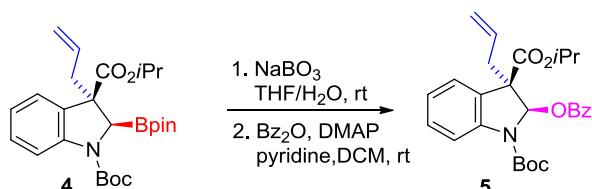
24.8, 21.7, 21.5, 14.0; HRMS (ESI) calcd for $C_{27}H_{41}BNO_8$ ($[M+H]^+$): 518.2920, found: 518.2927. The enantiopurity was determined by HPLC analysis (Daicel Chiralpak ODH column, Hexane/*i*PrOH = 90:10, flow rate = 1.0 mL/min., wavelength = 254 nm, t_R = 6.65 (minor), 13.54 (major).



12. NOESY spectrum of allylated 2-borylindoline 4



13. Oxidation of 2-borylindoline 4

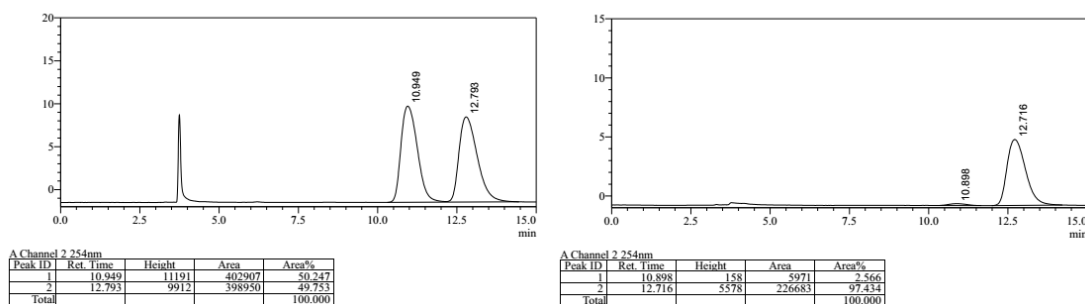


The oxidation of 2-borylindoline was adapted from literature procedures.⁷ To a 25-mL flask charged with 4 (95 mg, 0.20 mmol), THF (2 mL) and H₂O (2 mL) was added NaBO₃ 4H₂O (123.2 mg, 0.80 mmol, 4.0 equiv) at room temperature. The

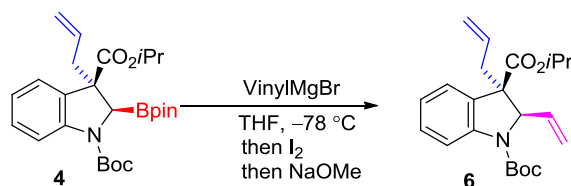
7. F. Meng, K. P. McGrath and A. H. Hoveyda *Nature* 2014, **513**, 367.

resulting mixture was allowed to stir at same temperature for 2 h. The reaction was then quenched with saturated Na₂S₂O₃ (5 mL) and diluted with water (20 mL). The biphasic solution was extracted with DCM 3 times (3 X 10 mL). The combined organic phase was then concentrated. The residue was dissolved in anhydrous DCM (5 mL) and cooled to 0 °C. Bz₂O (68.0 mg, 0.30 mmol) and DMAP (2.7 mg, 0.02 mmol) were then introduced followed by slow addition of pyridine (24 μL, 0.30 mmol). The resulting mixture was then continued to stir at room temperature for 2 hours. After removal of the solvent, the residue was purified by column chromatography on silica gel using PE/EtOAc (10:1) as the eluent to afford corresponding indoline **5** as colorless oil (55.8 mg, 60% yield).

95% *ee*, $[\alpha]_D^{25} = +82.7$ (*c* 0.38, CHCl₃). ¹H NMR (400 MHz, CDCl₃) δ 7.65-7.93 (m, 2H), 7.53-7.49 (m, 2H), 7.37-7.26 (m, 3H), 7.16-7.10 (m, 2H), 5.62-5.55 (m, 1H), 5.12-5.05 (m, 2H), 4.88-4.85 (m, 1H), 2.97-2.92 (m, 1H), 2.55-2.50 (m, 1H), 1.45 (s, 9H), 1.17 (d, *J* = 6.0 Hz, 3H), 0.92 (d, *J* = 5.6 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 168.8, 164.2, 151.1, 141.2, 134.5, 133.3, 131.2, 130.6, 129.8, 129.6, 128.9, 128.7, 128.4, 126.8, 122.8, 119.9, 114.7, 87.0, 82.5, 69.2, 59.7, 42.4, 28.2, 21.7, 21.3; HRMS (ESI) calcd for C₂₀H₂₆NO₄ ([M-OBz]⁺): 344.1862, found: 344.1849. The enantiopurity was determined by HPLC analysis (Daicel Chiralpak IE column, Hexane/*i*PrOH = 99:1, flow rate = 1.0 mL/min., wavelength = 254 nm, *t*_R = 10.90(minor), 12.72 (major))



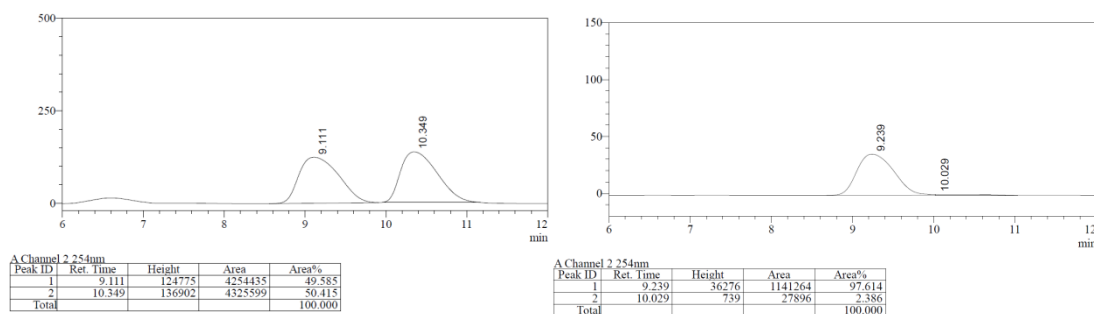
14. Stereospecific vinylation of 2-borylindoline **4**.



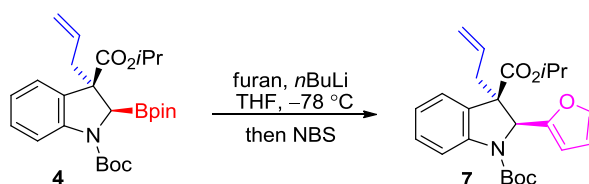
This reaction was adapted from the literature procedures.⁸ To a 25-mL flame-dried Schlenk tube charged with **4** (110 mg, 0.23 mmol) and THF (2 mL) was added vinylMgBr (0.35 mL, 1 M in THF, 0.35 mmol, 1.5 equiv) at room temperature. The resulting mixture was allowed to stir at same temperature for 0.5 h. Methanolic

8. R. P. Sonawane, V. Jheengut, C. Rabalakos, R. Larouche-Gauthier, H. K. Scott and V. K. Aggarwal, *Angew. Chem., Int. Ed.* **2011**, 50, 3760.

solution of I₂ (292 mg, 1.15 mmol, 3 mL MeOH) was then introduced slowly to the reaction mixture at -78 °C. The reaction was then allowed to stir at this temperature for additional 0.5 h. Methanolic solution of NaOMe (62.1 mg, 1.15 mmol, 3 mL MeOH) was then added slowly at -78 °C. The resulting mixture was then warmed to room temperature and continued to stir at this temperature for 1 h. Saturated aqueous Na₂S₂O₃ (5 mL) was then added to quench the reaction. After dilution with H₂O (20 mL), the mixture was extracted with EtOAc 3 times (3 X 20 mL). The combined organic phase was dried over anhydrous Na₂SO₄. After removal of the solvent, the residue was purified by column chromatography on silica gel using PE/EtOAc (20:1) as the eluent to afford corresponding 2-vinylindoline 6 as colorless oil (83.5 mg, 98% yield). 95% *ee*, $[\alpha]_D^{25} = +99.4$ (*c* 0.4, CHCl₃). ¹H NMR (400 MHz, CDCl₃) 7.84 (brs, 1H), 7.30-7.24 (m, 2H), 7.01-6.98 (m, 1H), 5.73-5.64 (m, 1H), 5.57-5.48 (m, 1H), 5.25-5.21 (m, 1H), 5.14-5.11 (m, 1H), 5.05-5.03 (m, 2H), 4.59 (brs, 1H), 2.82-2.79 (m, 1H), 2.53-2.48 (m, 1H), 1.53 (s, 9H), 1.26-1.22 (m, 6H); ¹³C NMR (100 MHz, CDCl₃) δ 170.4, 151.8, 141.0, 133.0, 132.2, 128.4, 126.8, 122.2, 119.3, 118.2, 115.0, 82.3, 70.5, 68.6, 59.1, 43.9, 28.3, 21.8, 21.7 (one *sp*² carbon signal does not show.) HRMS (ESI) calcd for C₂₂H₃₃N₂O₄ ([M+NH₄]⁺): 389.2435, found: 389.2429. The enantiopurity was determined by HPLC analysis (Daicel Chiralpak IE column, Hexane/*i*PrOH = 99:1, flow rate = 1.0 mL/min., wavelength = 254 nm, *t*_R = 9.23 (major), 10.03 (minor))



15. Stereospecific arylation of 2-borylindoline 4.

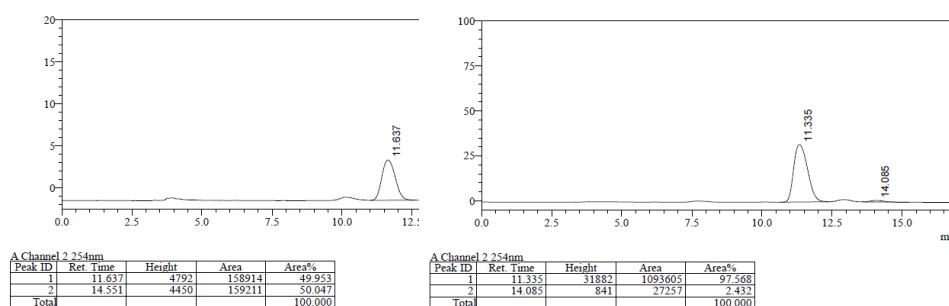


This reaction was adapted by the literature procedures.⁹ To a 25-mL Schlenk tube charged with 4 (94 mg, 0.20 mmol) and THF (2 mL) was added freshly prepared fur-2-yl lithium (0.20 mL, 1.0 M in THF/hexanes, 0.20 mmol) slowly at -78 °C. The resulting mixture was then allowed to stir at this temperature for 1.5 h. NBS (36 mg, 0.20 mmol) in THF (2 mL) was then added slowly to the reaction mixture at -78 °C. The reaction was continued to stir at -78 °C for 0.5 h. Saturated aqueous Na₂S₂O₃ (2.5

9. A. Bonet, M. Odachowski, D. Leonori, S. Essafi and V. K. Aggarwal, *Nat. Chem.* 2014, **6**, 584.

mL) was then introduced to quench the reaction at room temperature. The organic phase was separated and the rest was extracted with EtOAc 3 times (3 X 10 mL). The combined organic phase was then dried over Na₂SO₄. After removal of the solvent, the residue was purified by column chromatography on silica gel using PE/EtOAc (20:1) as the eluent to afford corresponding 2-(2-furyl)-indoline **7** as colorless oil (32.9 mg, 40% yield).

Colorless oil, 32.9 mg, 40% yield, 95% *ee*, $[\alpha]_D^{25} = +72.8$ (*c* 0.2, CHCl₃). ¹H NMR (400 MHz, CDCl₃) δ 7.90 (brs, 1H), 7.41-7.45 (m, 1H), 7.23-7.21 (m, 1H), 7.06-7.03 (m, 1H), 6.21 (s, 1H), 6.03 (brs, 1H), 5.62-5.52(m, 1H), 5.25 (s, 1H), 5.09-5.02(m, 2H), 4.83-4.78 (m, 1H), 2.86-2.81 (m, 1H), 2.64-2.58 (m, 1H), 1.41 (s, 9H), 1.09 (d, *J* = 6.0 Hz, 3H), 0.95 (d, *J* = 6.0 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 170.1, 152.8, 151.6, 141.1, 132.0, 128.5, 126.8, 122.3, 119.5, 114.7, 110.3, 106.9, 81.1, 68.5, 65.1, 45.1, 29.3, 28.2, 21.6, 21.3; HRMS (ESI) calcd for C₂₄H₃₃N₂O₅ ([M+NH₄]⁺): 429.2384, found: 429.2380. The enantiopurity was determined by HPLC analysis (Daicel Chiralpak IE column, Hexane/*i*PrOH = 99:1, flow rate = 1.0 mL/min., wavelength = 254 nm, *t*_R = 11.34 (major), 14.09 (minor).



16. Crystallographic data of compound **2s**

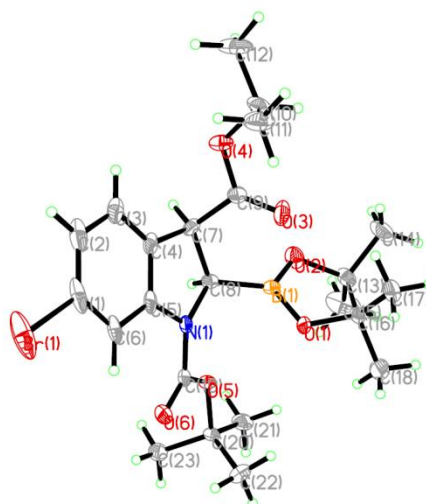


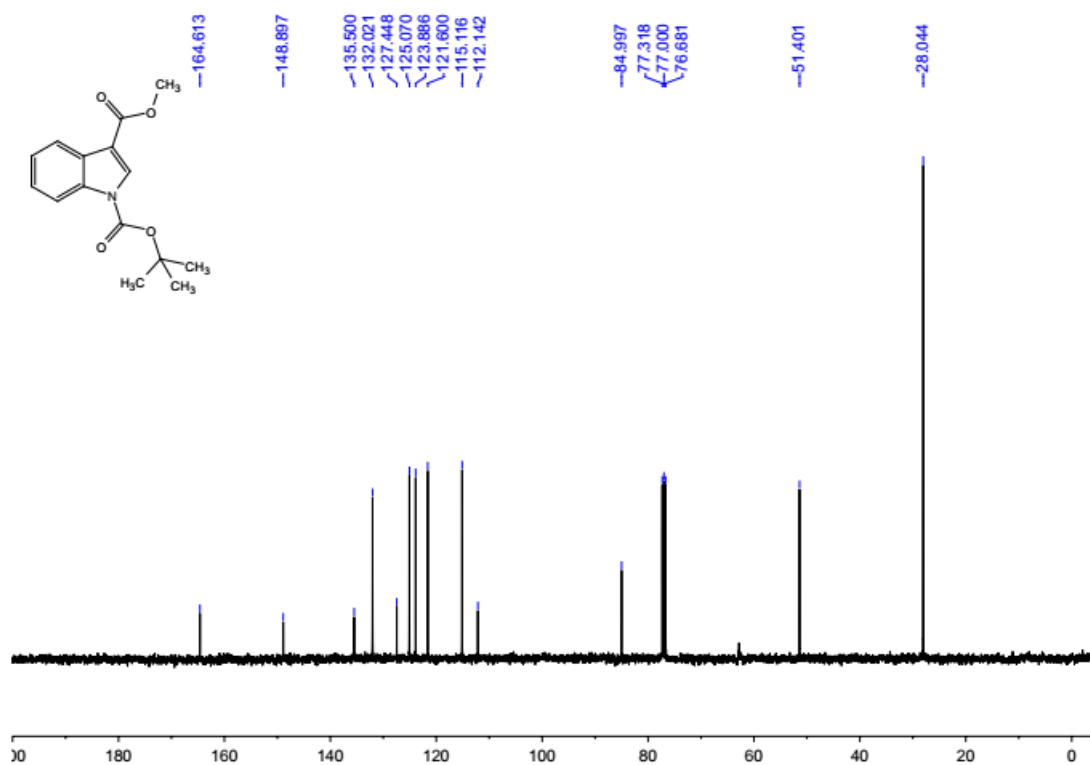
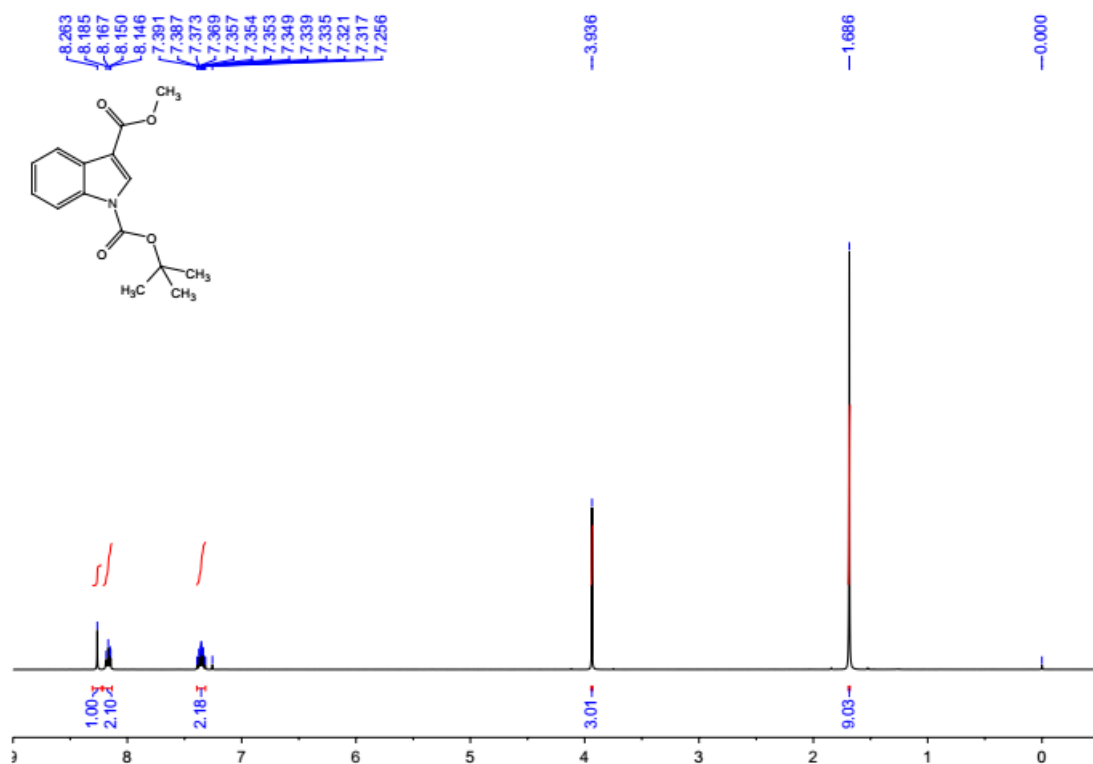
Table S2. Crystal data and structure refinement for **2s**.

Identification code	1
Empirical formula	C ₂₃ H ₃₃ B Br N O ₆

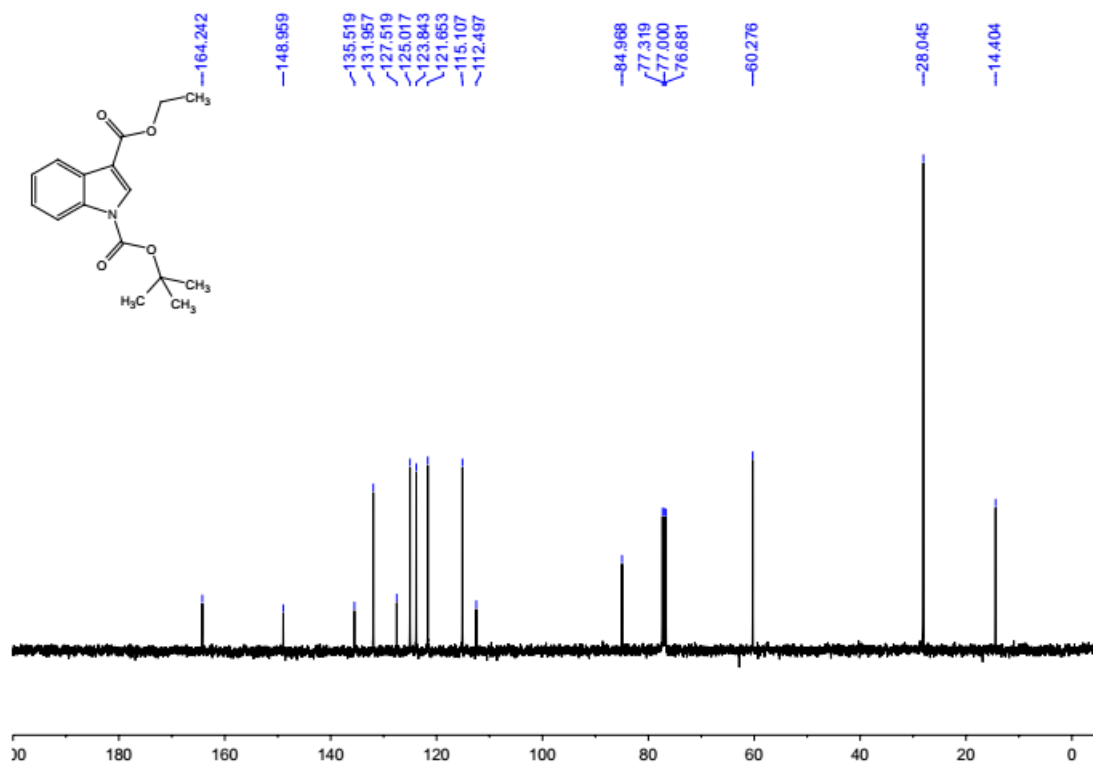
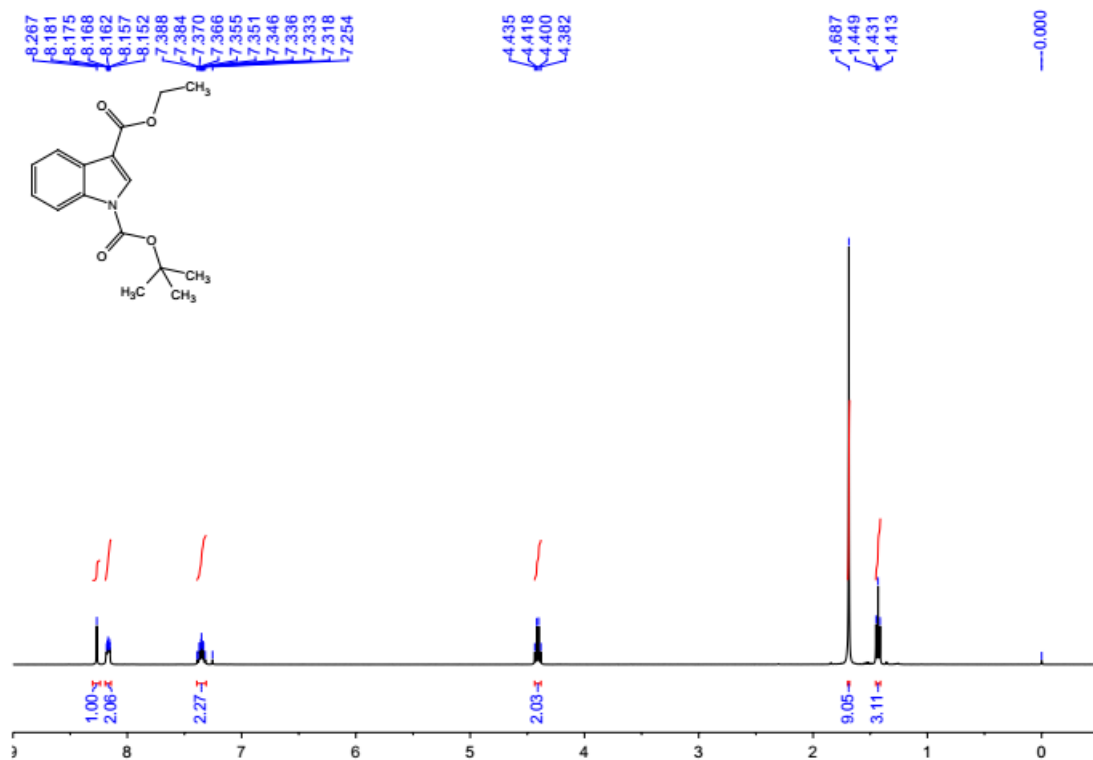
Formula weight	510.22	
Temperature	153(2) K	
Wavelength	0.71073 Å	
Crystal system	Orthorhombic	
Space group	P2(1)2(1)2(1)	
Unit cell dimensions	a = 10.0308(12) Å	$\alpha = 90^\circ$
	b = 12.6903(16) Å	$\beta = 90^\circ$
	c = 19.943(2) Å	$\gamma = 90^\circ$
Volume	2538.6(5) Å ³	
Z	4	
Density (calculated)	1.335 Mg/m ³	
Absorption coefficient	1.655 mm ⁻¹	
F(000)	1064	
Crystal size	0.30 x 0.20 x 0.20 mm ³	
Theta range for data collection	1.90 to 25.00 °	
Index ranges	-11<=h<=11, -15<=k<=11, -23<=l<=22	
Reflections collected	12877	
Independent reflections	4452 [R(int) = 0.0382]	
Completeness to theta = 25.00°	100.0 %	
Absorption correction	Semi-empirical from equivalents	
Max. and min. transmission	0.7331 and 0.6365	
Refinement method	Full-matrix least-squares on F ²	
Data / restraints / parameters	4452 / 0 / 298	
Goodness-of-fit on F ²	1.001	
Final R indices [I>2sigma(I)]	R1 = 0.0408, wR2 = 0.1040	
R indices (all data)	R1 = 0.0506, wR2 = 0.1091	
Absolute structure parameter	0.029(10)	
Largest diff. peak and hole	0.584 and -0.841 e.Å ⁻³	

17. NMR spectra of all new compounds

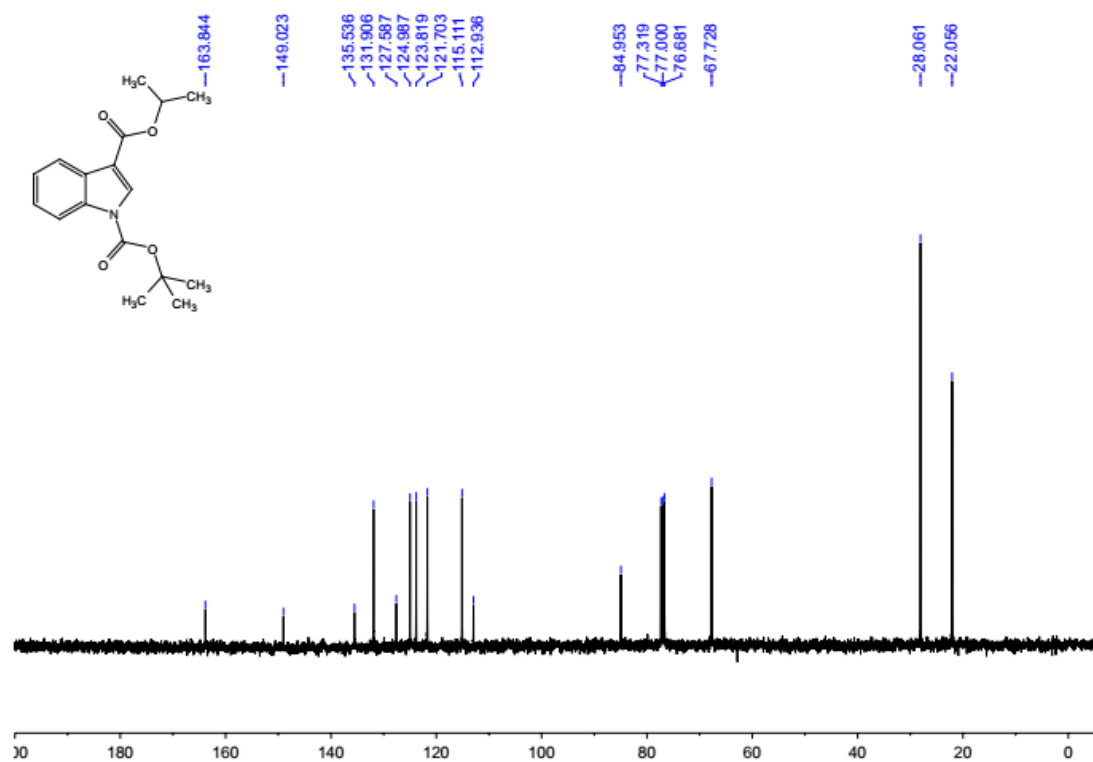
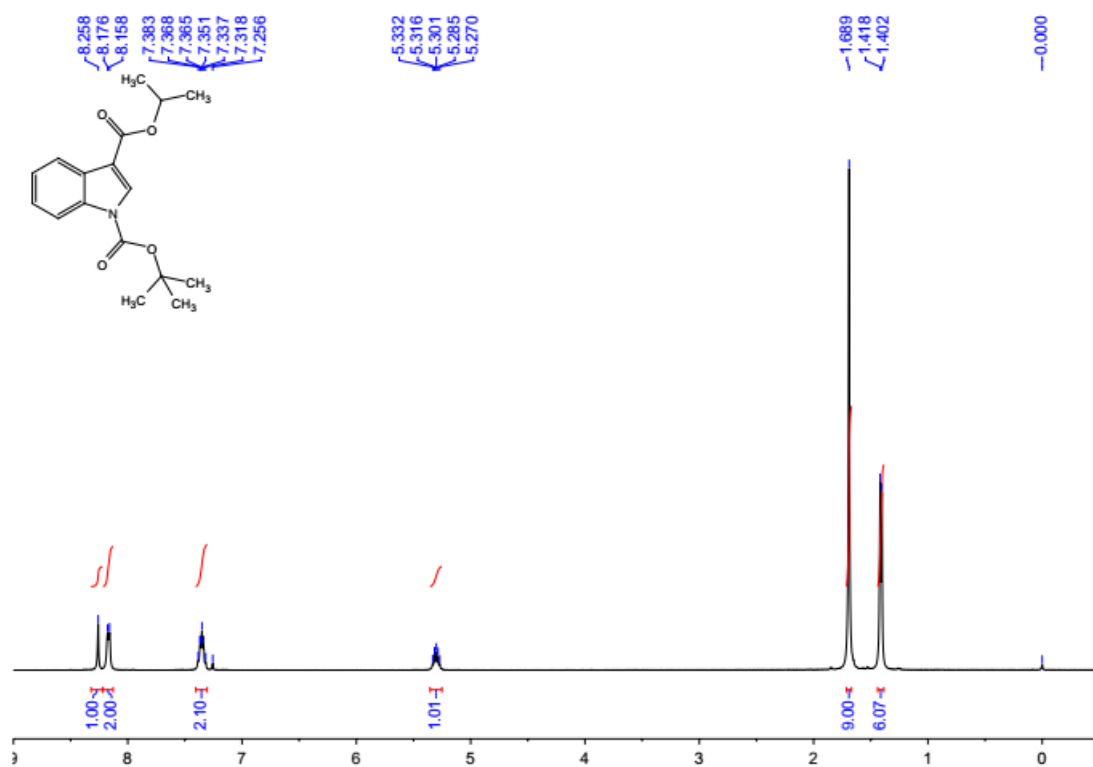
Compound 1a



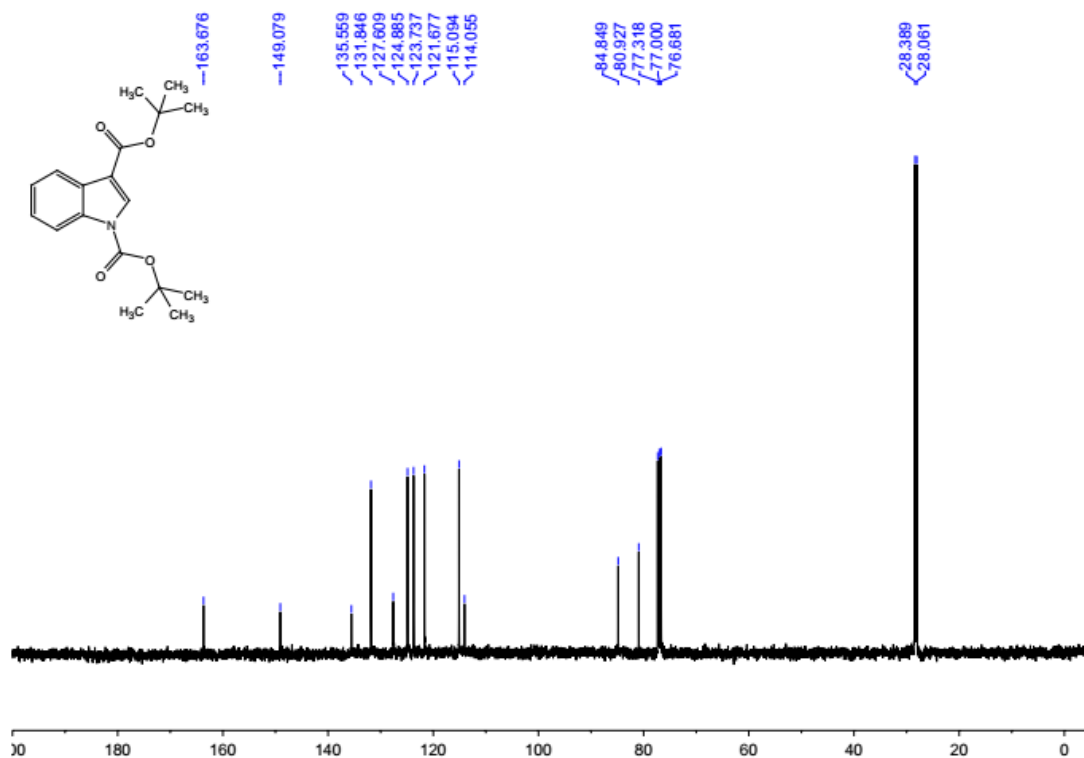
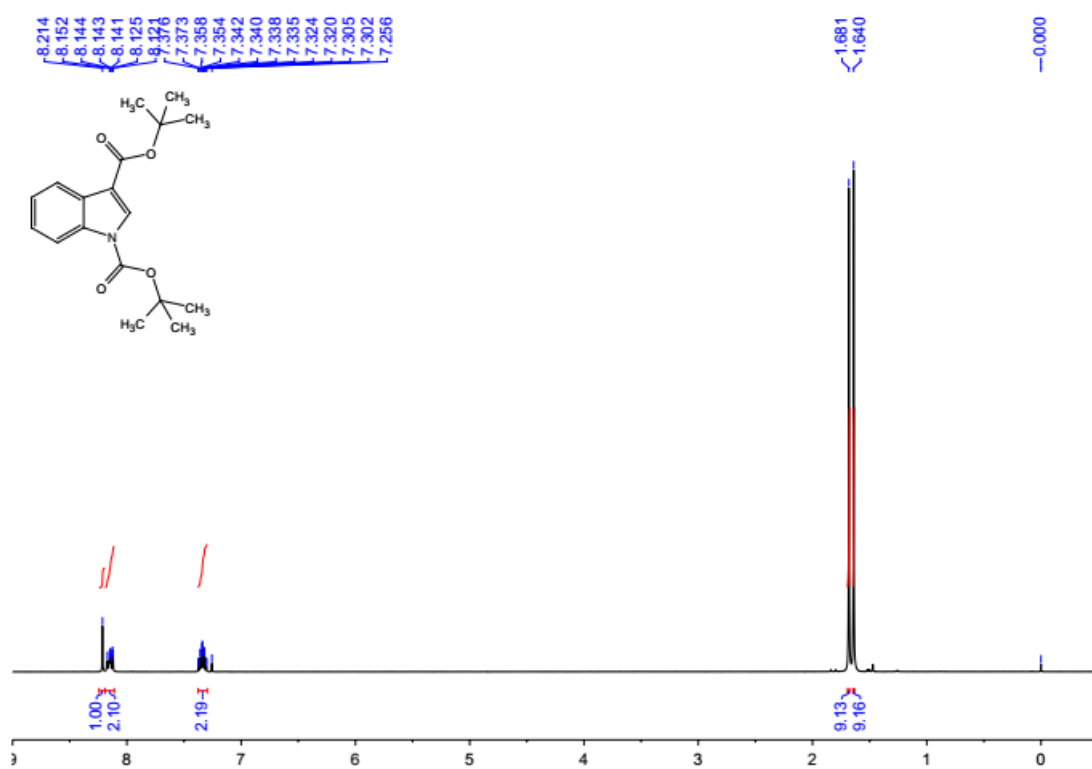
Compound 1b



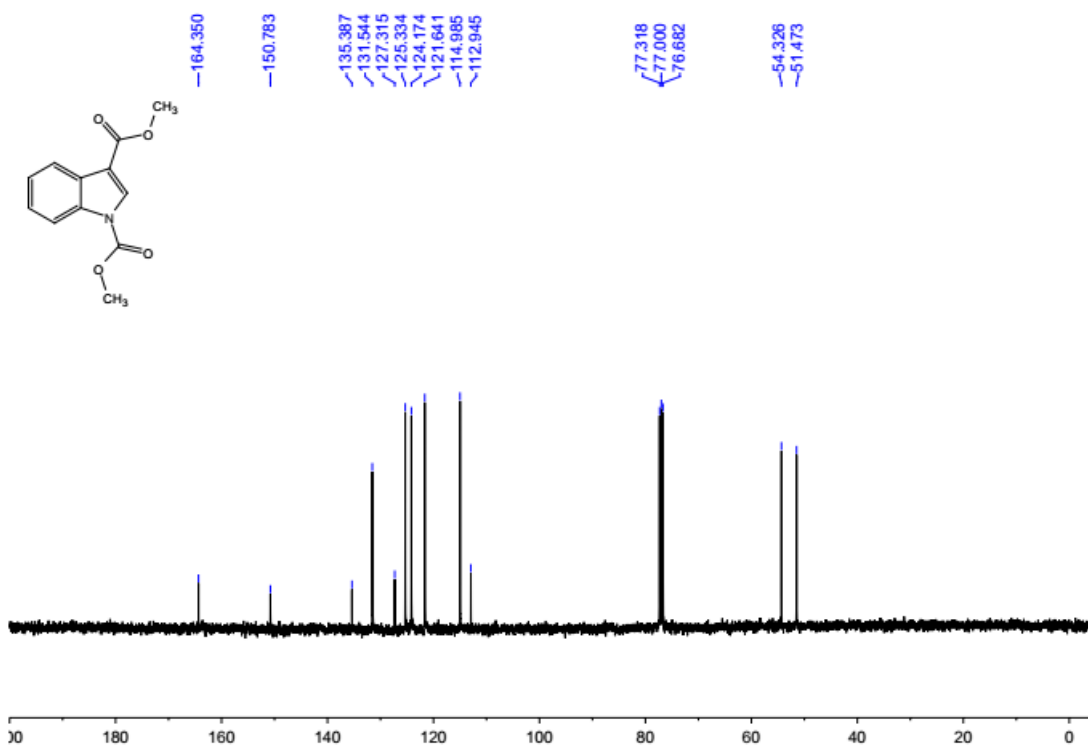
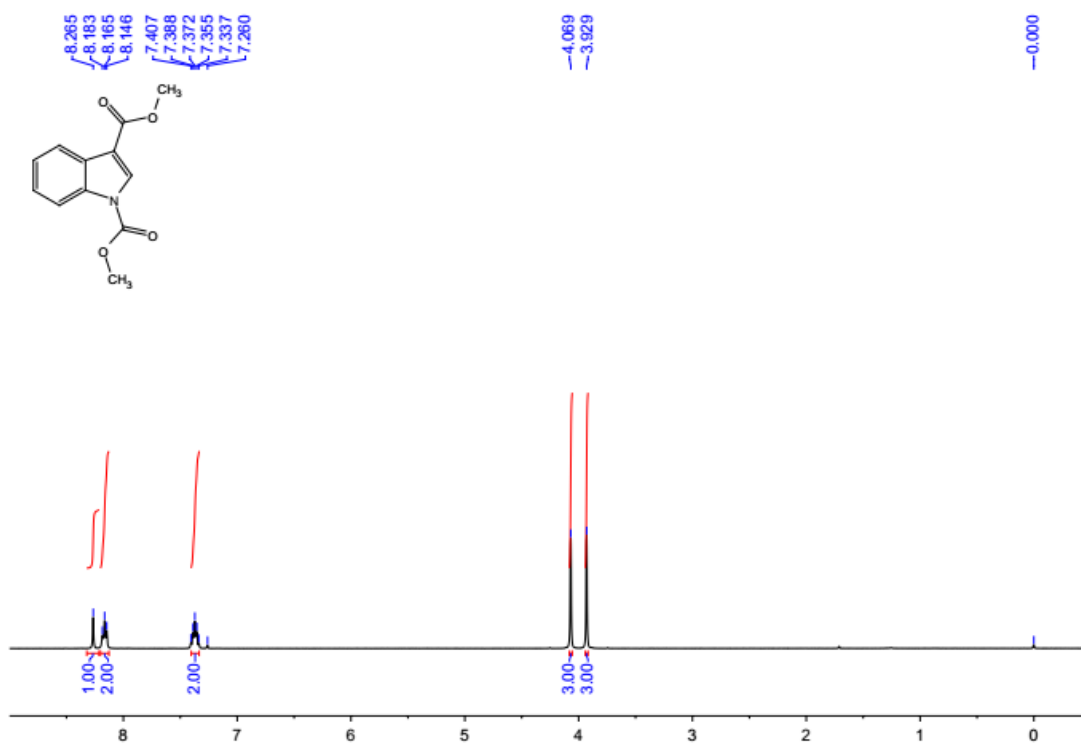
Compound 1c



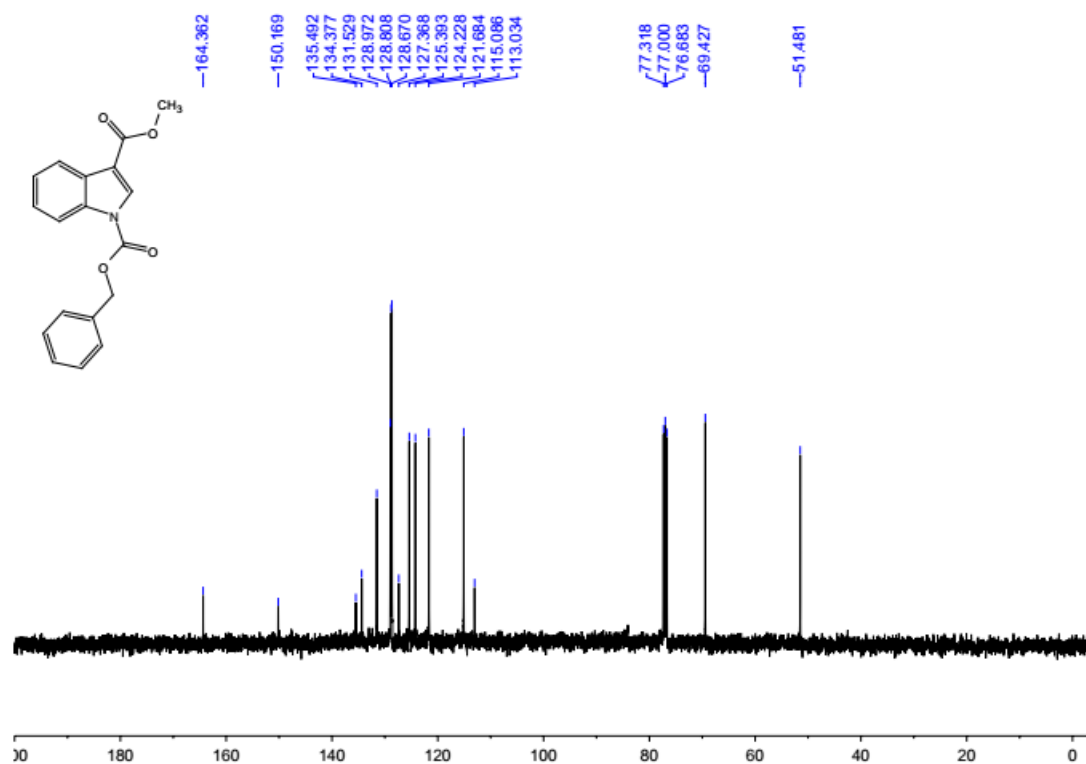
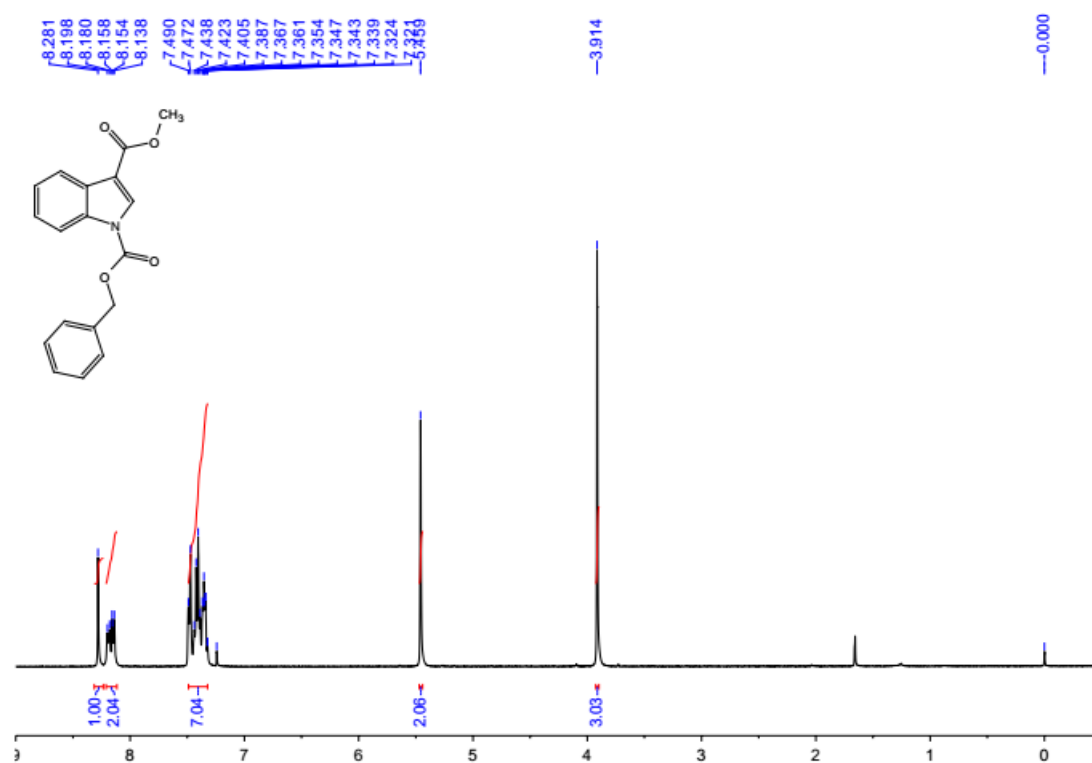
Compound 1d



