

Supplementary Tables

**Coverage and diagnostic yield of Whole Exome Sequencing for the Evaluation of Cases with Dilated and Hypertrophic Cardiomyopathy**

*Timothy Shin Heng Mak, Yee-Ki Lee, Clara S Tang, JoJo SH Hai, Xinru Ran, Pak-Chung Sham, Hung-Fat Tse*

Supplementary Table S1. Genes covered by three commercial panels for cardiomyopathy

Company	Admera Health <sup>1</sup>	GeneDx <sup>2</sup>	Invitae <sup>3</sup>	Ambry <sup>4</sup>
<b>Product</b>	CardioGxOne (cardiomyopathies general panel)	Cardiomyopathy panel	Cardiomyopathies	CMNEXT
<b>Diseases covered</b>	HCM, DCM, RCM, ACM, non-compaction cardiomyopathy, RASopathies, storage diseases, and congenital heart diseases	ARVC, DCM, HCM, LVNC, Noonan syndrome	ARVC, DCM, HCM, Fabry disease, LVNC, transthyretin amyloidosis	ARVD, DCM, HCM, LVNC, RCM
<b>Genes</b>	AARS2, ABCC9, ACAD9, ACADVL, ACTA1, ACTA2, ACTC1, ACTN2, AGK, AGL, AGPAT2, ALMS1, ANK2, ANKRD1, ATPAF2, BAG3, BRAF, BSCL2, CALR3, CASQ2, CAV3, CBL, COQ2, COX15, COX6B1, CRELD1, CRYAB, CSRP3, CTF1, CTNNA3, DES, DLD, DMD, DNAJC19, DOLK, DSC2, DSG2, DSP, DTNA, ELN, EMD, EYA4, FAH, FHL1, FHL2, FHOD3, FKR, FKTN, FLNA, FLNC, FOXD4, GAA, GATA4, GATA6, GATAD1, GFM1, GJA1, GJA5, GLA, GLB1, GNPTAB, GUSB, HCN4, HFE, HRAS, JAG1, JPH2, JUP, KCNH2, KCNJ2, KCNJ8, KCNQ1, KLF10,	ABCC9, ACTC1, ACTN2, ALMS1, ALPK3, ANKRD1, BAG3, BRAF, CAV3, CHRM2, CRYAB, CSRP3, DES, DMD, DOLK, DSC2, DSG2, DSP, DTNA, EMD, FHL1, FKR, FKTN, GATAD1, GLA, HCN4, HRAS, ILK, JPH2, JUP, KRAS, LAMA4, LAMP2, LDB3, LMNA, MAP2K1, MAP2K2, MIB1, MTND1, MTND5, MTND6, MTTD, MTTG, MTTT, MTTI, MTTK, MTTL1, MTTL2, MTTM, MTTQ, MTTT1, MTTT2, MURC, MYBPC3, MYH6, MYH7, MYL2, MYL3, MYLK2, MYOZ2, MYPN, NEBL, NEXN, NKX2-5, NRAS, PDLIM3, PKP2,	A2ML1, ABCC9, ACADVL, ACTC1, ACTN2, AGL, ALMS1, ANKRD1, BAG3, BRAF, CACNA1C, CALR3, CAV3, CBL, CHRM2, CPT2, CRYAB, CSRP3, CTF1, CTNNA3, DES, DMD, DNAJC19, DOLK, DSC2, DSG2, DSP, DTNA, ELAC2, EMD, EYA4, FHL1, FHL2, FKR, FKTN, FLNC, GAA, GATA4, GATA6, GATAD1, GLA, HCN4, HRAS, ILK, JPH2, JUP, KRAS, LAMA4, LAMP2, LDB3, LMNA, LRRC10, MAP2K1, MAP2K2, MTO1, MYBPC3, MYH6, MYH7, MYL2, MYL3, MYLK2, MYOM1, MYOZ2, MYPN, NEBL, NEXN,	ABCC9, ACTC1, ACTN2, ANKRD1, BAG3, CRYAB, CSRP3, DES, DMD, DSC2, DSG2, DSP, EMD, EYA4, FKTN, FXN, GATAD1, GLA, JPH2, JUP, LAMA4, LAMP2, LDB3, LMNA, MYBPC3, MYH6, MYH7, MYL2, MYL3, MYOZ2, MYPN, NEXN, NKX2-5, PKP2, PLN, PRKAG2, PTPN11, RAF1, RBM20, RYR2, SCN5A, TAZ, TBX20, TCAP, TGFB3, TMEM43, TMPO, TNNC1, TNNI3, TNNT2, TPM1, TTN, TTR, TXNRD2, VCL, ZASP

