Stabilization of Insulin as a Therapeutic Protein Assembly via Enhanced Aromatic-Aromatic Interactions

Nischay K. Rege, Nalinda P. Wickramasinghe, Alisar N. Tustan, Nelson B. Phillips, Vivien C. Yee, Faramarz Ismail-Beigi, & Michael A. Weiss

Purpose of Supplement

We provide 12 Supplemental Figures and 12 Supplemental Tables as cited in the main text. The information pertains to five aspects: (i) tabulation of *ab initio* and molecular mechanics calculations; (ii) details of the crystal structure of Trp^{B26} , Orn^{B29} -insulin, corresponding NMR studies, and comparison with WT insulin; (iii) additional control SEC studies; (iv) additional rat studies; and (iv) details regarding the structural properties of insulin as exploited in protein engineering. The figures and tables are as follows

Table of Contents

Supporting Information (Rege, N.K. *et al*)

Figure S1. Structural summary of rapid-acting insulin analogs. (*A*) Representation of modifications to the sequence of insulin to create rapid-acting insulin analogs. Insulin lispro (developed by analogy to insulin-like growth factor-I [IGF-I]), highlighted the Pro^{B28}/Gly^{D23} dimer contact (highlighted in *sky blue* in panels *B* and *C*) [\(1\)](#page-32-0). Other designs introduced polar or charged substitutions at the dimer interface. Among these were substitution of Thr^{B27} , Typ^{B26} , or Typ^{B16} by Glu [\(2,](#page-32-1)[3\)](#page-32-2) (shown as *black circles* in *A*). Clinical analogs insulin glulisine (Apidra®) and insulin aspart (Novolog®) contain acidic residues at positions B29 (*green*) and B28 (*yellow*), respectively [\(4\)](#page-32-3). These residues were hypothesized to repel residue Glu^{D21}, impairing formation of dimers and higher-order oligomers [\(3\)](#page-32-2). (*B*) The native residues of the positions listed above are highlighted in a 3D structure of an insulin dimer (PDB entry 5INS with color code as above). (*C*) Expanded view of panel *B* with residue positions as labeled.

Figure S2. Past strategies to create basal insulin analogs (see also Table S10). (*A*) Sites in insulin modified in long-acting insulins. Insulin glargine (Lantus®; purple) represents the most successful product [\(5\)](#page-32-4) with pI shifted towards neutrality due to a basic extension of the B chain. Such pIshifted analogs are soluble as formulated at acidic but precipitate in the neutral environment of the SQ space to create a long-lived depot [\(6\)](#page-32-5). A second class of basal insulins are derivatized with long acyl chains at Lys^{B29} [\(7\)](#page-32-6). These include insulin detemir (Levemir[®]; *green circles*) and degludec (Tresiba®; *magenta circles*) [\(8\)](#page-32-7), which bind to albumin as a "circulating depot [\(7\)](#page-32-6)." Insulin degludec forms an acyl-linked dimer of zinc hexamers in its formulation but undergoes a switch to form an acyl-bridged, extended multihexameric linear complex in the SQ depot [\(8](#page-32-7)[,9\)](#page-32-8). These mechanisms are orthogonal to efforts to enhance the intrinsic stability of the insulin hexamer itself. Amino acid residues forming the dimer and hexamer interfaces of insulin in *orange* and *light blue*, respectively. His^{B10}, which coordinates with divalent zinc ions in hexamers, is highlighted in *pink*. (*B*) Dimer- and hexamer interfaces in the zinc insulin hexamer (T_6 ; colorcoded as above). Analogs, designated "hydrophobic insulins," were engineered to contain hydrophobic substitutions at dimer or hexamer interfaces in an effort to stabilize the hexamer. Yet another class of analogs introduced hydrophobic substitutions to the surface of insulin hexamers to create non-specific multi-hexameric precipitates. However, the poor solubility of such analogs

limited their clinical utility [\(5\)](#page-32-4). (*C*) Electrostatic stabilization of the insulin hexamer: view of the negatively charged ring formed by the six Glu^{B13} side chains (highlighted in *green*). An unobstructed view is shown in panel *D*. The most successful effort to stabilize the insulin hexamer involved substitution of Glu^{B13} by Gln, which eliminated repulsion within the Glu residues; however, the Gln^{B13} analog had reduced biological activity $(3,10)$ $(3,10)$.