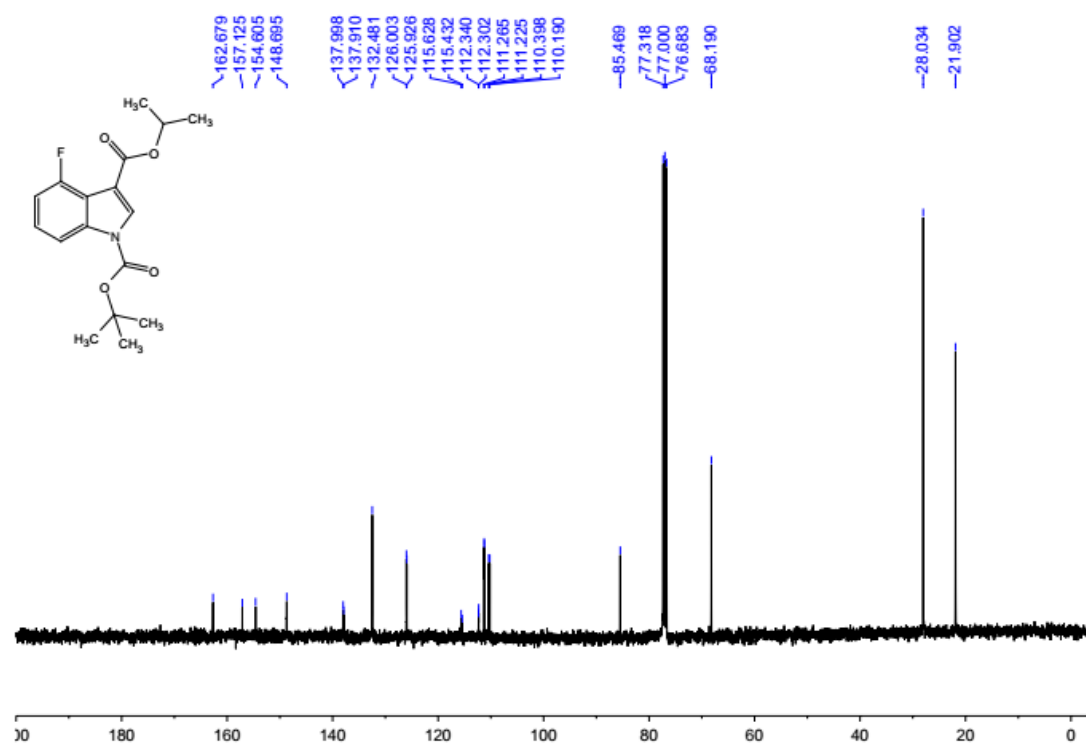
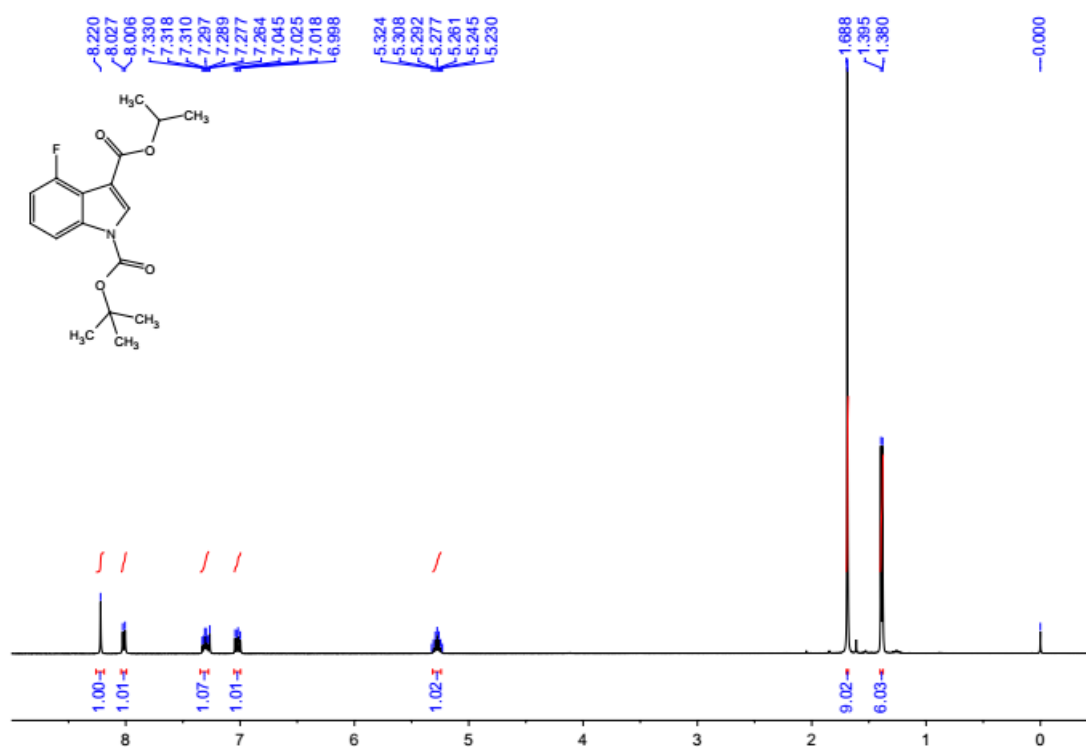
Compound 1e



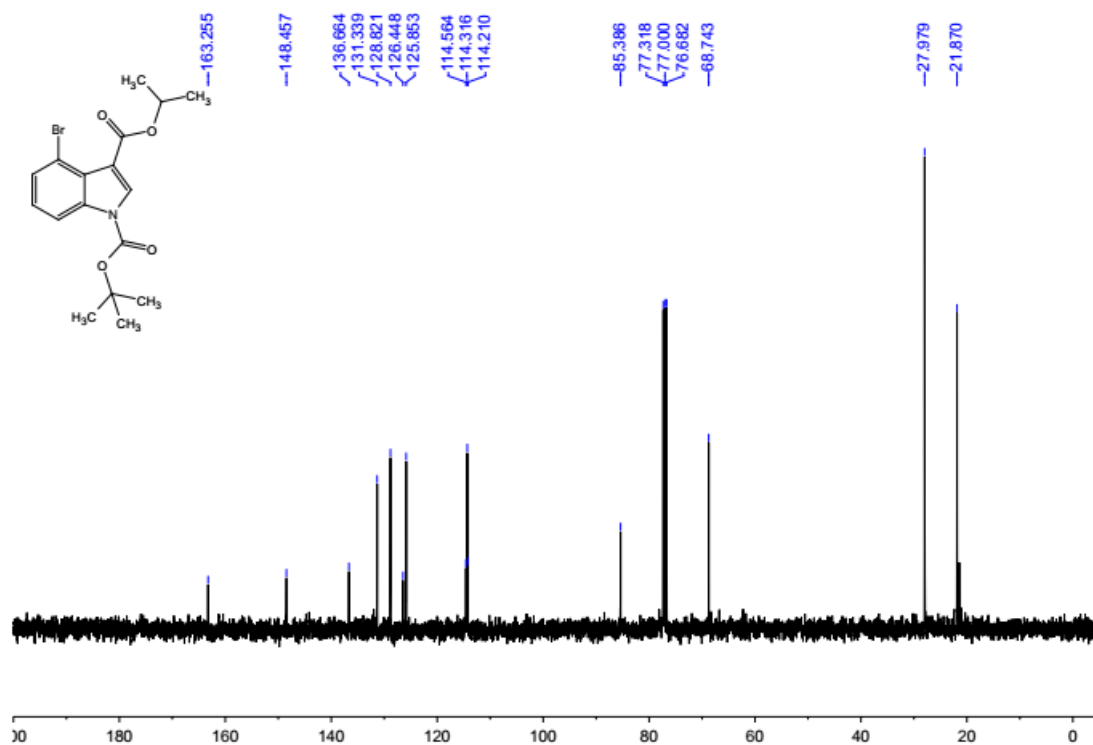
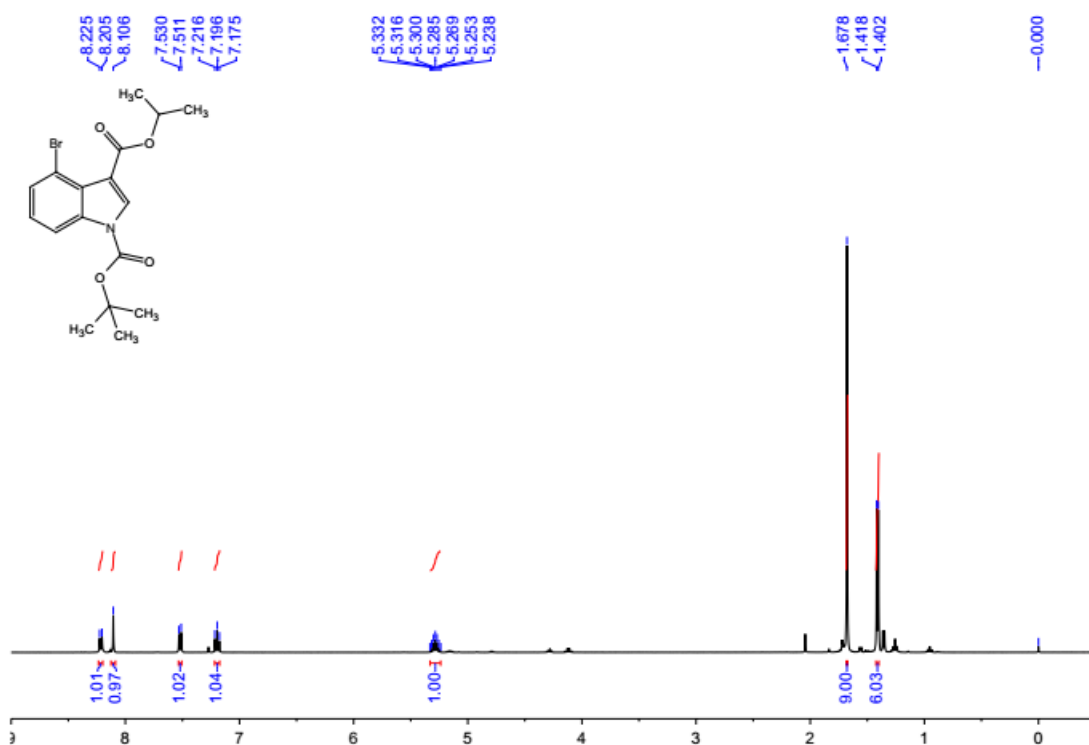
Compound 1f



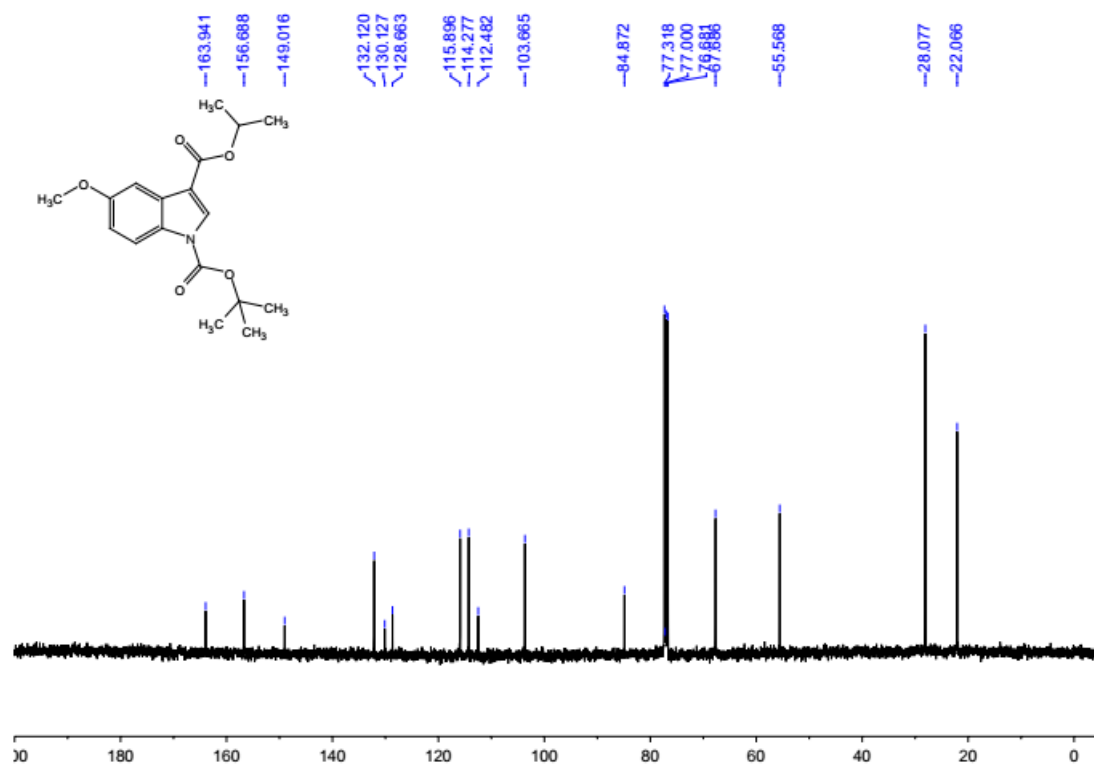
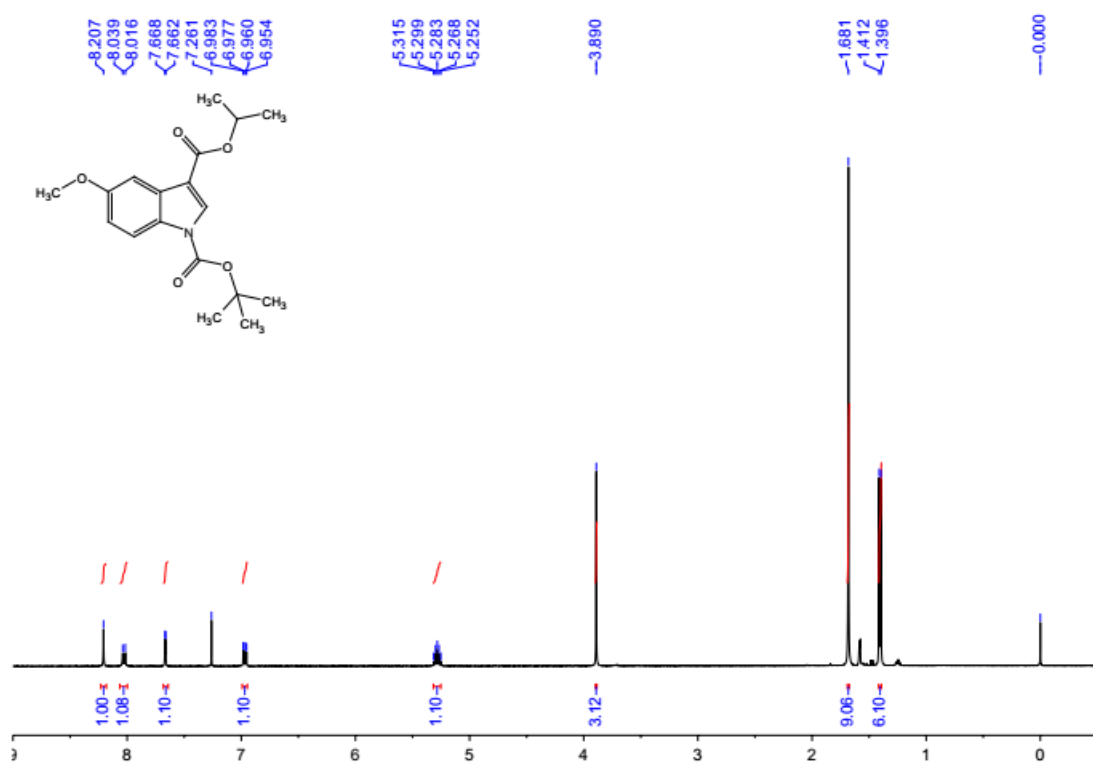
Compound 1g



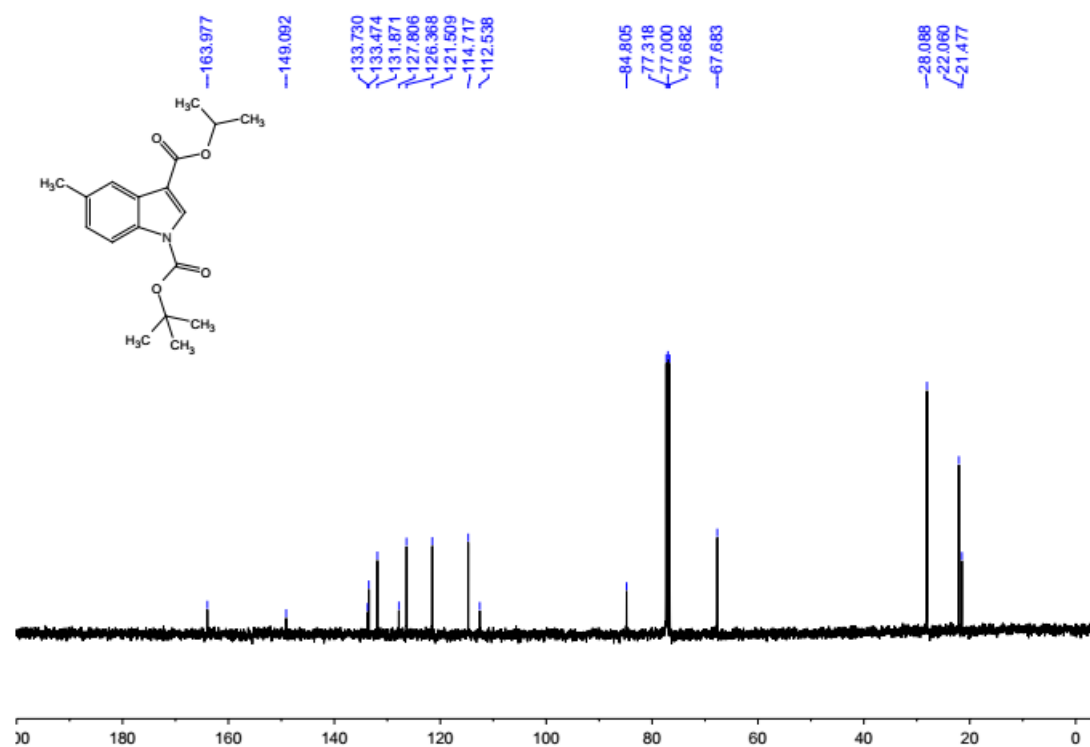
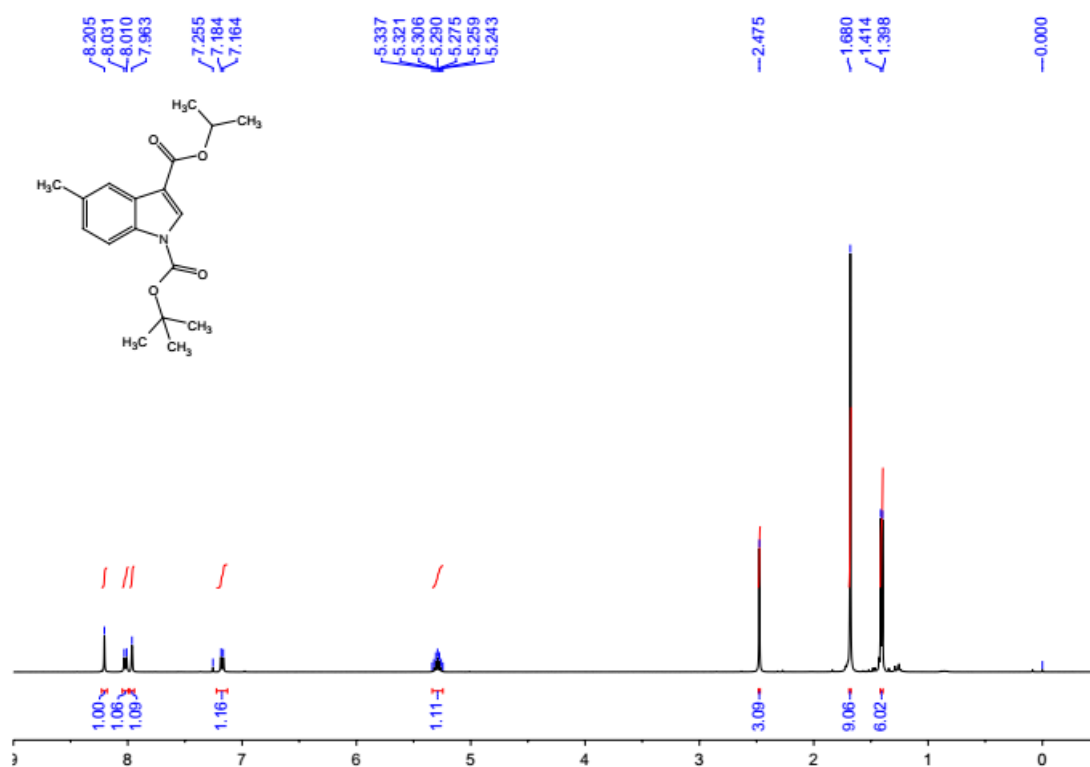
Compound 1h



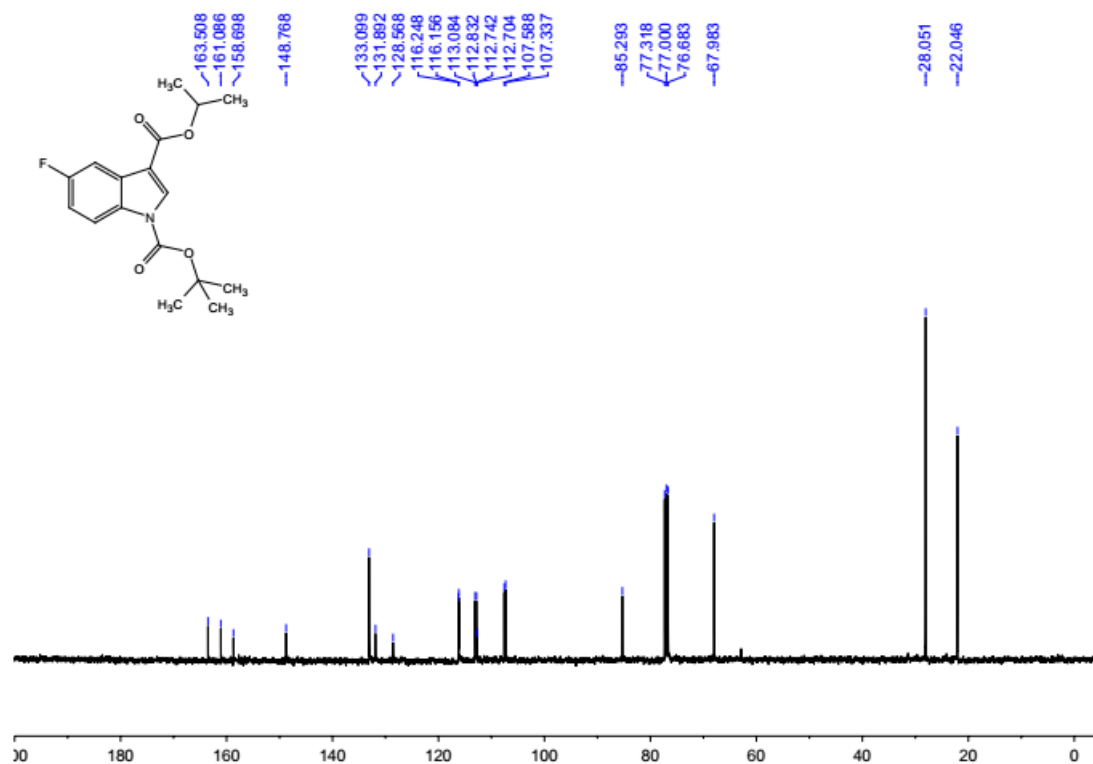
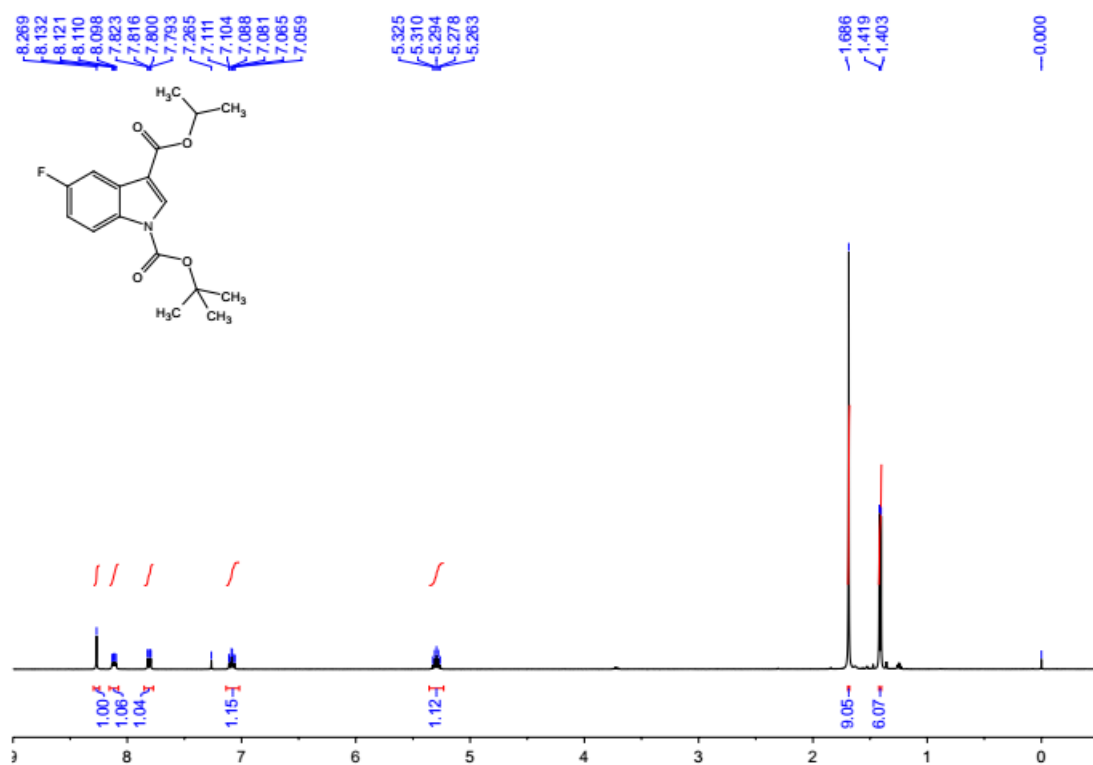
Compound 1i



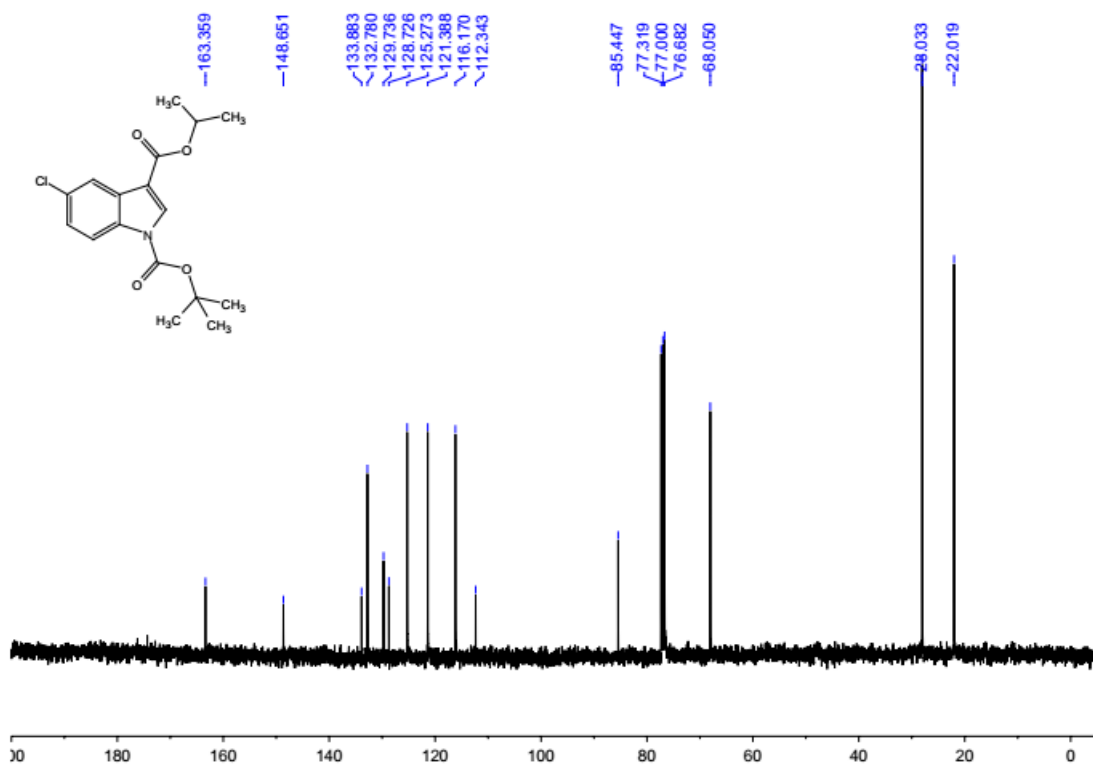
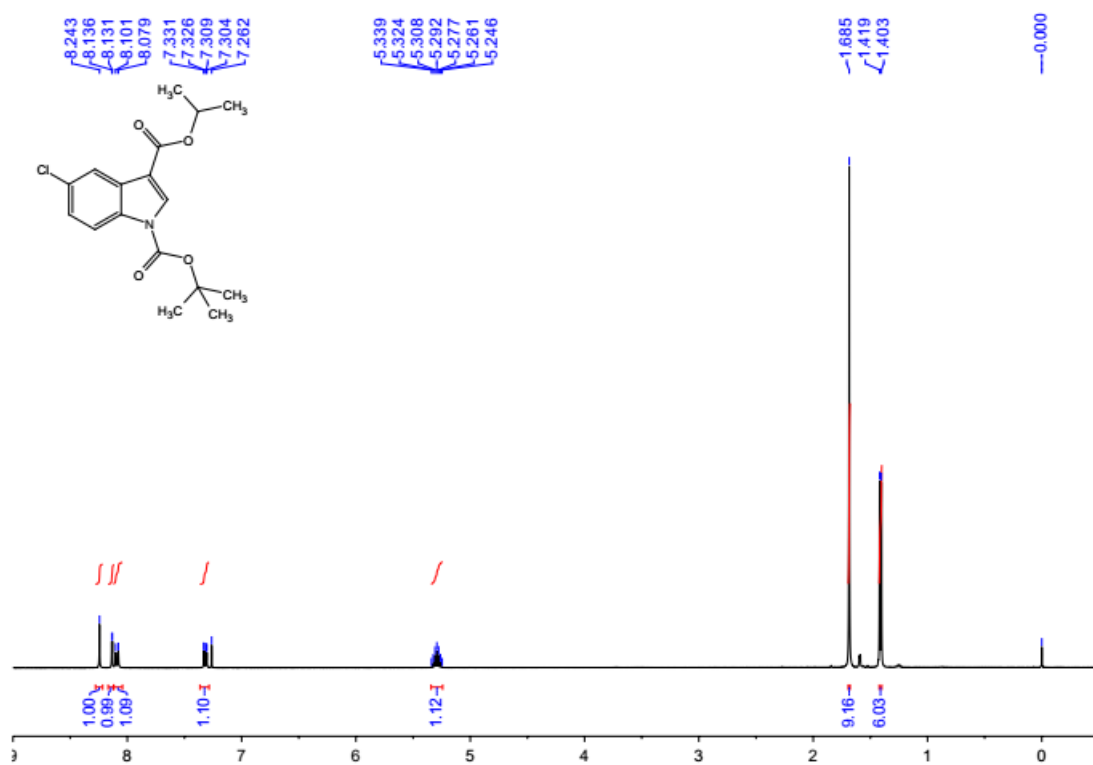
Compound 1j



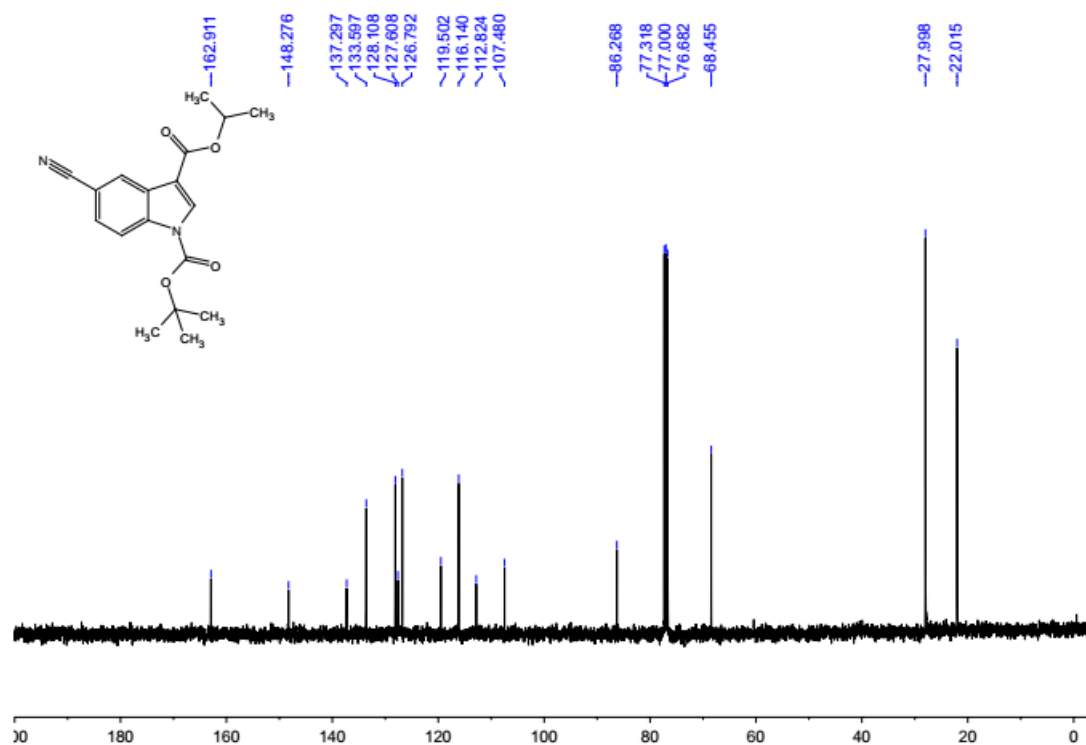
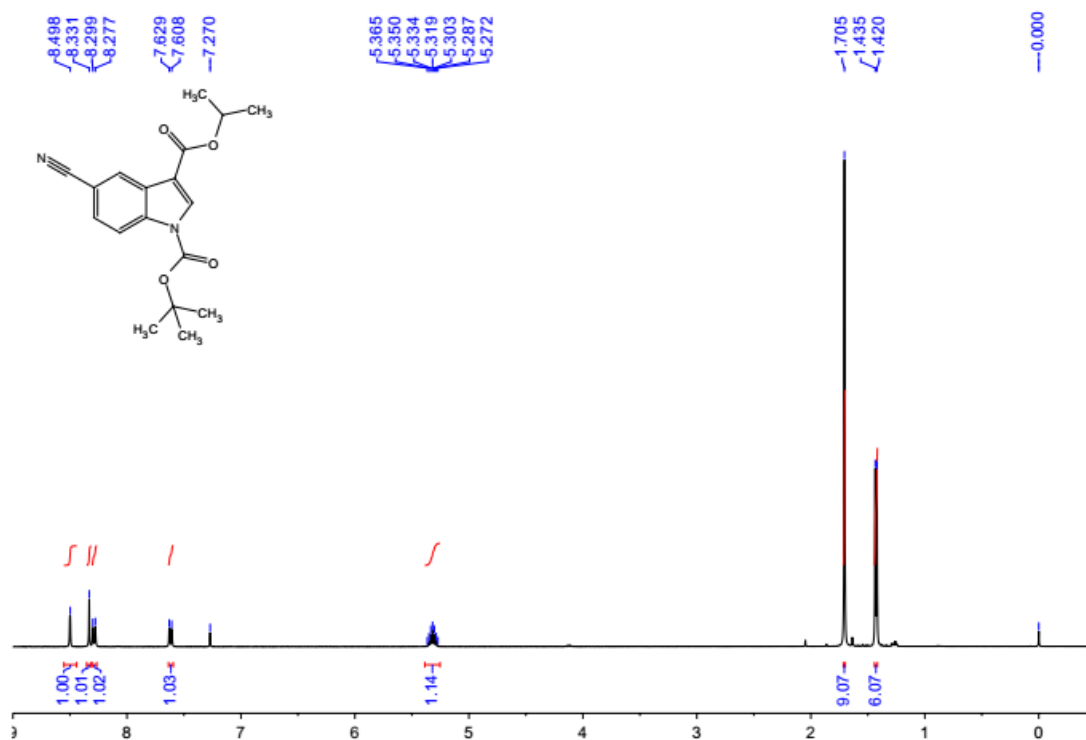
Compound 1k



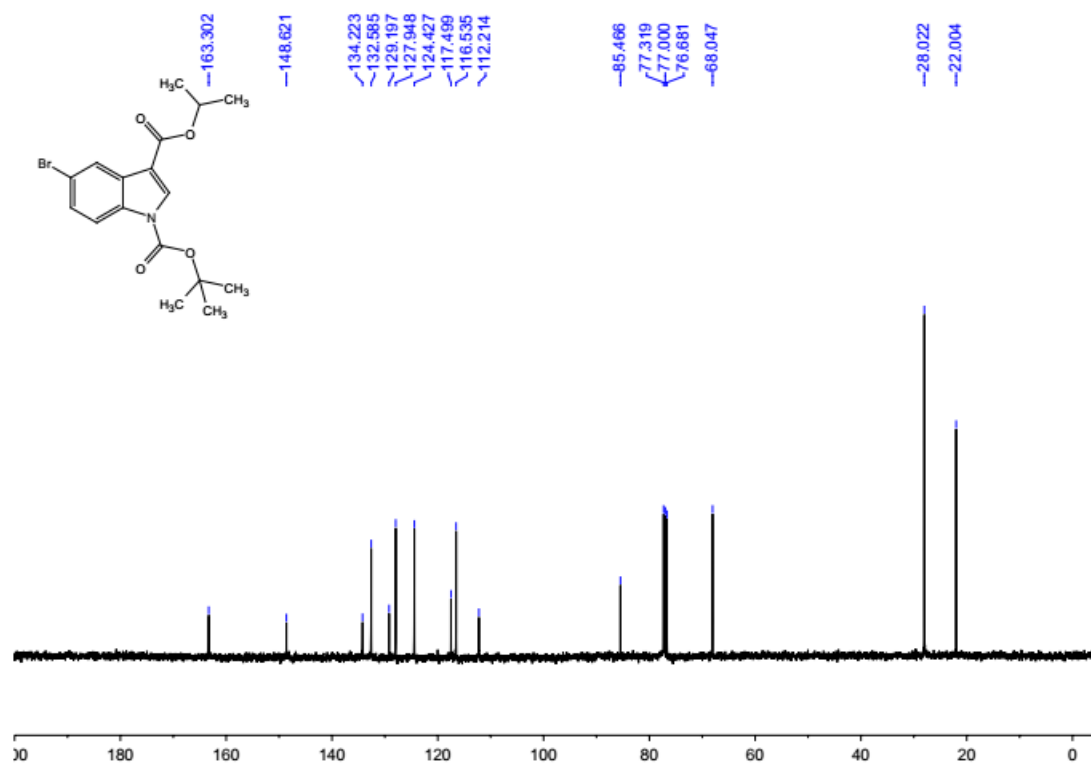
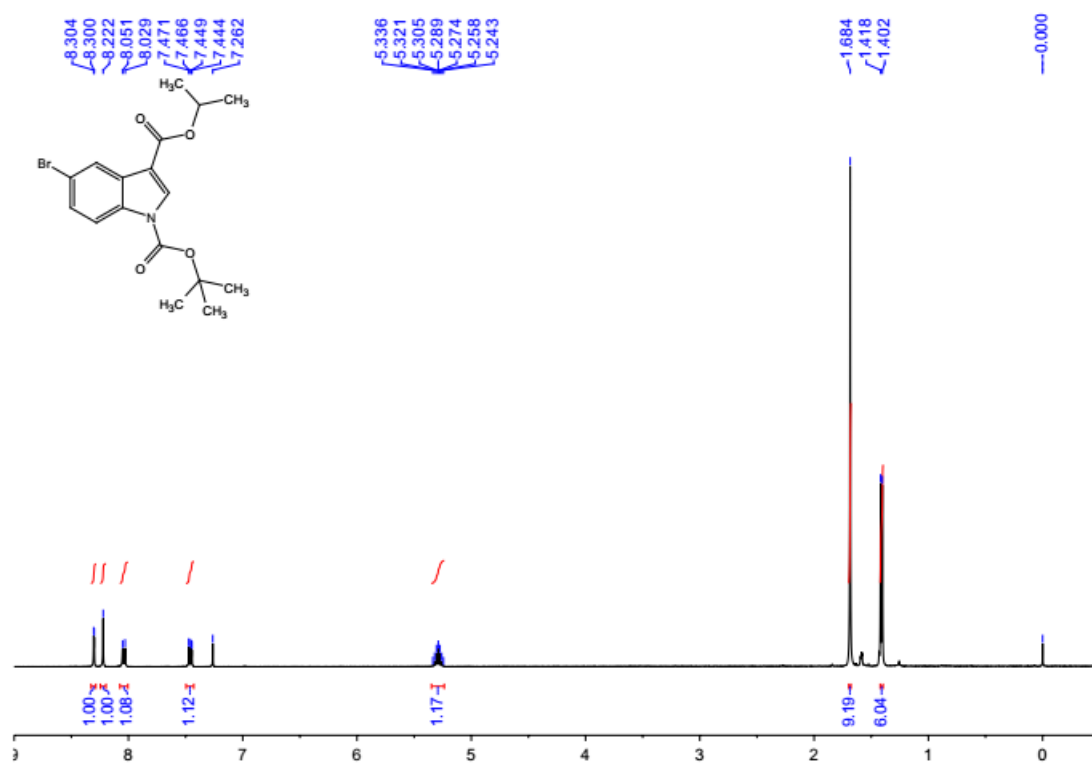
Compound 11



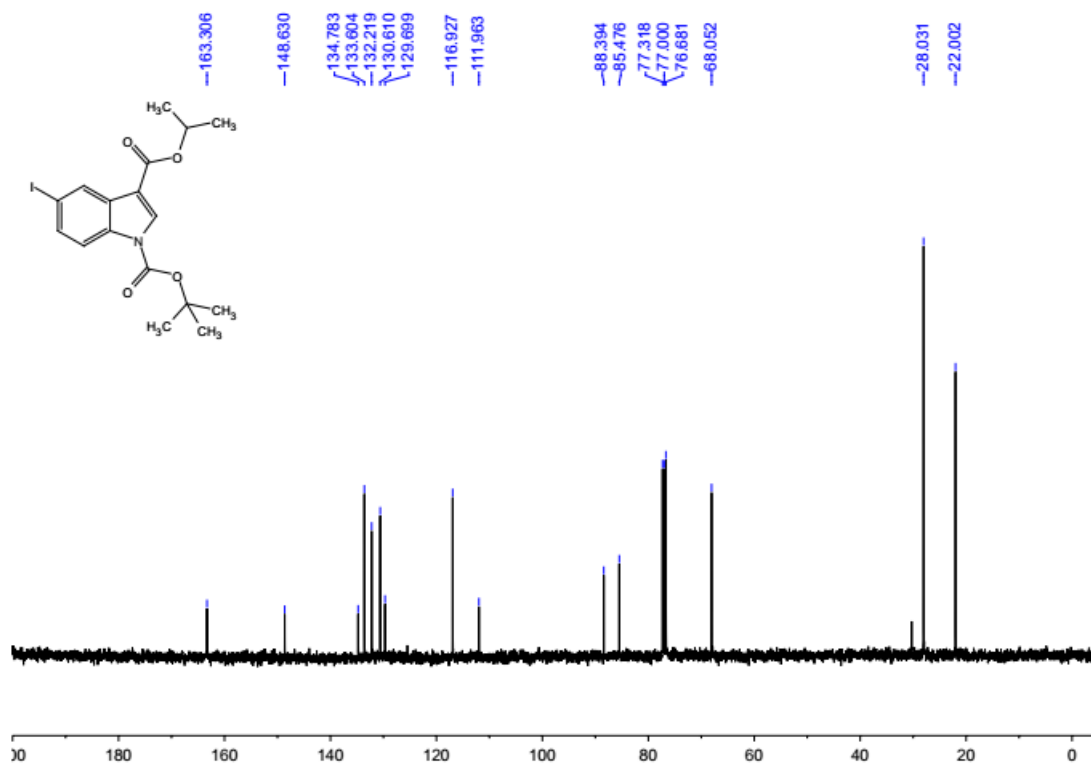
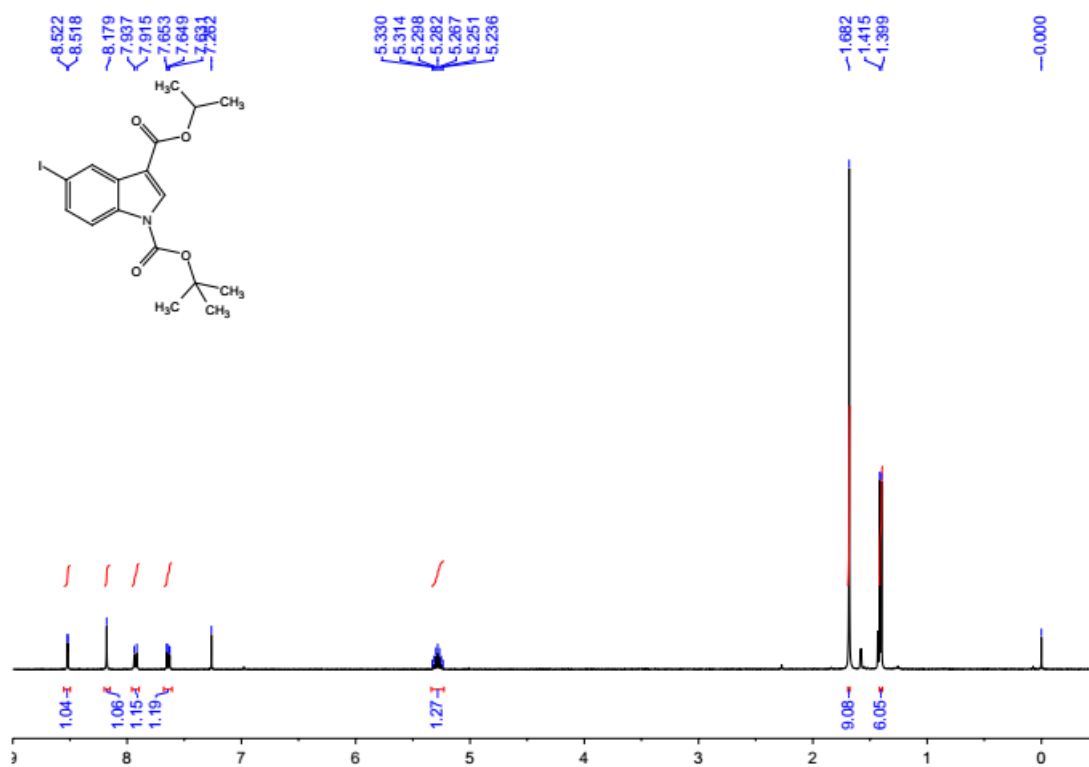
Compound 1m



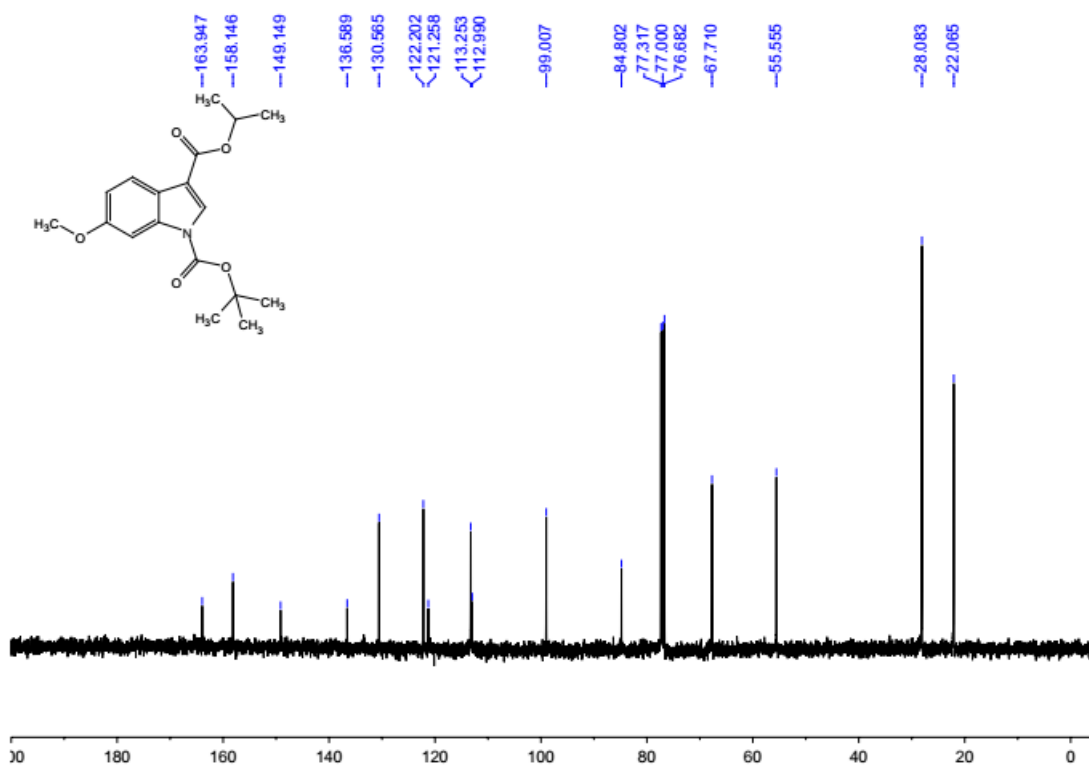
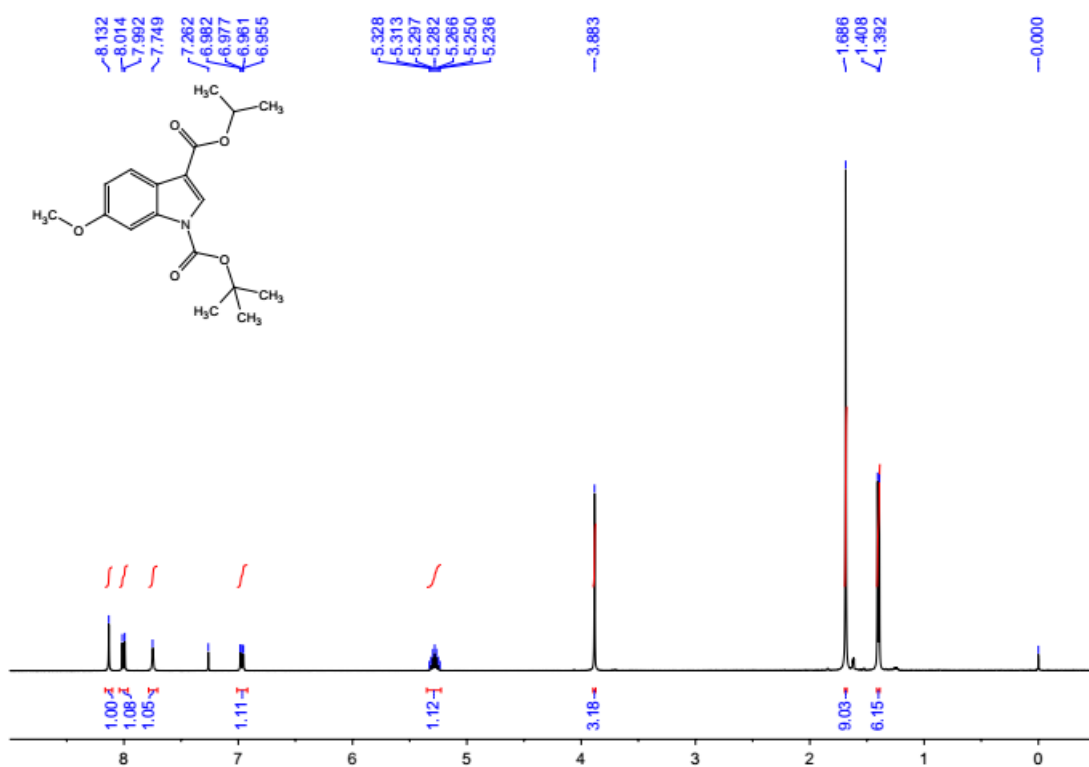
Compound 1n