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<p>KRAS, LAMA2, LAMA4, LAMP2,  LDB3, LIAS, LMNA, MAP2K1,  MAP2K2, MIB1, MLYCD, MRPL3,  MRPS22, MTO1, MURC, MYBPC3,  MYH11, MYH6, MYH7, MYL2, MYL3,  MYLK2, MYOT, MYOZ2, MYPN,  NEBL, NEXN, NKX2-5, NOTCH1,  NRAS, OBSL1, PDHA1, PDLIM3,  PHKA1, PITX2, PKP2, PLN, PMM2,  PRDM16, PRKAG2, PSEN1, PSEN2,  PTPN11, RAF1, RBM20, RYR2,  SCN5A, SGCA, SGCB, SGCD,  SHOC2, SLC22A5, SLC25A4,  SMAD3, SOS1, SPRED1, SURF1,  TAZ, TBX1, TBX20, TBX5, TCAP,  TGFB3, TMEM43, TMEM70, TMPO,  TNNC1, TNNI3, TNNT2, TPM1,  TRIM63, TSFM, TTN, TTR, TXNRD2,  VCL (n=149)</p>	<p>PLN, PRDM16, PRKAG2, PTPN11,  RAF1, RBM20, RIT1, RYR2,  SCN5A, SGCD, SOS1, TAZ, TCAP,  TGFB3, TMEM43, TMPO, TNNC1,  TNNI3, TNNT2, TPM1, TTN, TTR,  TXNRD2, VCL (n=91)</p>	<p>NF1, NKX2-5, NPPA, NRAS,  PDLIM3, PKP2, PLEKHM2, PLN,  PRDM16, PRKAG2, PTPN11,  RAF1, RASA1, RBM20, RIT1,  RRAS, RYR2, SCN5A, SDHA,  SGCD, SHOC2, SLC22A5, SOS1,  SOS2, SPRED1, TAZ, TCAP,  TGFB3, TMEM43, TMEM70,  TMPO, TNNC1, TNNI3, TNNT2,  TPM1, TTN, TTR, TXNRD2, VCL  (n=105)</p>	<p>(n=56)</p>
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1. <http://www.admerahealth.com/cardiogxone/cardiomyopathies-general-panel/>
2. <https://www.genedx.com/test-catalog/available-tests/cardiomyopathy-panel/>
3. <https://www.invitae.com/en/physician/tests/02251/#test-order>
4. <http://www.ambrygen.com/tests/cmne>

Supplementary Table S2. Methods for applying the ACMG guidelines

<b>Evidence of pathogenicity</b>	<b>ACMG definition</b>	<b>Our working definition</b>
<b>Very strong</b>	PVS1: Null variant (nonsense, frameshift, canonical +1 1 or 2 splice sites, initiation codon, single or multiexon deletion) in a gene where Truncating is a known mechanism of disease	Truncating variants in genes whose truncating variants are significantly associated with HCM/DCM according to Walsh et al <sup>6</sup> (after correction for multiple testing).
<b>Strong</b>	PS1: Same amino acid change as a previously established pathogenic variant regardless of nucleotide change	A previously established pathogenic variant was defined as one which had a Pathogenic call from the ACMG guideline.
	PS2: De novo in a patient with the disease and no family history	based on literature review
	PS3: Well-established in vitro or in vivo functional studies supportive of a damaging effect on the gene or gene product	based on literature review
	PS4: The prevalence of the variant in affected individuals is significantly increased compared with the prevalence in controls	Not applicable.
<b>Moderate</b>	PM1: Located in a mutational hot spot and/or critical and well-established functional domain without benign variation	Mutational hotspots for 190 genes were given in the Supplementary material of Maxwell et al <sup>7</sup>
	PM2: Absent from controls in Exome sequencing project, 1000 Genomes Project, or Exome Aggregation Consortium	Absent in the Exac database, the 1000 Genome database, and a cohort of 712 local DDD patients
	PM3: For recessive disorders, detected in trans with a pathogenic variant	Not applicable
	PM4: Protein length changes as a result of in-frame deletions/insertions in a nonrepeat region or stop-loss variants	nonframeshift variants
	PM5: Novel missense change at an amino acid residue where a different missense change determined to be pathogenic has been seen before	Like PS1, a previously established pathogenic variant was defined as one which had a Pathogenic call from the ACMG guideline.
	PM6: Assumed de novo, but without confirmation of paternity and maternity	based on literature review
<b>Supporting</b>	PP1: Cosegregation with disease in multiple affected family members in a gene definitively known to cause the disease	If evidence found in Clinvar.