Figure S3. Comparison of *Ab initio* electrostatics and CHARMM parameters of Tyr and Trp side chains. (*A*) Isosurface representation of electron density and molecular electrostatic potential (MEP) map of Tyr (*left*) and Trp (*right*). Electron density and MEP were calculated using B3LYP method and 6-31G(d) basis set using Gaussian utility Cubegen on Gaussian 09 [\(11\)](#page-32-10). The isosurface map was then generated using Jmol [\(12\)](#page-32-11). (*B*) Ball-and-stick models of Tyr (*left*) and Trp (*right*) side chains. Point charges of each atom as implemented in CHARMM22 are indicated $(13,14)$ $(13,14)$.

Figure S4. Solvent exclusion chromatography (SEC). Elution times of insulin in monomeric and hexameric states in SEC provide context for species identified in Figure 4. (*A*) SEC profile of monomeric lispro (Zn^{2+}) -and phenolfree). The molecular weight (MW; or molecular mass) of the species was 3.1 kDa (calculated as in Fig. 4). (*B*) WT insulin formulated with 0.3 mM ZnCl₂ and phenol was run in a mobilephase containing 50 mM cyclohexanol and 0.3 mM ZnCl₂. The sample eluted as a hexamer (Calculated molecular mass 48 kDa) with dissociation intermediates constituting the "tail" of the peak. (*C*) Calibration plot of SEC column with mobile phase used in panel *B*. Linear fit (*red line*) of $log(MW)$ to V_e/V_0 of MW standards (*blue squares*). The equation of the line is $log(MW) = -1.79$ (V_e/V₀) +6.83 (R²) $= 0.986$). The following standards were used for calibration: thyroglobulin $(669 \text{ kDa}, \text{V}_0)$, ovalbumin (45 kDa), carbonic anhydrase (29 kDa), elastase (26 kDa), ribonuclease A (13.7 kDa), cytochrome C (12.3 kDa) and synthetic peptides (3.6 and 1.2 kDa).

Figure S5. Effects of Zn^{2+} on PD profile of WT insulin. (*A*) Time course of [blood glucose] after SQ injection of WT insulin formulated in absence of Zn^{2+} (*blue*; N=8) or in presence of 0.3 mM ZnCl2 (*black*; N=9). (*B*) Normalized curves from panel *A*.

Figure S6. (*A*) Time course of [glucose] after IV injection of parent pI-shifted insulin analog (blue; N=6) and its Trp^{B26} derivative (red; N=6). (*B*) Bar graph showing area over curve (AOC) of curves from panel *A*. *Black bars* indicate S.E.M. The Trp^{B26} derivative displayed 82 \pm 6% potency relative to the parent analog but is a complete agonist on injection of higher doses

Figure S7. (*A*) Time course of [blood glucose] after SQ injection of zinc-free parent pI-shifted insulin analog (*blue*; N=6) and Trp^{B26} derivative (*red*; N=6). Normalized data are shown in *B*. (*C*-*D*) Corresponding plots of analogs administered after formulation in presence of 0.3 mM ZnCl₂, color-coded as above (N=6).

Figure S8. Comparison of the structure of Trp^{B26}, Orn^{B29}-insulin to a collection of WT insulin structures. (A) R^f -state protomer of Trp^{B26} , Orn^{B29} -insulin. The A chain is shown in *red*, and the B chain in *blue* (B1-B8) or *green* (B9-B30). (*B*) R^f-state protomer of Trp^{B26}, Orn^{B29}-insulin (*sticks*, color coded as above) in relation to extensive set of crystal structures of insulin and insulin analogs (PDB entries: 1BEN, 1G7A, 1RWE, 1EV3, 1EV6, 1MPJ, 1TRZ, 1TYL, 1MPJ, 1ZEG, 1ZNJ and 1ZNI; *gray sticks*). Structures are aligned with respect to the main-chain atoms of residues A1- A21 and B3- B28. RMSD values are given in Table S3. (*C-D*) Corresponding representations of the T-state protomer of Trp^{B26} , Orn^{B29} -insulin, color code as above. PDB entries used for alignment are as follows: 1APH, 1DPH, 1BEN, 1MPJ, 1TRZ, 1TYL, 1TYM, 1RWE, 1G7A, 1ZNI, 2INS and 4INS. RMSD values are given in Table S4.

Figure S9. *Ab initio* calculations of energy of interaction between pairs of isolated aromatic molecules: phenol-phenol (*top*), phenol-benzene (*middle*), and phenol-indole (*bottom*). The phenol-indole pair was determined to form the most stable ETF interaction as a result of Van der Waals forces. Interaction energies were calculated using MP2 method and aug-cc-pVDZ basis set using Gaussian 09 [\(15\)](#page-33-1).