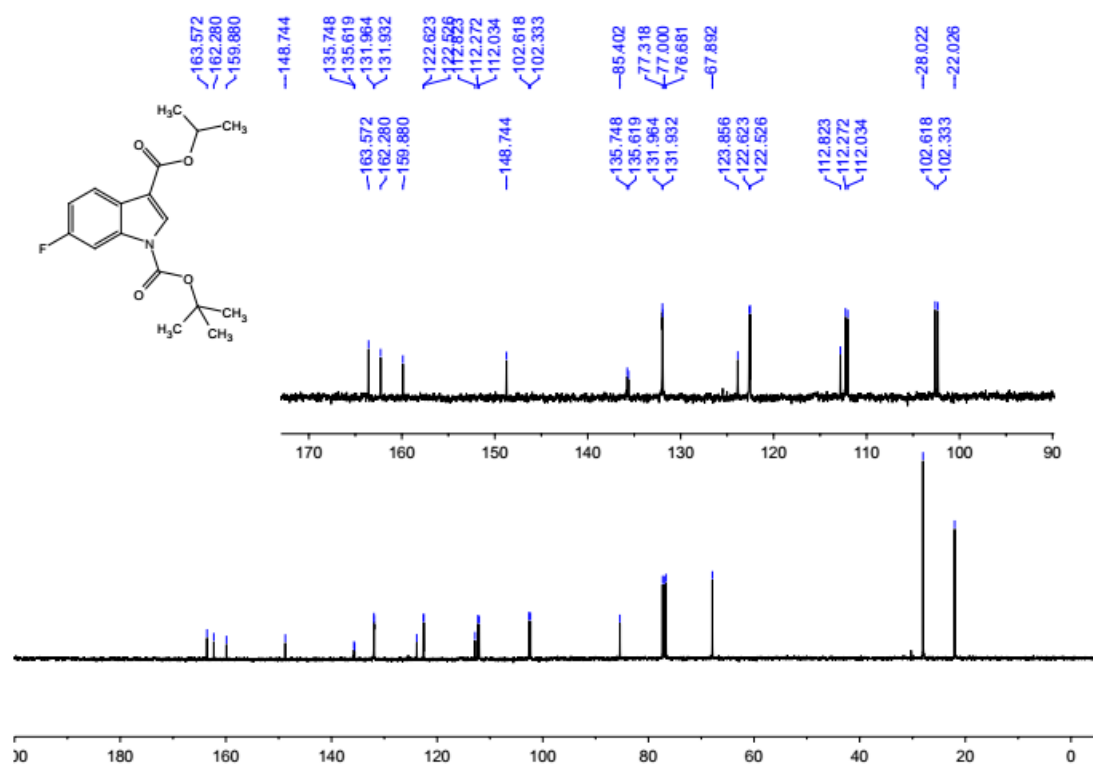
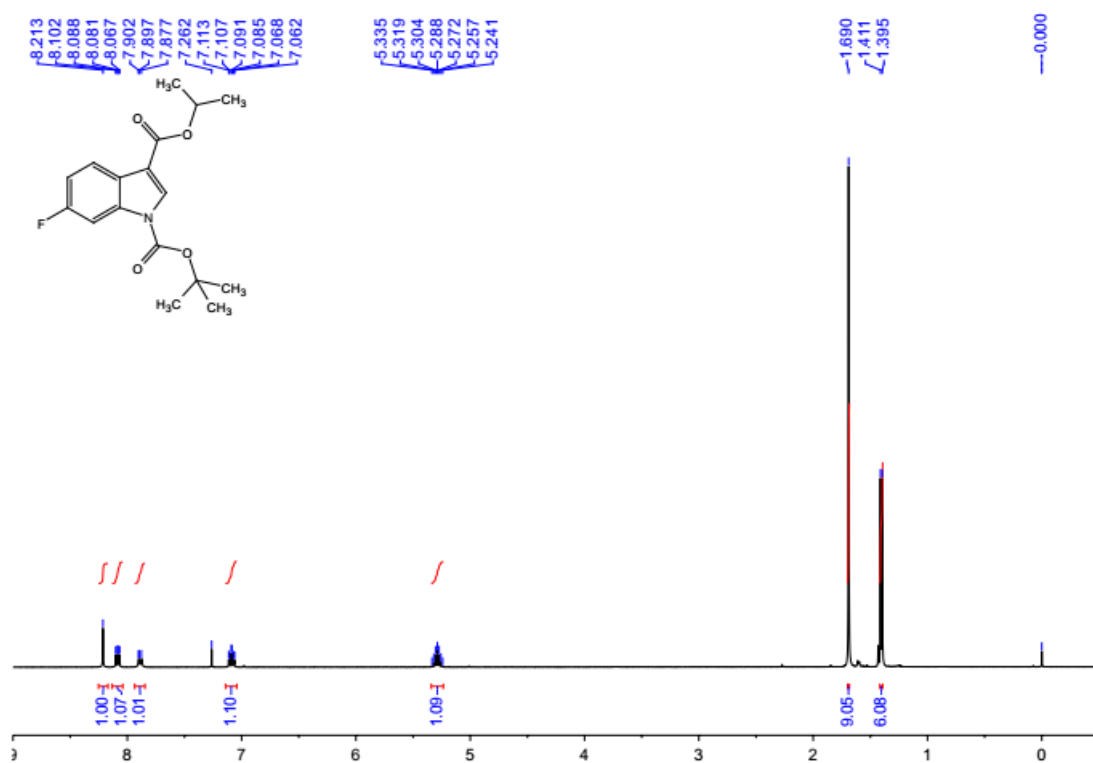
Compound 1o



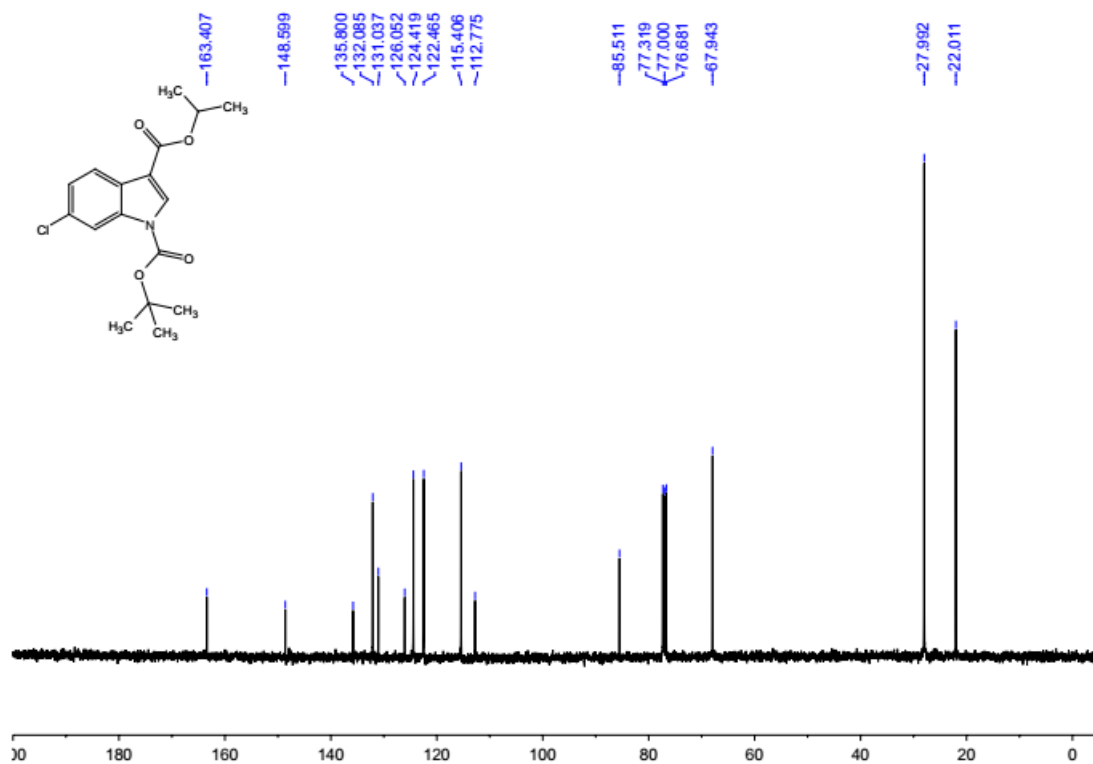
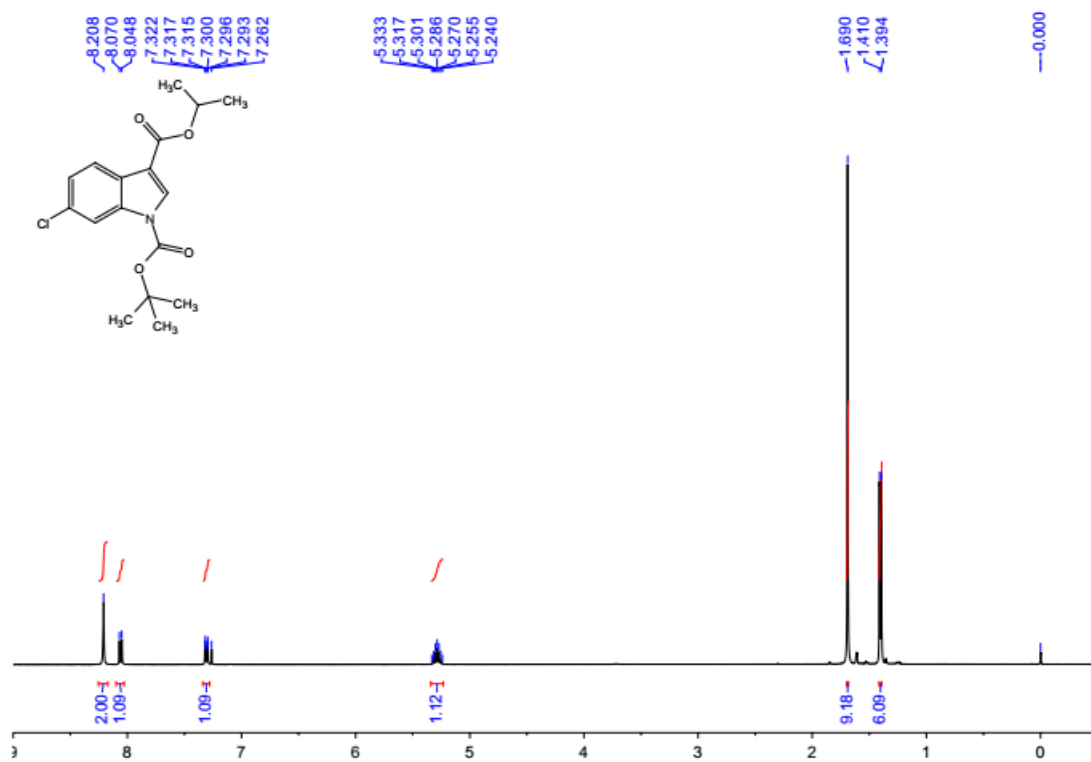
Compound 1p



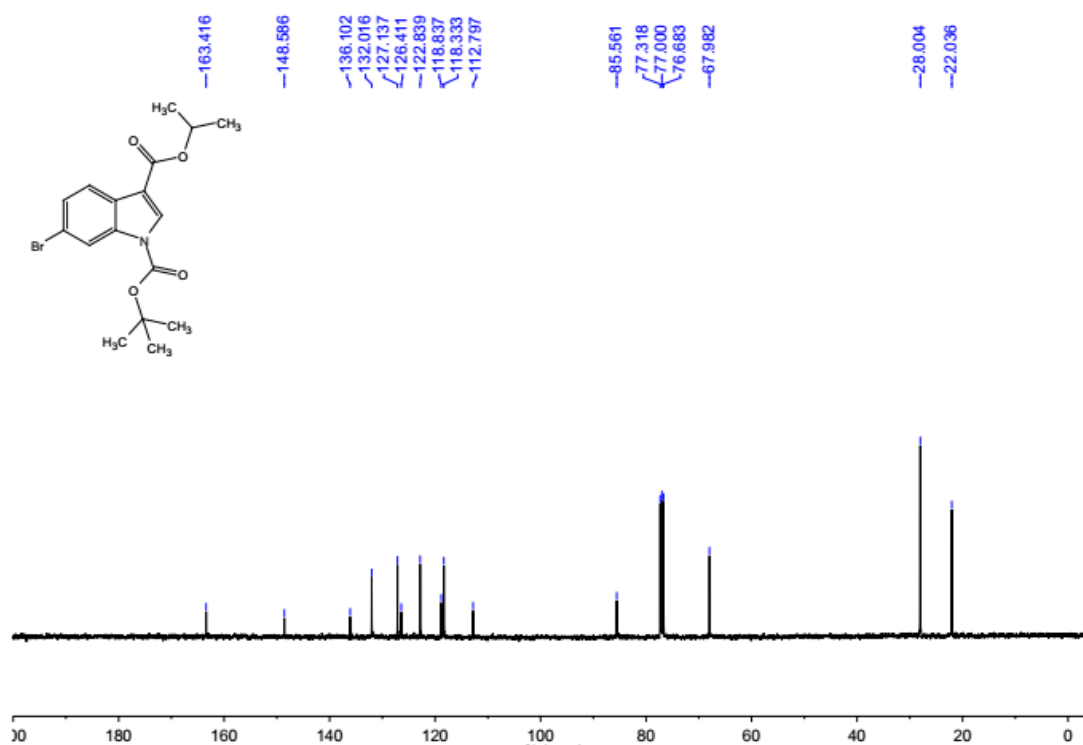
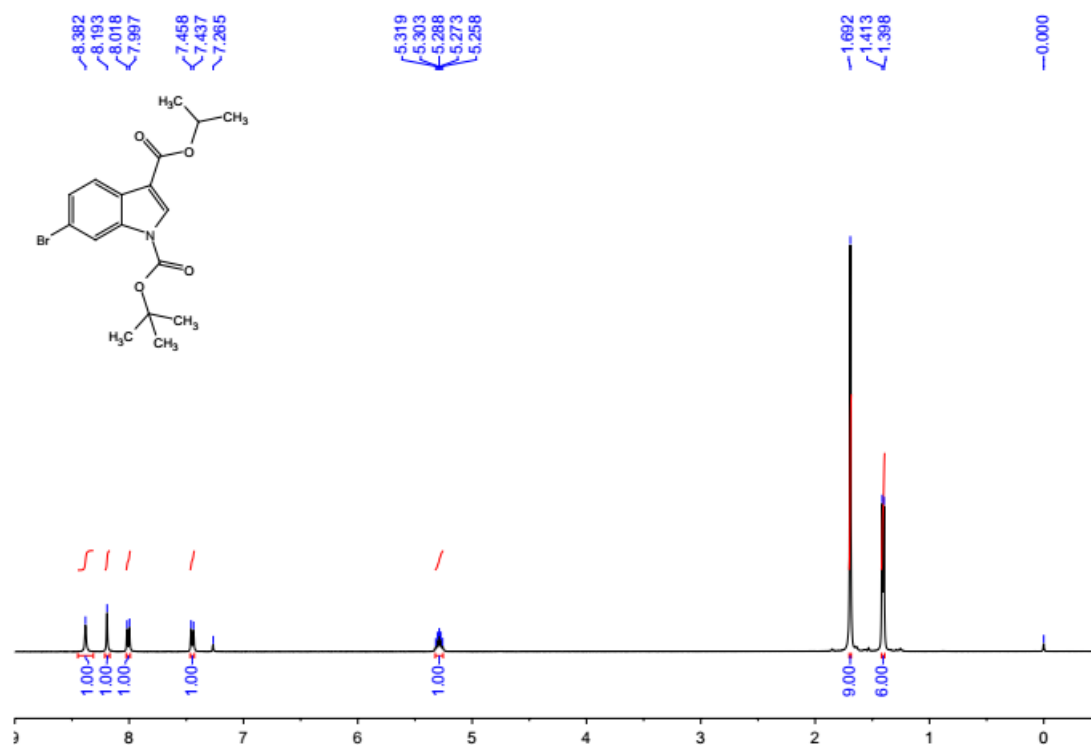
Compound 1q



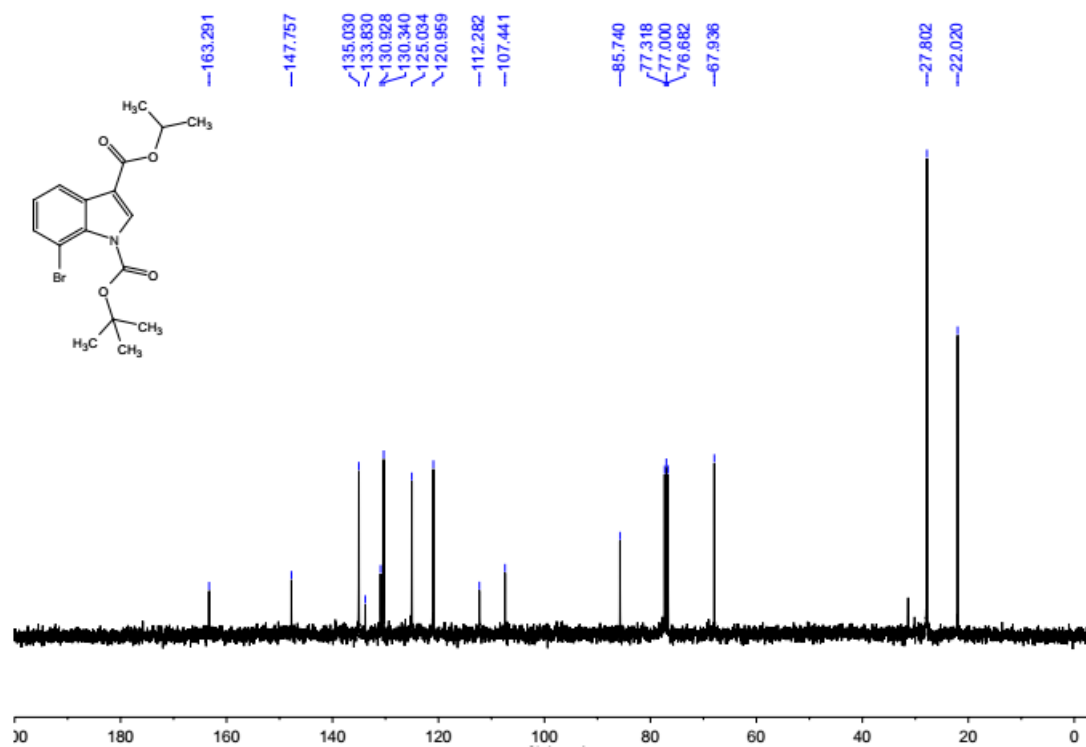
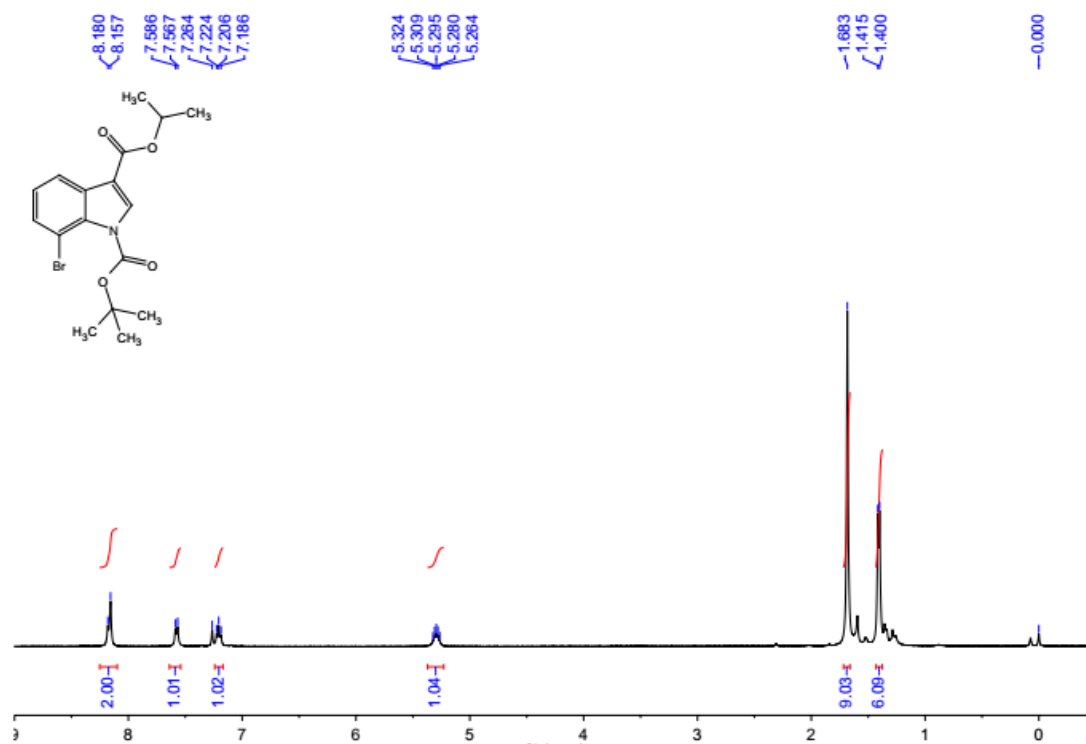
Compound 1r



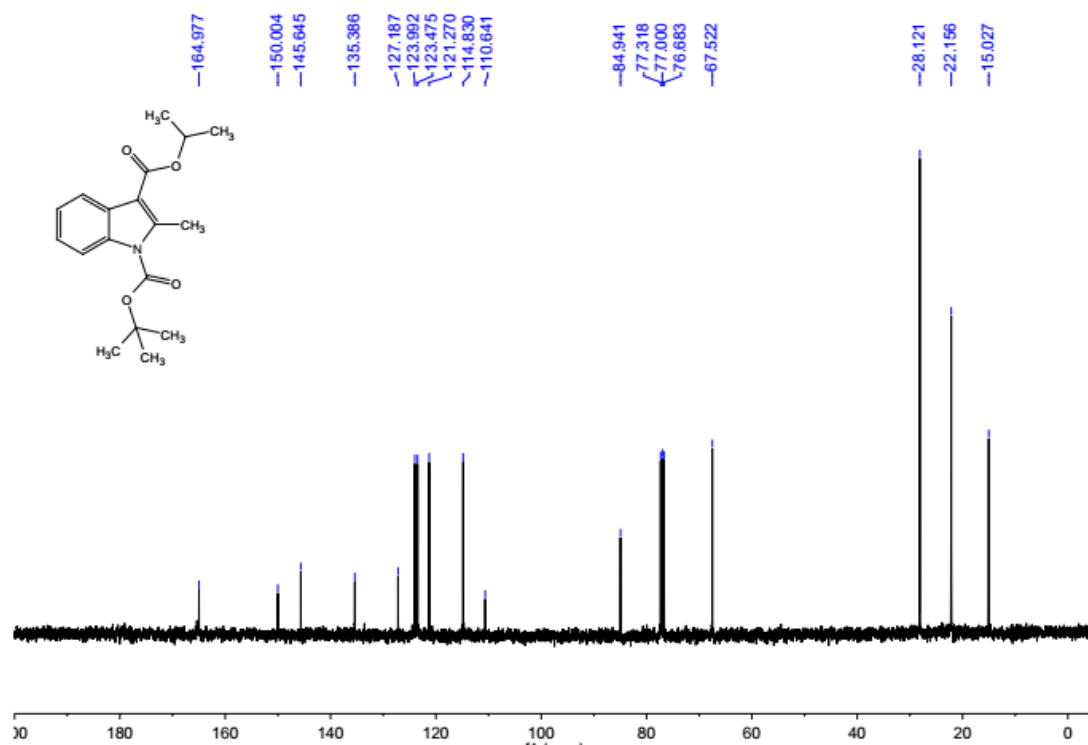
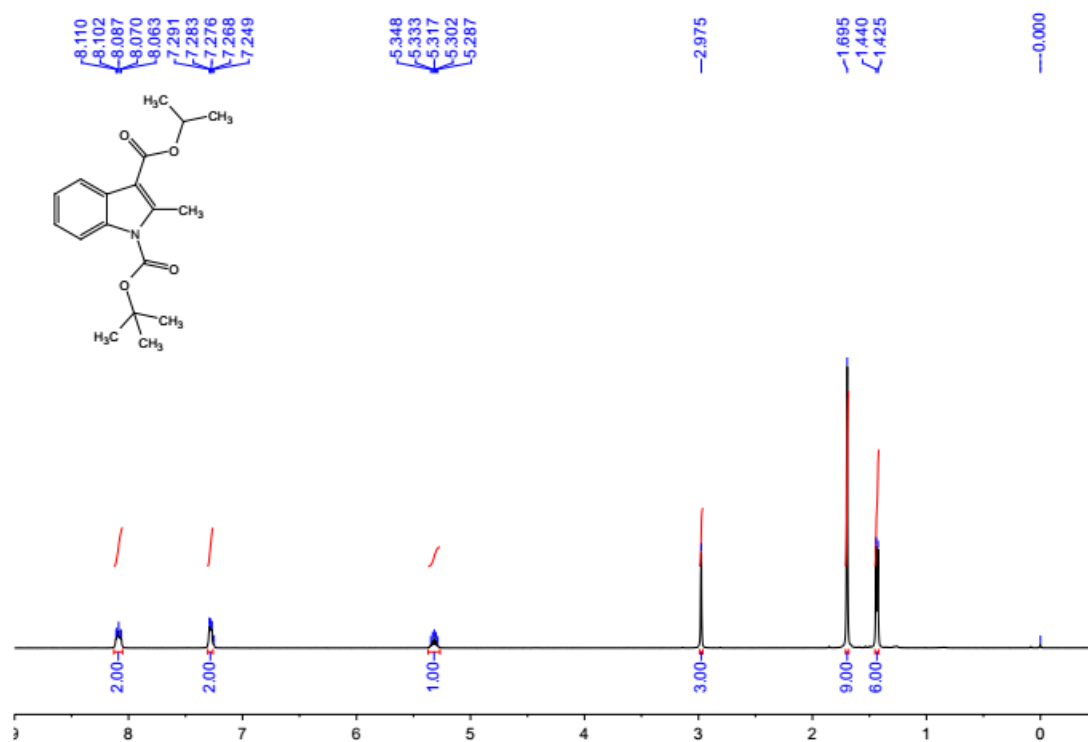
Compound 1s



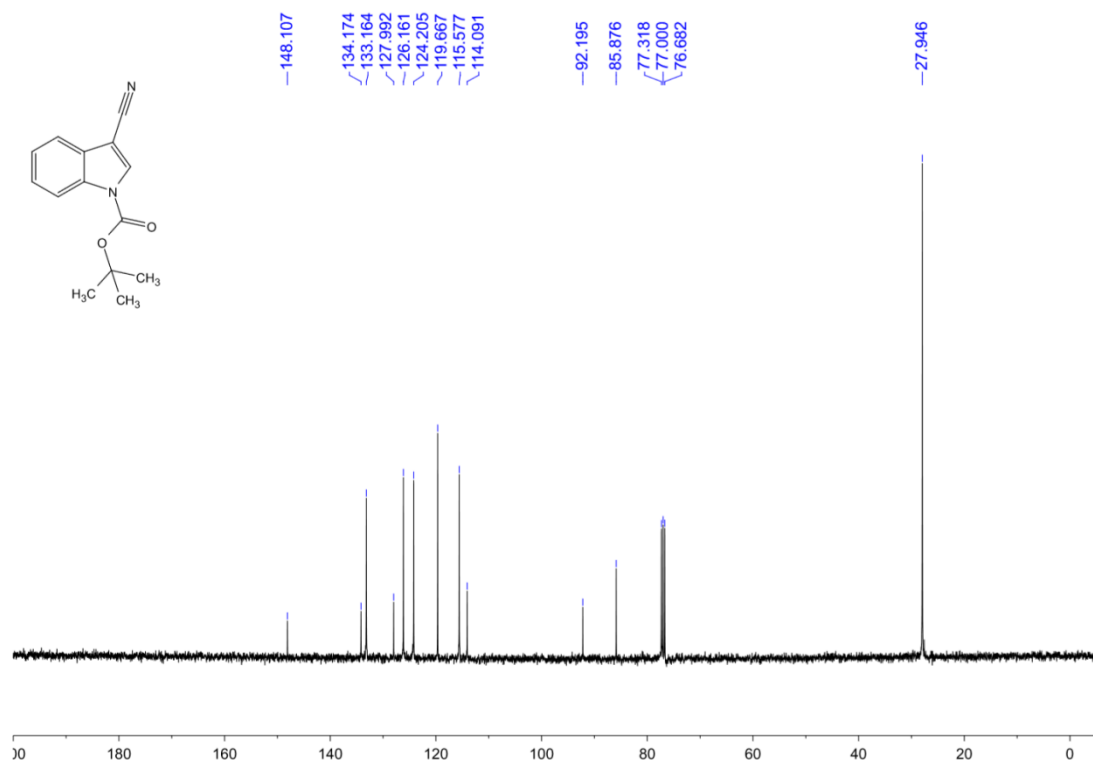
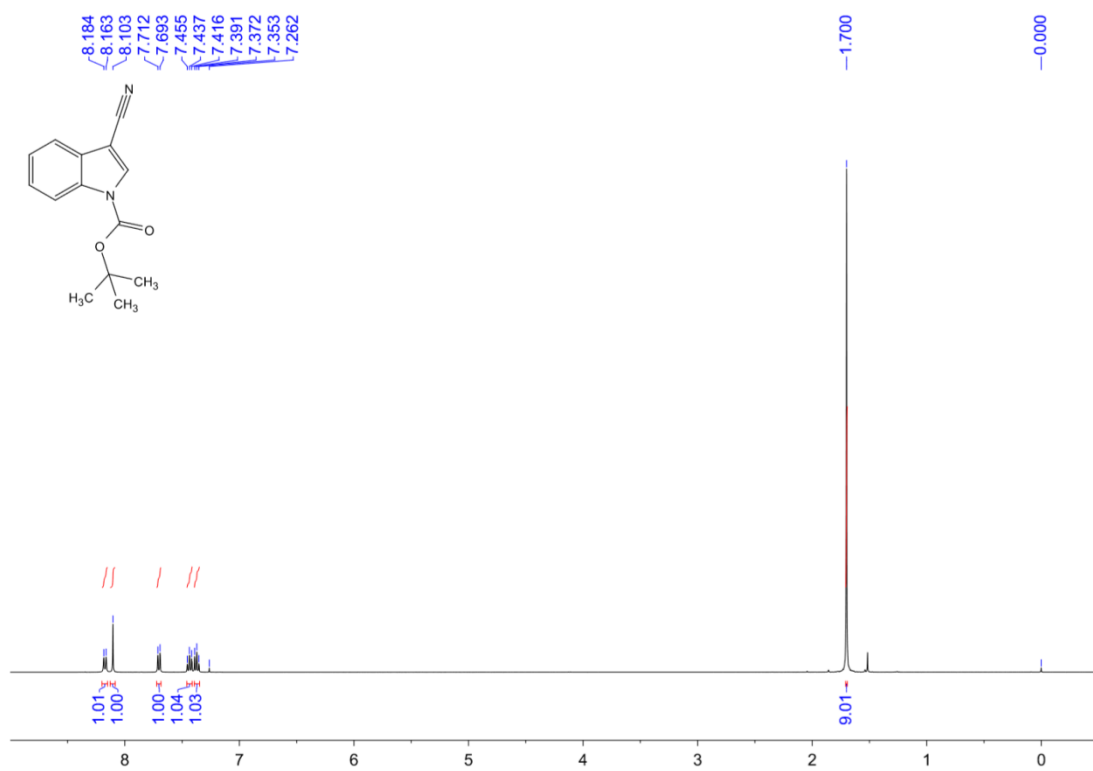
Compound 1t



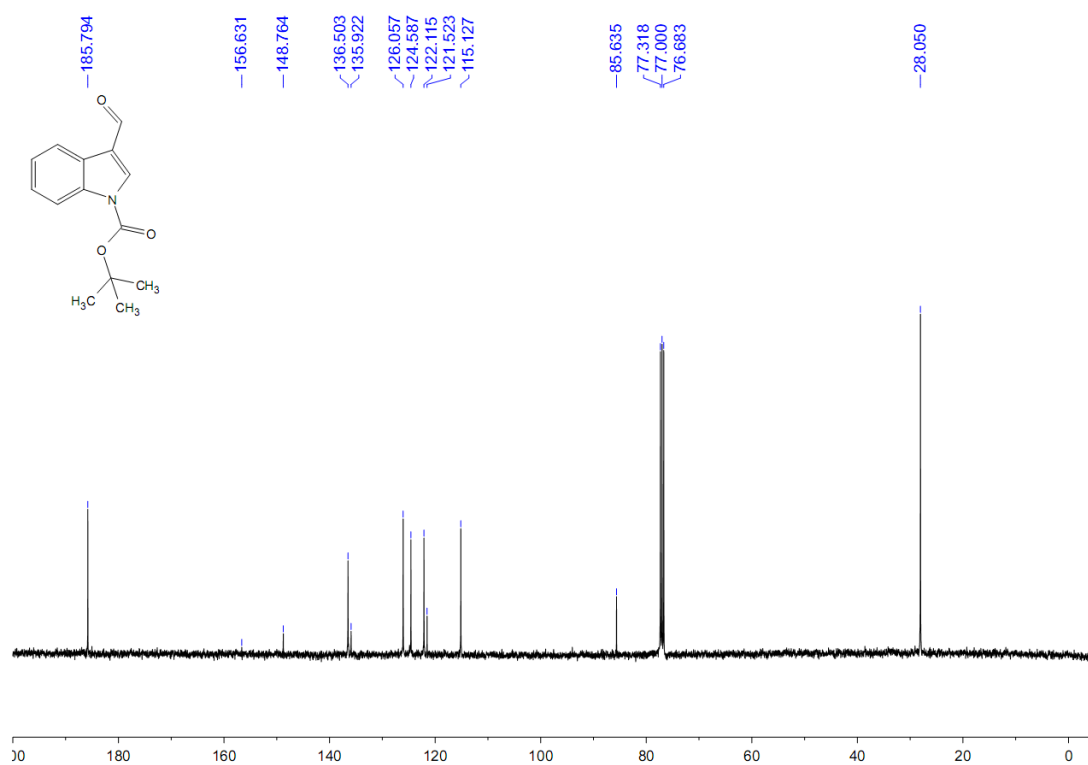
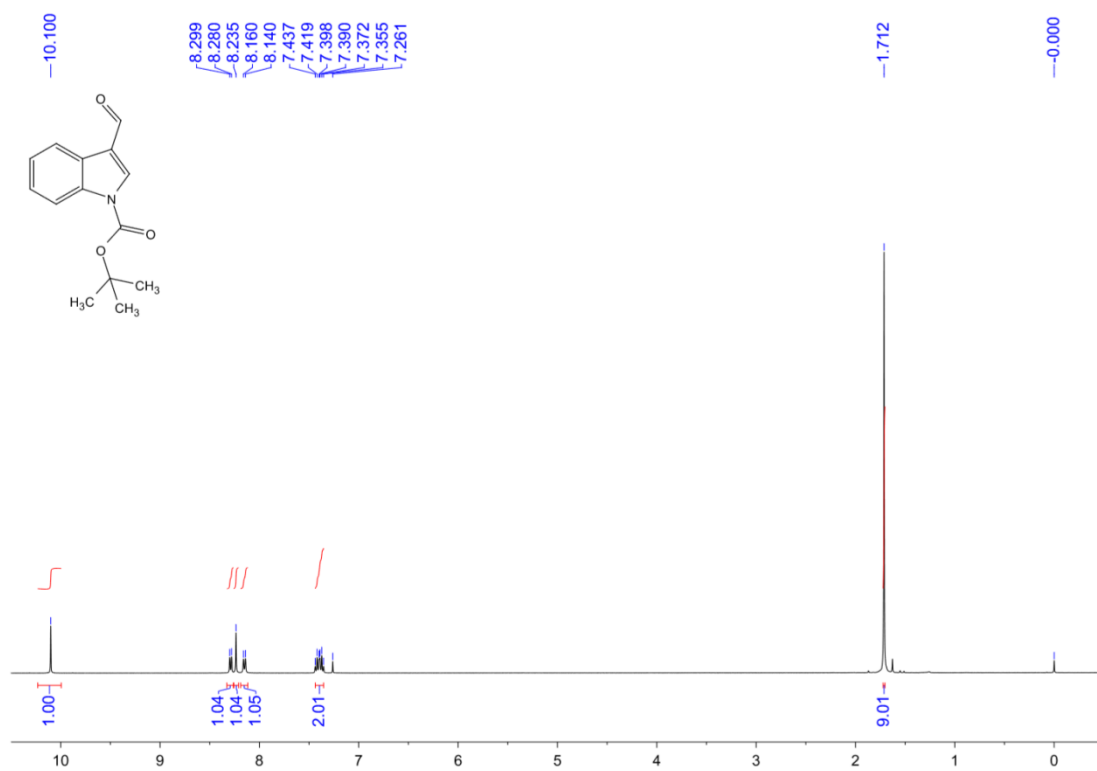
Compound 1u



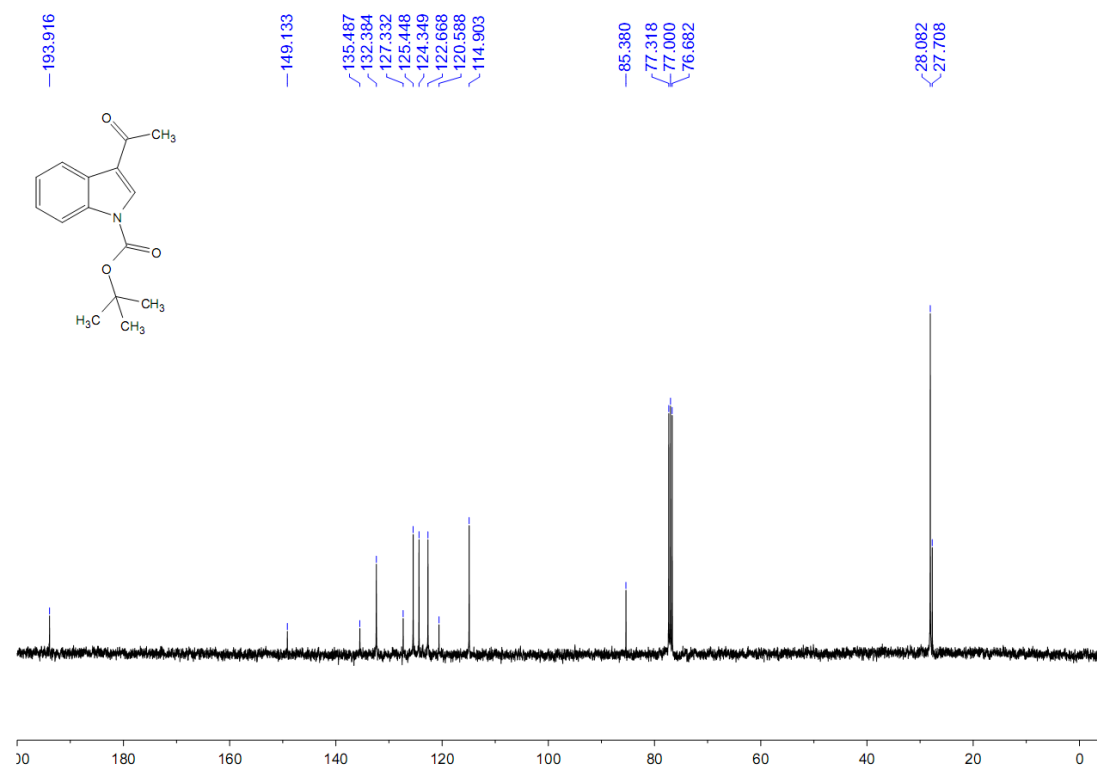
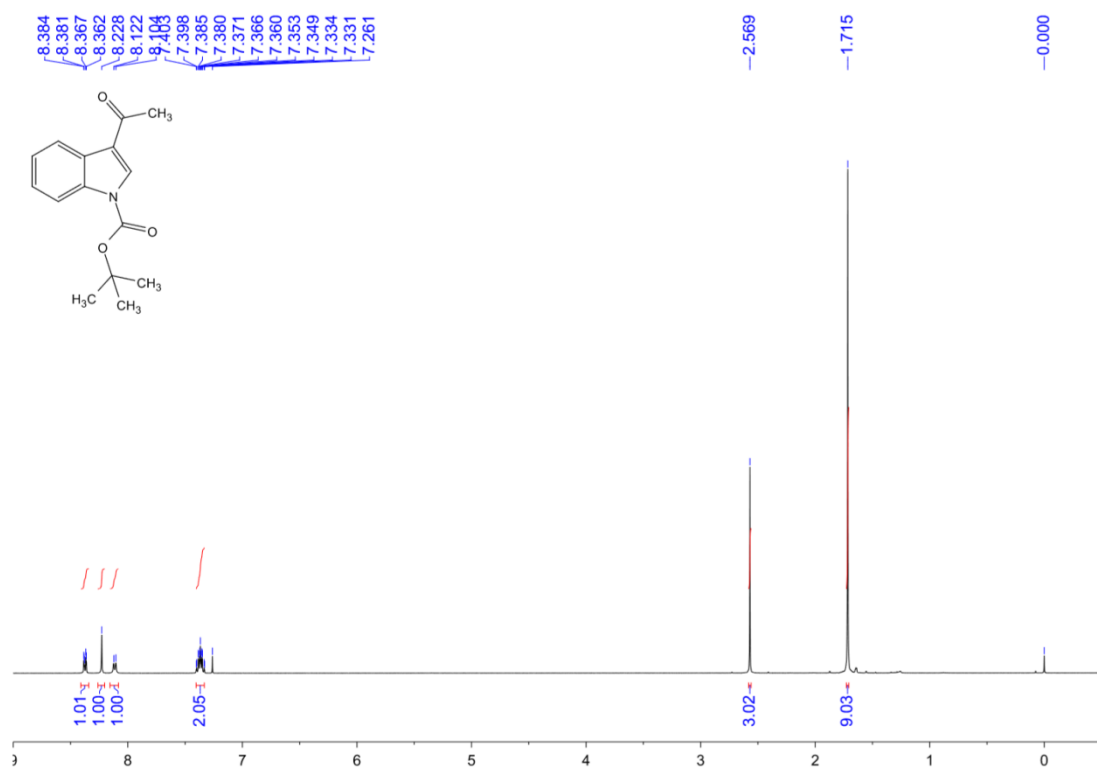
Compound 1v



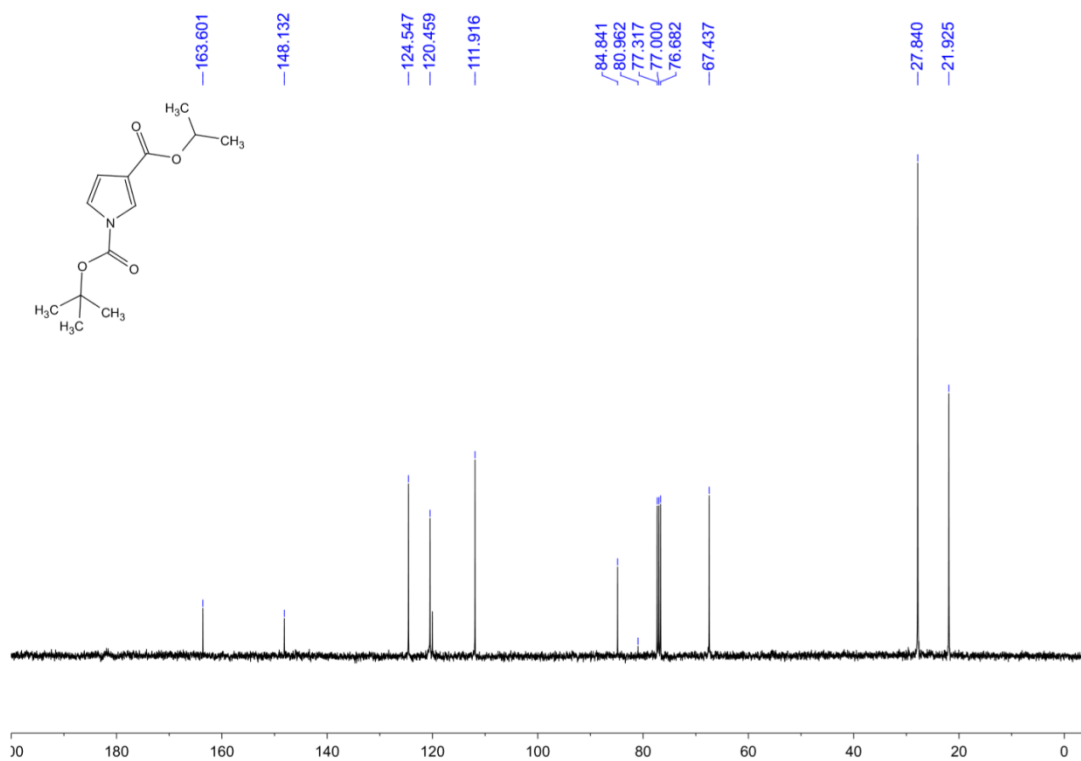
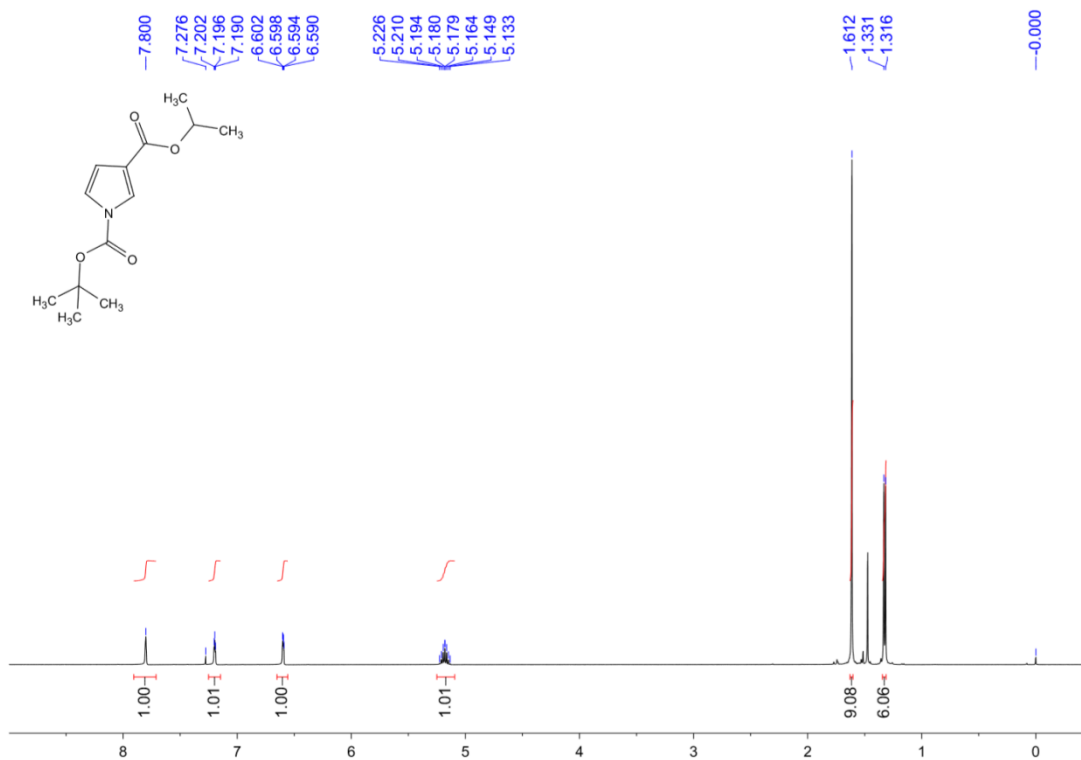
Compound 1w