Evidence of pathogenicity	ACMG definition	Our working definition
<b>Benign (stand-alone)</b>	<p>PP2: Missense variant in a gene that has a low rate of benign missense variation and in which missense variants are a common mechanism of disease</p> <p>PP3: Multiple lines of computational evidence support a deleterious effect on the gene or gene product</p> <p>PP4: Patient's phenotype or family history is highly specific for a disease with a single genetic etiology</p> <p>PP5: Reputable source recently reports variant as pathogenic, but the evidence is not available to the laboratory to perform an independent evaluation</p> <p>BA1: Allele frequency is &gt;5% in Exome sequencing project, 1000 Genome project, or Exome Aggregation consortium</p>	<p>non-truncating variant found in gene whose RVIS (Residual Variation Intolerance Score) <sup>1</sup> is in the most intolerant 10%, or</p> <p>Truncating variant found in gene whose pLI (probability of Loss-of-function intolerant) <sup>2</sup> score is in the most intolerant 10%.</p> <p>Predicted to be pathogenic by Polyphen (HDIV), Polyphen (HVAR)<sup>3</sup>, deleterious by SIFT (Sorting Intolerant From Tolerant) <sup>4</sup>,and Mutation Taster<sup>5</sup>.</p> <p>Not applicable</p> <p>Not applicable</p> <p>Not applicable</p>
<b>Benign (Strong)</b>	<p>BS1: Allele frequency is greater than expected for disorder</p> <p>BS2: Observed in a healthy adult individual with full penetrance expected at an early age</p> <p>BS3: Well-established in vitro or in vivo function studies show no damaging effect on protein function or splicing</p> <p>BS4: Lack of segregation in affected members of a family</p>	<p>MAF &gt; 0.1%</p> <p>Not applicable. Cardiomyopathy not fully penetrant at early age</p> <p>Literature review</p> <p>Literature review</p>
<b>Benign (Supporting)</b>	<p>BP1: Missense variant in a gene for which primarily truncating variants are known to cause disease</p> <p>BP2: Observed in trans with a pathogenic variant for a fully penetrant dominant gene/disorder or observed in cis with a pathogenic variant in any inheritance pattern</p> <p>BP3: In-frame deletions/insertions in a repetitive region without a known function</p> <p>BP4: Multiple lines of computational evidence suggest no impact on gene or gene product</p>	<p>Missense variants in TTN or SCN5A (according to Walsh et al)</p> <p>Not applicable. Information not available.</p> <p>Not applicable. Information not available.</p> <p>Predicted to be benign by Polyphen (HDIV), Polyphen (HVAR)<sup>4</sup>, deleterious by SIFT<sup>5</sup>,and Mutation Taster<sup>3</sup>.</p>

<b>Evidence of pathogenicity</b>	<b>ACMG definition</b>	<b>Our working definition</b>
	<p>BP5: Variant found in a case with a an alternate molecular basis for disease</p> <p>BP6: Reputable source recently reports variant as benign</p> <p>BP7: A synonymous variant for which splicing prediction algorithms predict no impact to the splice consensus sequence nor the creation of a new splice site and the nucleotide is not highly conserved</p>	<p>awarded to variants found in patients with variants classified as "Pathogenic"</p> <p>One "Benign" or "Likely benign" classification in Clinvar</p> <p>Not applicable. Synonymous variants are not considered</p>

## References:

1. Petrovski, S., Wang, Q., Heinzen, E. L., Allen, A. S. & Goldstein, D. B. Genic Intolerance to Functional Variation and the Interpretation of Personal Genomes. *PLoS Genet.* **9**, (2013).
2. Lek, M. *et al.* Analysis of protein-coding genetic variation in 60,706 humans. *Nature* **536**, 285–291 (2016).
3. Adzhubei, I. A. *et al.* A method and server for predicting damaging missense mutations. *Nat. Methods* **7**, 248–249 (2010).
4. Kumar, P., Henikoff, S. & Ng, P. C. Predicting the effects of coding non-synonymous variants on protein function using the SIFT algorithm. *Nat. Protoc.* **4**, 1073–1081 (2009).
5. Schwarz, J. M., Cooper, D. N., Schuelke, M. & Seelow, D. MutationTaster2: mutation prediction for the deep-sequencing age. *Nat. Methods* **11**, 361–2 (2014).
6. Walsh, R. *et al.* Reassessment of Mendelian gene pathogenicity using 7,855 cardiomyopathy cases and 60,706 reference samples. *Genet. Med.* 41111 (2016). doi:10.1038/gim.2016.90
7. Maxwell, K. N. *et al.* Evaluation of ACMG-Guideline-Based Variant Classification of Cancer Susceptibility and Non-Cancer-Associated Genes in Families Affected by Breast Cancer. *Am. J. Hum. Genet.* **98**, 801–817 (2016).