Figure S10. Structural representation of the classical T→R transition of the zinc insulin hexamer. (A) T₆ insulin hexamer. The eight N-terminal residues of the B chain (*orange*) are in an extended conformation (*red box*). (*B*) R₆ hexamer is stabilized by bound phenolic ligands (blue). Residues B1-B8 form an α-helix (*red box*). (*C*) Gly^{B8} serves as the pivot point of the transition between the R- (*blue*) and T-states (*orange*). (*D*) GlyB8 adopts a right-handed conformation (*i.e*., with positive φ angle) in the T-state (*orange sticks*) and a left-handed conformation (negative φ angle) in the Rstate (*blue sticks*).

Figure S11. Ribbon model displaying the orientation of Phe^{B25} in the insulin dimer (PDB 4INS) in two views (*A, B*). The A chain is shown as dark gray ribbon, and the B chain is shown as *green ribbon.* Phe^{B25} (*blue sticks*) is peripheral to the aromatic network formed by Tyr^{B16} , Phe^{B24}, Tyr^{B26} and their symmetry-related mates (*orange sticks*).

Figure S12. Comparison of B26 crevice within TR^f dimers of Trp^{B26} , Orn^{B29}-insulin to WT (1TRZ). (*A*) Stereo view of Tyr^{B26} (*sticks*) of 1TRZ T-state protomer within an electrostatic potential surface (generated using APBS plug-in to $Pymol^{\circledR}(16)$ $Pymol^{\circledR}(16)$) formed by the surrounding residues (Fig. 1*D*, *E*). Positively-charged surfaces are represented in *blue*, negatively charged surfaces in *red*, and neutral surfaces in *white*. (*B, C*) Two possible orientations of the Trp^{B26} that retain γ_1 and γ_2 angles of the WT structure shown in panel A. Due to the asymmetric structure of the Trp side chain, two possible χ_2 angles of correspond in principle to the native Tyr. However, TrpB26 encounters a steric clash with residues in the core of insulin in the orientation shown in *C*. (*D*) Trp^{B26} of the R^f-state protomer from the Trp^{B26} , Orn^{B29} -insulin crystal structure depicted within an electrostatic potential surface formed by surrounding residues. (E) Depiction of Trp^{B26} from panel *D* within the B26 crevice from panel *A* (WT). The Trp^{B26} side chain does not encounter a steric clash. (*F*) Alignment of the naïve model of Trp^{B26} from panel *C* (*green sticks*) to the Trp^{B26} ,

Orn^{B29}-insulin structure (*orange sticks*). Residues are depicted within the WT crevice (*panels A*, *B, C,* and *E*). The steric clash predicted in the naïve model is mitigated in the Trp^{B26} , Orn^{B29}insulin structure by (a) a local shift (0.2 Å) in the backbone of the C-terminal B-chain and (b) a slight difference in the χ_1 torsion angle of Trp^{B26}.

Table S1a. Interaction energy between the B26 residue of an energy minimized model of Trp^{B26} insulin relative to native Tyr^{B26} in the context of a TR^f dimer.

Residue	Interacting Residue	Total Energy of Interaction (kcal/mol)	
		$1TRZ$ Tyo ^{B26}	$1TRZ \overline{\text{Tro}}^{\overline{\text{B26}}}$
B26	Phe^{B24}	-0.358	-0.503
	Phe ^{B25}	-0.090	-0.138
	Tyr^{B16}	0.006	0.009
	Phe ^{D24}	-0.882	-1.501
	Phe ^{D25}	-0.054	-0.095
	Tyr^{D26}	0.000	0.001
	Tyr^{D16}	-1.416	-1.527
D26	Phe ^{B24}	-0.821	-1.478
	Phe ^{B25}	0.005	-0.039
	Tyr^{B26}	0.000	-0.001
	Tyr^{B16}	-0.938	-0.209
	Phe ^{D24}	-0.566	-0.708
	Phe ^{D25}	-0.049	-0.061
	Tyr^{D16}	0.006	0.003
		-6.531	-7.438

Table S1b. Local energy-minimized model of Trp^{B26} insulin relative to native Tyr^{B26} where polar atoms of Tyrosine and Tryptophan rings are replaced with nonpolar synthetic residues.

^a Tabulation of non-bonded interaction energy at the insulin dimer interface in the energy-minimized naïve model of $Tyr^{B26} \rightarrow Trp$ displays interactions most improved by the substitution.