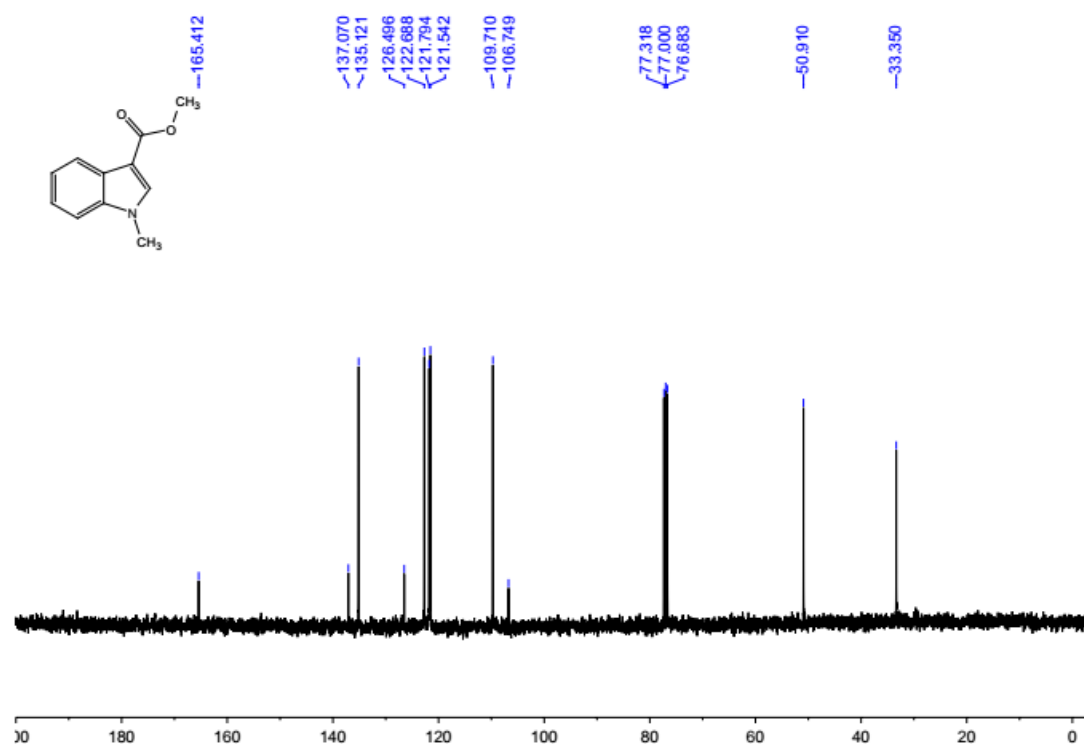
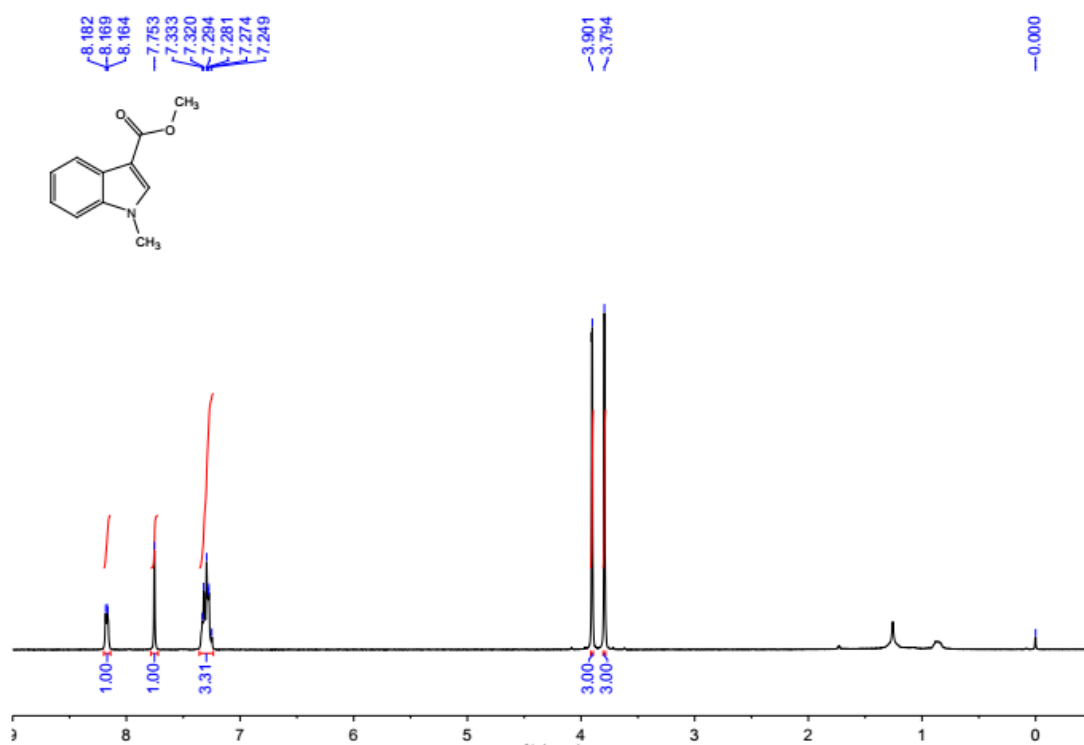
Compound 1x



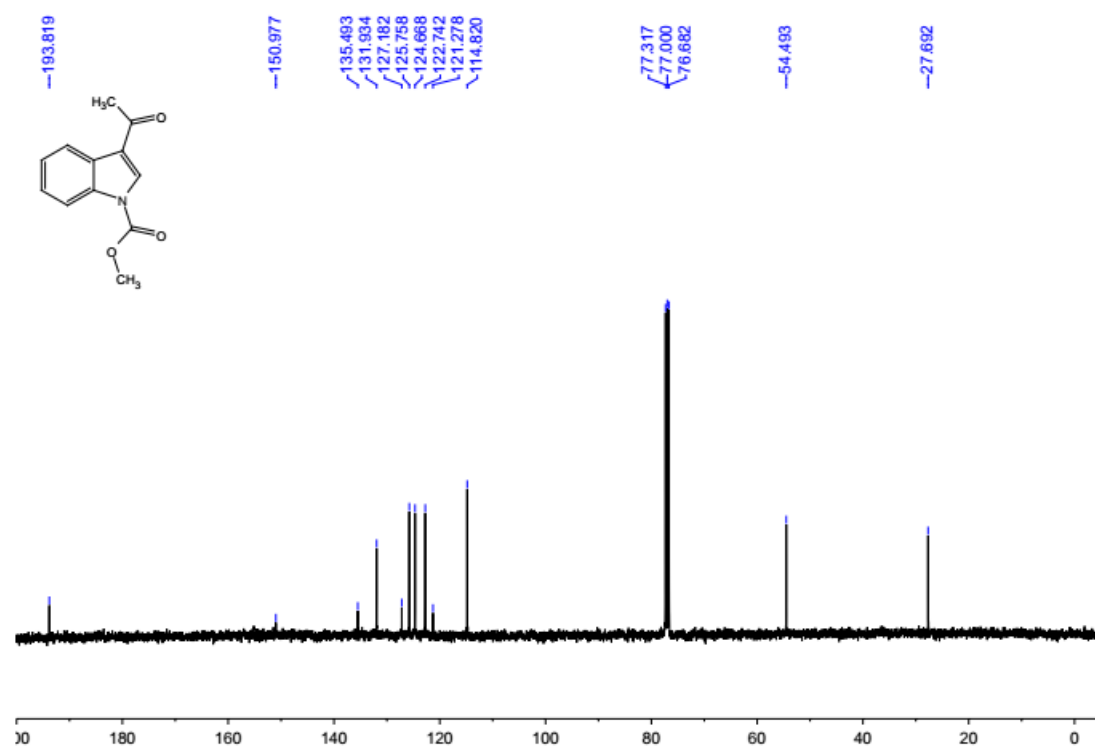
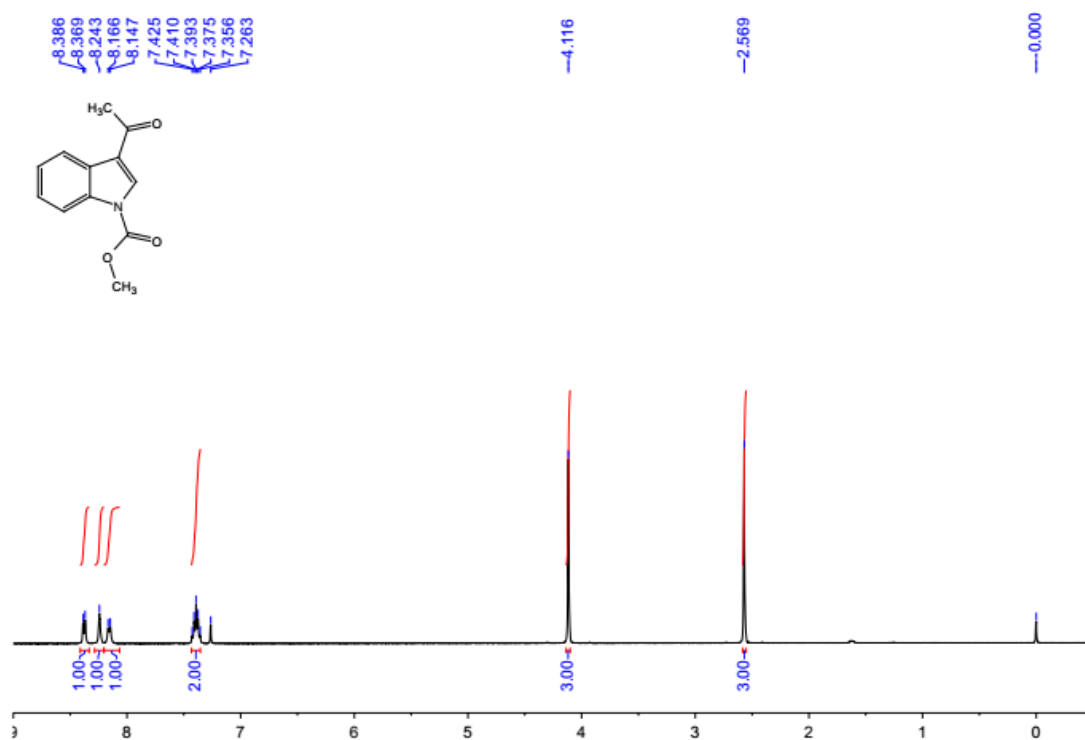
Compound 1y



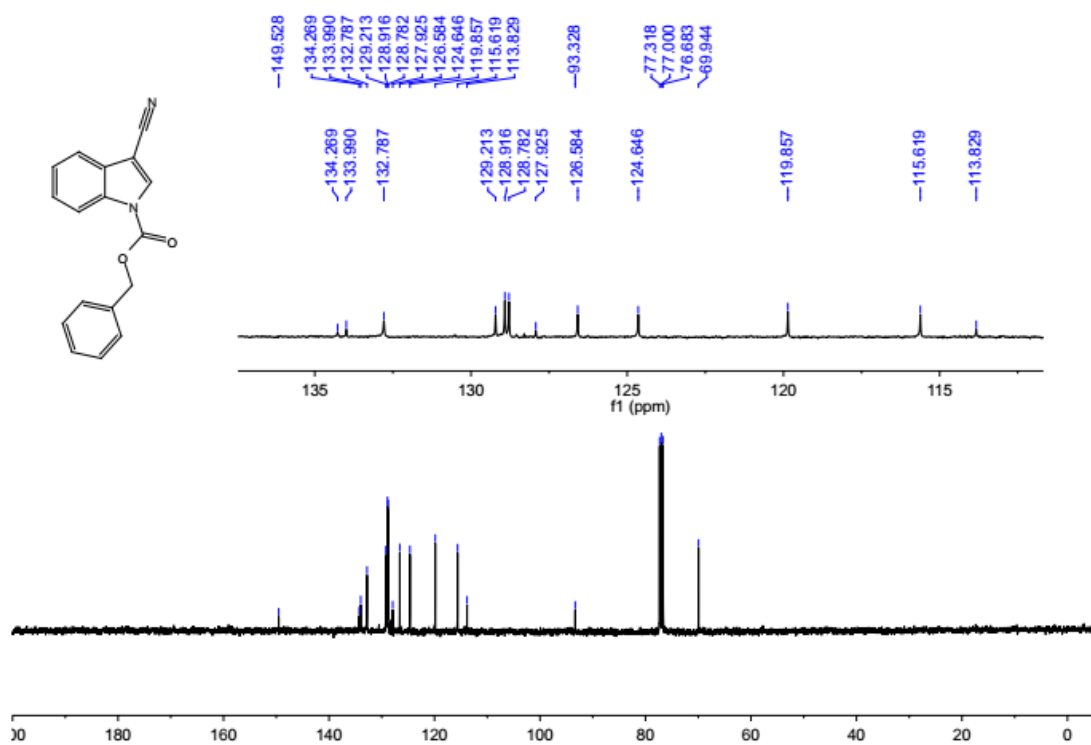
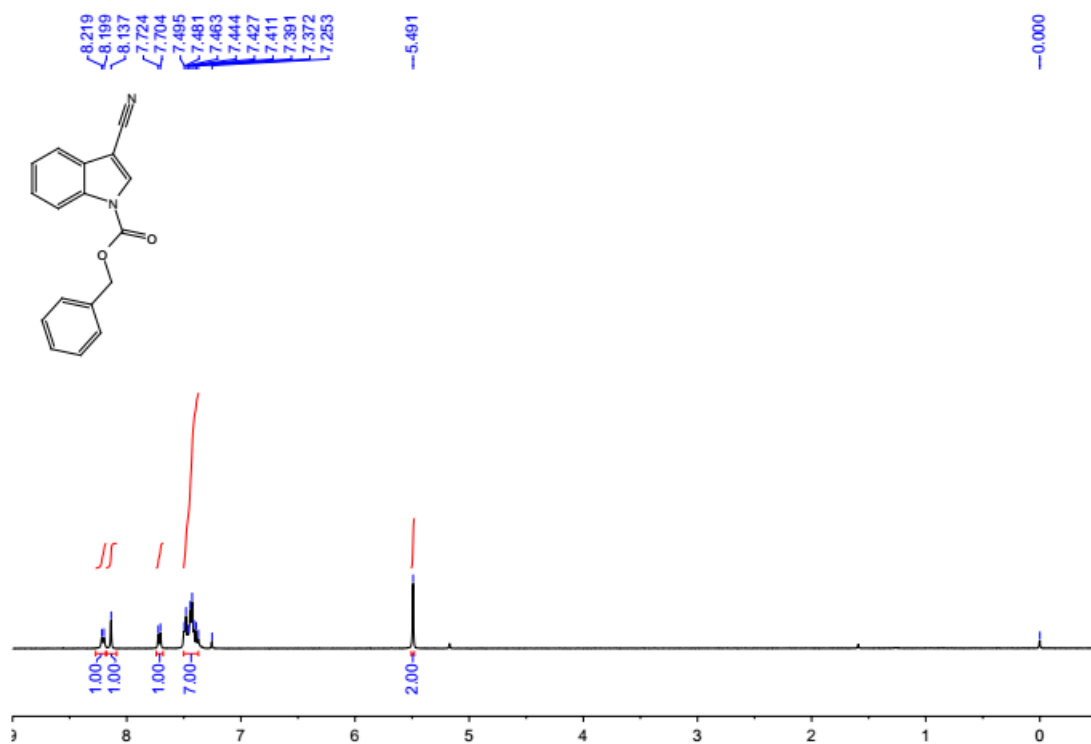
Compound S2



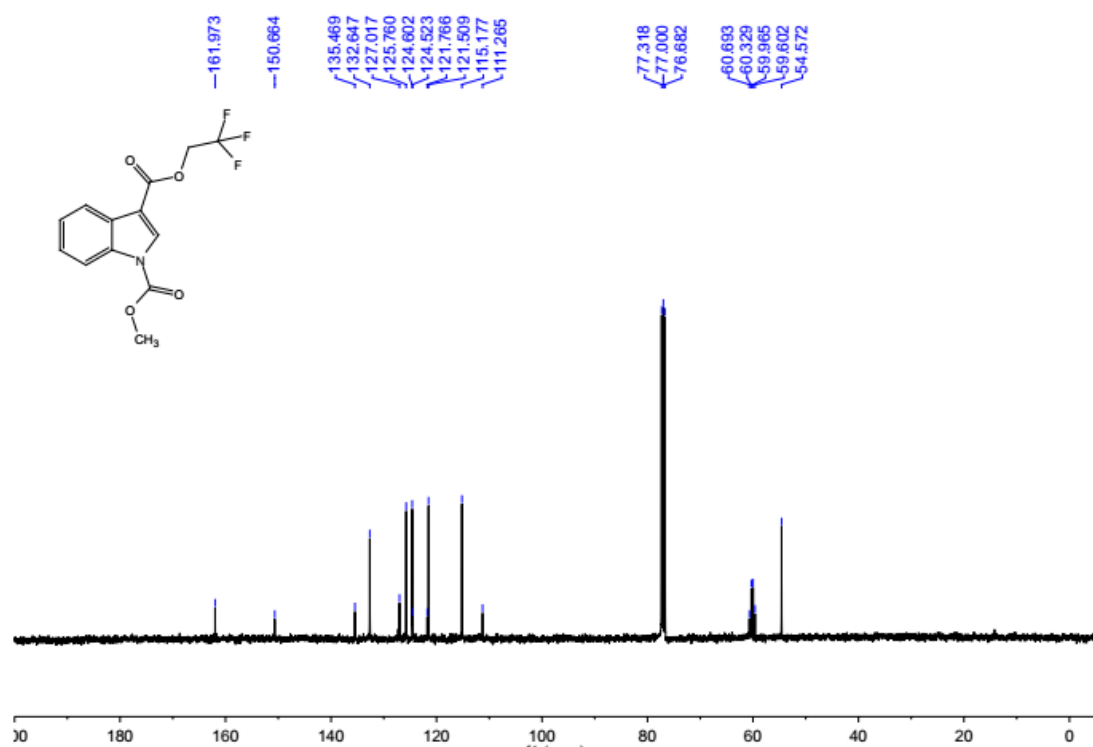
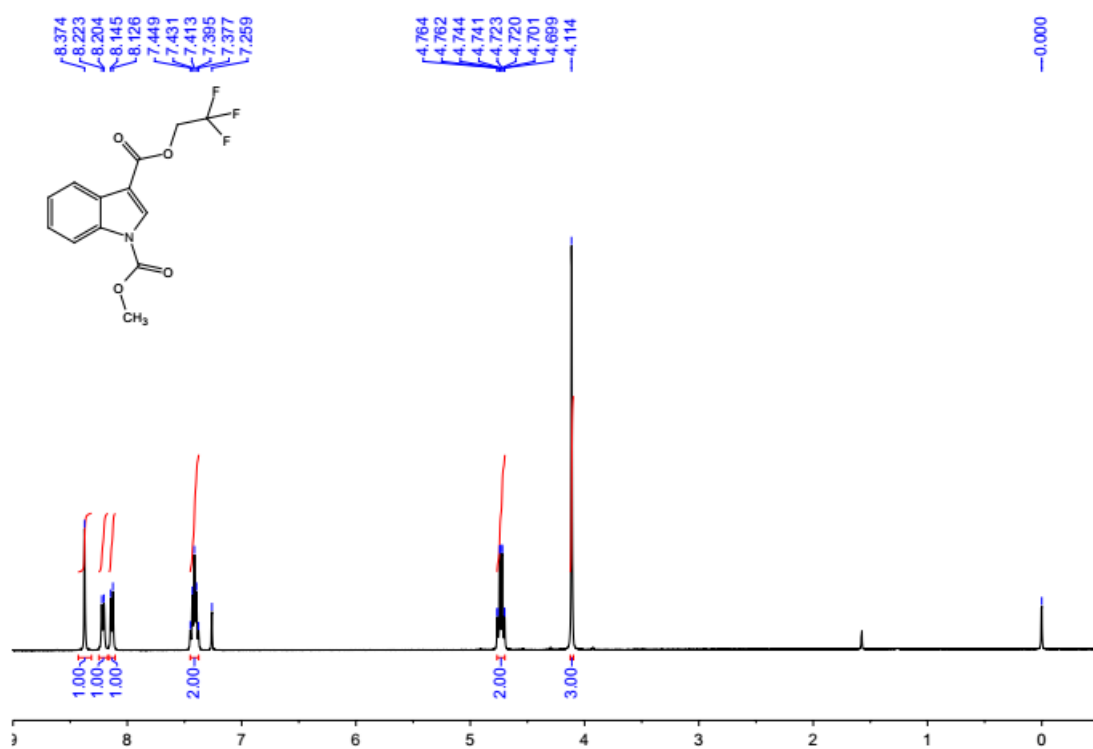
Compound S3



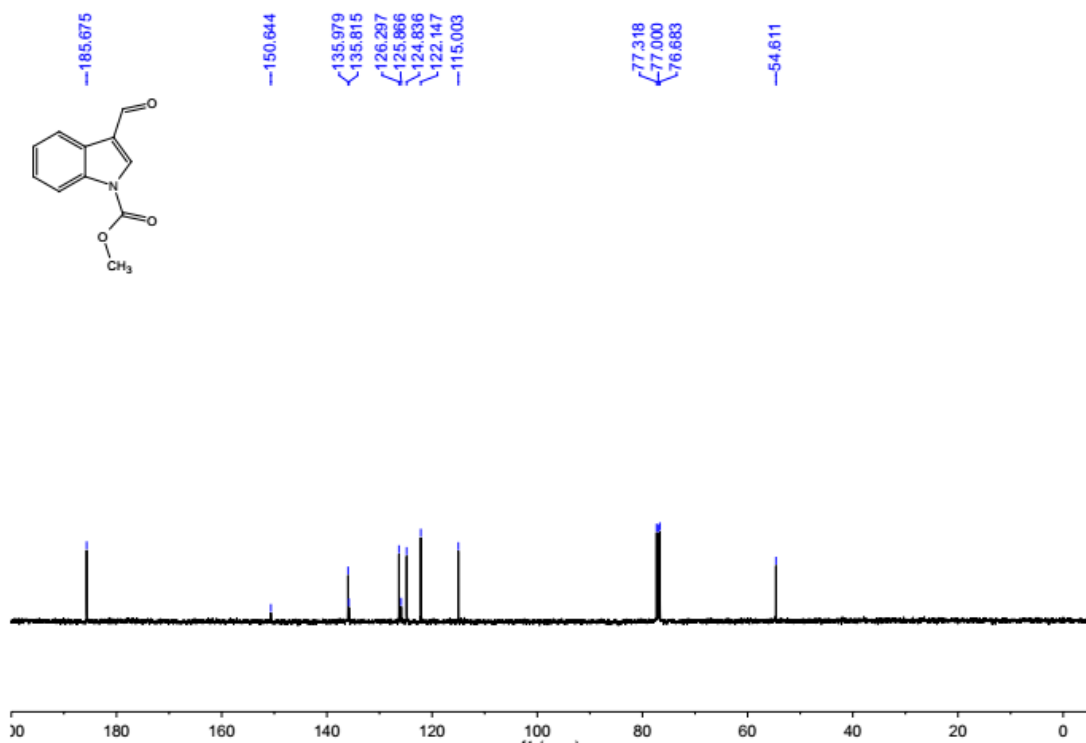
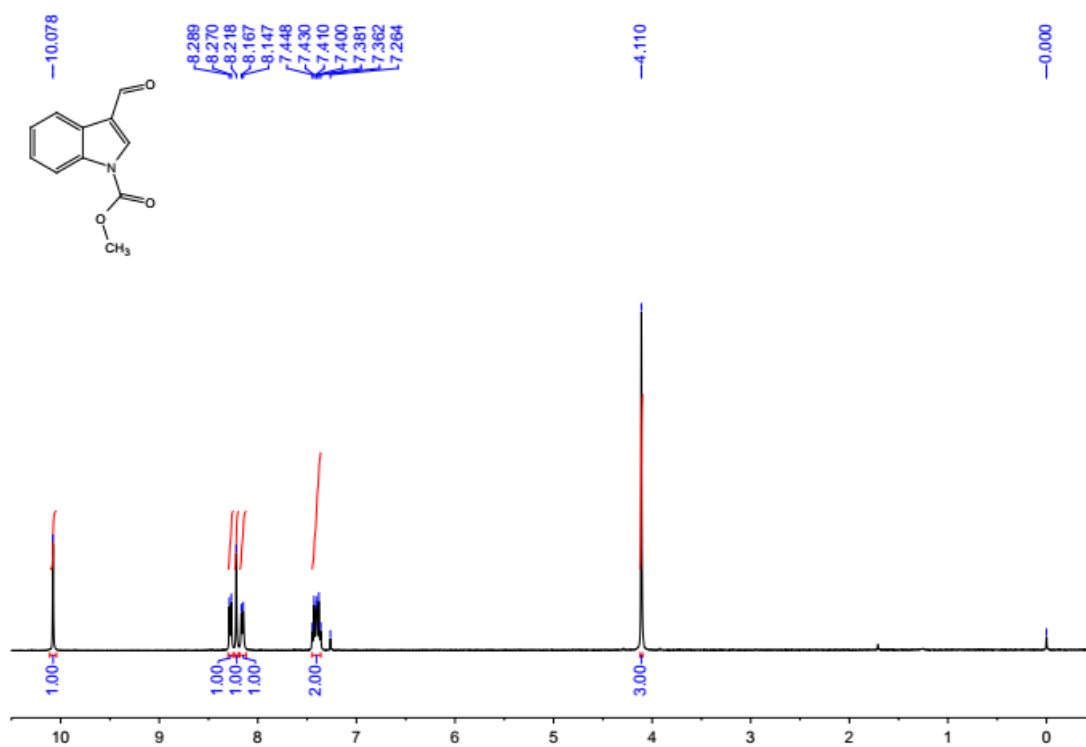
Compound S4



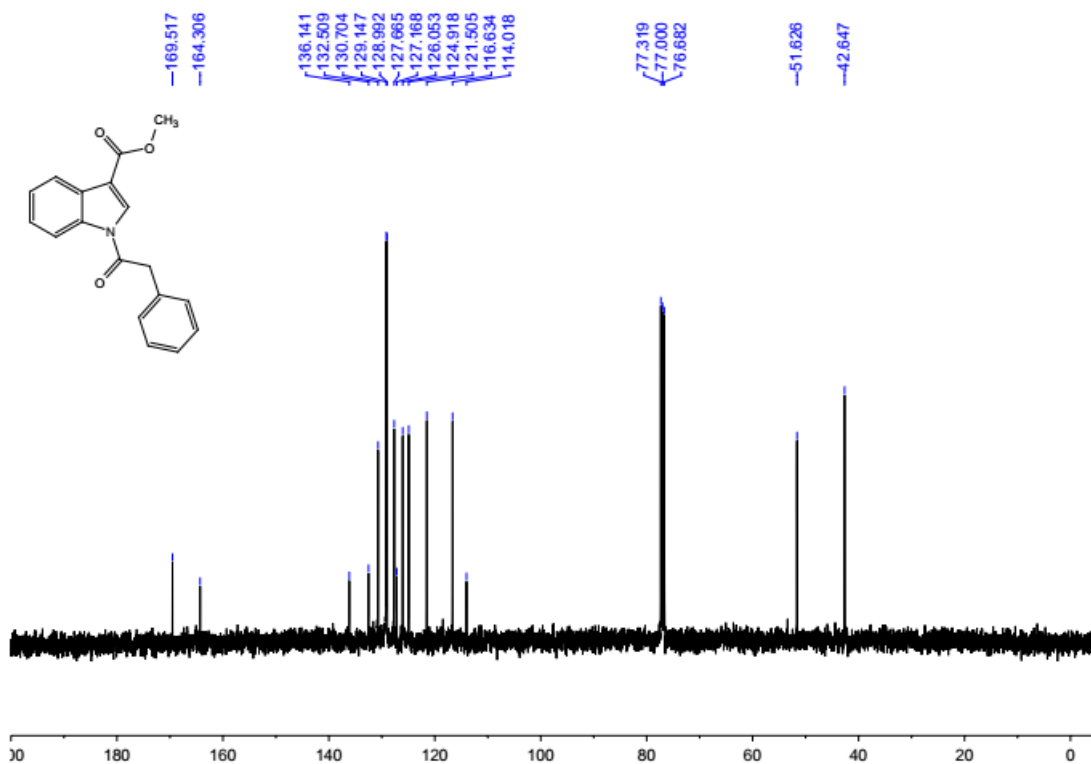
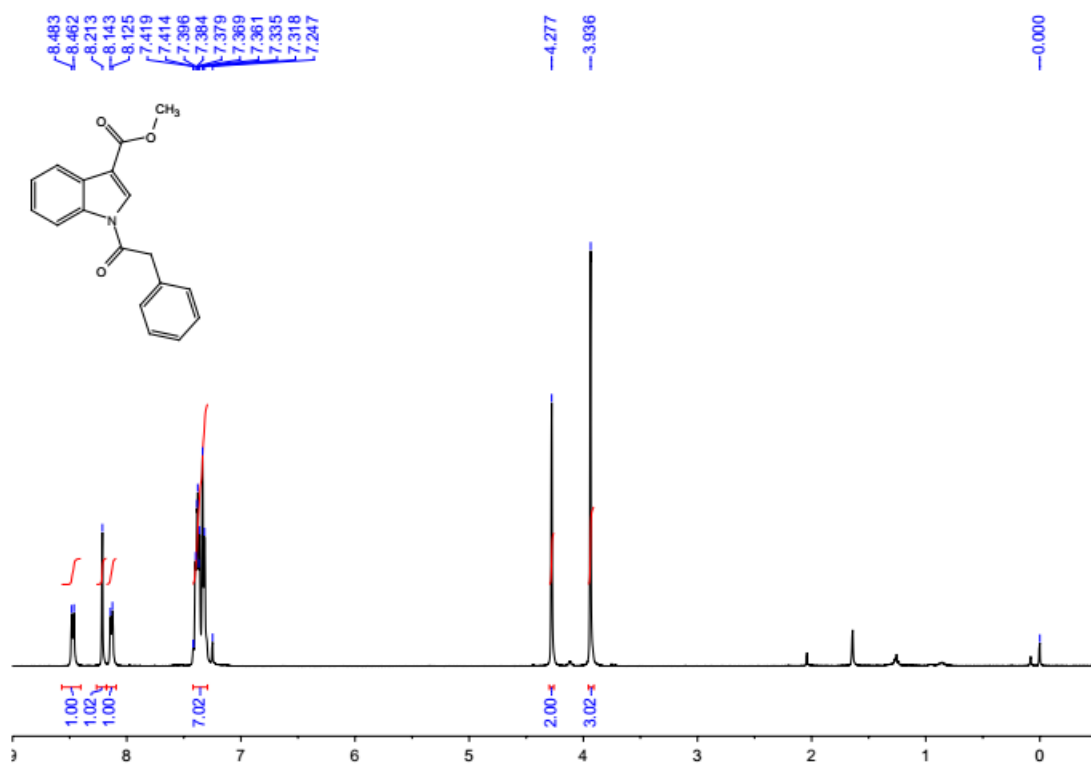
Compound S5



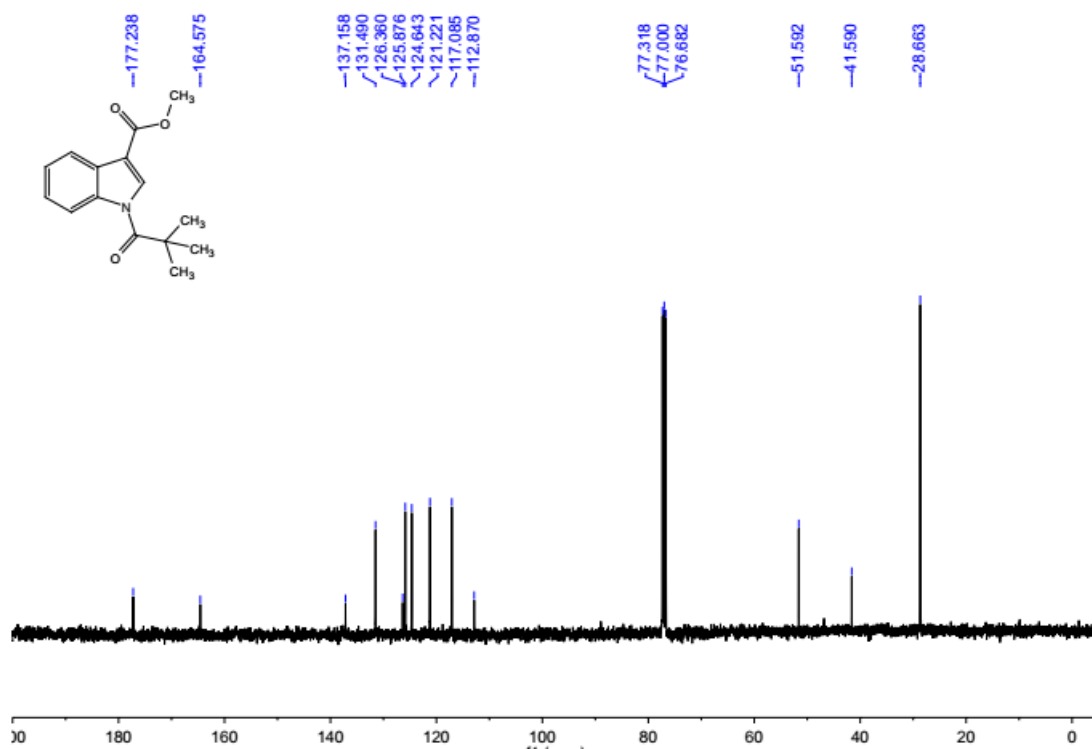
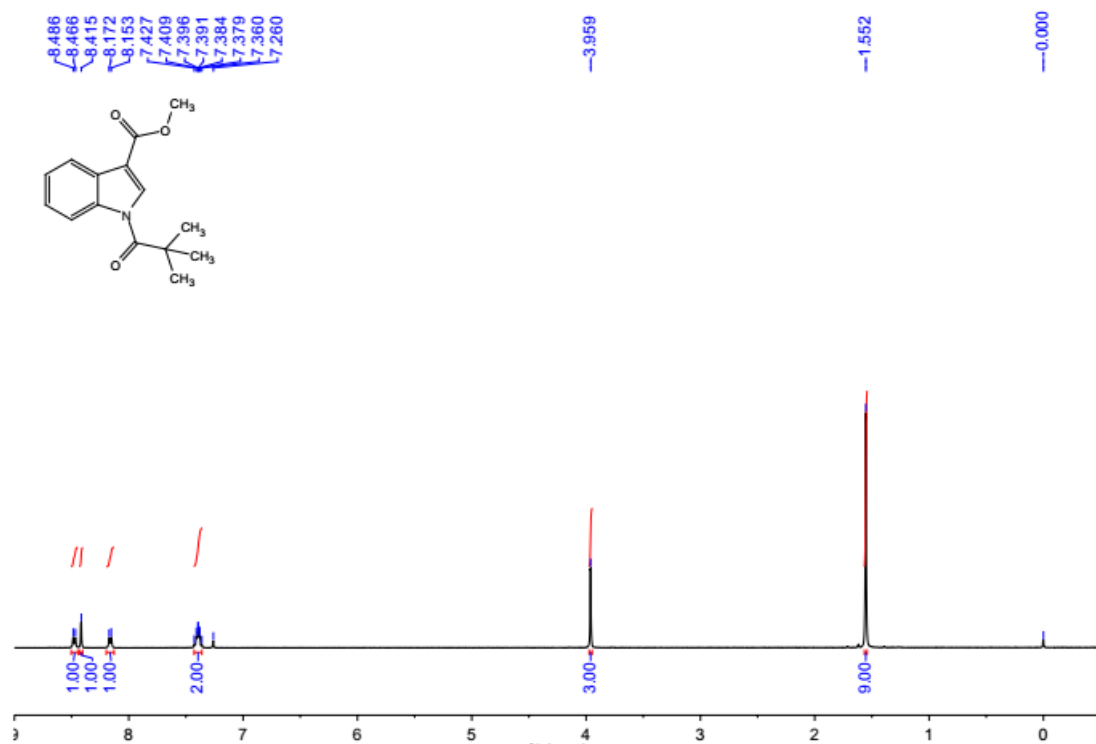
Compound S6



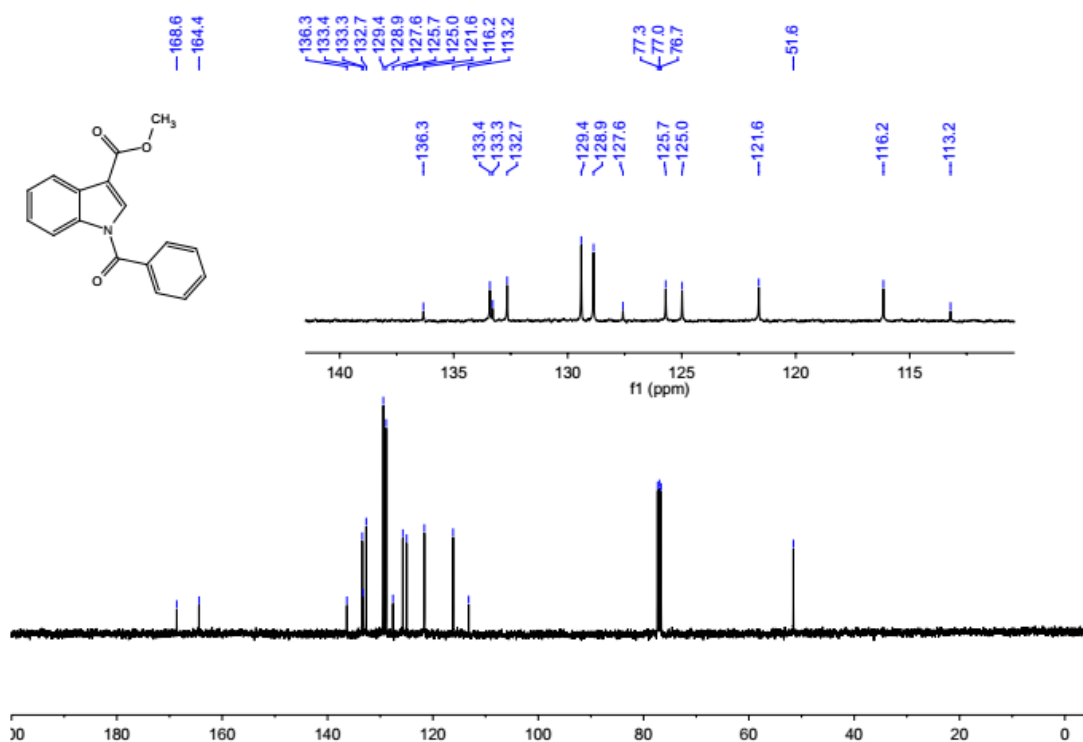
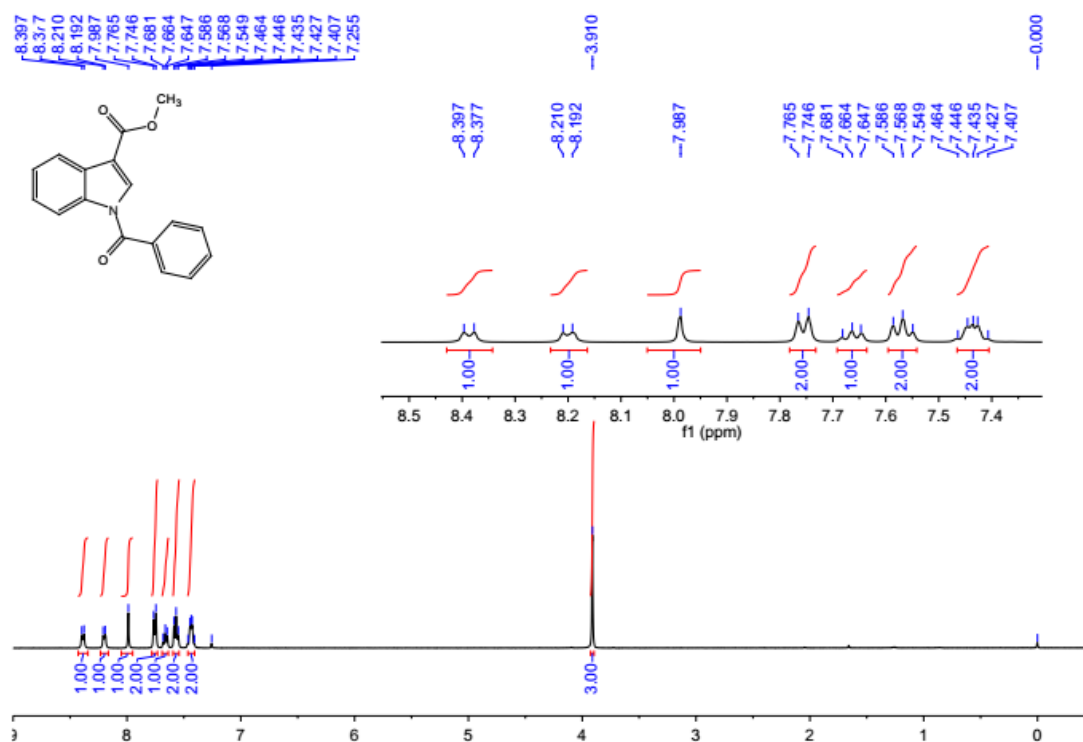
Compound S7



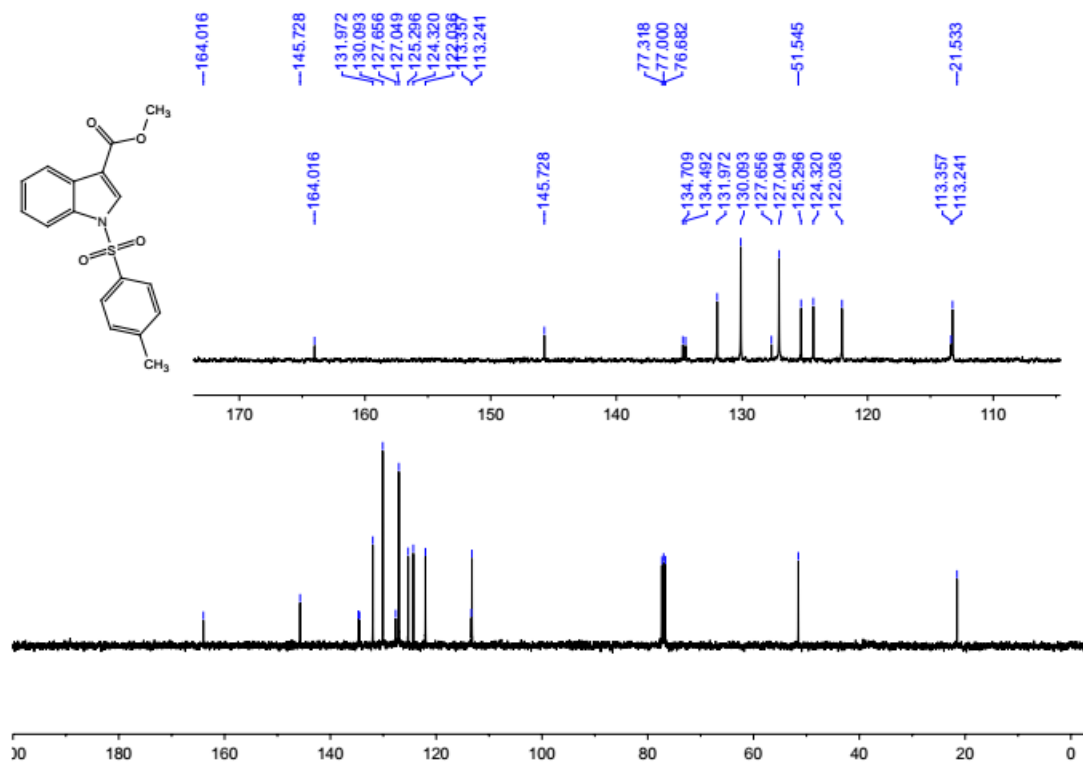
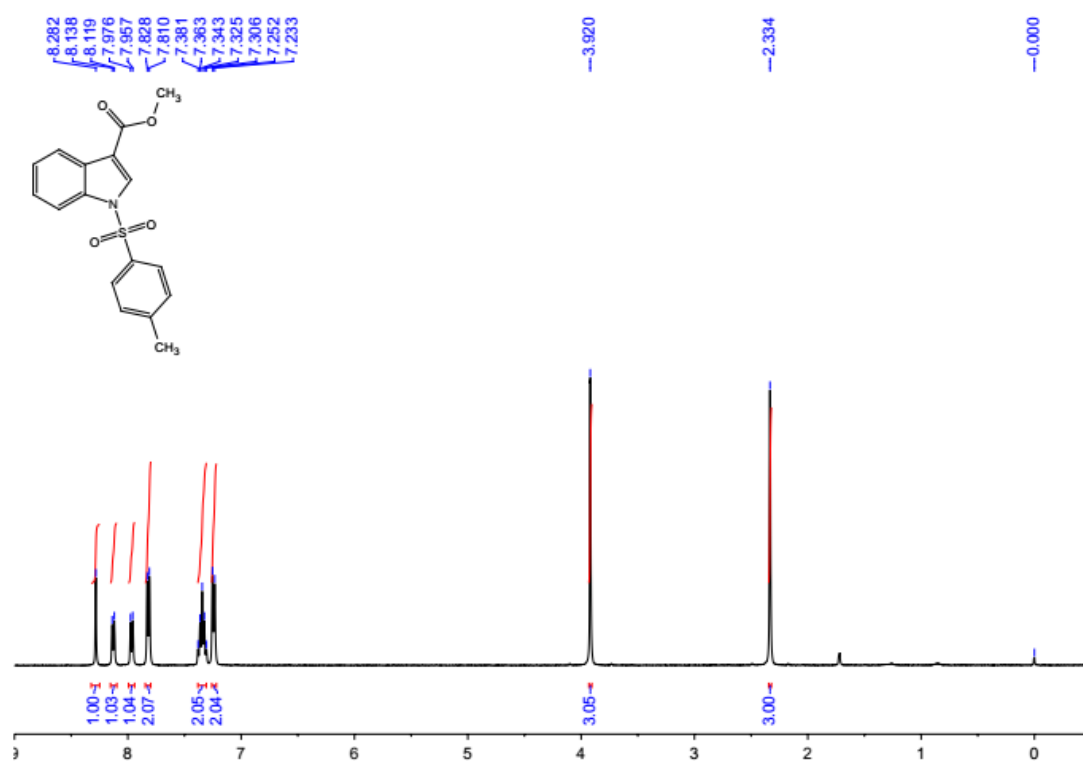
Compound S8