^b This table demonstrates the impact of π - π interactions on non-bonded interaction energy across the insulin dimer interface. Polar oxygen and nitrogen atoms are replaced, *in silico*, with non-polar synthetic atoms eliminating the confounding effects of polar interactions involving those atoms on free-energy calculations.

Residue	Interacting Residue	Total Energy of Interaction (kcal/mol)		
		$1ZNJ$ Tyr B26	1ZNJ Trp ^{B26}	
B26	$Phe^{\overline{B24}}$	-0.501	-0.571	
	Phe ^{B25}	-0.142	-0.146	
	Tyr^{B16}	0.008	0.008	
	Phe ^{D24}	-1.032	-1.308	
	Phe ^{D25}	-0.002	-0.027	
	Tyr/Trp ^{D26}	-0.011	-0.003	
	Tyr^{D16}	-1.569	-1.996	
D26	Phe ^{B24}	-0.738	-1.283	
	Phe ^{B25}	0.004	-0.034	
	Tyr/Trp^{B26}	-0.011	-0.003	
	Tyr^{B16}	-0.613	-1.326	
	Phe ^{D24}	-0.683	-0.635	
	Phe ^{D25}	-0.120	-0.145	
	Tyr^{D16}	0.004	0.008	
Total		-5.406	-7.461	

Table S1c. Interaction energy between the B26 residue of a local energy-minimized model of Trp^{B26} insulin relative to native Tyr^{B26} in the context of an R_2 dimer.

Residue	Interacting Residue	Total Energy of Interaction (kcal/mol)		
		4INS Tyr ^{B26}	4INS Trp ^{B26}	
B26	Phe ^{B24}	-0.397	-0.549	
	Phe ^{B25}	-0.027	-0.058	
	Tyr^{B16}	0.003	0.006	
	Phe ^{D24}	-1.405	-1.181	
	Phe ^{D25}	-0.008	-0.017	
	Tyr/Trp^{D26}	-0.008	-0.054	
	Tyr^{D16}	-1.643	-1.868	
D26	Phe ^{B24}	-1.242	-0.679	
	Phe ^{B25}	-0.016	-0.022	
	Tyr/Trp^{B26}	-0.016	-0.054	
	Tyr^{B16}	-2.710	-1.261	
	Phe ^{D24}	-0.365	-0.466	
	Phe ^{D25}	-0.049	-0.140	
	Tyr^{D16}	0.004	0.006	
Total		-7.879	-6.337	

Table S1d. Interaction energy between the B26 residue of a local energy-minimized model of Trp^{B26} insulin relative to native Tyr^{B26} in the context of an T₂ dimer.

Table S2. Data collection and refinement statistics pertaining to the crystal structure of Trp^{B26}, Orn^{B29}-insulin.

PDB ID	main-chain RMSD $\rm(\AA)$	Side-chain RMSD $\rm(\AA)$
1RWE R^f $(T_3R_3)^a$	0.70	1.25
1BEN R^f (T ₃ R ₃)	0.39	0.97
1TRZ R^f (T ₃ R ₃)	0.43	0.95
1TYL R^f (T ₃ R ₃)	0.46	1.04
1 TYM R^f (T ₃ R ₃)	0.44	1.10
1ZNI R^f (T ₃ R ₃)	0.45	0.89
1MPJ $R^f(T_3R_3)$	0.45	0.91
$3MTHRf(T_3R_3)$	0.52	1.18
1LPH $R^f(T_3R_3)$	0.53	1.07
$1G7A R1f$ (rhom T ₃ R ₃)	0.61	1.06
$R2^f$	0.79	1.53
1EV3 R1 (rhom R_6) ^b	0.71	1.59
R ₂	0.56	1.32
1ZEG R1 (rhom R_6)	0.52	1.35
R ₂	0.61	1.82
1ZEH R1 (rhom R_6)	0.55	1.25
R ₂	0.66	1.85
1ZNJ R1 (monoclinic R_6) ^c	0.62	1.37
R ₂	0.56	1.66
R ₃	0.76	1.79
R ₄	0.61	1.18
R ₅	0.63	1.47
R ₆	0.66	1.23

Table S3. RMSD values of main-chain and side-chains of the R-state protomer in the crystal structures of Trp^{B26} , Orn^{B29} -insulin and selected WT R_6 and T_3R_3 hexamers.

Continued on next page

 $a R^f$ or "R-frayed" indicates that the first three residues of the insulin B chain (B1-B3) are in a coil conformation rather than part of the B1-B16 helix, as in standard R-state structures.