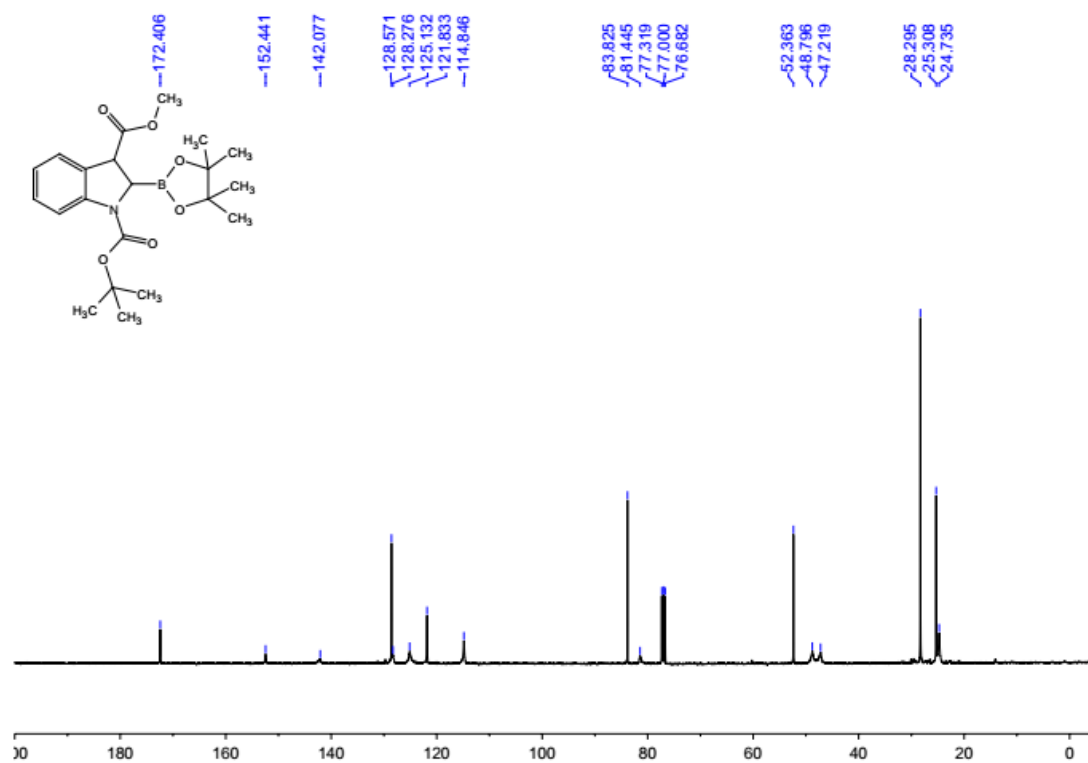
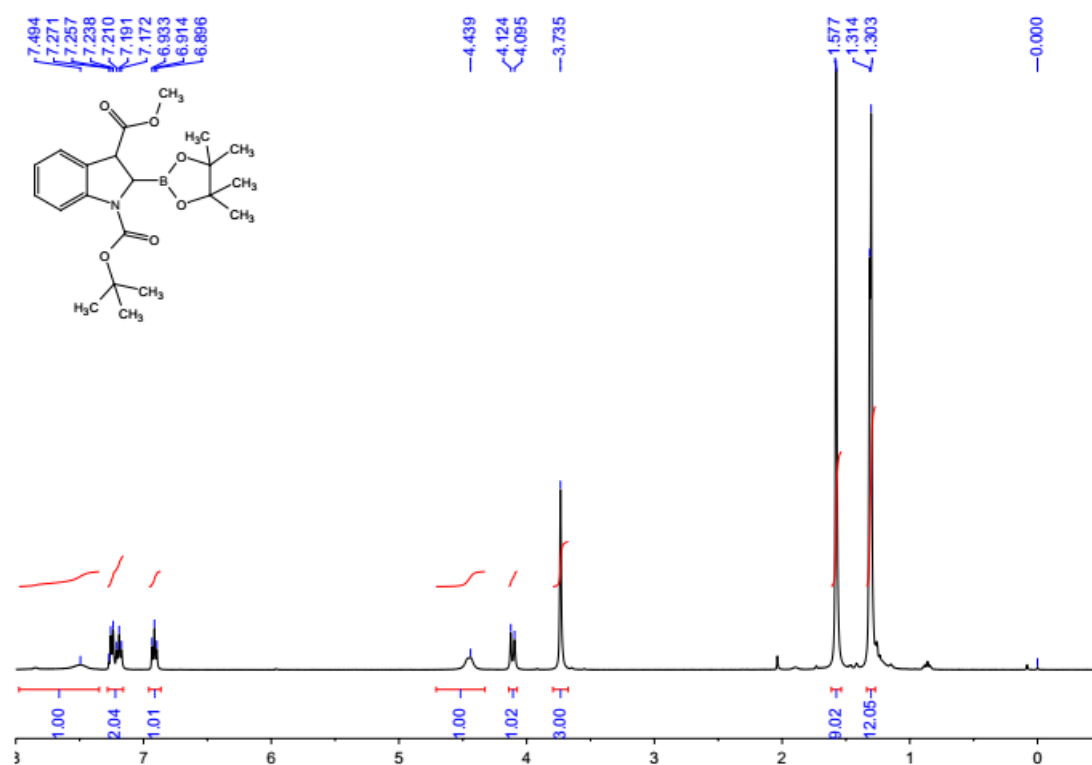
Compound S9

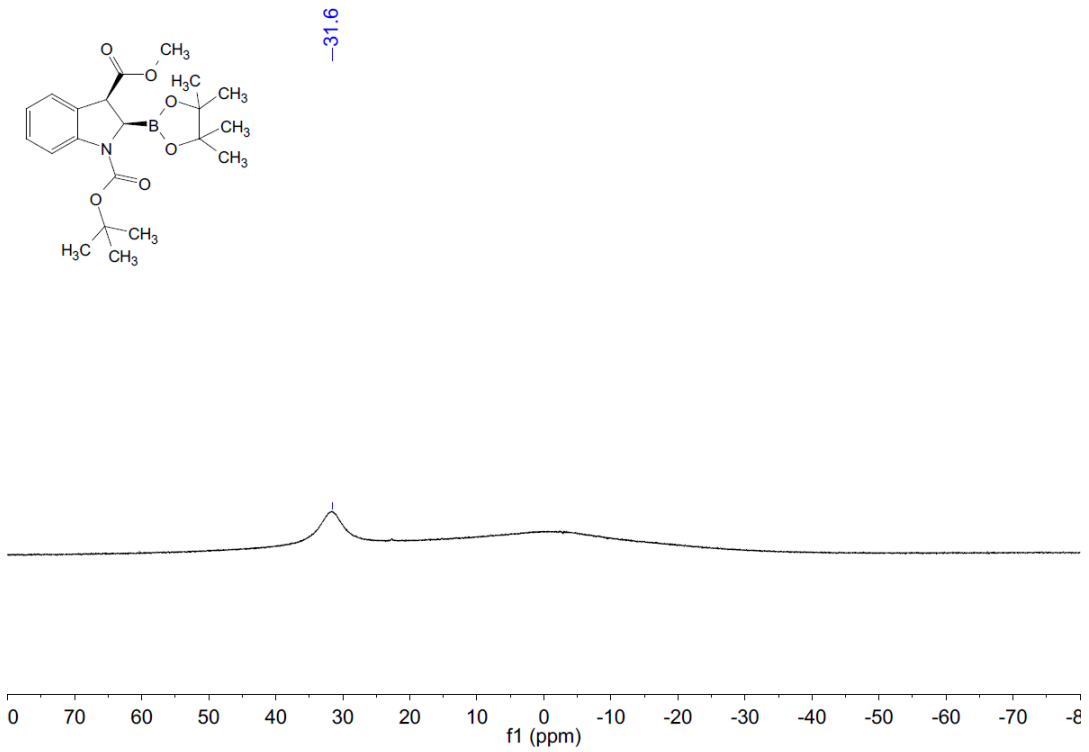


Compound S10

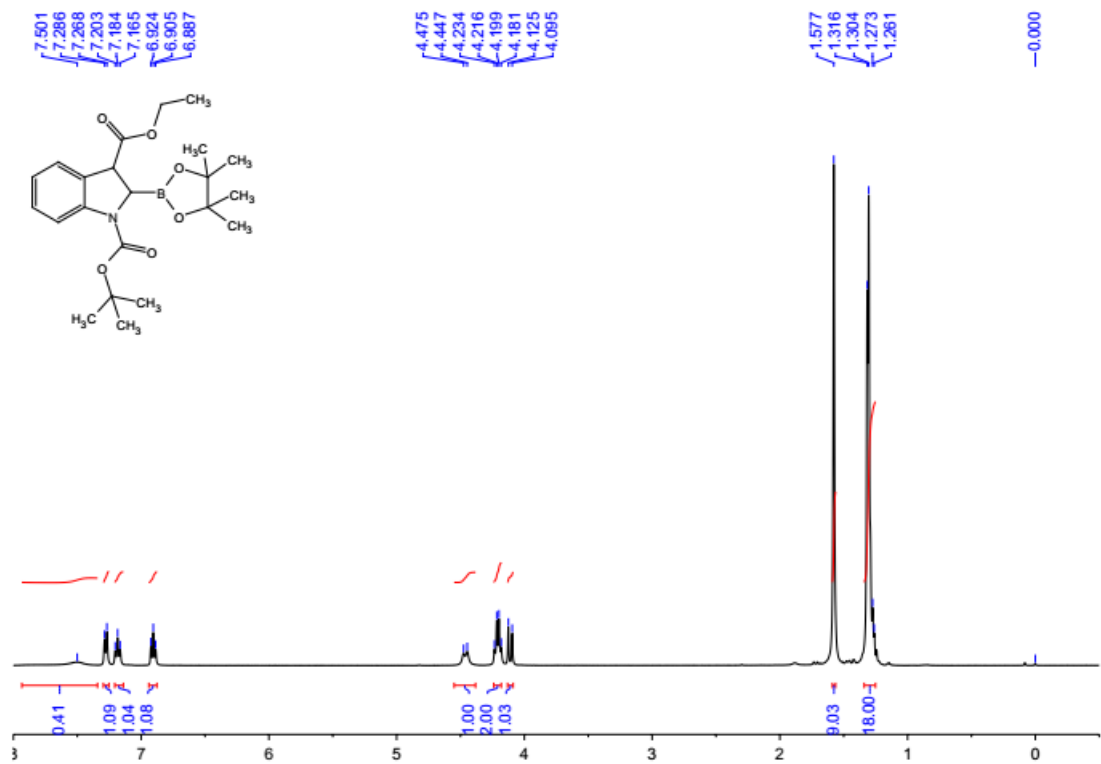


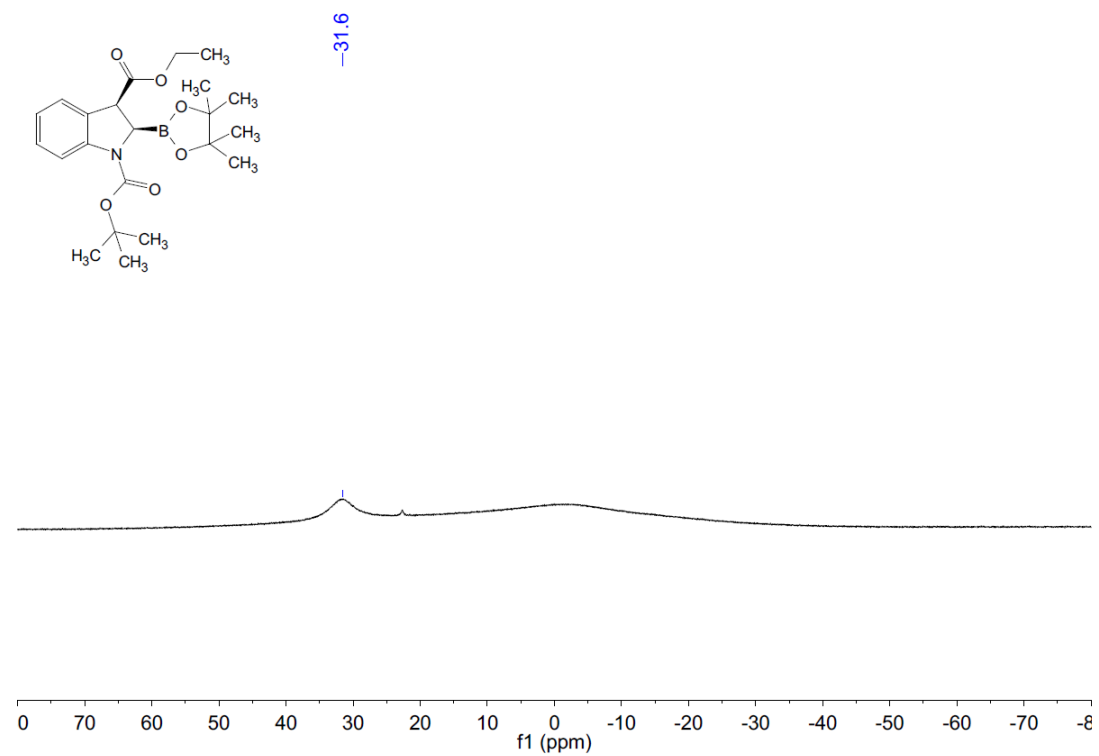
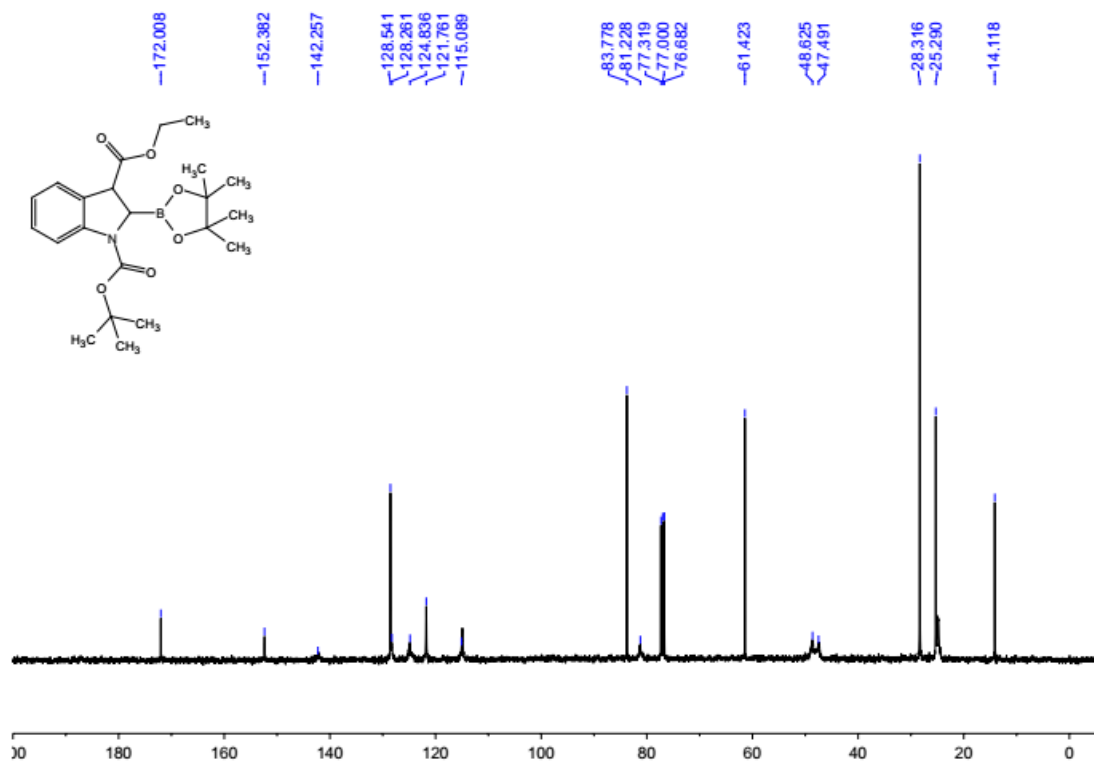
Compound 2a



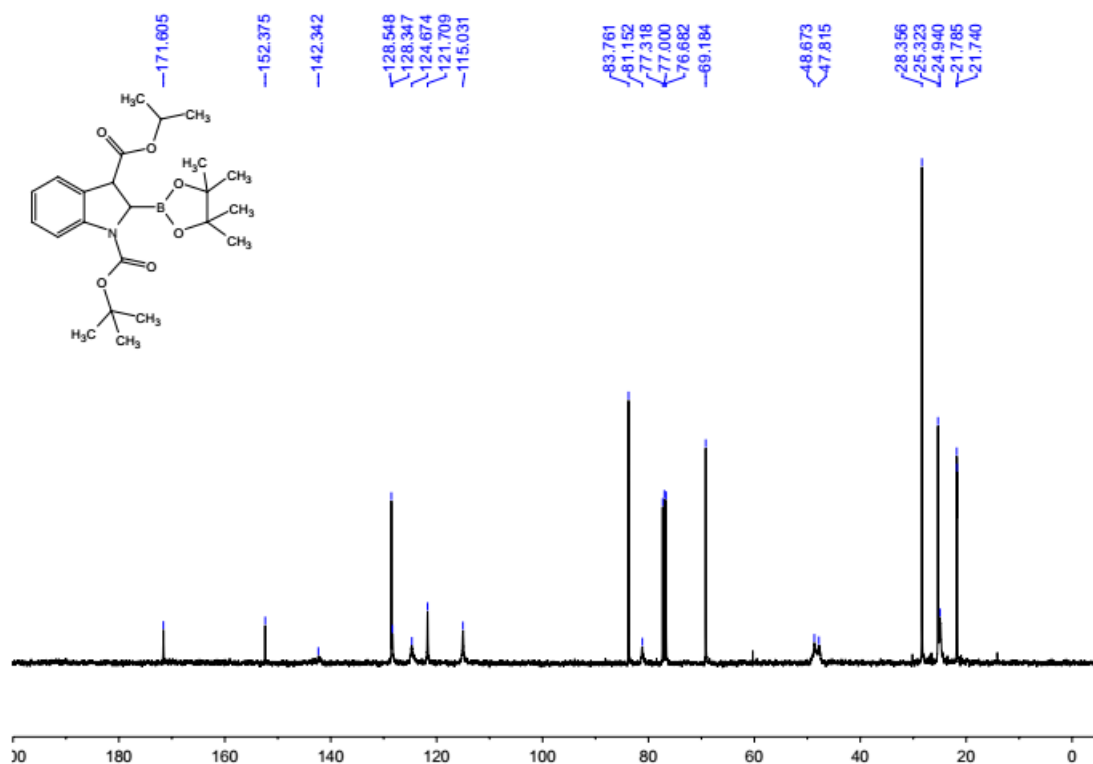
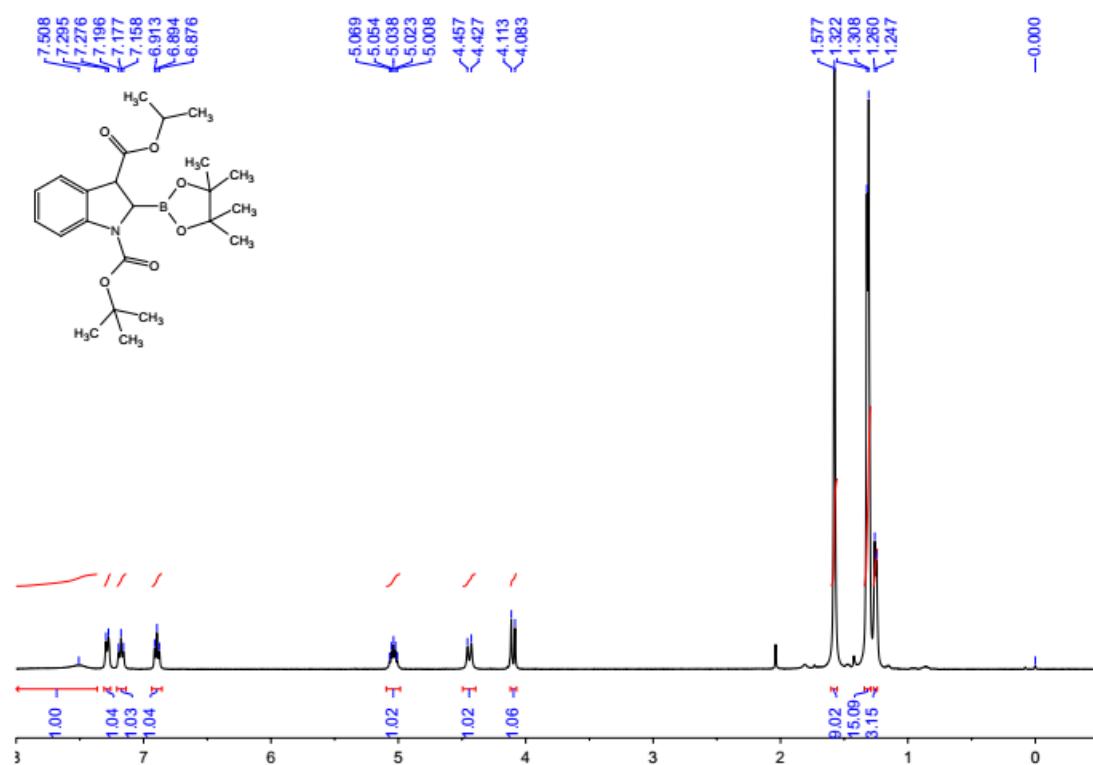


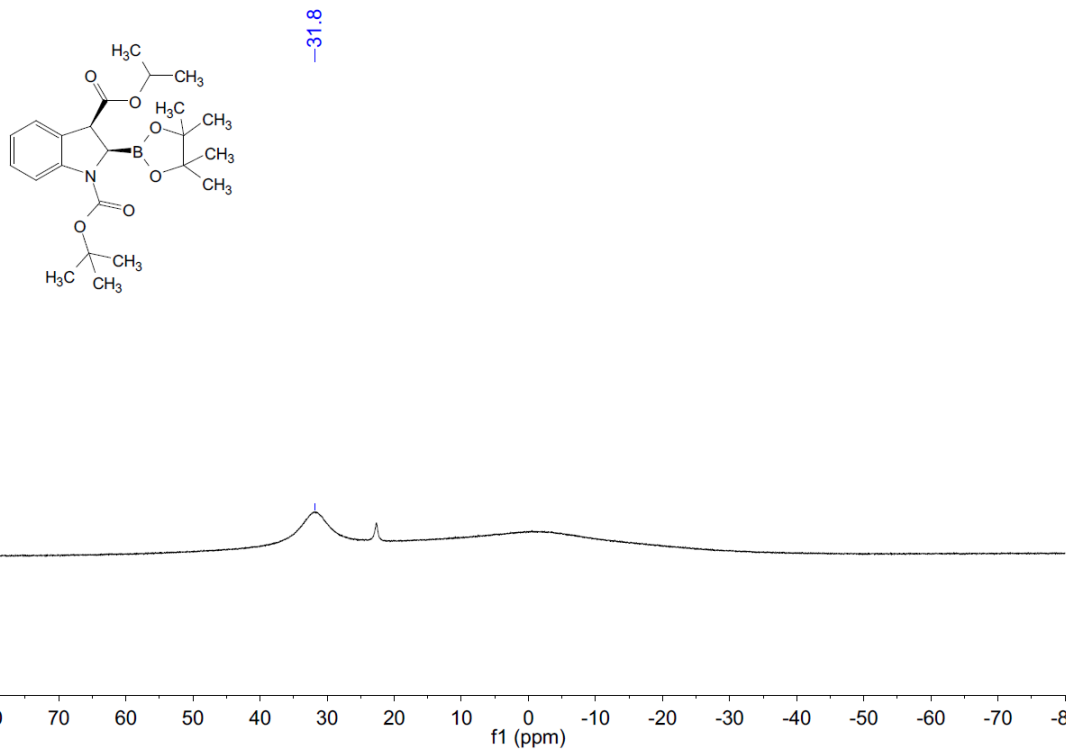
Compound 2b



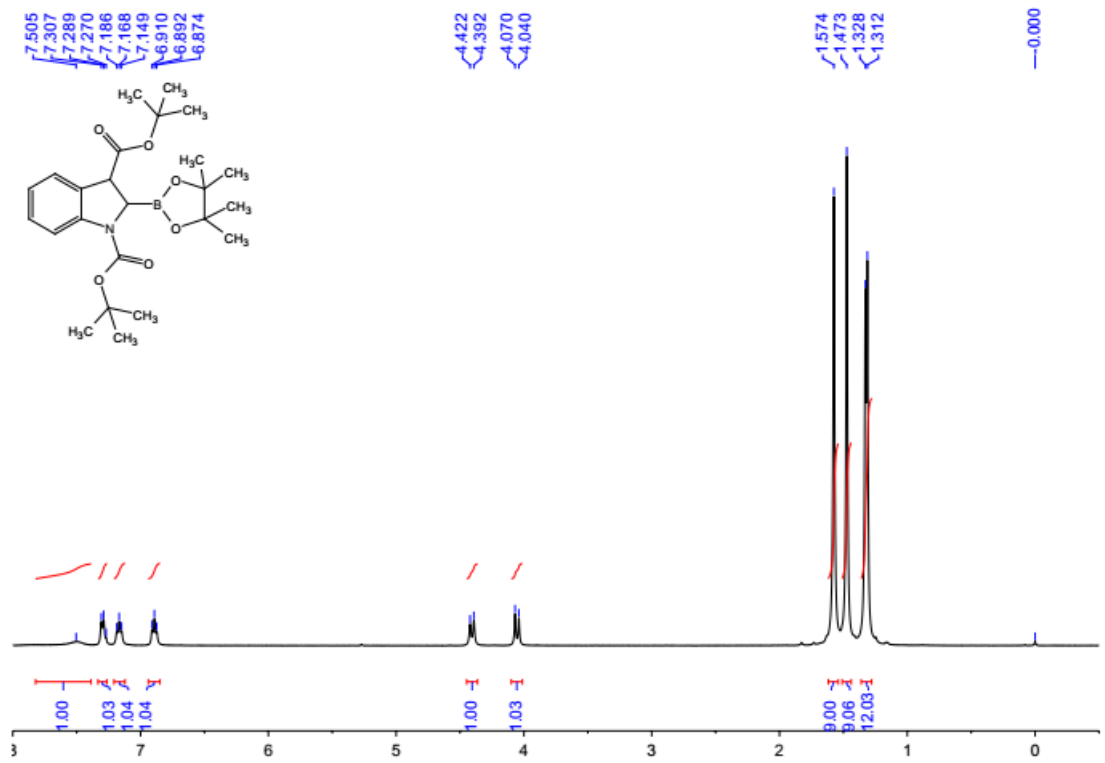


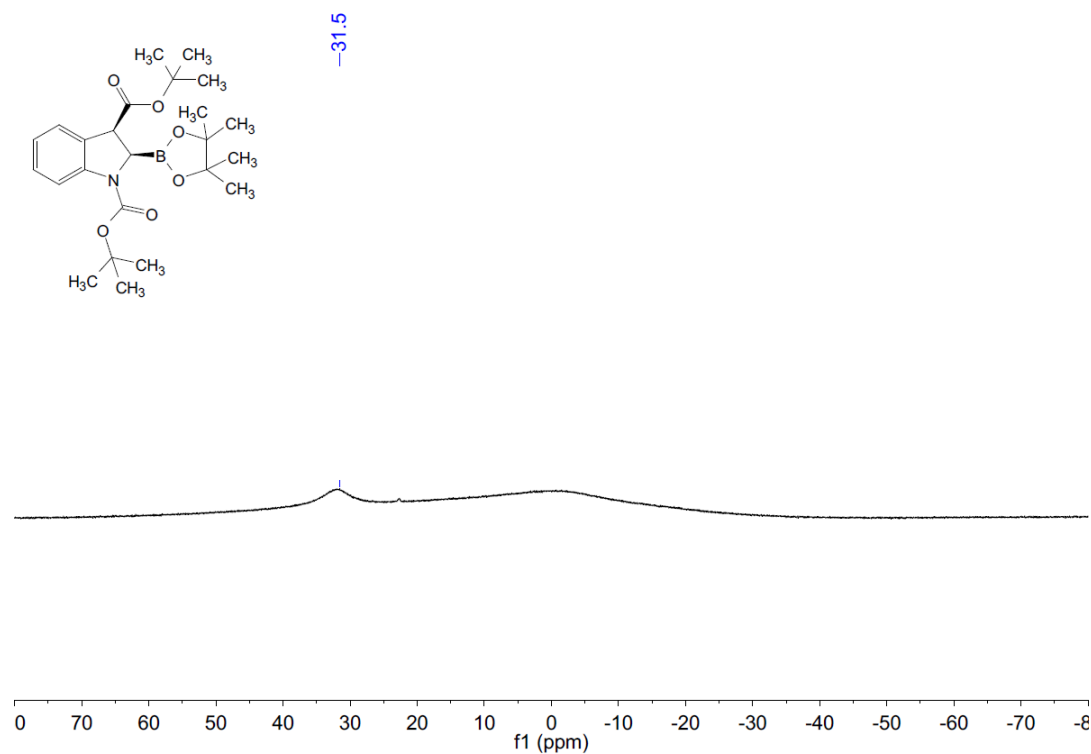
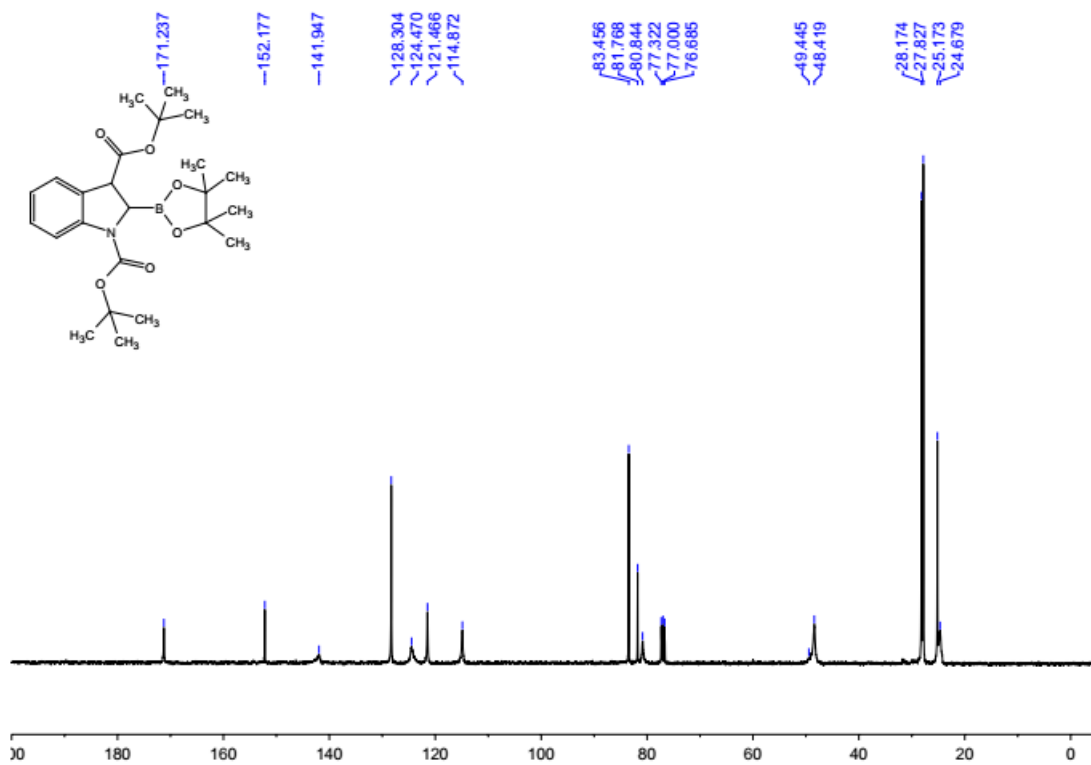
Compound 2c



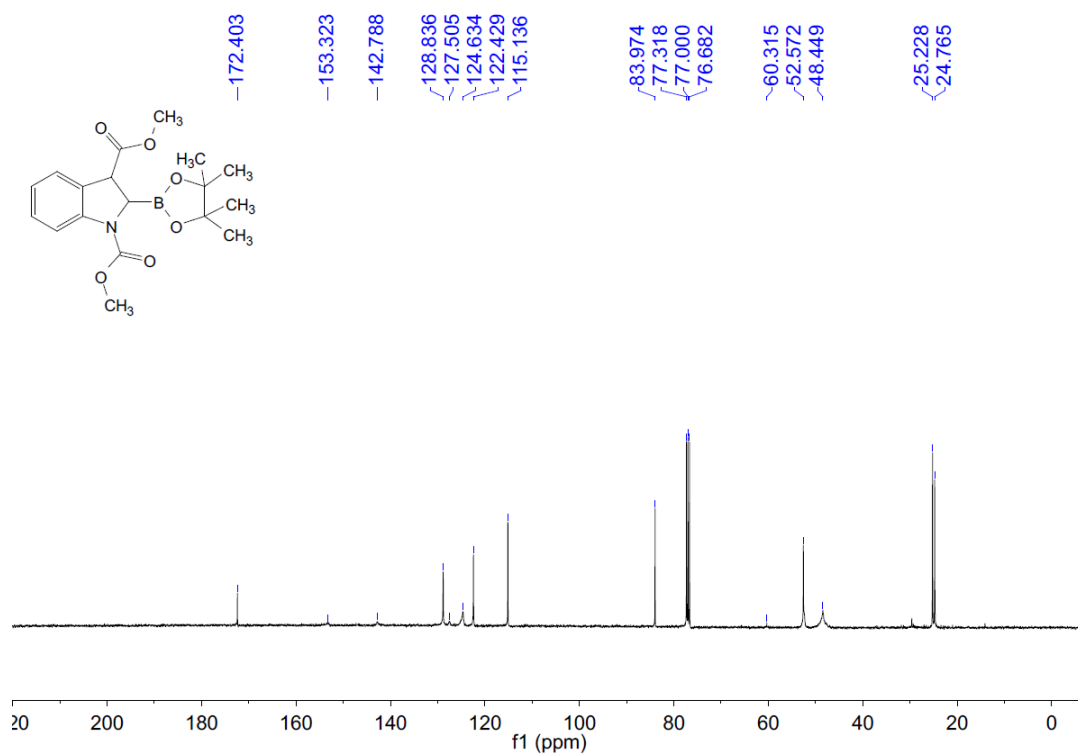
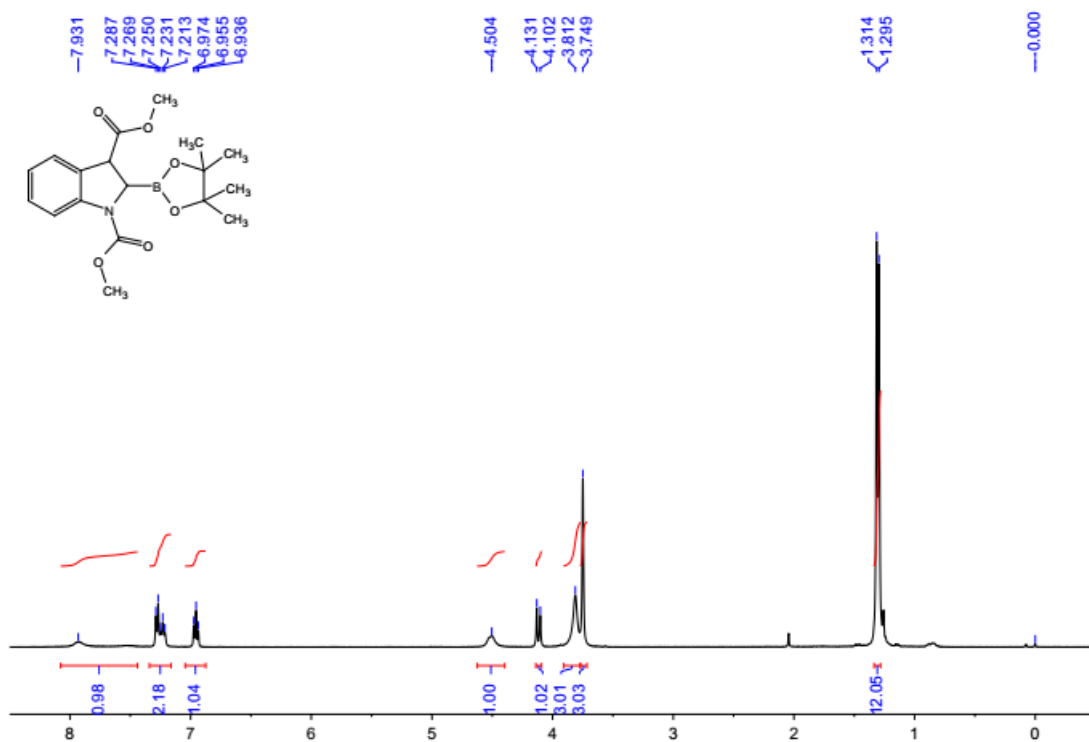


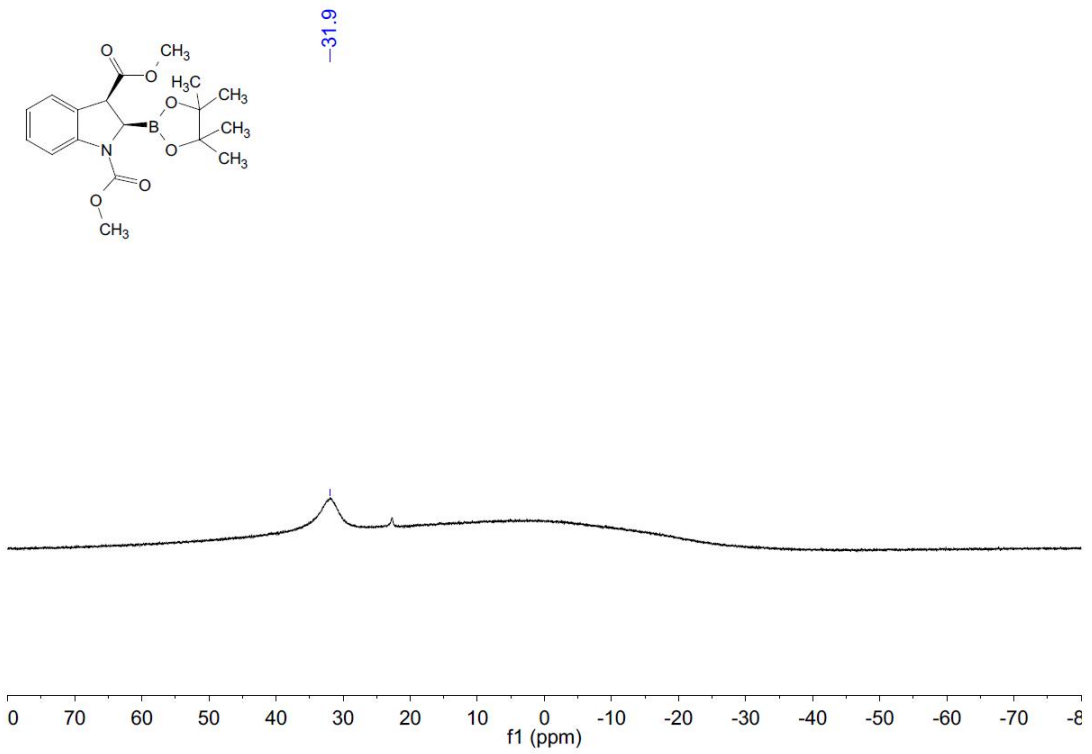
Compound 2d



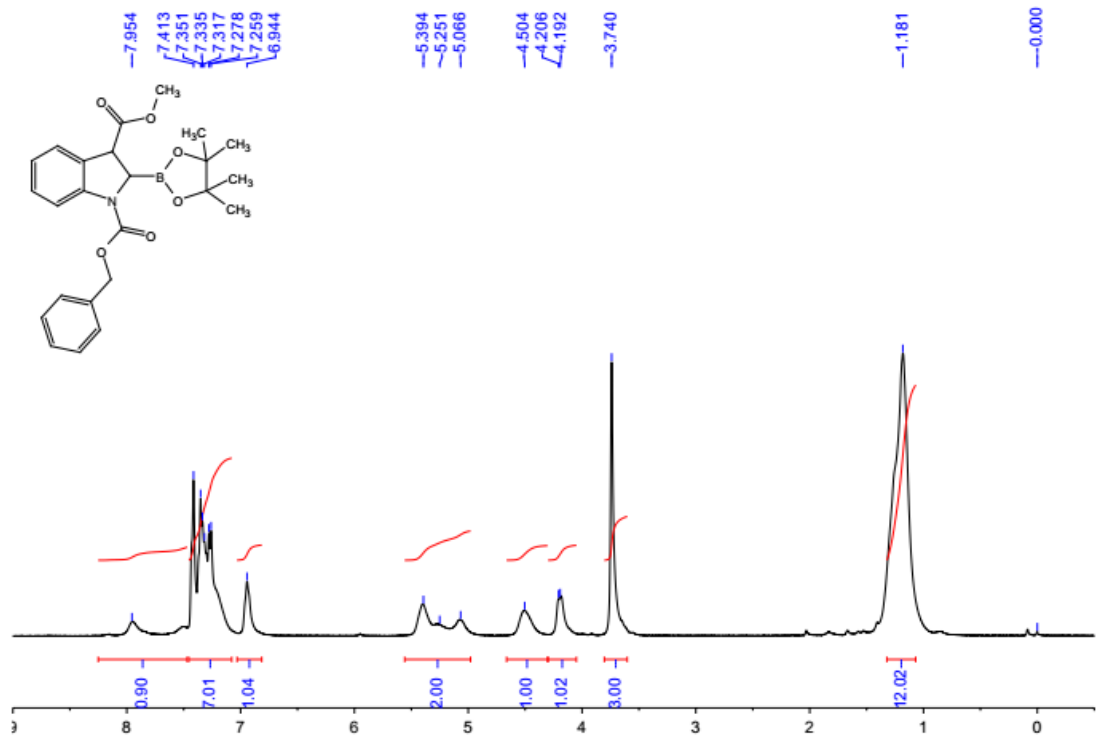


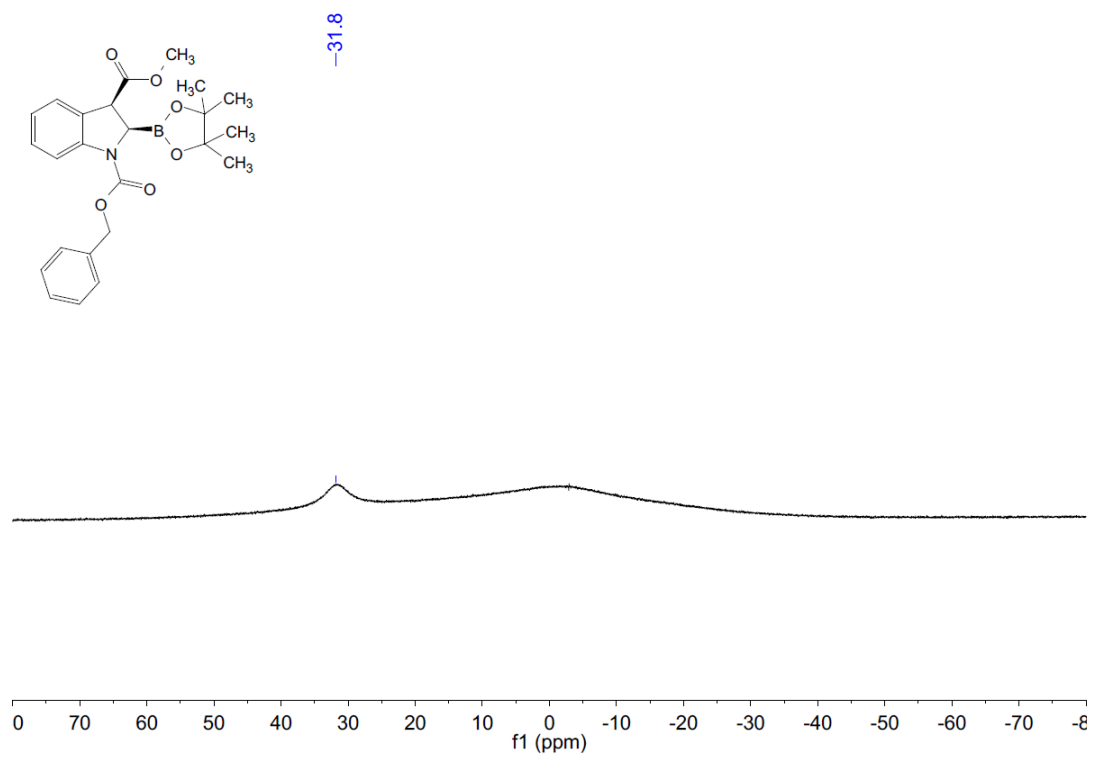
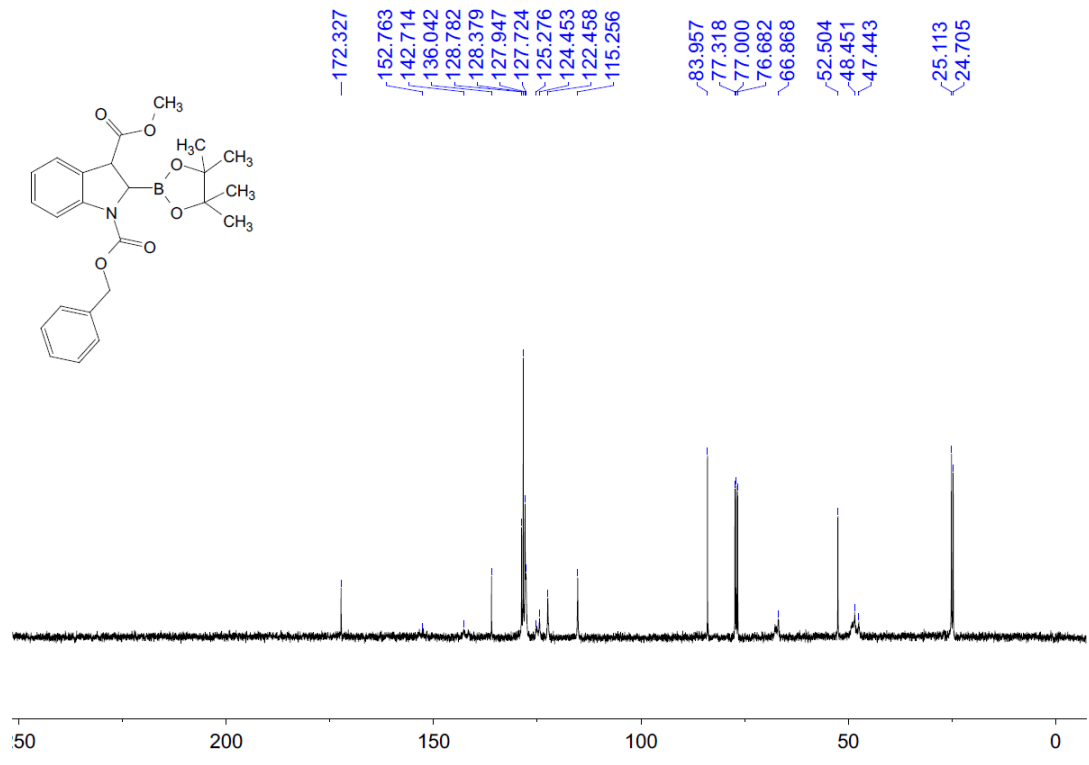
Compound 2e



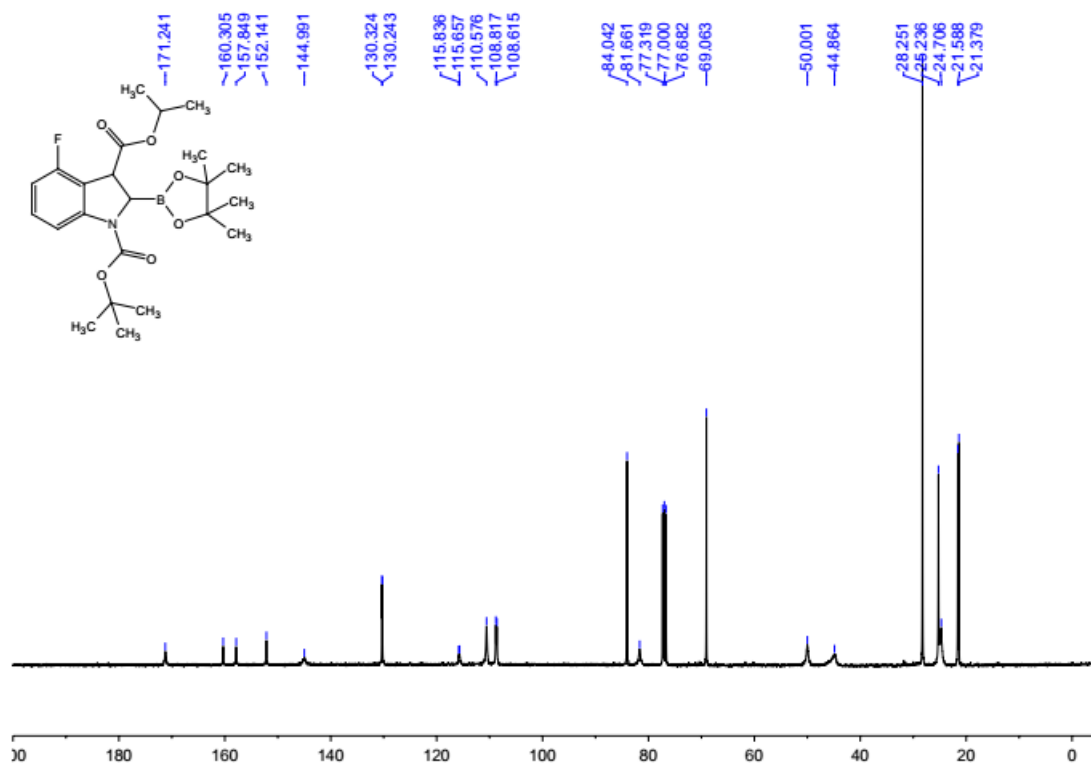
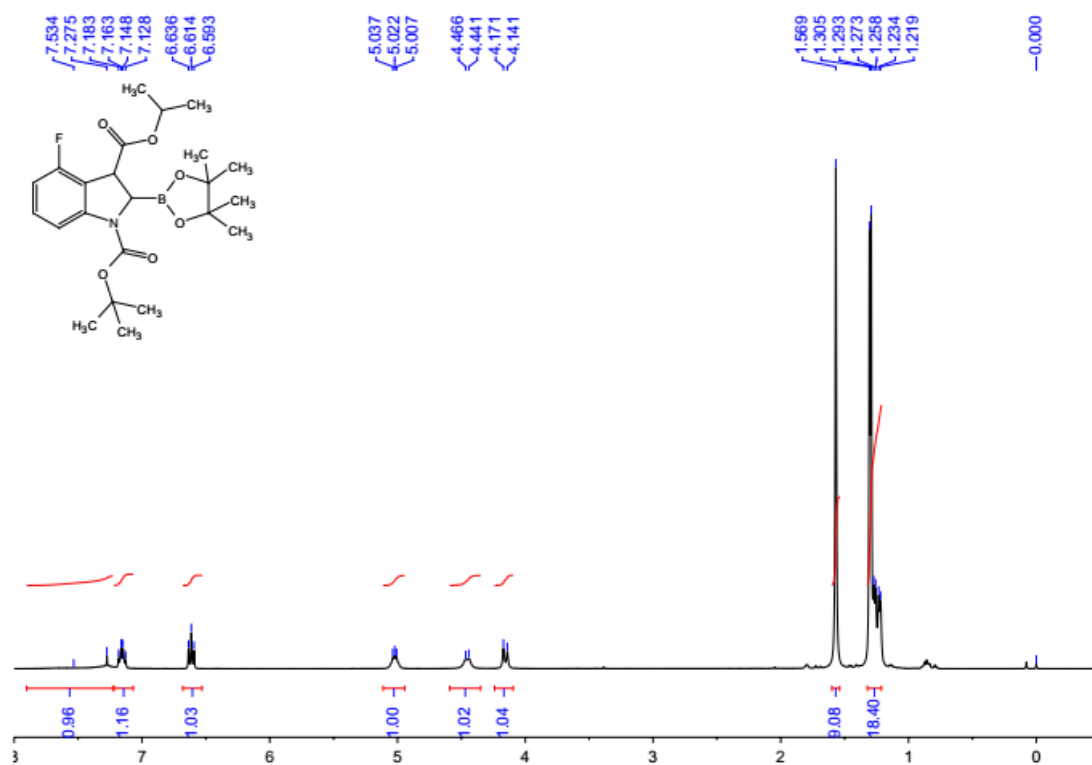


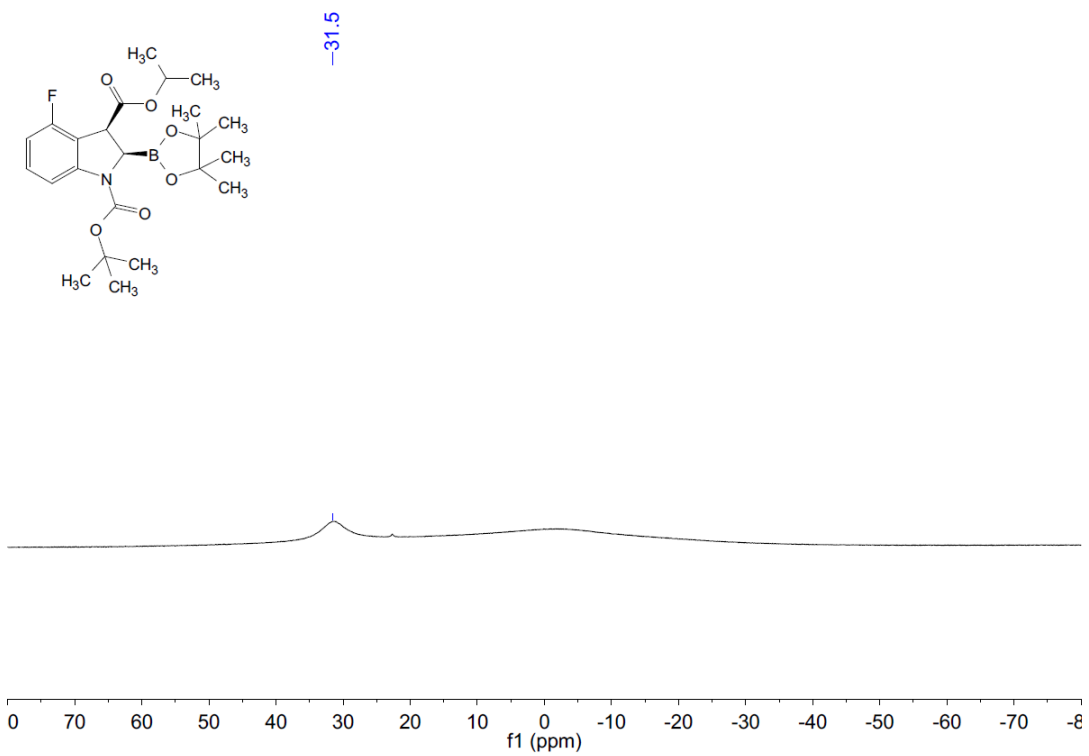
Compound 2f



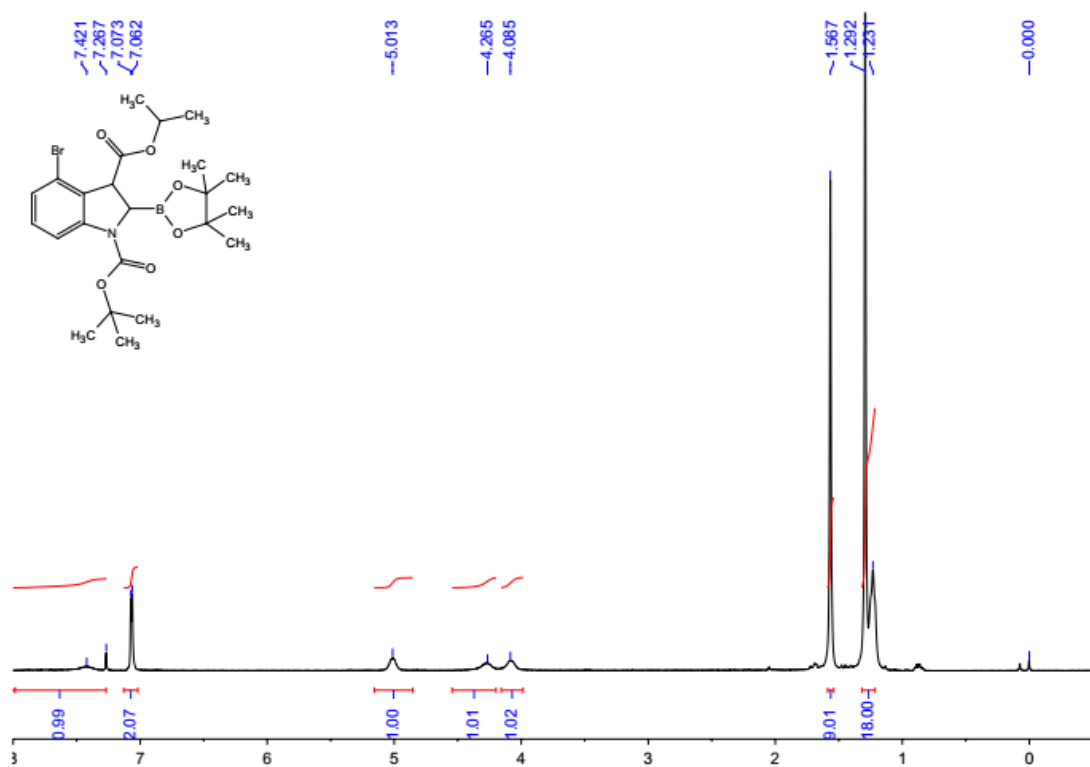


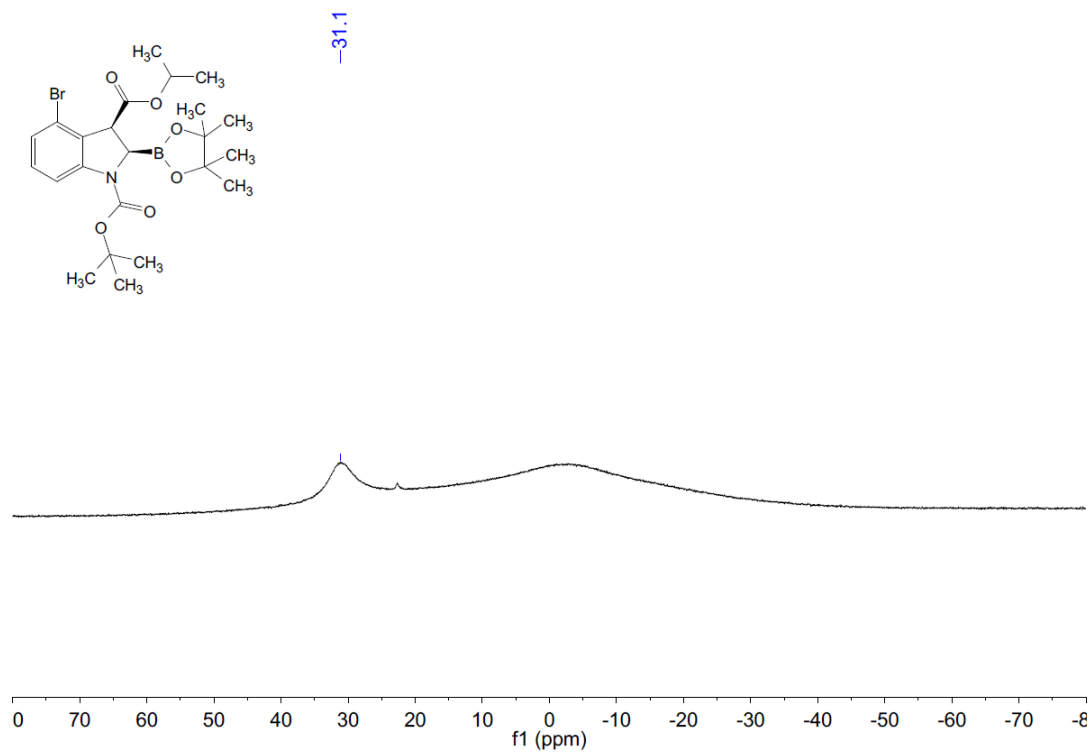
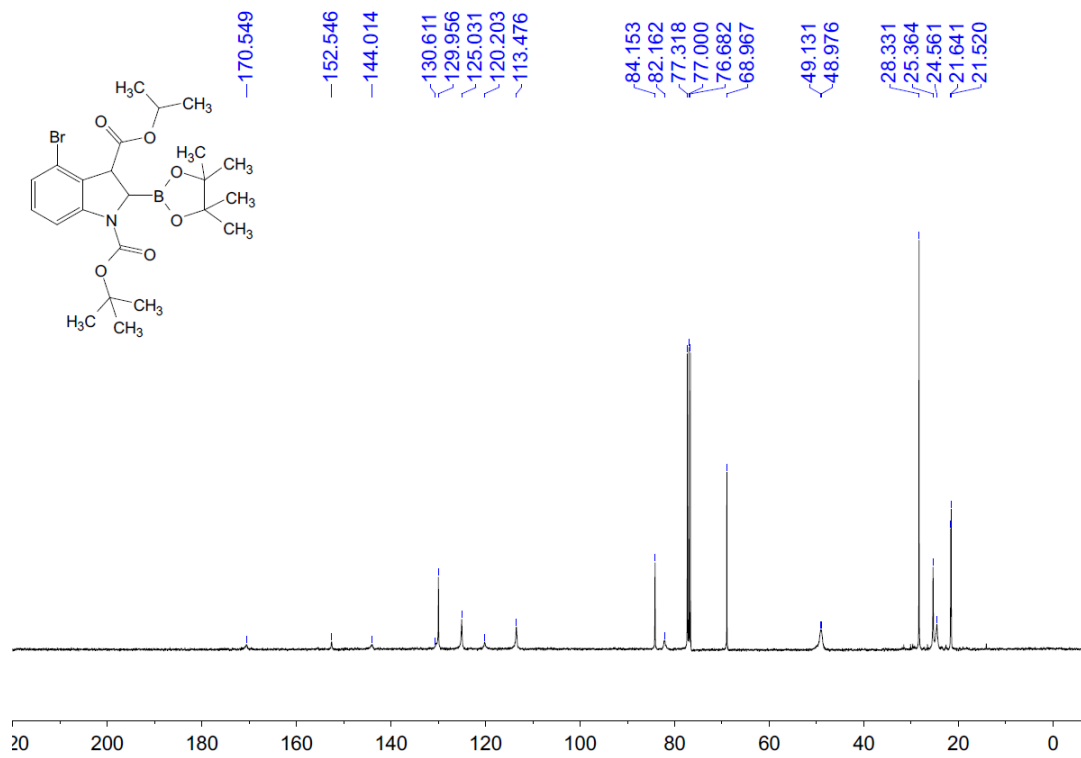
Compound 2g



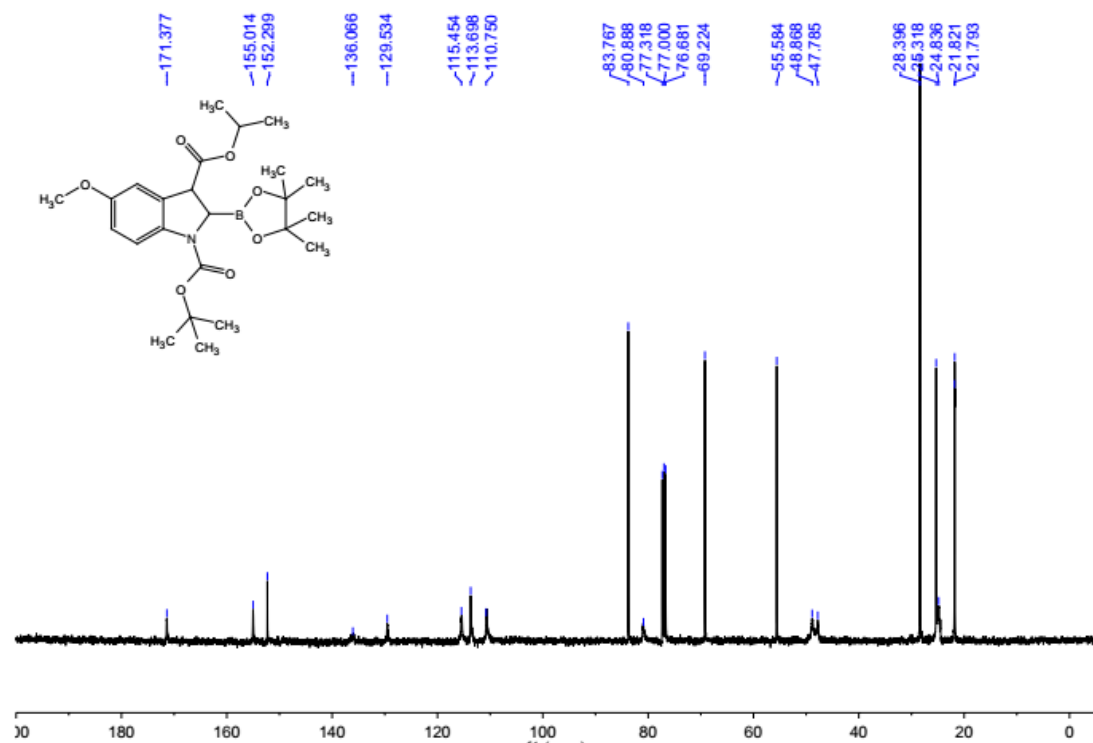
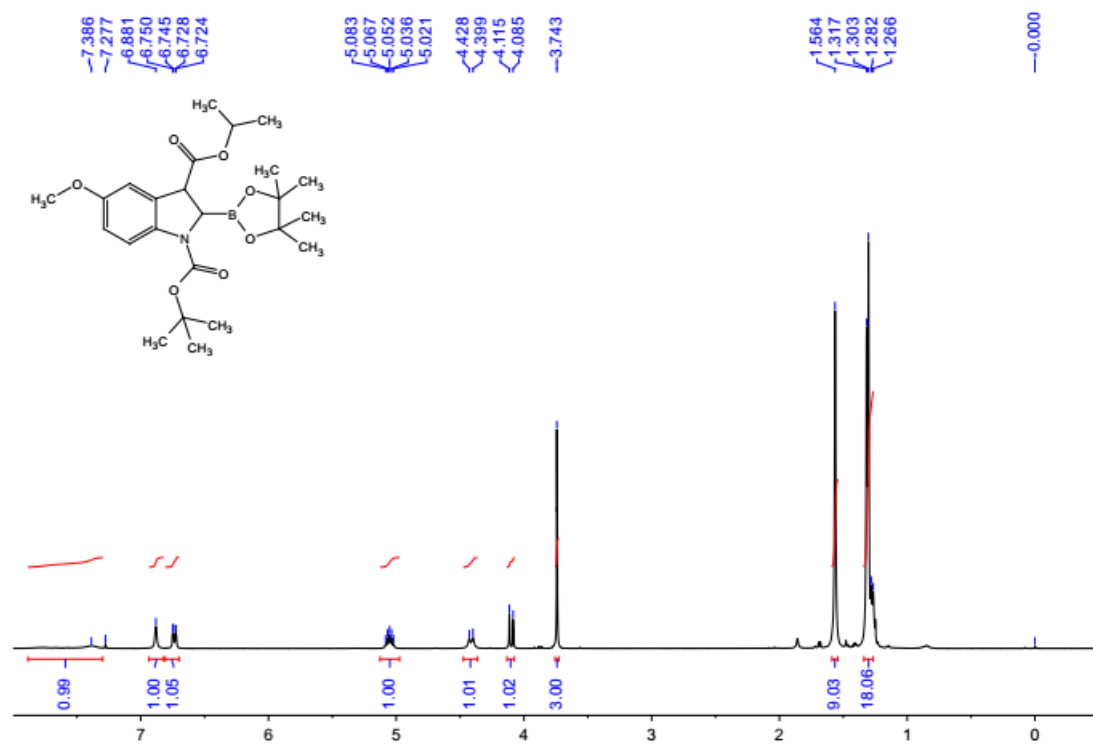


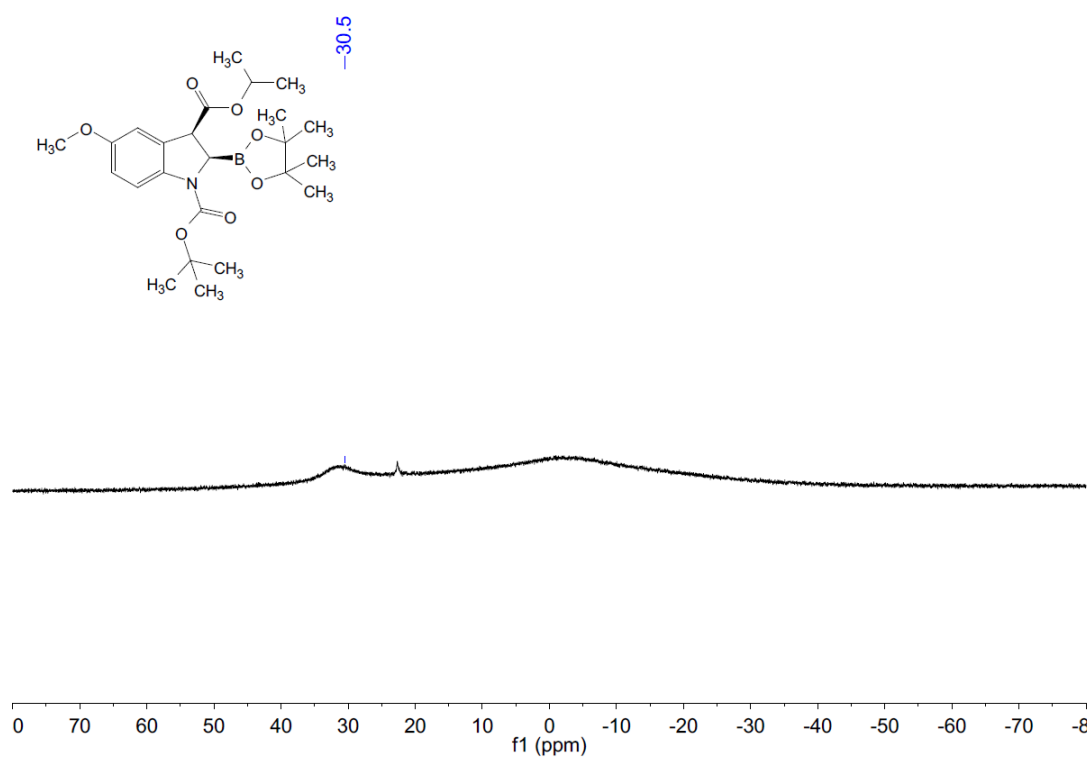
Compound 2h



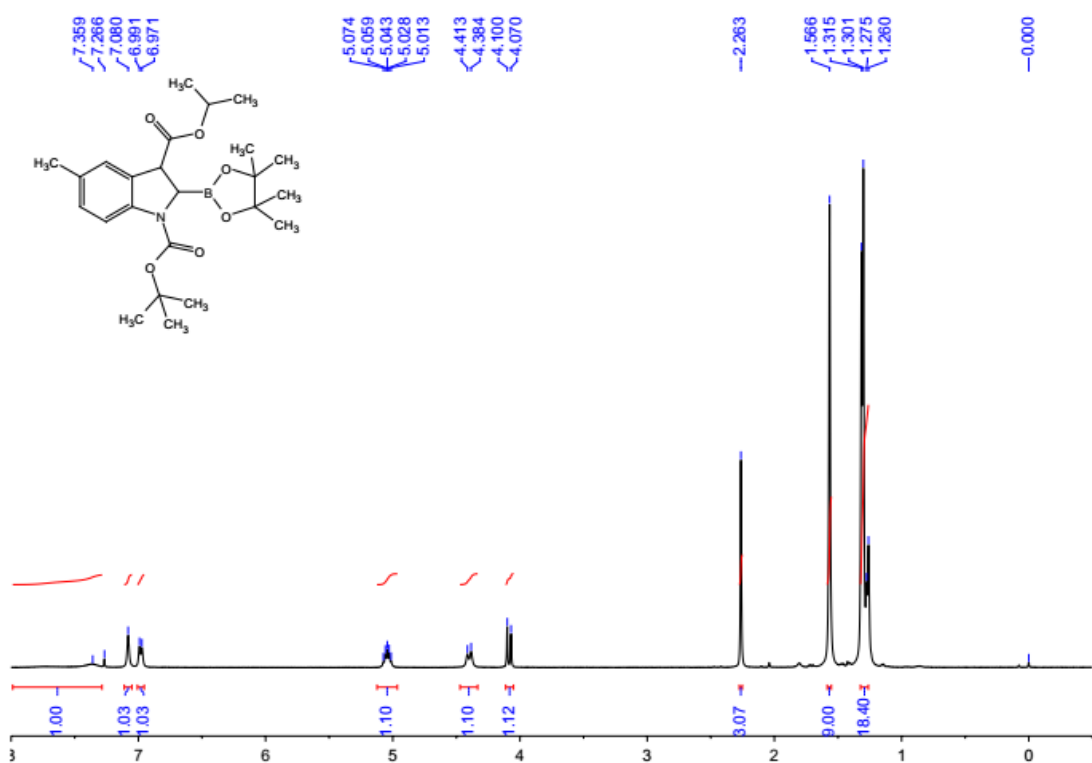


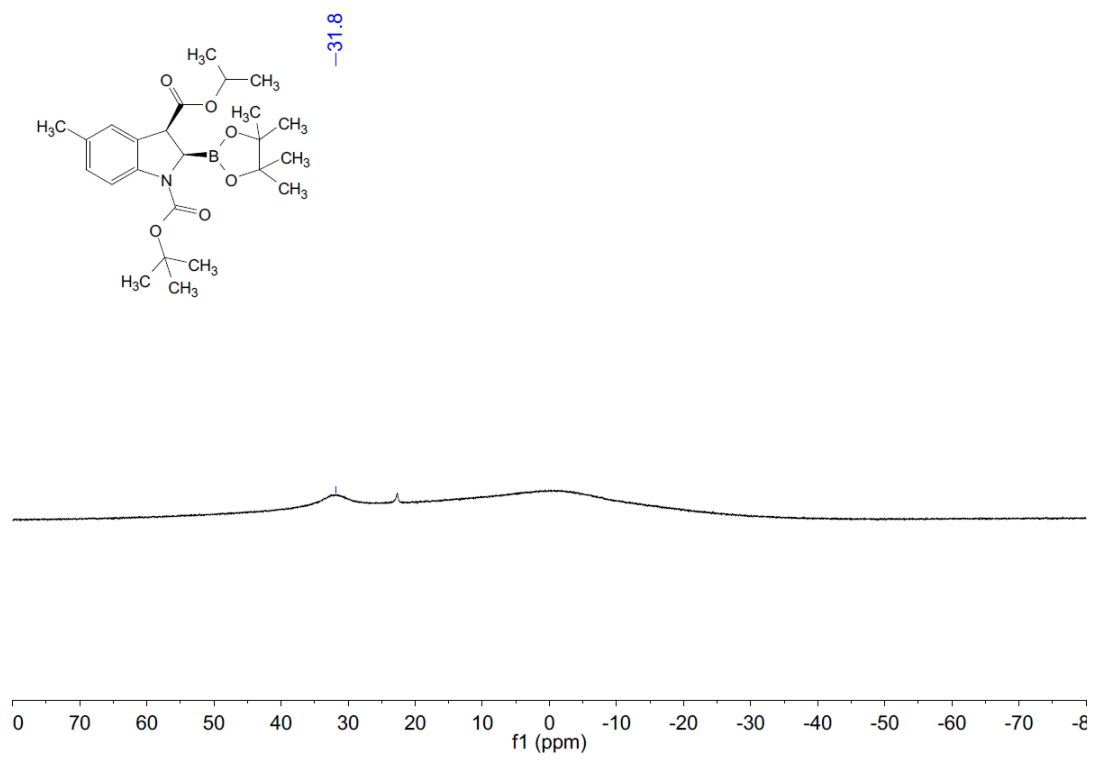
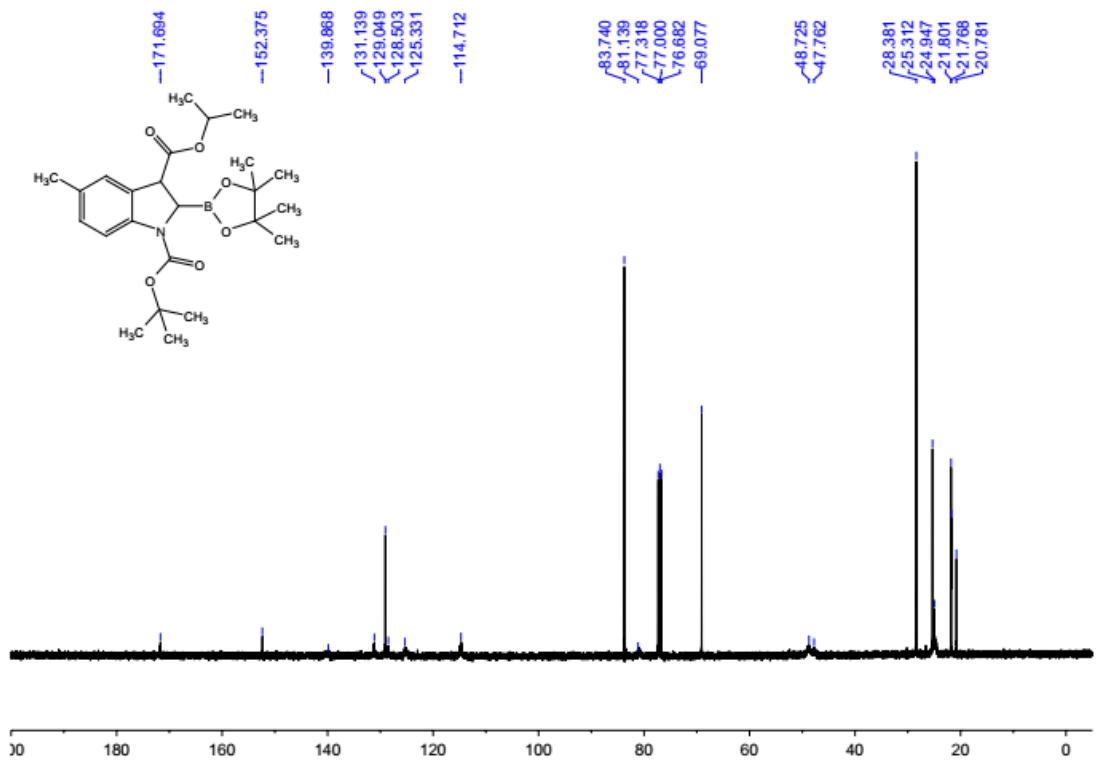
Compound 2i



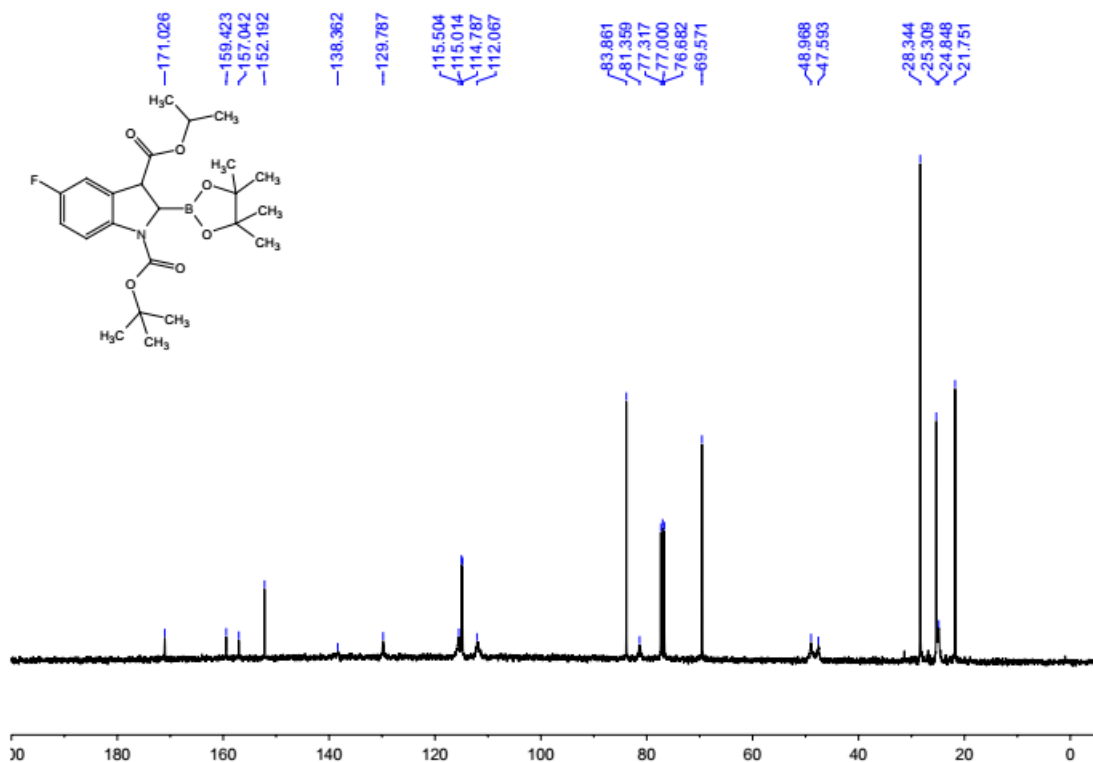
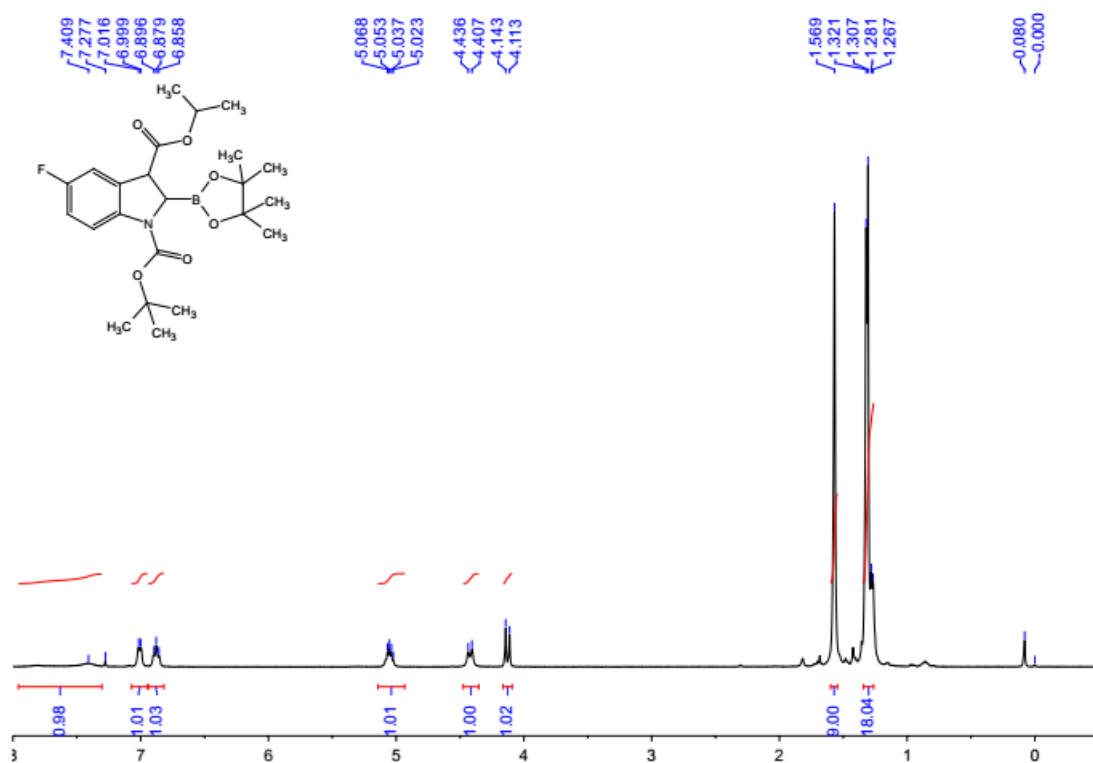


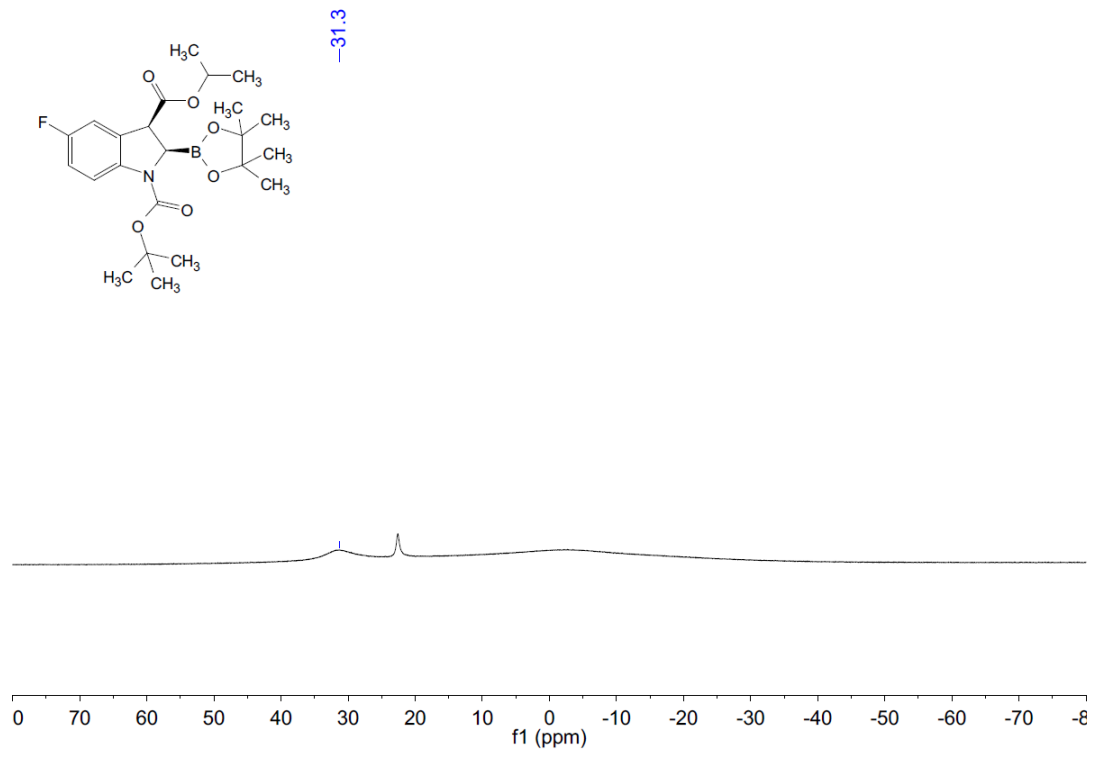
Compound 2j



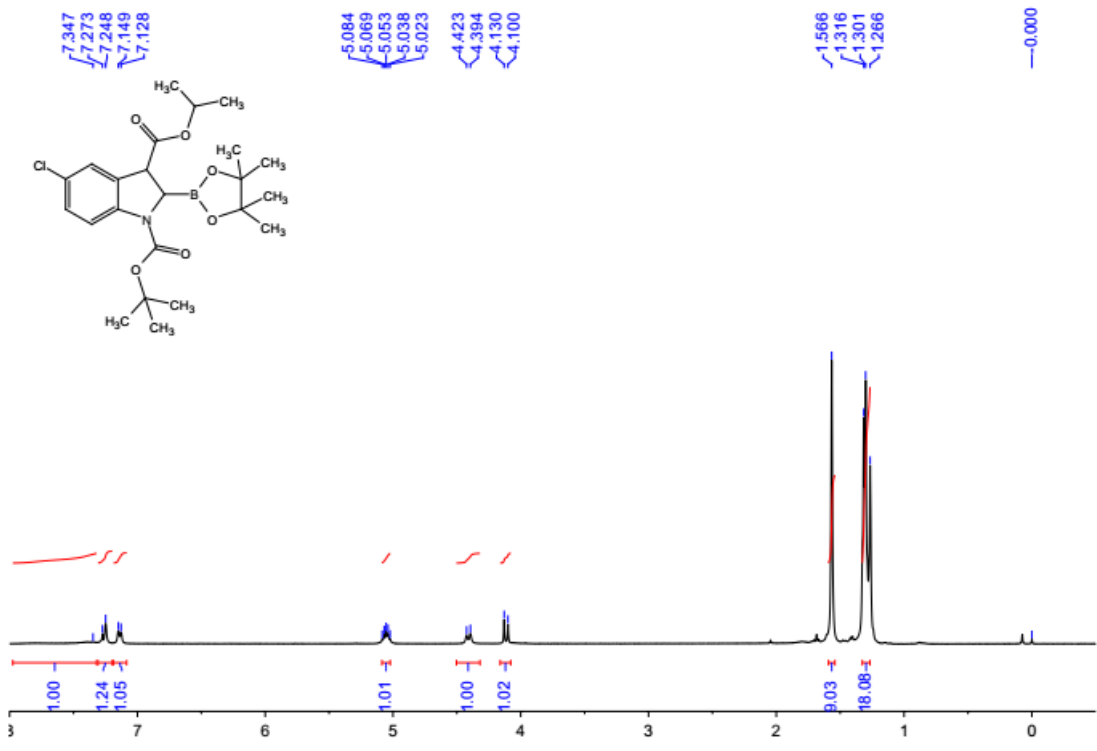


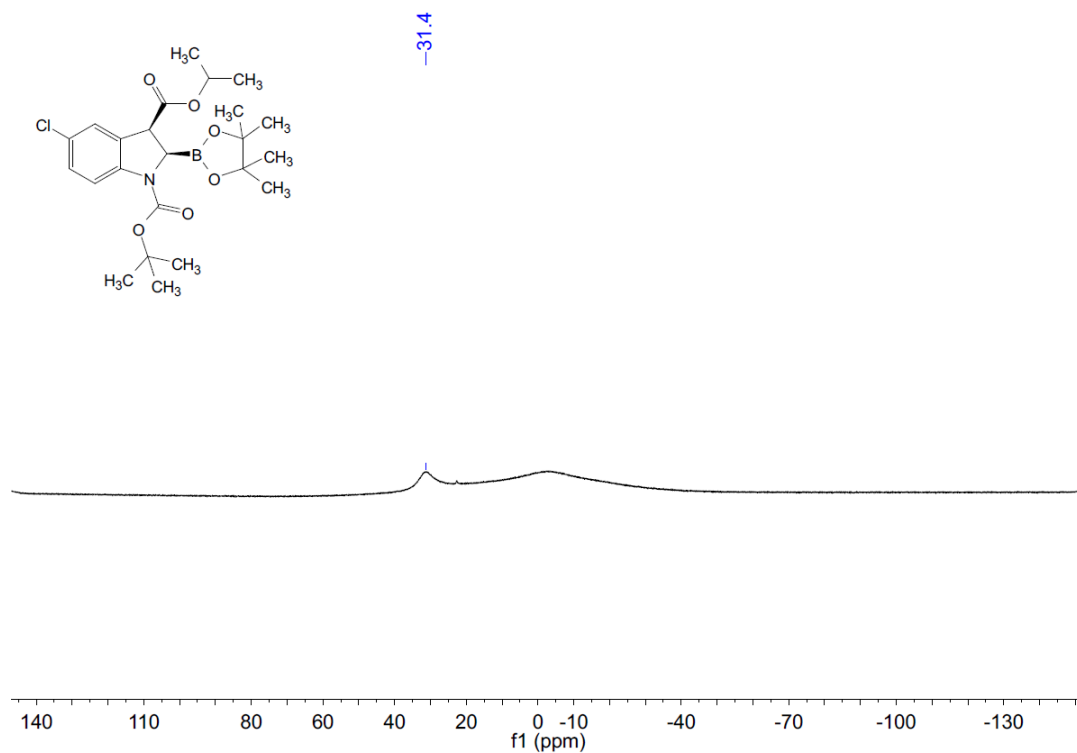
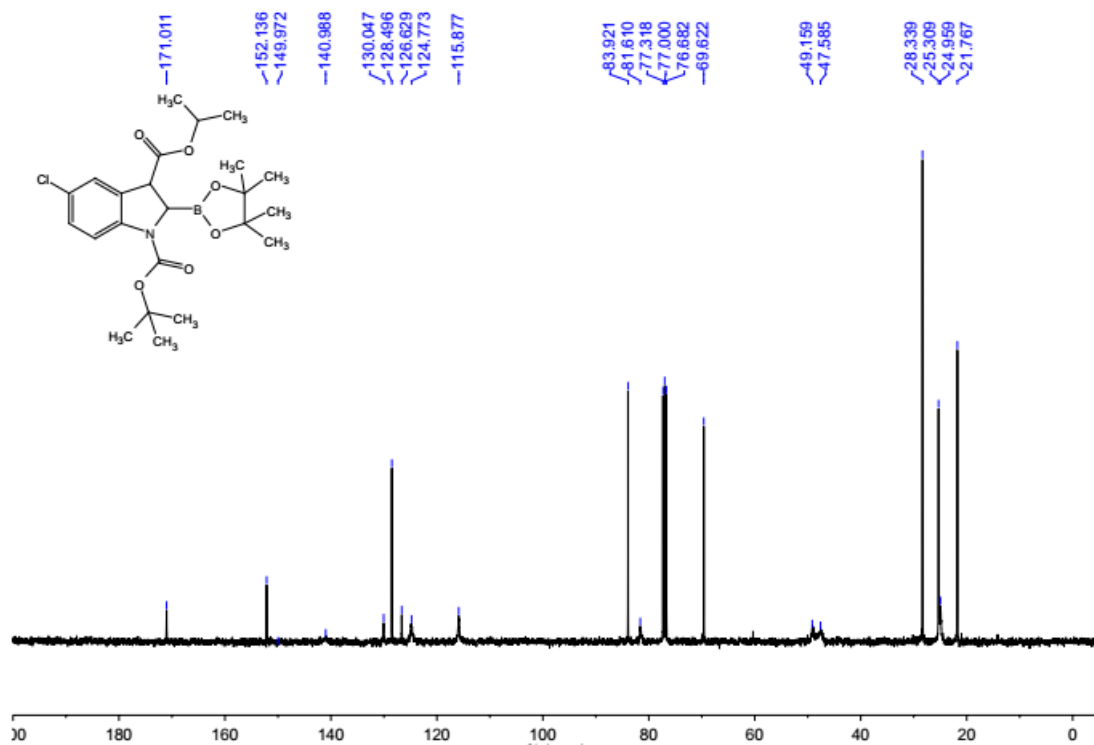
Compound 2k



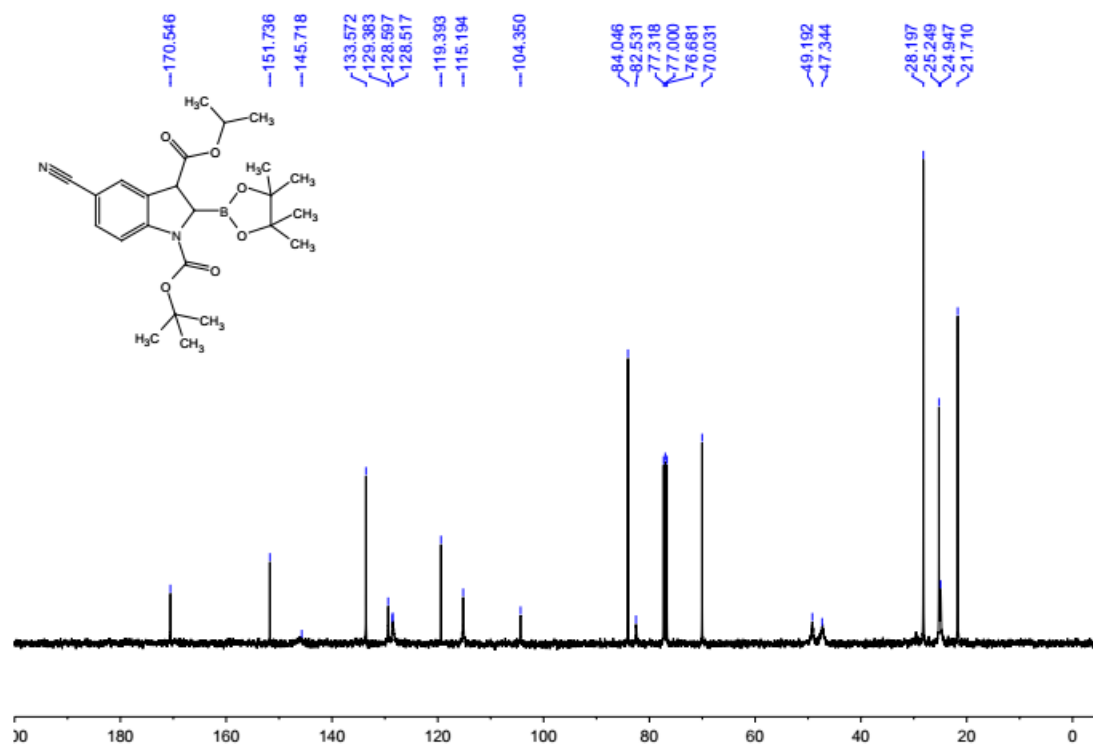
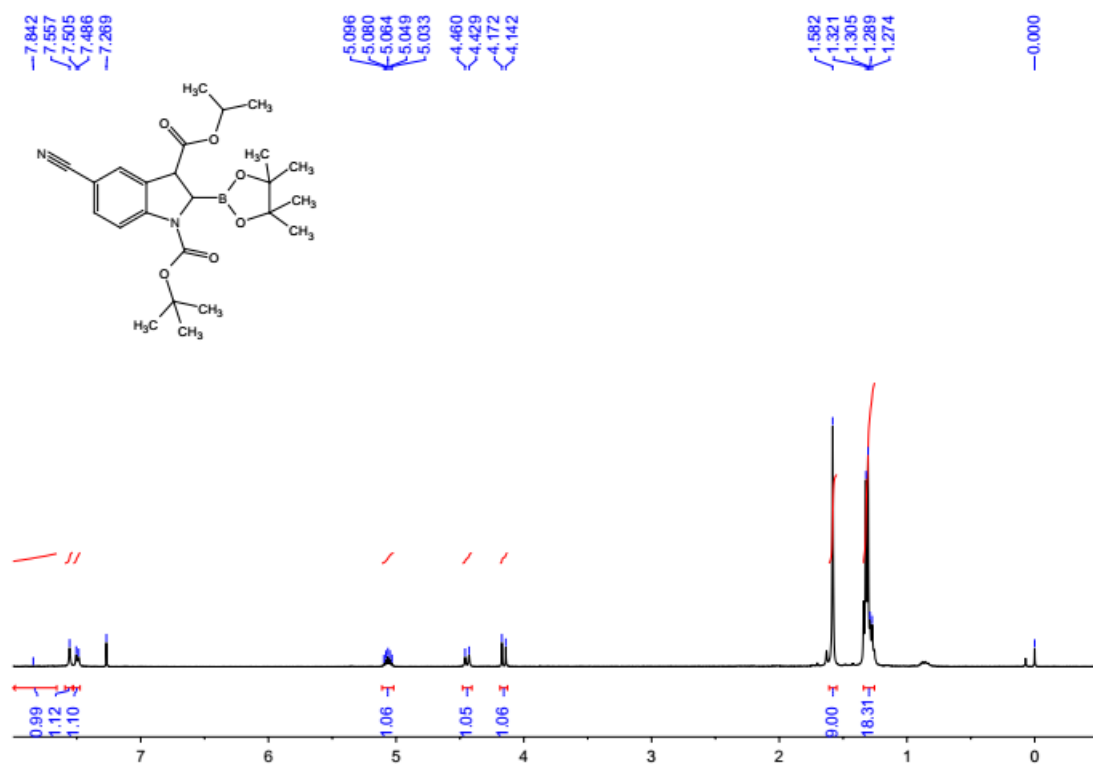