^b Rhombohedral crystals contain two unique R-state protomers in the asymmetrical unit that are related by 3-fold symmetry to the two other R-state dimers comprising an R_6 hexamer.

 c Monoclinic R₆ crystals contain 6 individual R-state protomers in the asymmetrical unit.

PDB ID	Main-chain RMSD $\rm(\AA)$	side-chain RMSD (Å)
1RWE T(T ₃ R ₃)	0.302	1.18
1BEN T (T_3R_3)	0.289	0.67
$1TRZT(T_3R_3)$	0.316	1.09
1 TYL T (T_3R_3)	0.319	1.38
1 TYM T (T_3R_3)	0.295	1.07
1ZNI T(T ₃ R ₃)	0.283	0.68
1MPJ T (T_3R_3)	0.382	0.84
$3MTHT(T_3R_3)$	0.318	0.94
$1LPH T (KP T3R3)$	0.486	0.95
1G7A T1 (rhom $T_3R_3)^a$	0.307	0.75
T ₂	0.334	1.16
4INS T1 (2ZN T ₆)	1.127	1.94
T ₂	0.686	1.49
1APH (cubic T_2) ^b	0.620	1.00
1BPH (cubic T_2)	0.653	1.12
1CPH (cubic T ₂)	0.667	1.32
1DPH (cubic T_2)	0.659	1.21
Average	0.47 ± 0.11	1.11 ± 0.30

Table S4. RMSD values of main-chain and side-chains of the T-state protomer of the crystal structures of Trp^{B26} , Orn^{B29} -insulin and selected WT T_6 and T_3R_3 hexamers

^a Rhombohedral crystals contain two unique T-state protomers in the asymmetrical unit that are related by 3-fold symmetry to the two other TR dimers comprising an T_3R_3 hexamer.

 b Cubic T₂ crystals contain two T-state protomers forming an insulin dimer. They are zinc-free crystals.

PDB ID	chain	χ_1	χ_2
Trp^{B26} , Orn ^{B29} -insulin	B	167.131	-108.457
1TRZ	\boldsymbol{B}	-179.531	78.679 ^b
4INS	B	170.768	66.198
	D	175.602	-113.722
2INS	B	170.867	63.894
	D	170.708	-111.824
1ZNI	D	178.670	67.486
1BEN	\boldsymbol{B}	178.543	67.578
	D	172.629	60.183
3MTH	D	178.321	70.378

Table S5. Comparison of χ_1 and χ_2 dihedral-angle values of the T protomer of the Trp^{B26}, Orn^{B29}insulin crystal structure to the B26 side chains of WT T_6 or T_3R_3 insulin crystal structures^a

^a Tabulation of χ_1 and χ_2 angles demonstrates the native-like orientation of the Trp^{B26} ring of the T protomomer of the Trp^{B26}, Orn^{B29}-insulin crystal structure.

b Due to the symmetric structure of the Tyr side chain, χ_2 and χ_2 -180° are equivalent. For example, 78.679° is equivalent to -101.321°.

		$ -$	
PDB ID	chain	χ_1	χ_2
WB26	$\mathbf D$	-179.108	-102.751
1TRZ	D	179.520	81.505^{b}
1ZNJ	\bf{B}	-173.898	82.727
	D	168.865	-90.454
	$\mathbf F$	-176.674	69.837
	H	176.540	74.677
	$\bf J$	-173.848	80.139
	L	175.389	72.440
	B	-178.950	77.319
1BEN	D	-162.232	84.877
1MPJ	B	-177.306	-89.456
3MTH	$\, {\bf B}$	-173.822	-87.442

Table S6. Comparison of χ_1 and χ_2 dihedral angle values of the R protomomer of Trp^{B26}, Orn^{B29}insulin crystal structure to the B26 side chains of WT R₆ or R^f insulin crystal structures^a

^a Tabulation of χ_1 and χ_2 angles demonstrates the native-like orientation of the Trp^{B26} ring of the R protomomer of the Trp^{B26}, Orn^{B29}-insulin crystal structure.

^b Due to the symmetric structure of the Tyr side chain, χ_2 and χ_2 -180° are equivalent.