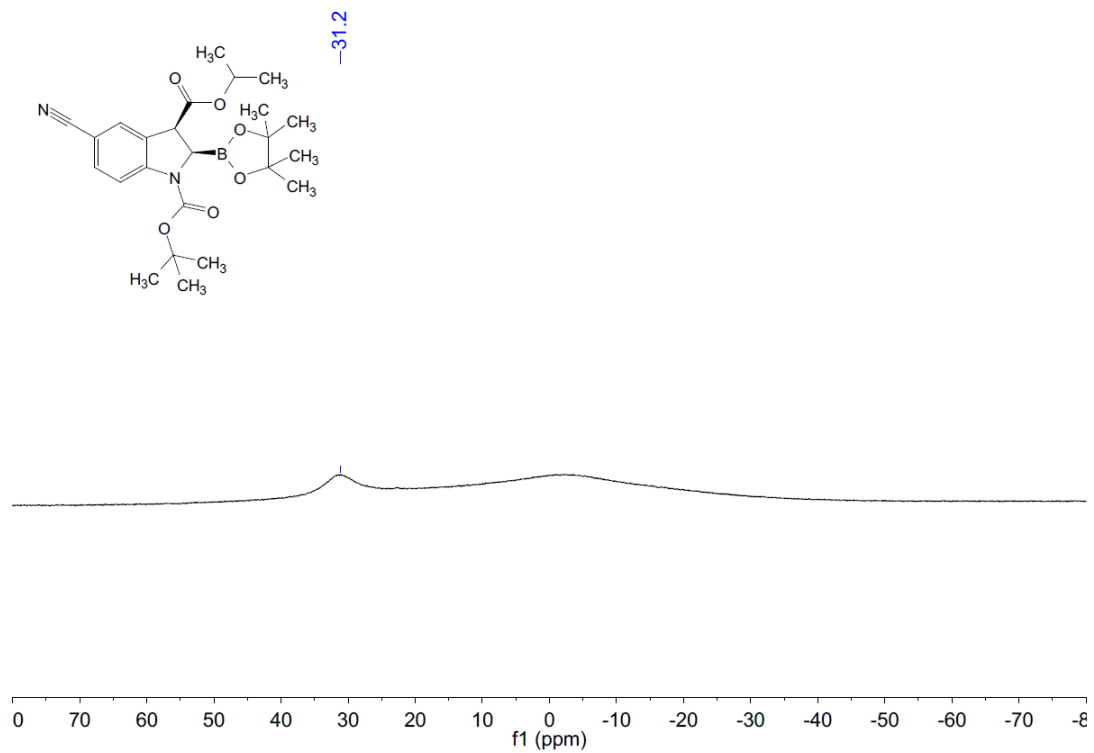
Compound 2l



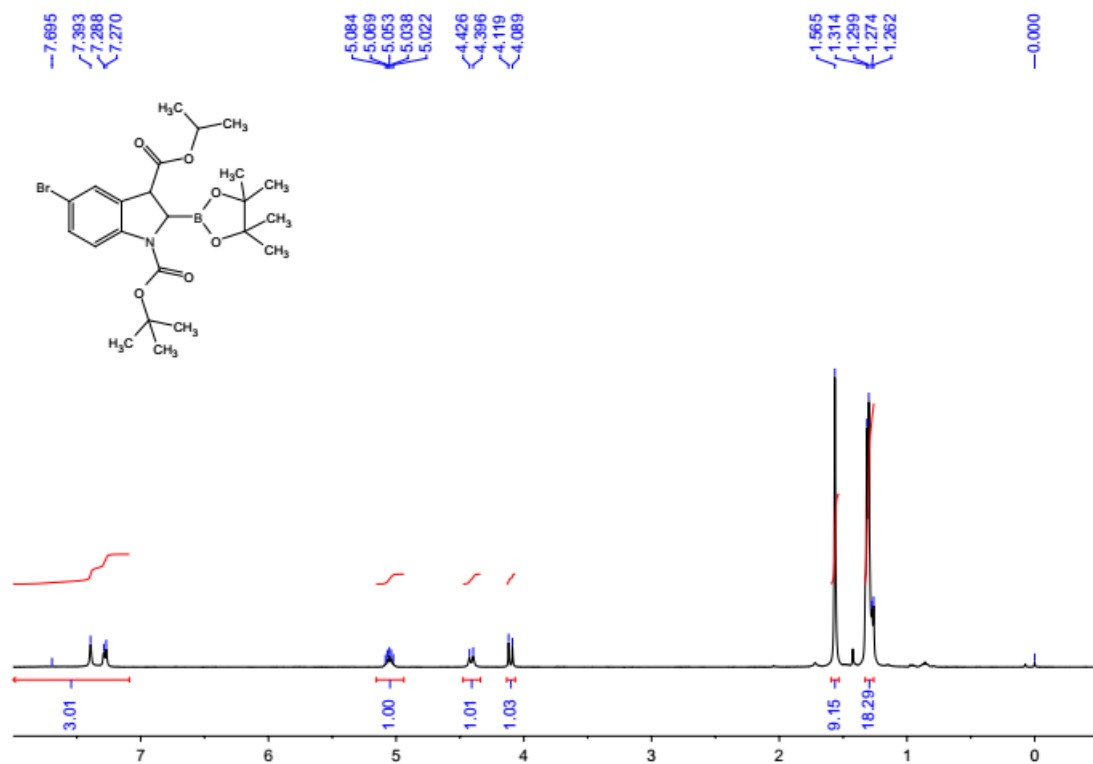


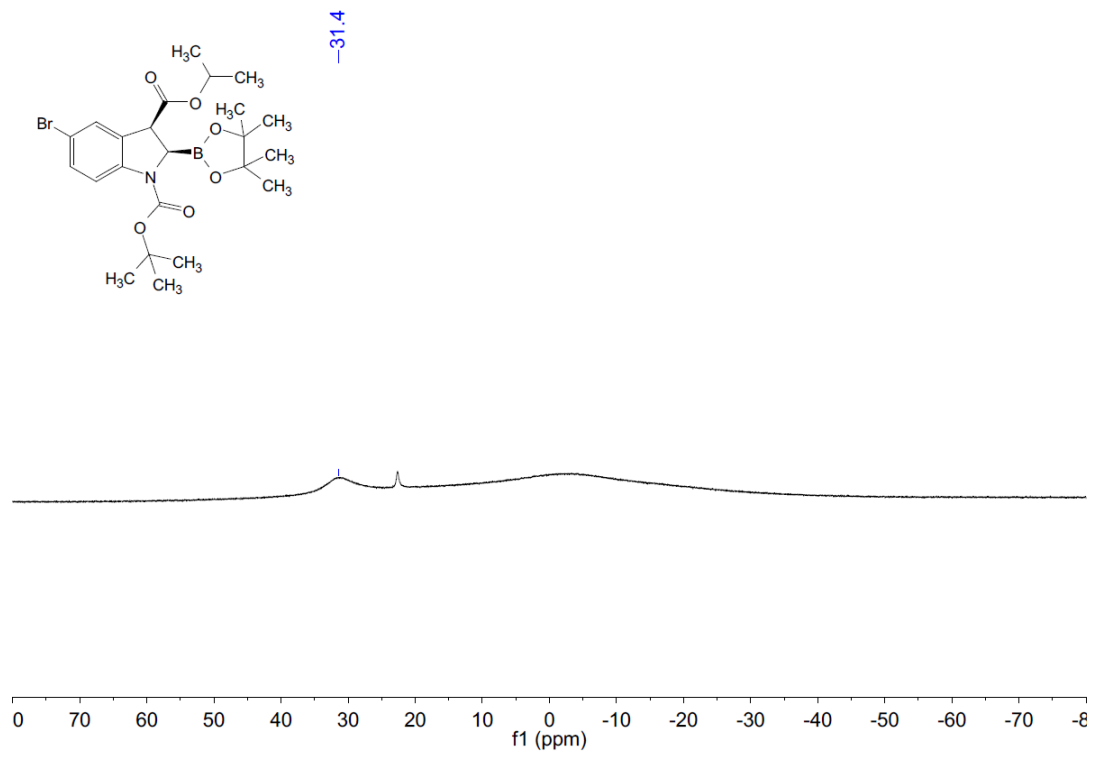
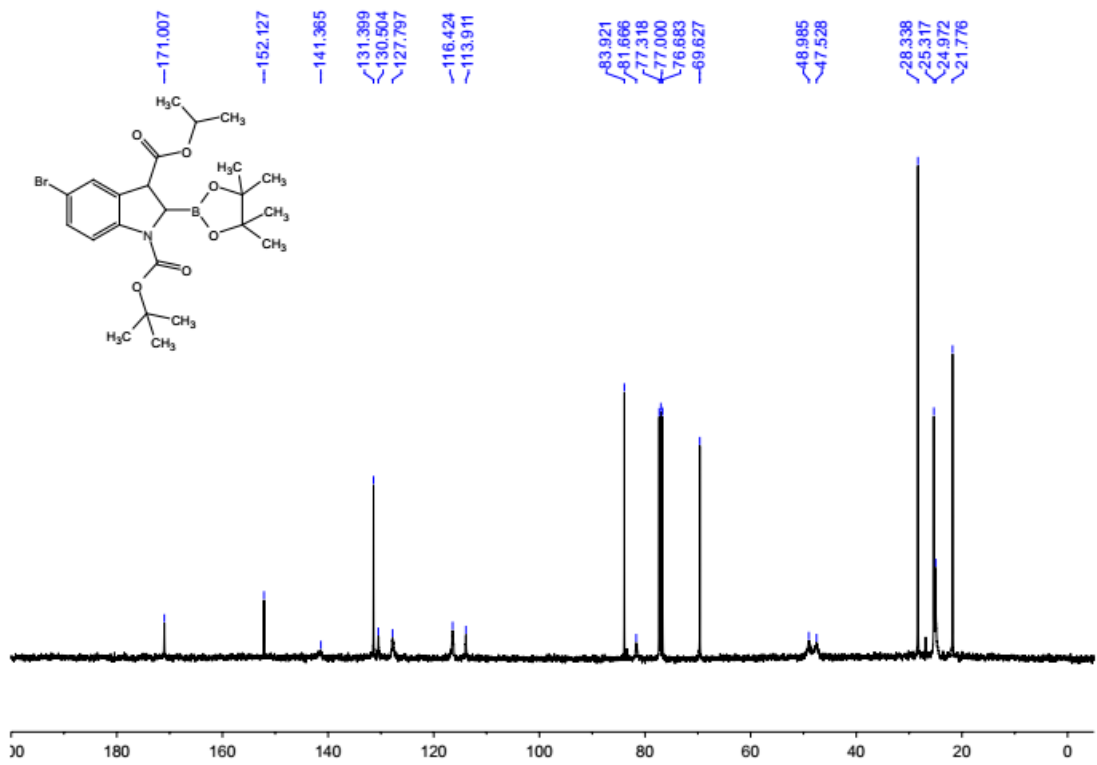
Compound 2m



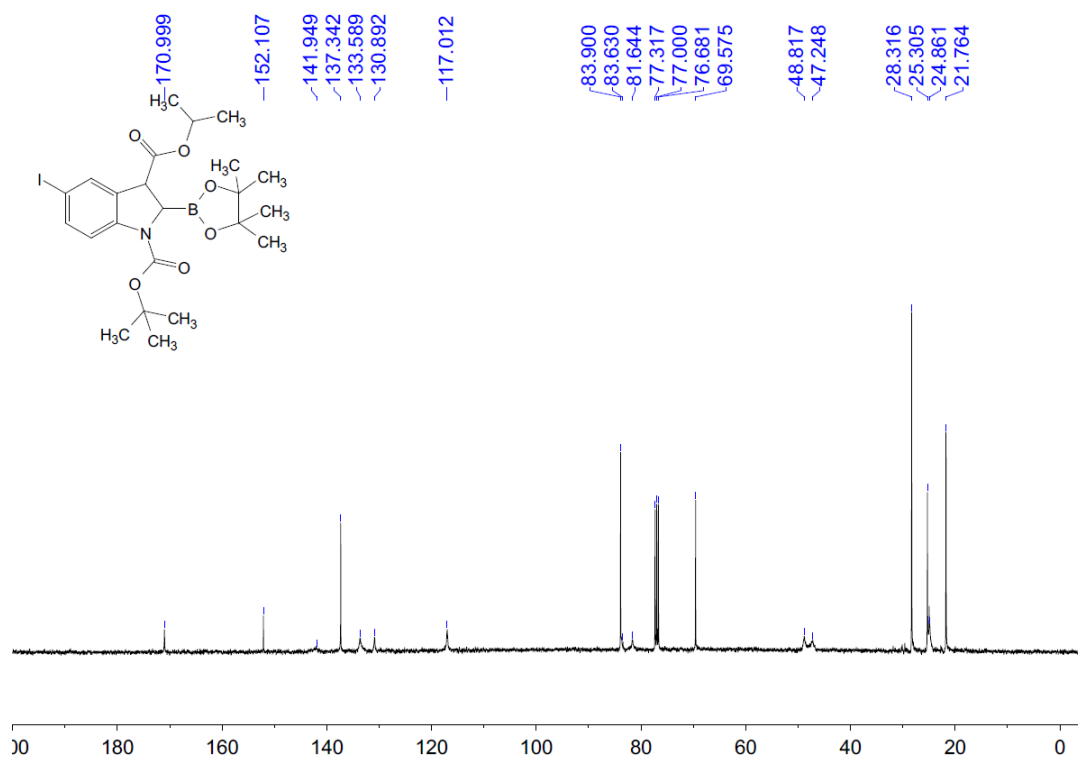
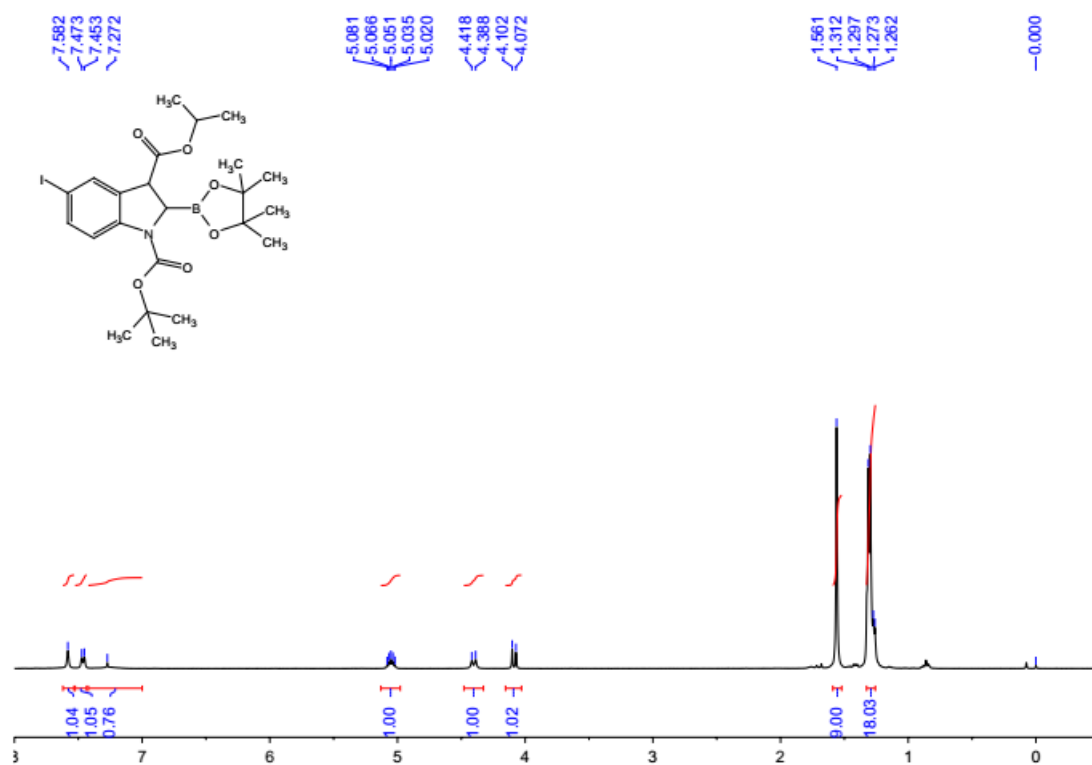


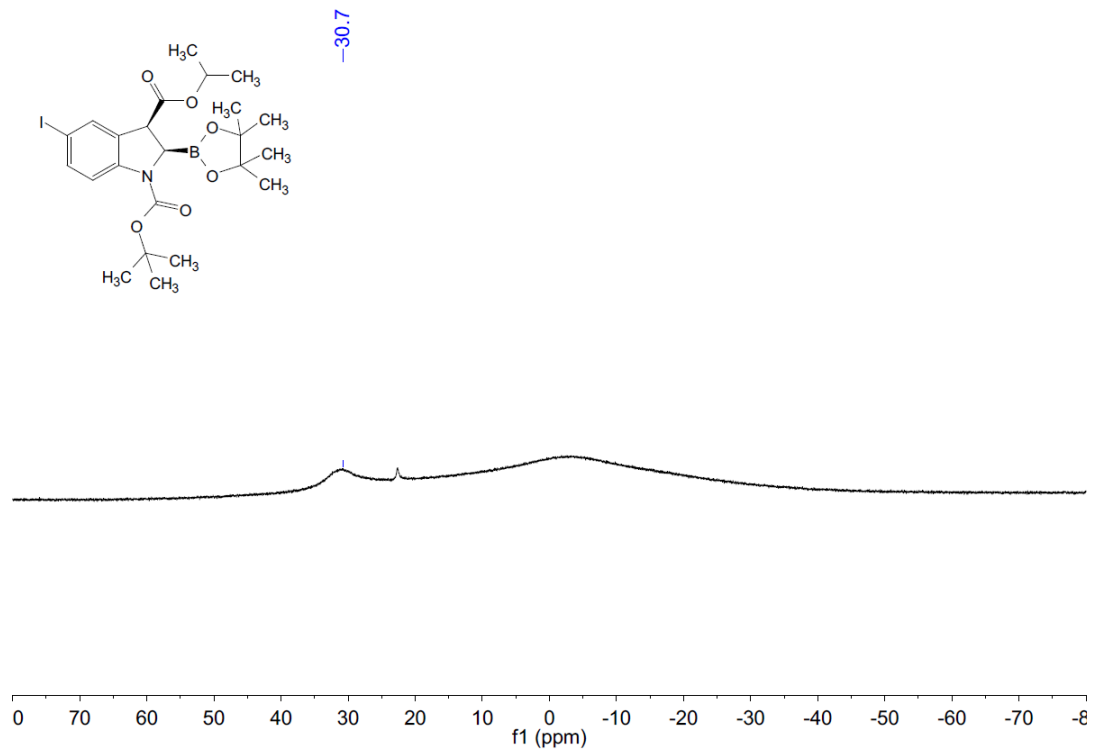
Compound 2n



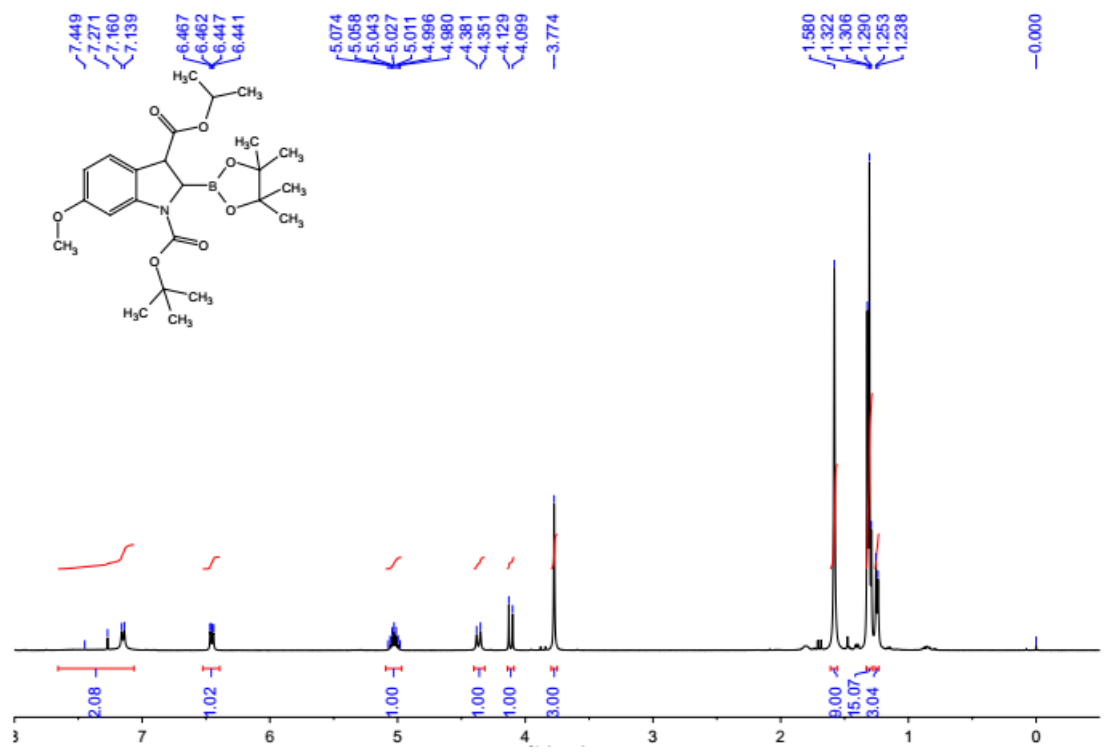


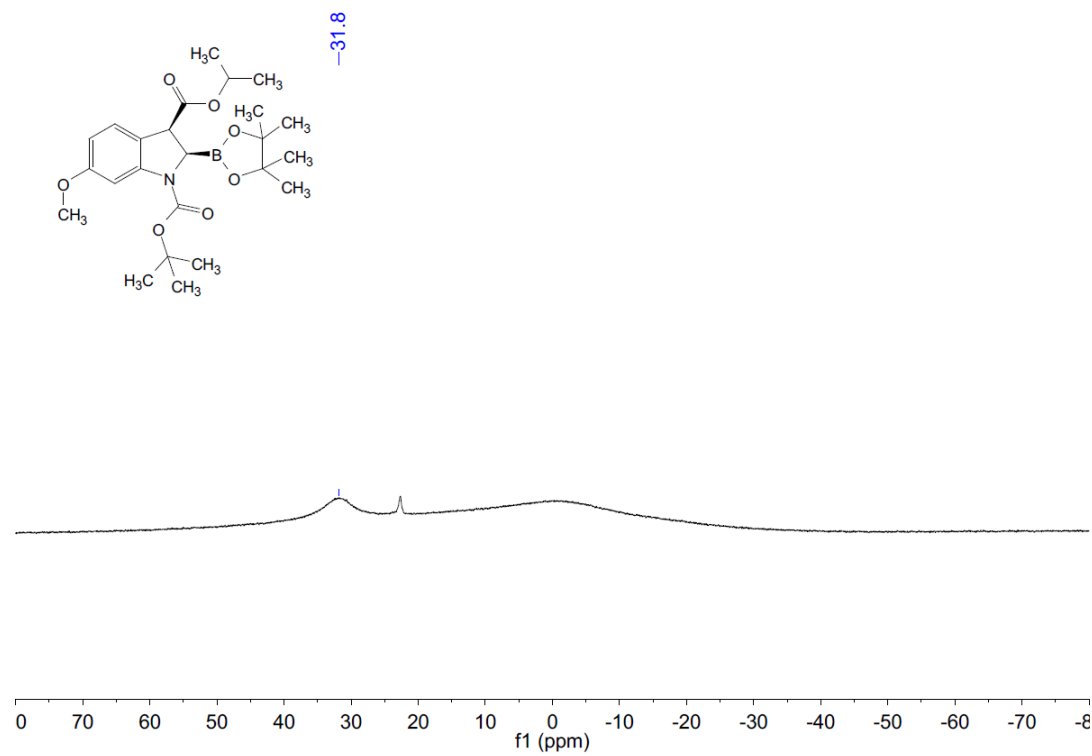
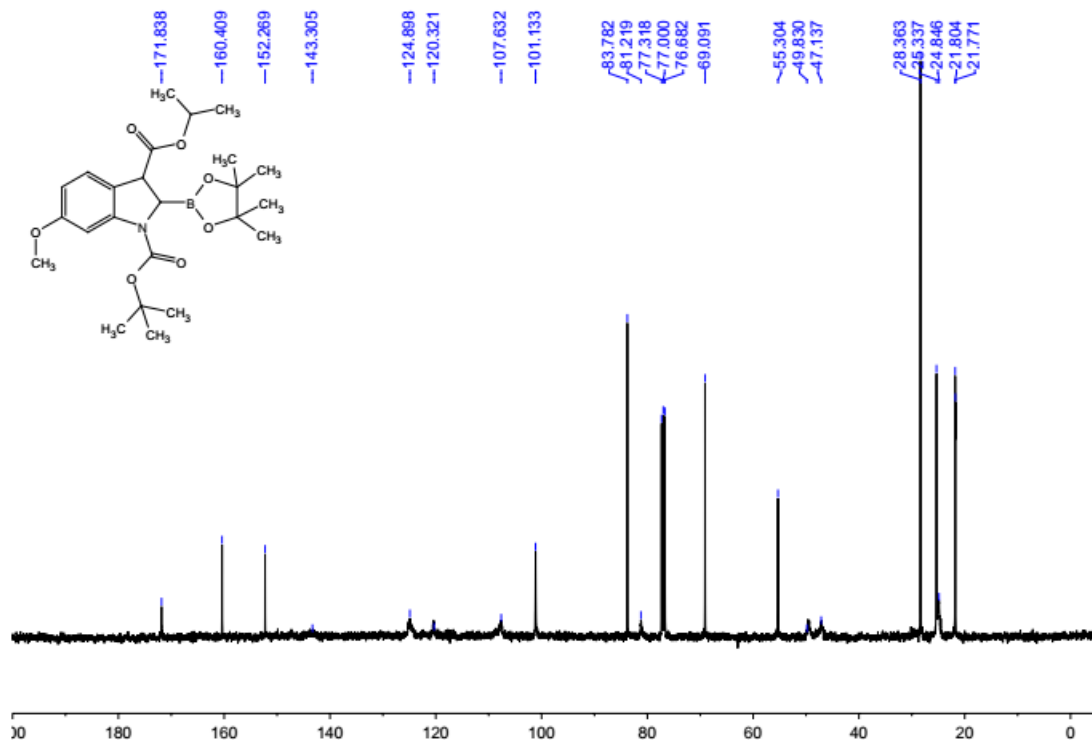
Compound 2o



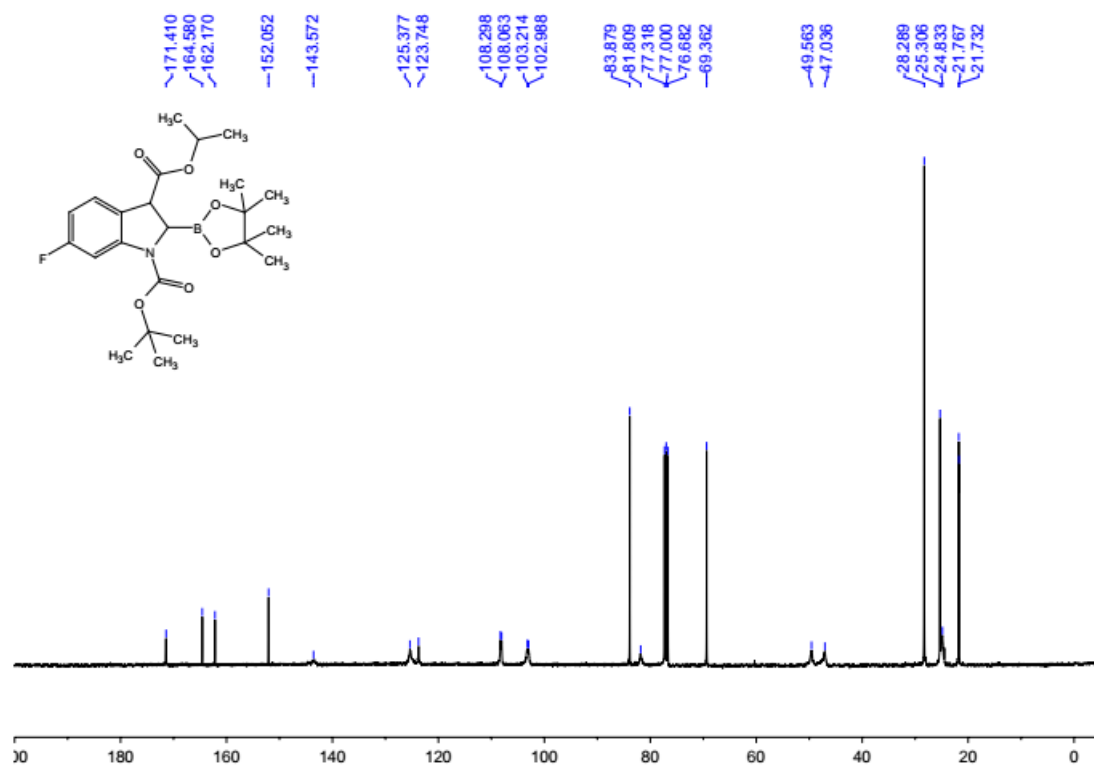
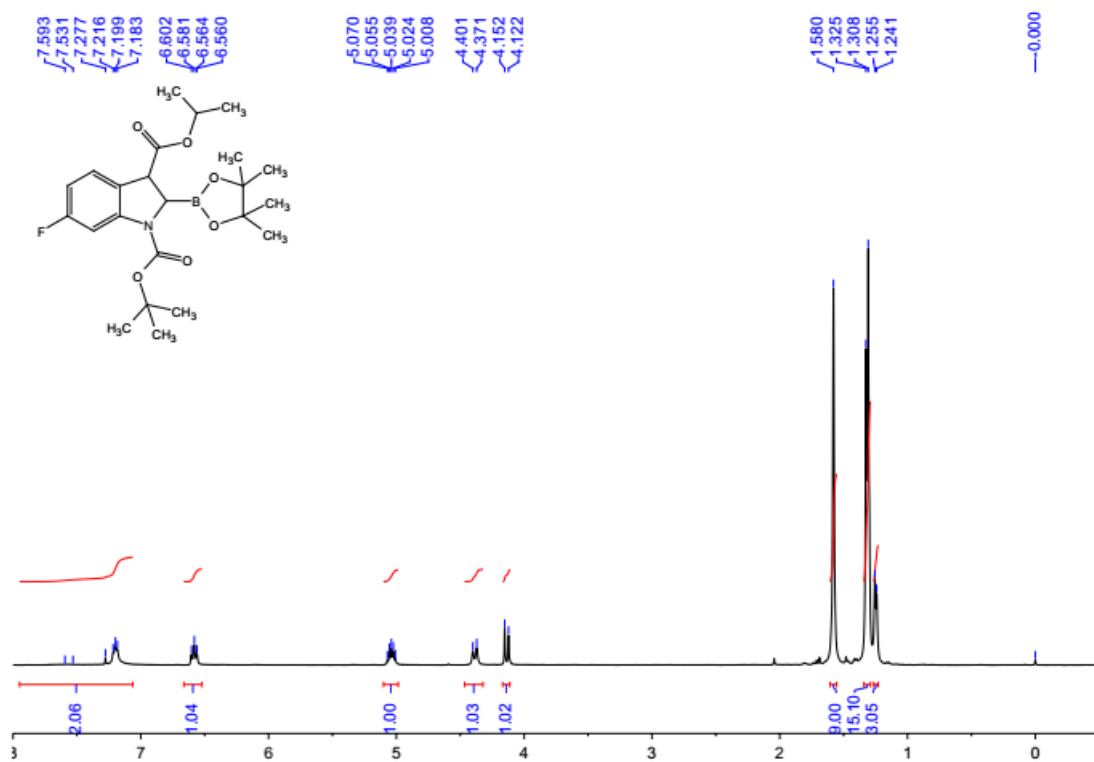


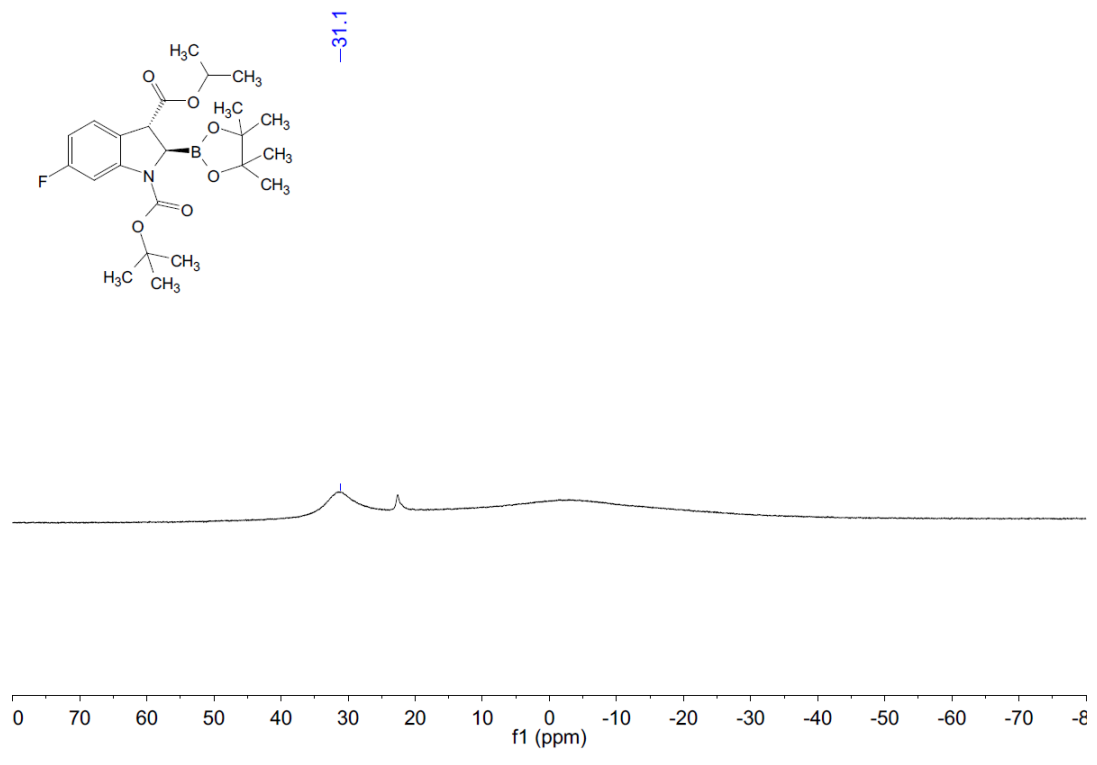
Compound 2p



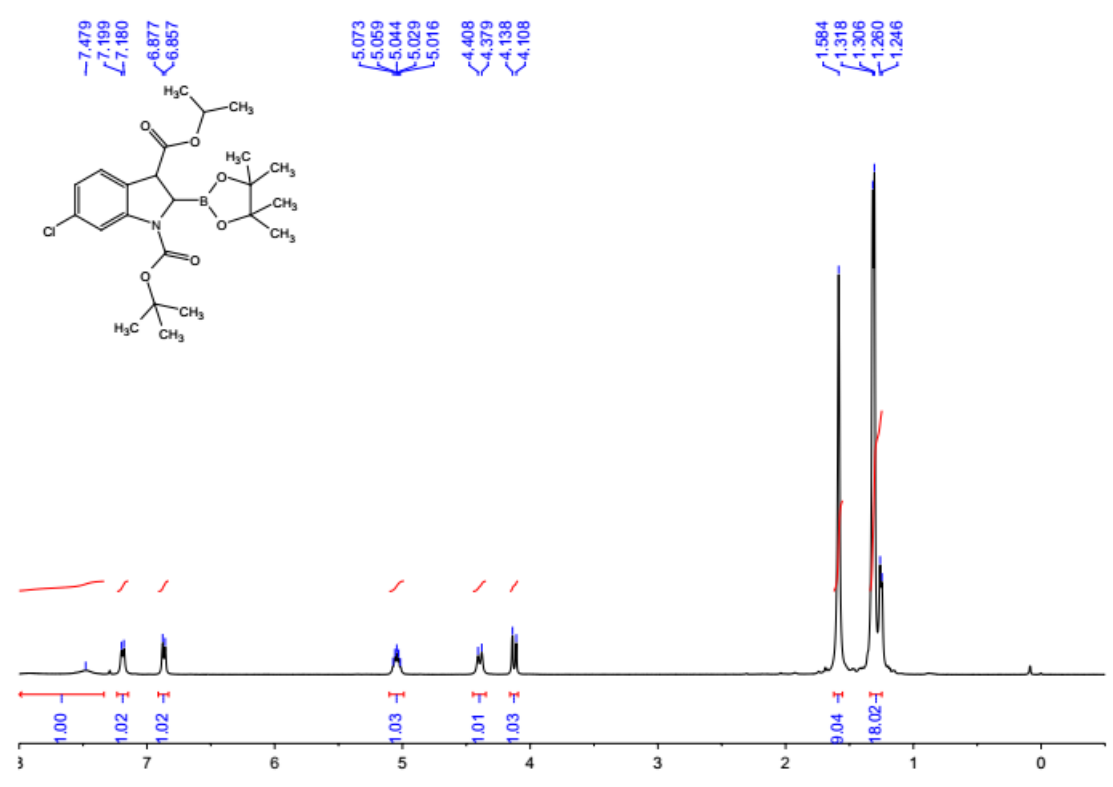


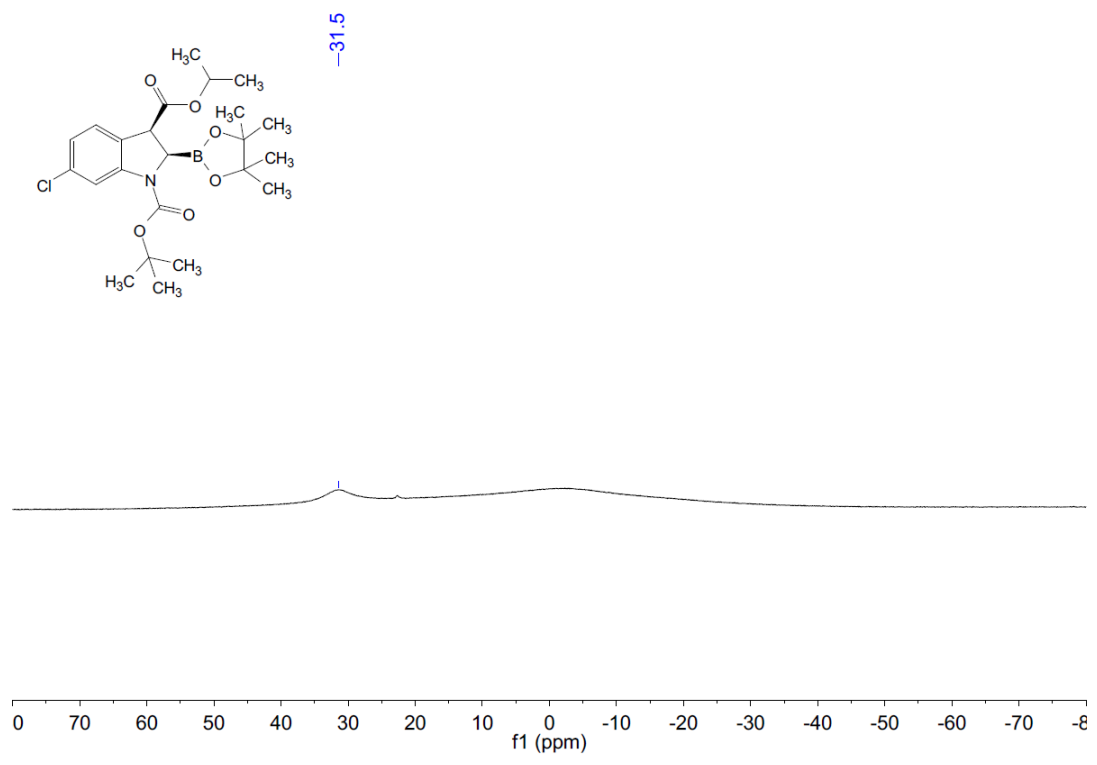
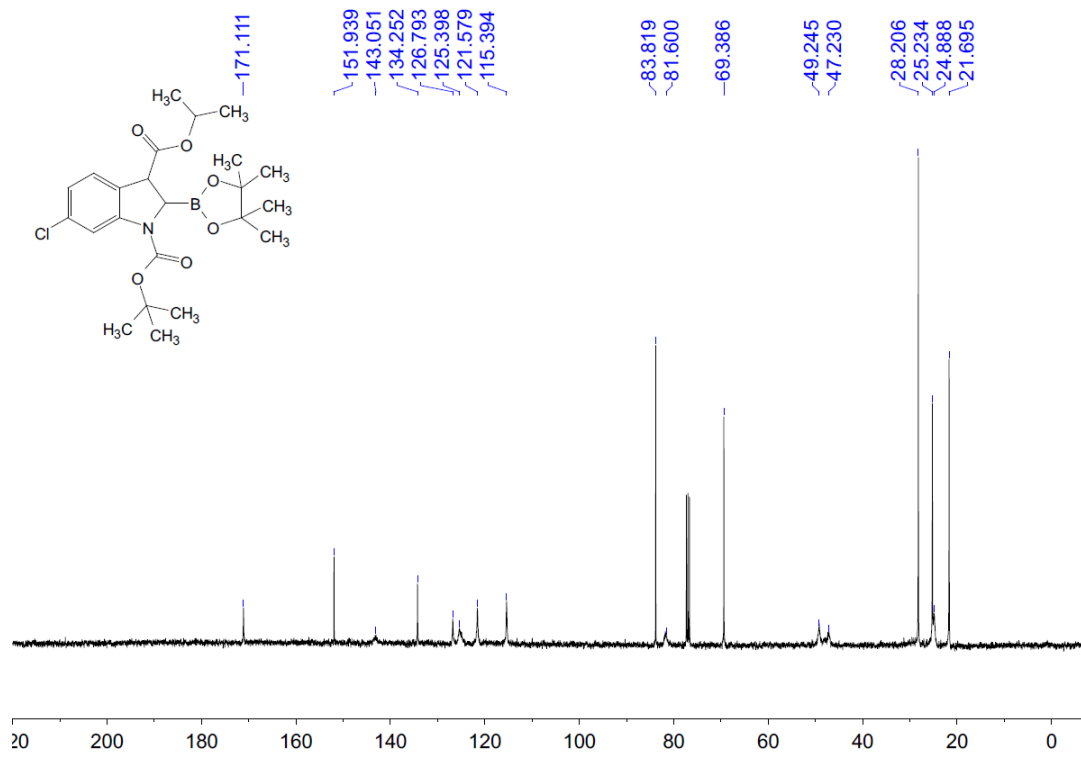
Compound 2q



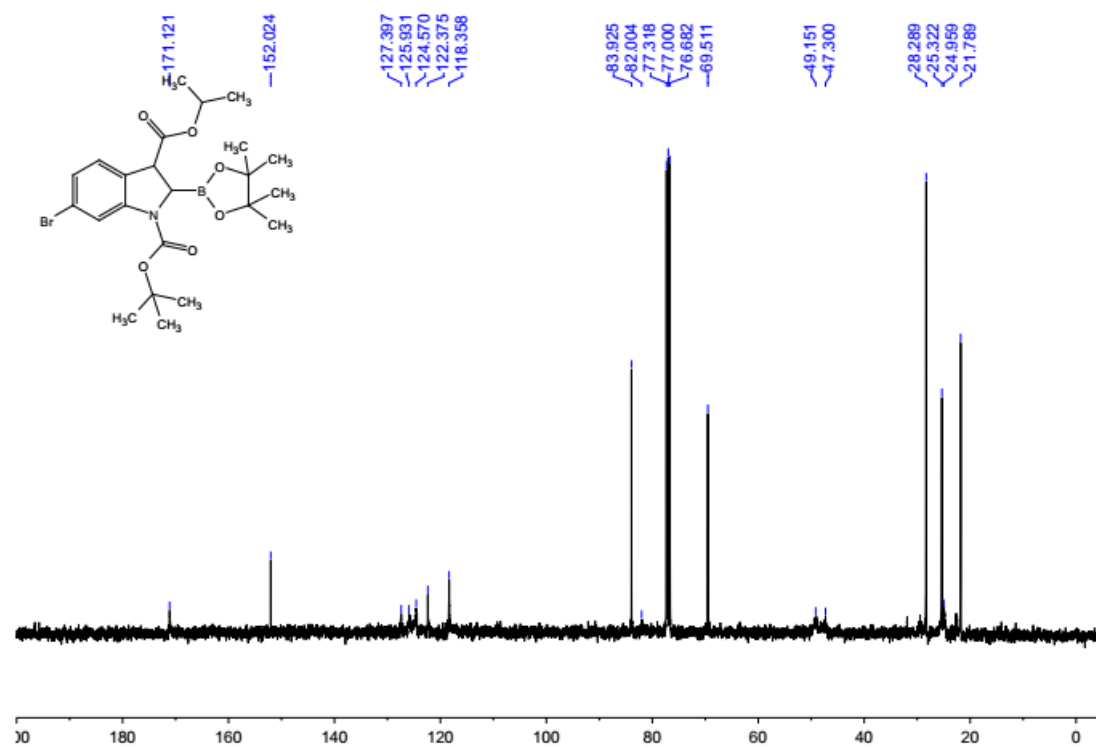
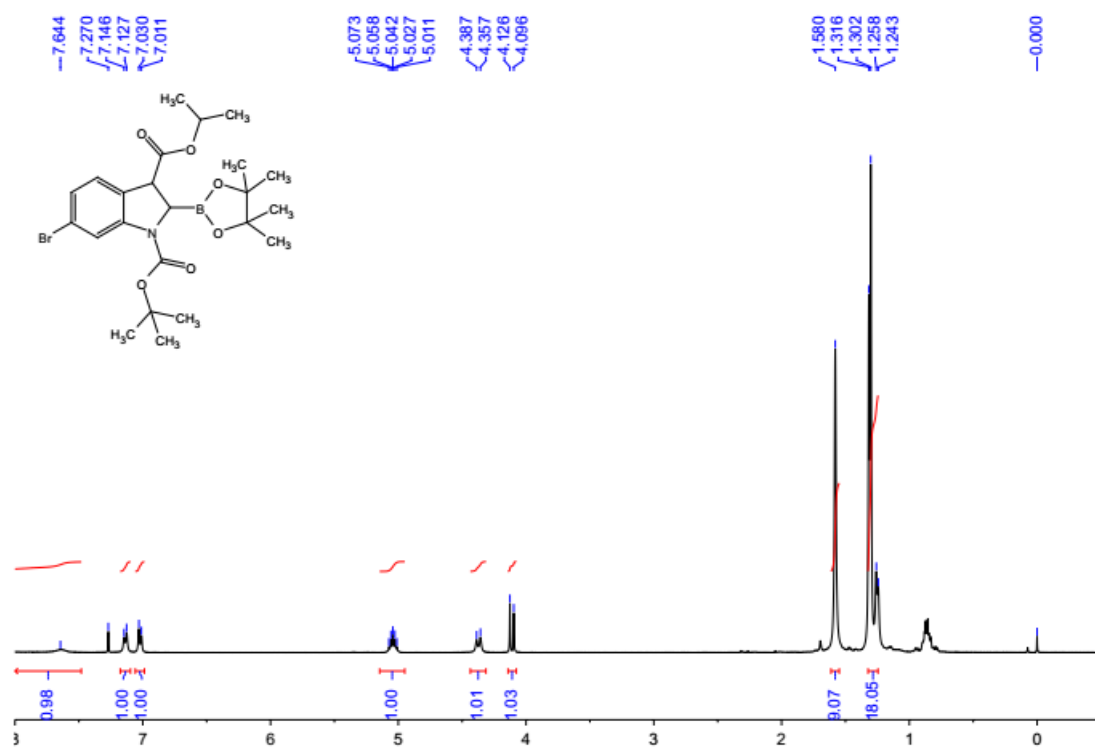


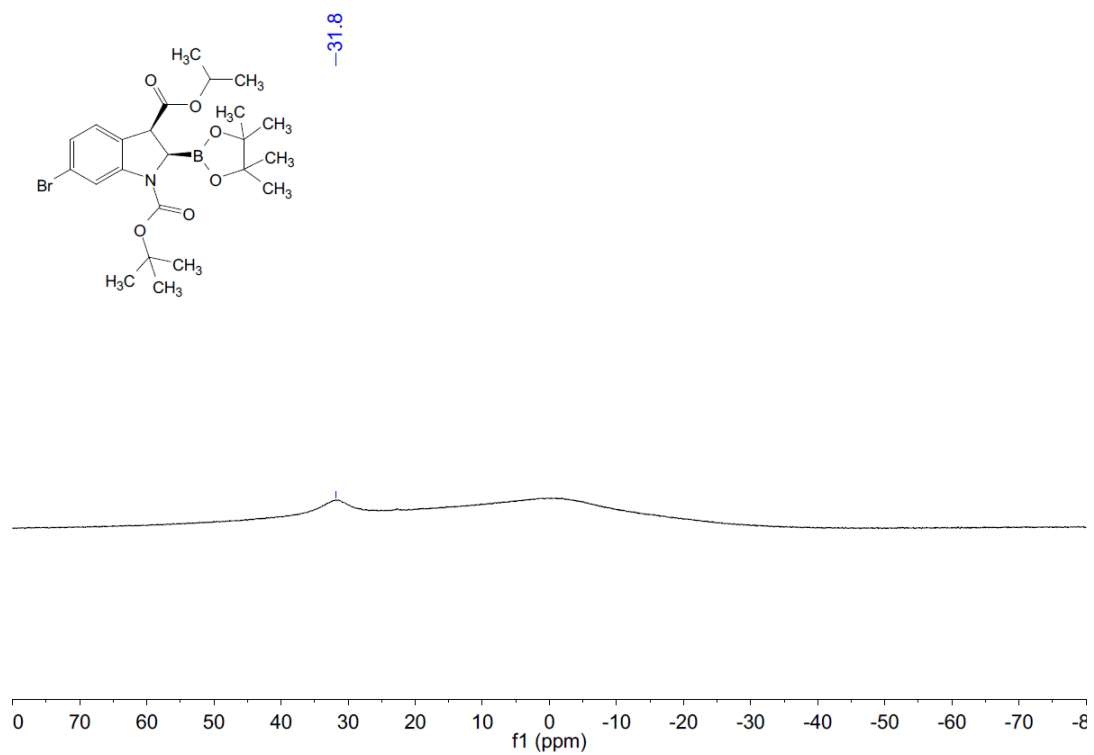
Compound 2r



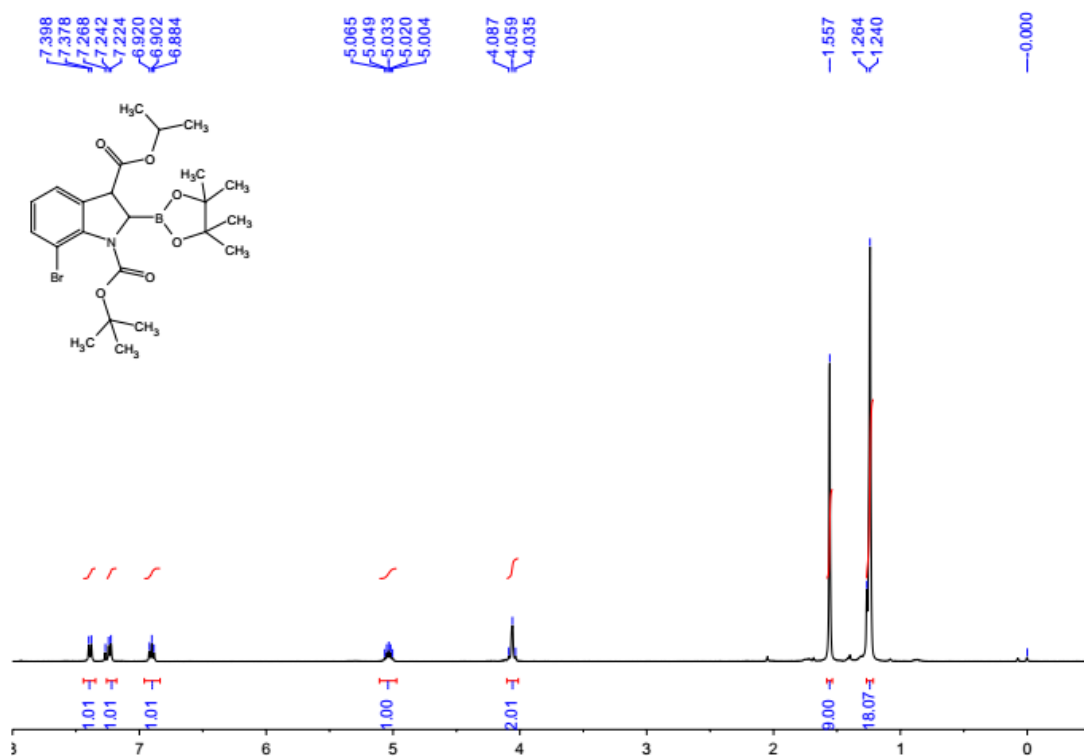


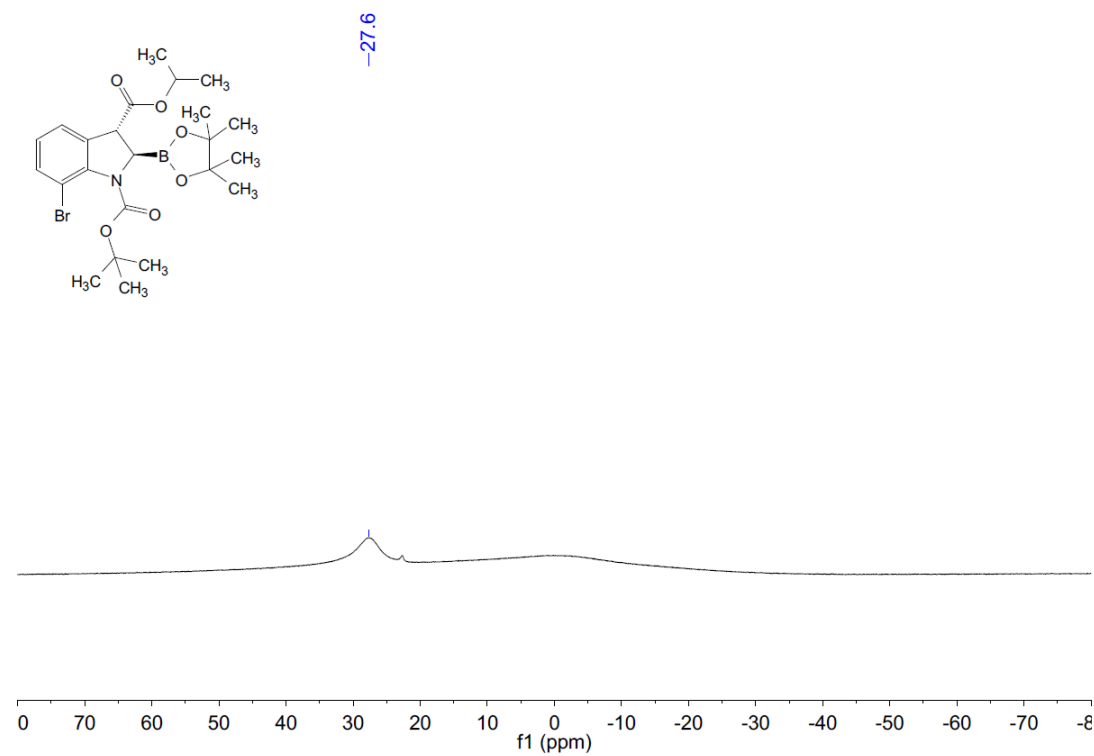
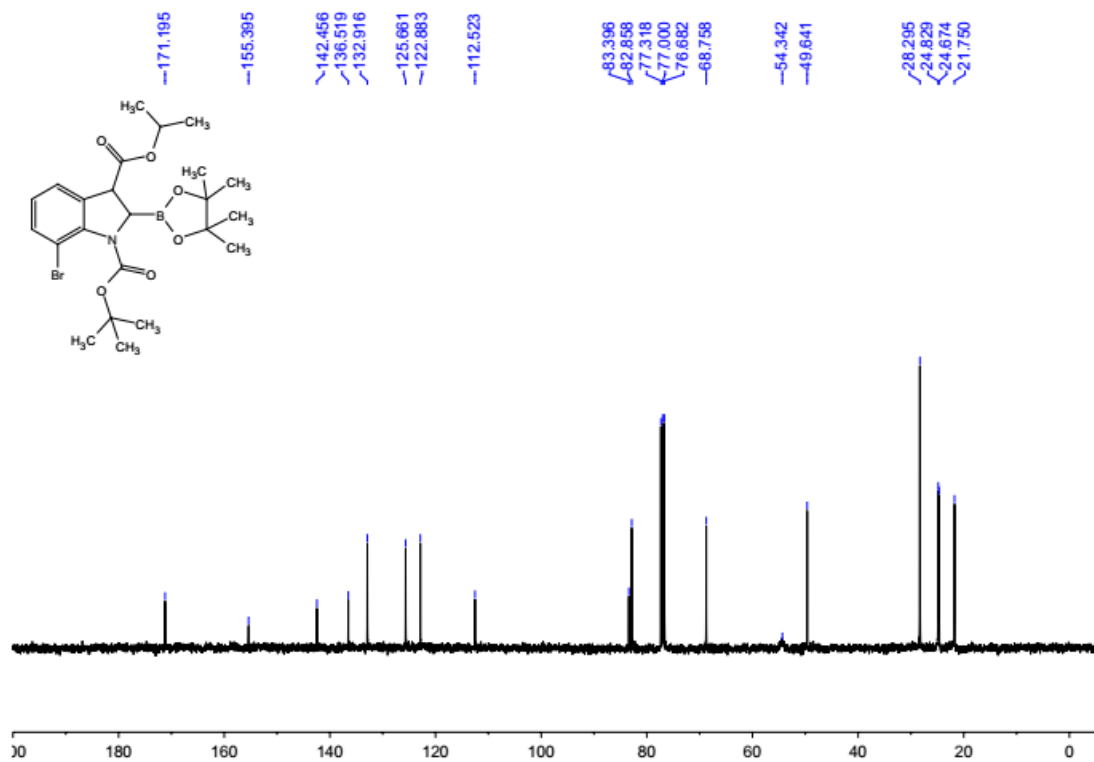
Compound 2s



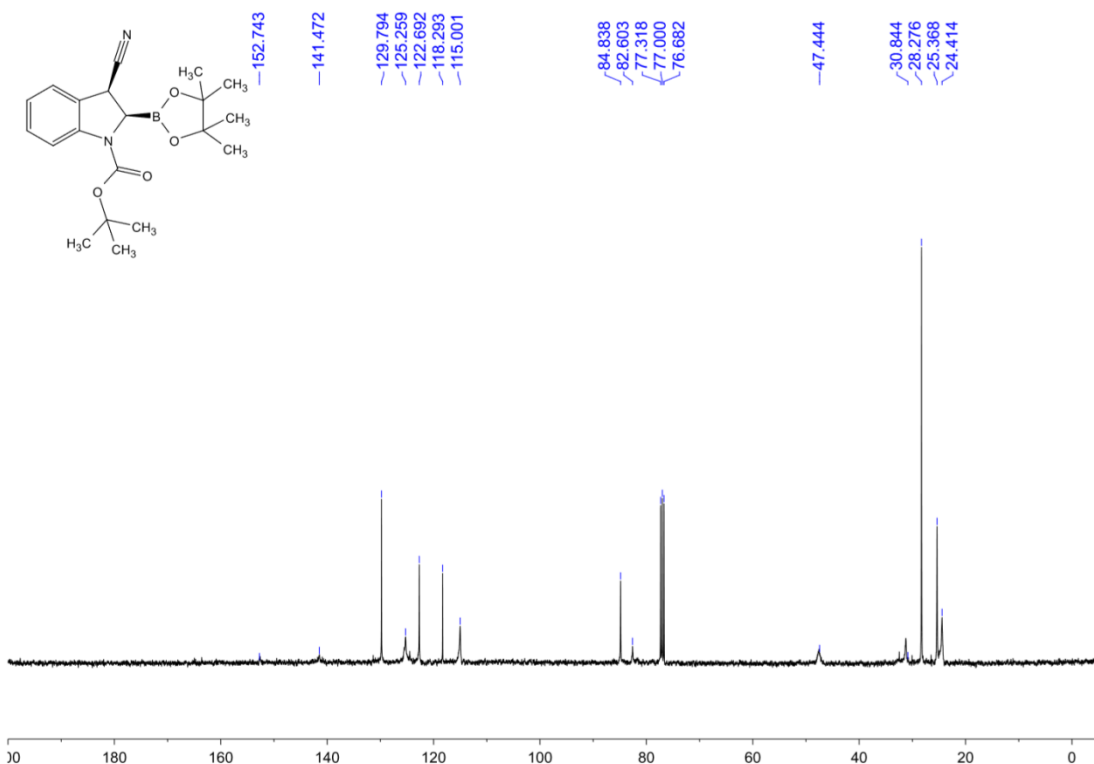
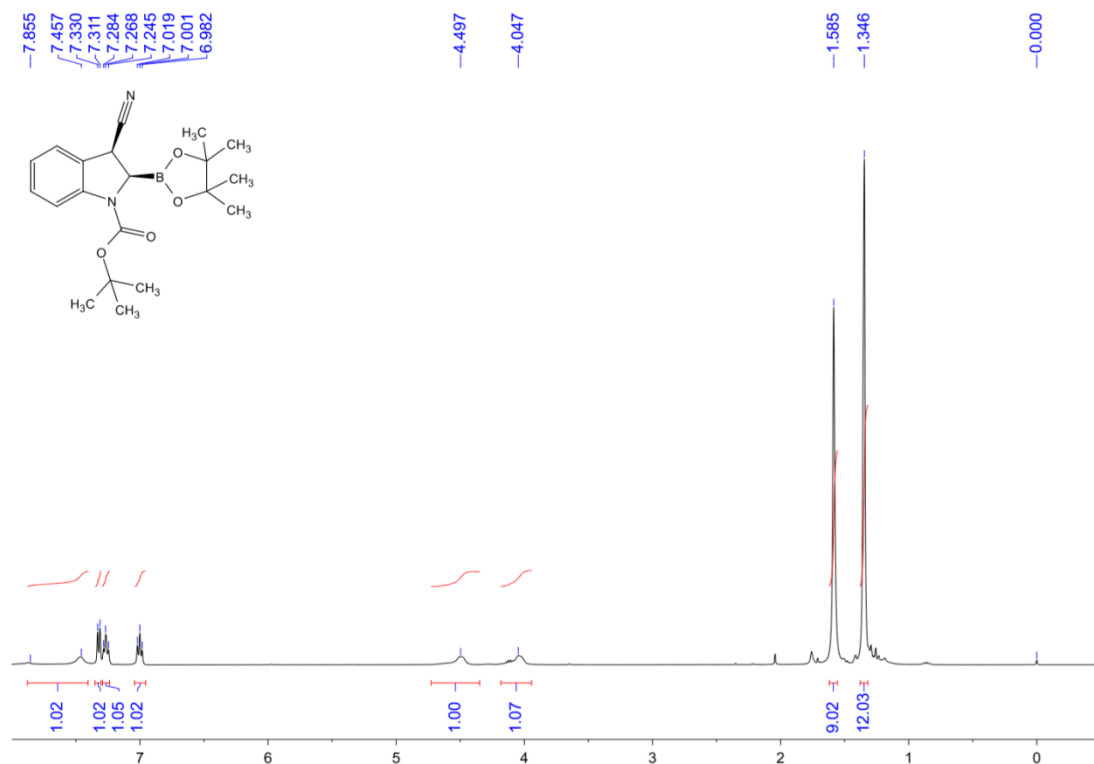


Compound 3t

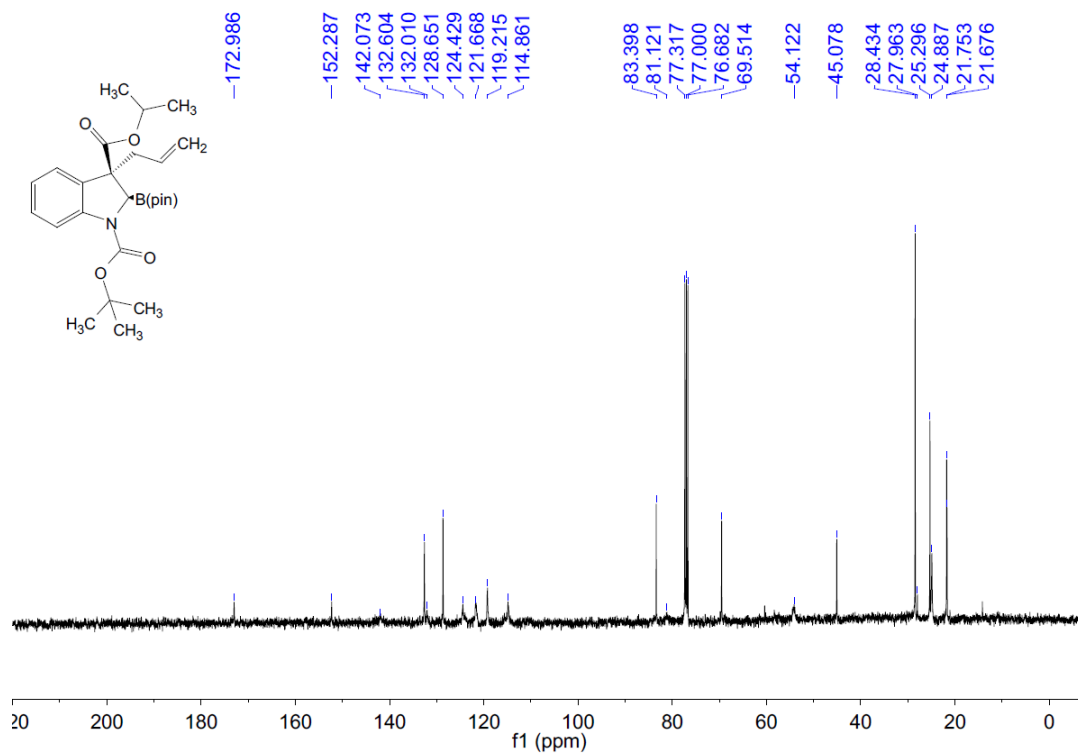
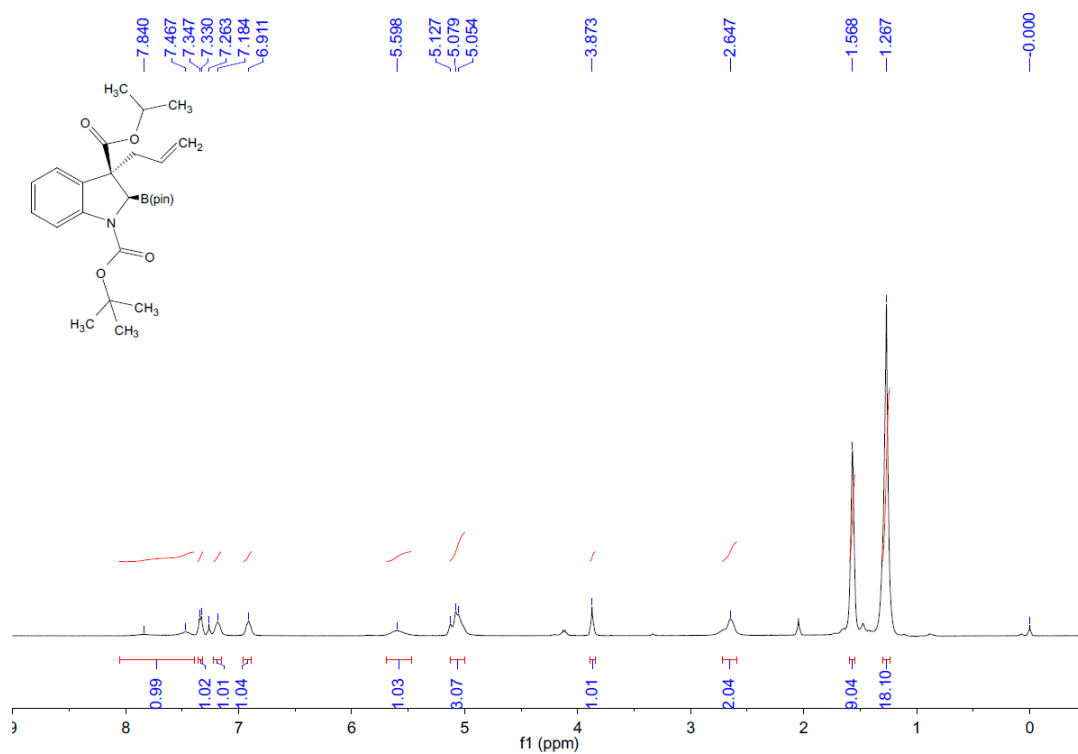




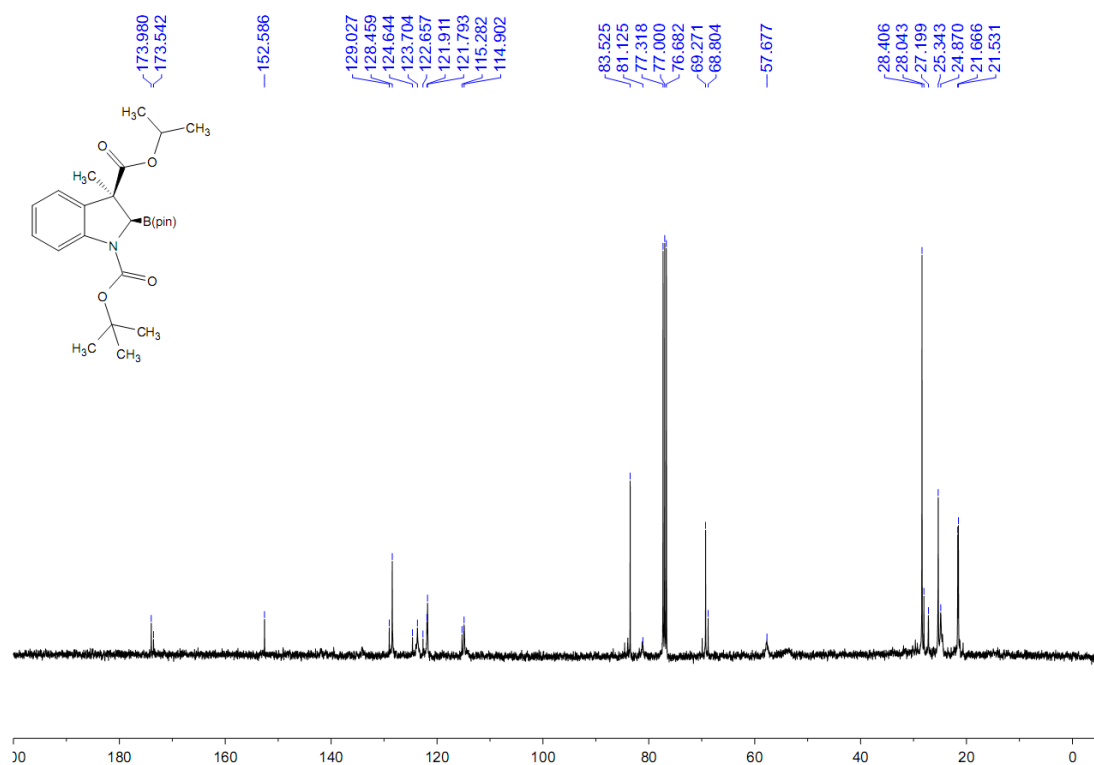
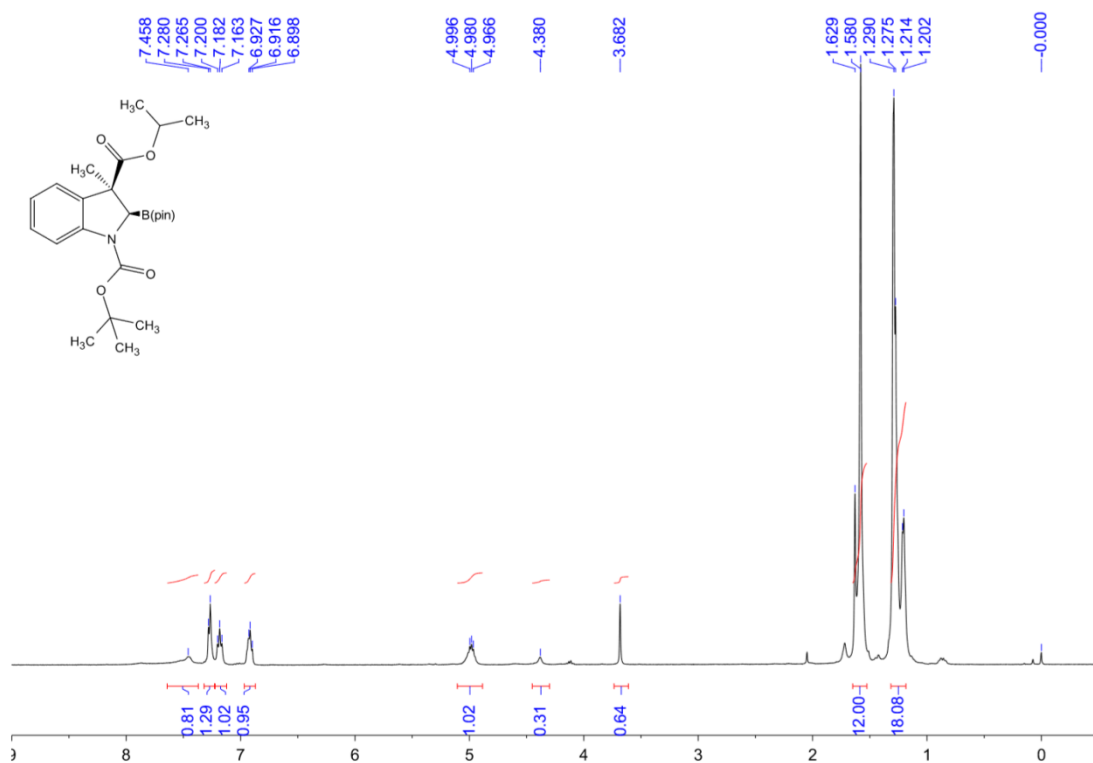
Compound 2v



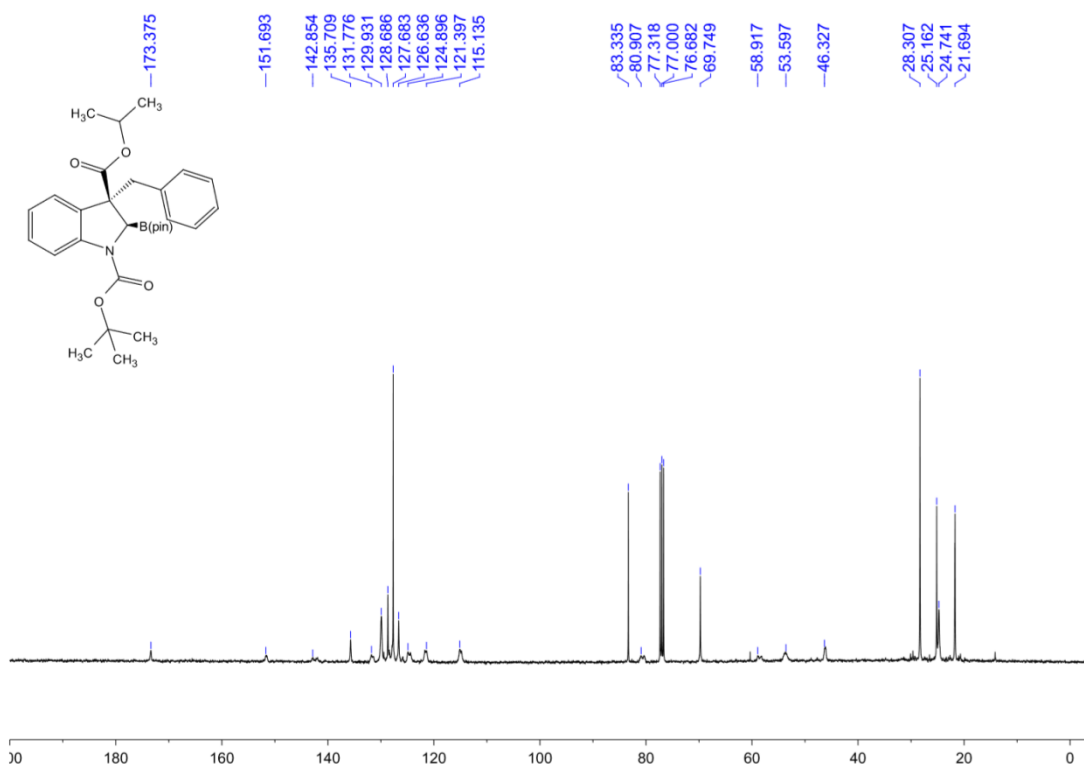
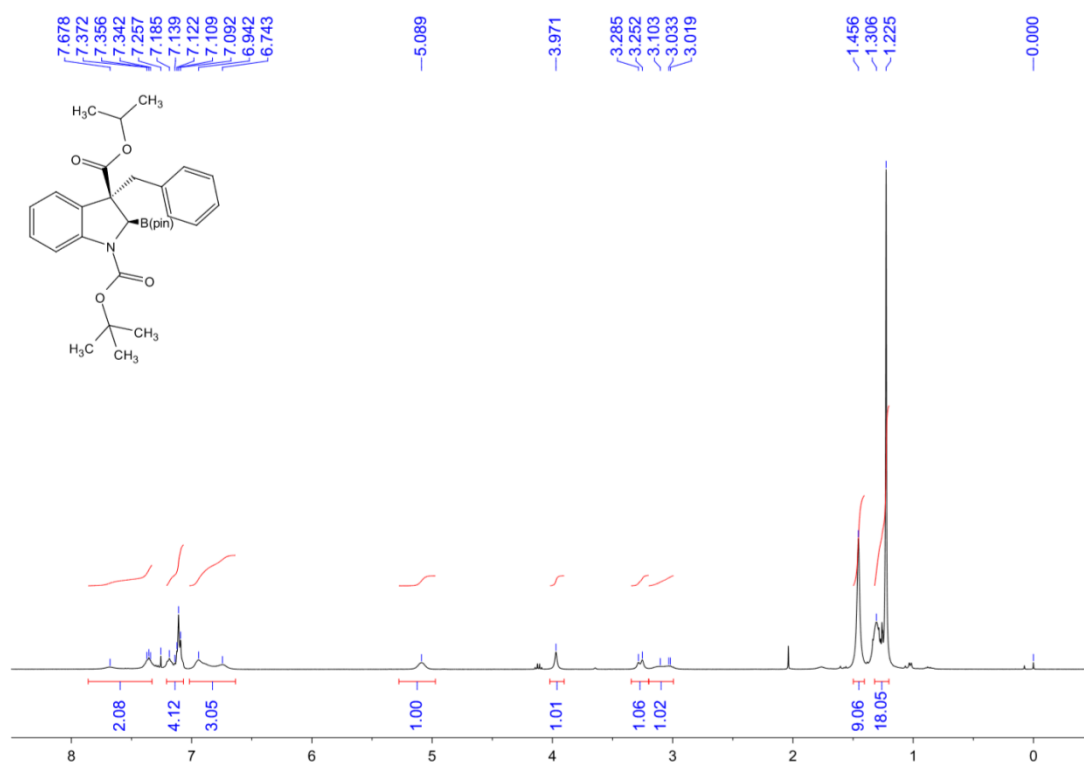
Compound 4a



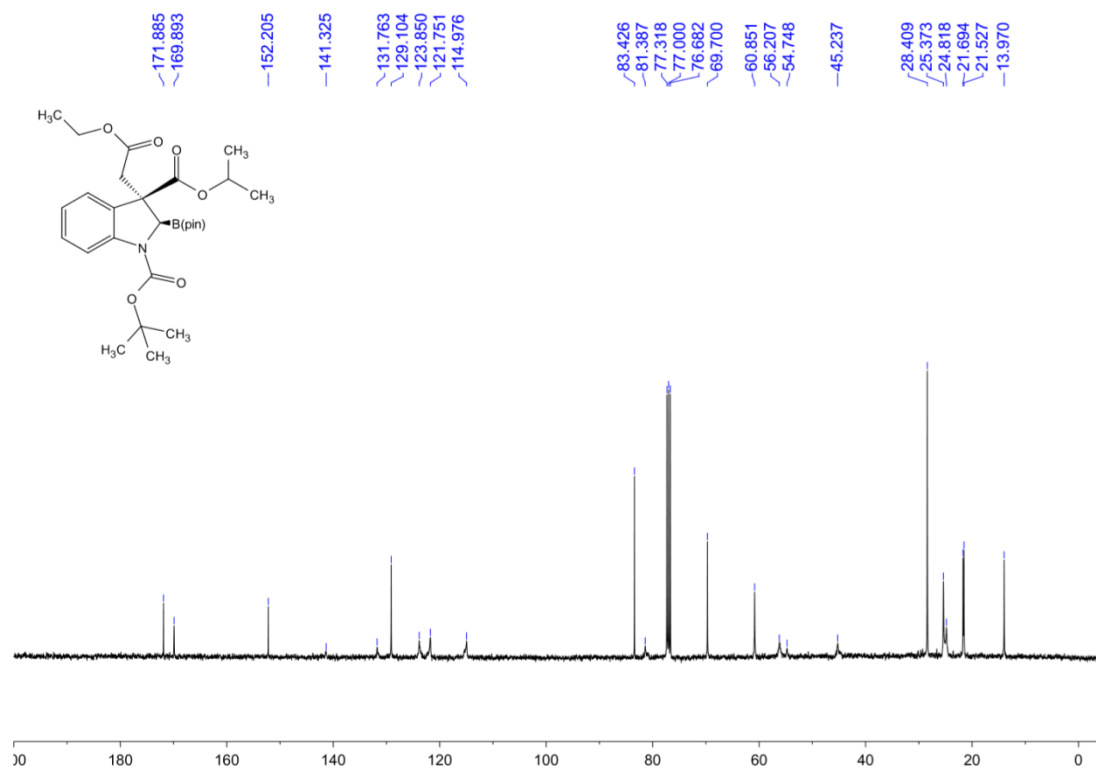
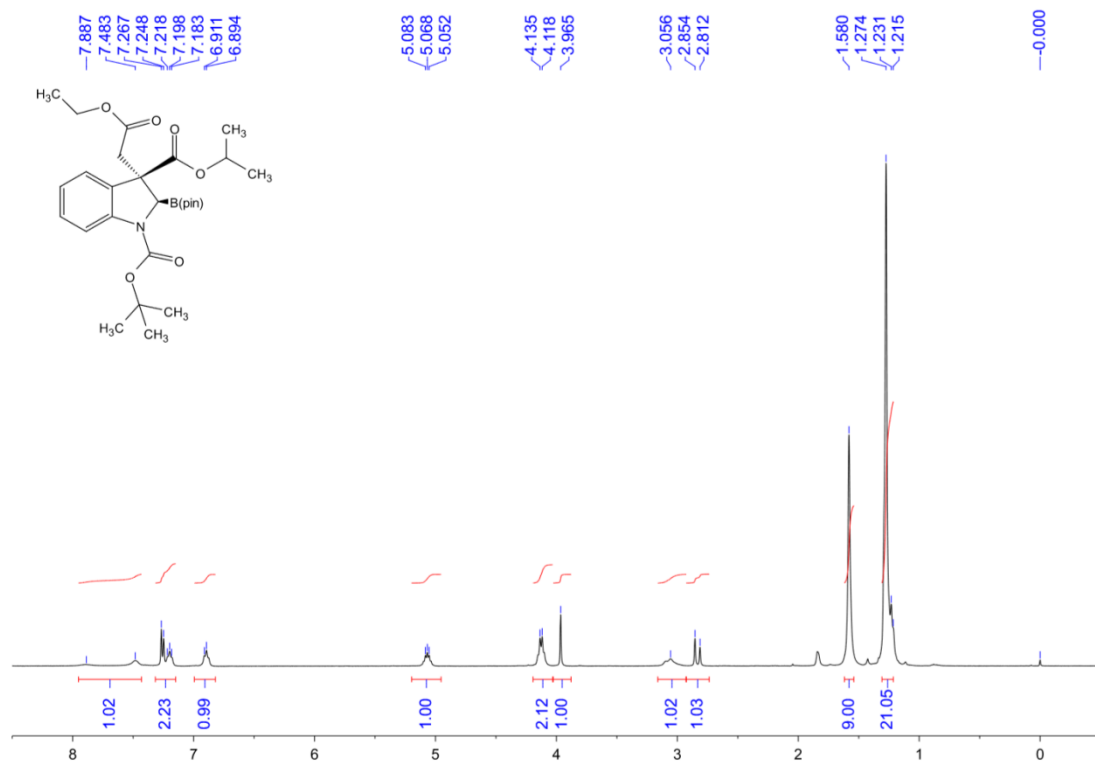
Compound 4b



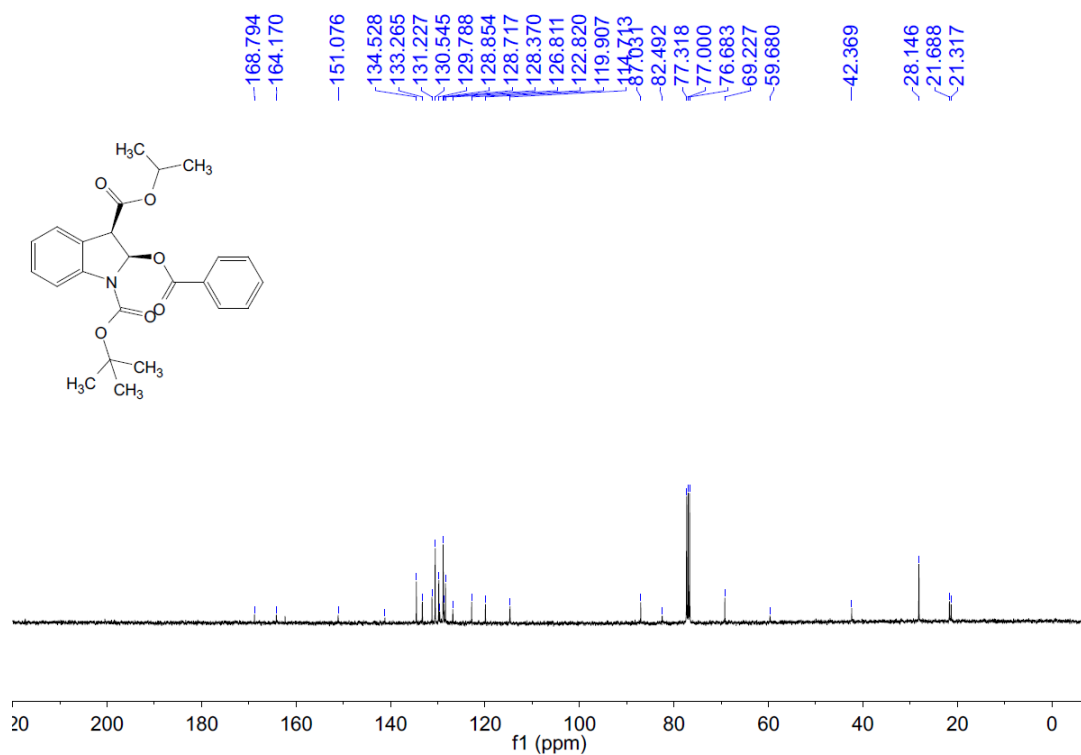
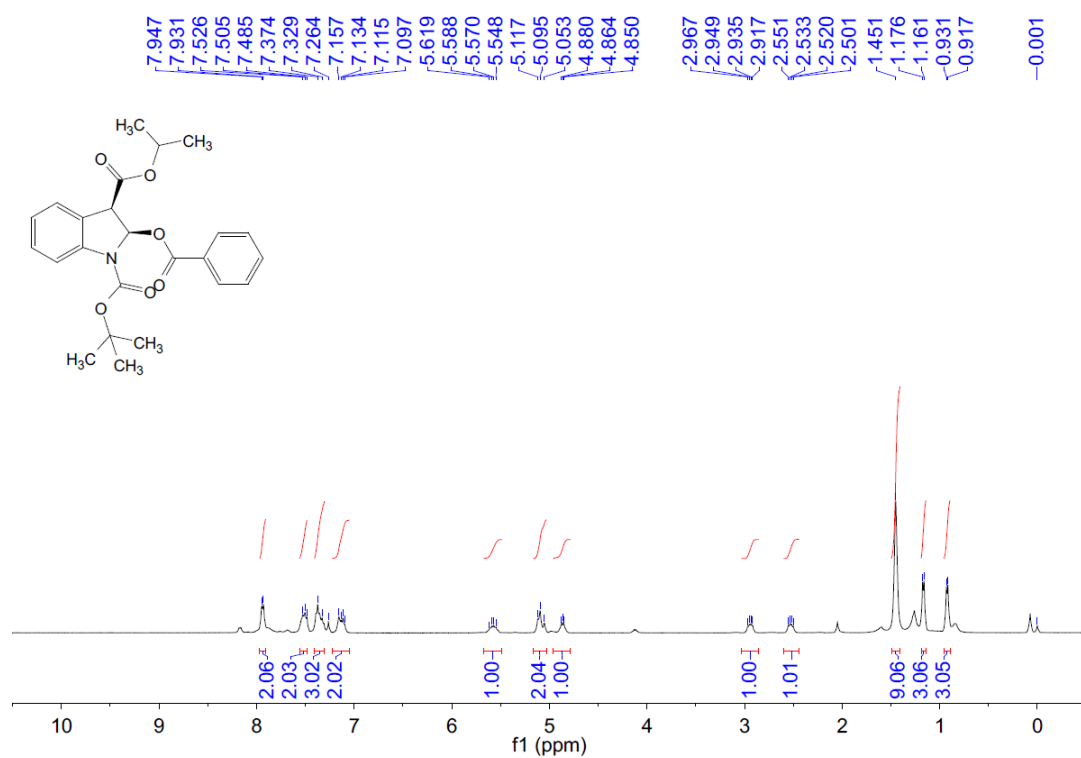
Compound 4c



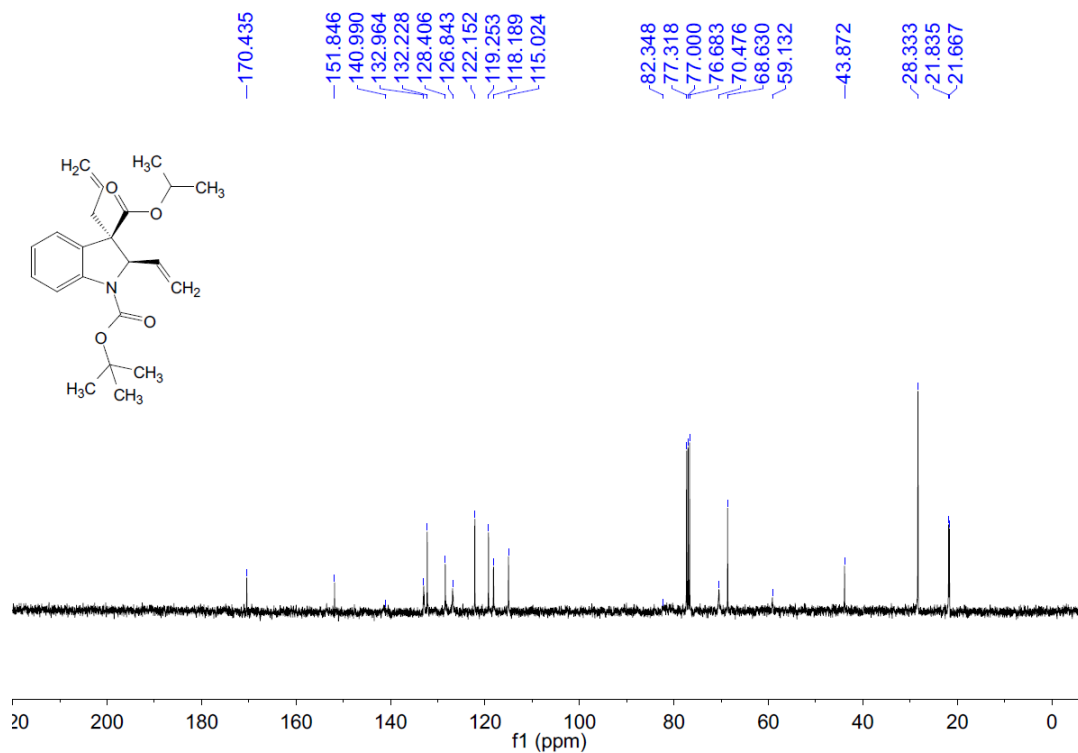
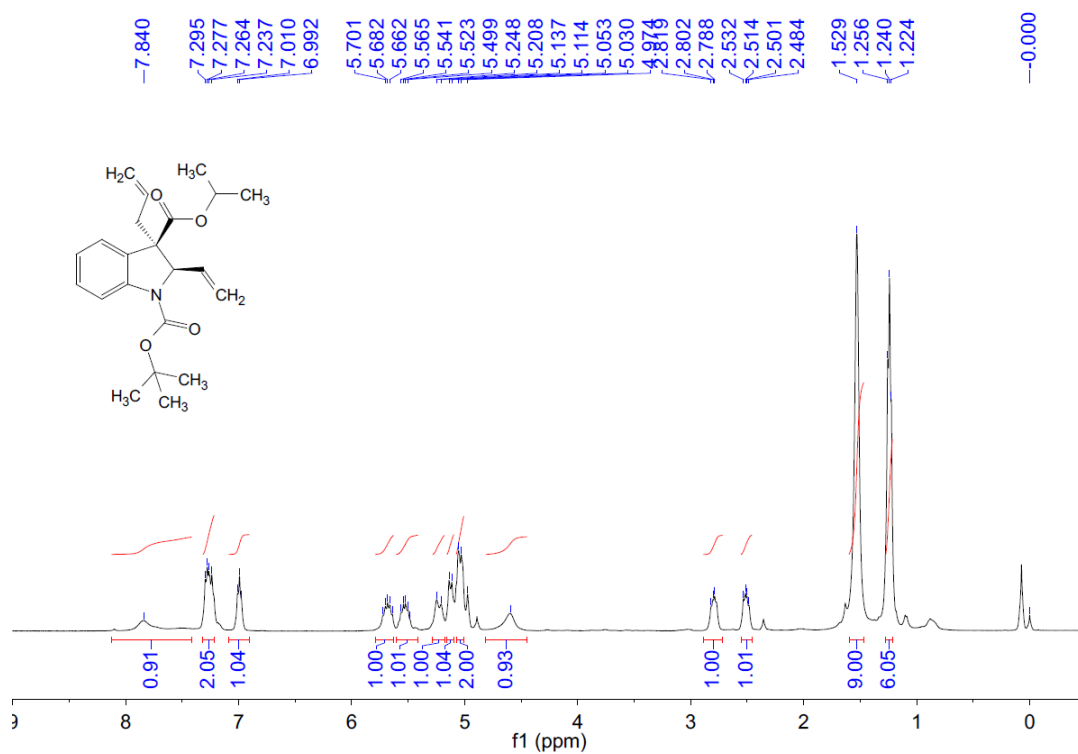
Compound 4d



Compound 5



Compound 6



Compound 7