Trp Proton	Residue	Proton	Predicted Distance ^a (\AA)	NOE Strength ^b
$H - \epsilon 3$	$\text{I} \text{I} \text{e}^{\overline{\text{A2}}}$	$H-81$	3.82	W
$H-$ ζ3	Val ^{B12}	$H-\gamma1^c$	6.93	S
H-ζ3	Val^{B12}	$H-\gamma$ 2	4.61	${\bf S}$
$H-\eta$ 2	Val ^{B12}	$H-\gamma$ 1	7.61	m
$H - \eta$ 2	Val^{B12}	H-γ2	4.68	m
$H-\zeta^2$	Val^{B12}	$H-\gamma$ 1	7.38	W
$H-\zeta^2$	Val^{B12}	H-γ2	4.40	W
$H - \epsilon 3$	Val ^{B12}	$H-\gamma$ 1	8.85	W
$H - \epsilon 3$	Val ^{B12}	$H-\gamma$ 2	4.24	W
$H - \epsilon 3$	Leu^{B15}	$H-81$	3.24	W
$H - \epsilon 3$	Leu^{B15}	$H-\delta 2$	6.04	W
H-ζ3	Leu^{B15}	$H-81$	4.74	W
$H-$ ζ3	Leu^{B15}	$H-\delta 2$	6.85	W

Table S7. Comparison of predicted proton-proton distances involving Trp^{B26} from Trp^{B26} , Orn^{B29}-insulin crystal structure and Trp^{B26} -associated NOEs in Trp^{B26} lispro

^aPredicted distances were obtained from the AB (T-state) protomer of the Trp^{B26} , Orn^{B29} -insulin crystal structure.

^b NOEs were categorized as strong (s, <4.0 Å), medium (m, 4.4-5.5 Å), and weak (w, >5.5 Å). ^c Predicted proton-proton distances are provided for protons at equivalent carbon positions although associated NOEs are difficult to differentiate.

Tyr Proton	Residue	Proto $\mathbf n$	Predicted Distance ^a (\AA)	NOE Strength ^b
$H\text{-}\epsilon$	$I1e^{A2}$	$H-\delta 1$	5.17	W
H - ε	Val^{A3}	$H-\gamma1^c$	6.82	W
H -E	Val^{A3}	$H-\gamma 2$	4.05	W
H- δ	Val ^{A3}	$H-\gamma 1$	4.12	W
$H-\delta$	$\mathrm{Val}^{\mathrm{A3}}$	$H-\gamma$ 2	6.25	W
H -ε	Leu^{B11}	$H-\beta$	4.61	W
H -E	Leu^{B11}	$H-\gamma$	3.13	W
H - ε	Leu ^{B11}	$H-\delta 1$	5.42	W
$\operatorname{H-\epsilon}$	Leu ^{B11}	$H-\delta 2$	3.67	W
H- δ	Leu^{B11}	$H-\delta 1$	5.75	W
$H-\delta$	Leu^{B11}	$H-\delta 2$	5.24	W
H - ε	Val ^{B12}	$H-\gamma$ 1	7.21	W
H - ε	Val ^{B12}	$H-\gamma$ ²	4.72	W
$H\text{-}\epsilon$	Leu^{B15}	$H-81$	5.12	W
H - ε	Leu^{B15}	$H-\delta 2$	7.80	W
$H-\delta$	Leu^{B15}	$H-\delta$ 1	3.14	${\bf m}$
H- δ	Leu^{B15}	$H-\delta 2$	6.14	$\mathbf W$
H- δ	Phe ^{B24}	$\rm H\text{-} \epsilon$	4.55	W
$H-\delta$	Thr^{B27}	$H-\gamma$ 2	6.88	W

Table S8. Comparison of predicted proton-proton distances involving Tyr^{B26} from WT crystal structure and Tyr^{B26}-associated NOEs from lispro NMR spectrum.

^a Predicted distances were obtained from the AB (T-state) protomer of WT structure 1TRZ.

^b NOEs were categorized as strong (s, <4.0 Å), medium (m, 4.4-5.5 Å) or weak (w, >5.5 Å). c Predicted proton-proton distances are provided for protons at equivalent carbon positions although associated NOEs are difficult to differentiate.

Table S9a. Energy of non-bonded interaction of Tyr^{B26} with local aromatic residues^a

^a As calculated from WT structure 1TRZ.

Table S9b. Energy of non-bonded interaction of Trp^{B26} with neighboring aromatic residues^a

^a Table (calculated from Trp^{B26} Orn^{B29} structure) show specific aromatic-aromatic aromatic interactions improved in the TR dimer of Trp^{B26} Orn^{B29}-insulin in relation to WT insulin.

Table S10. Summary of previous designs of basal insulin analogs^a

 $^{\rm a}$ This table underscores the challenges associated with the development of insulin analogs with protracted PD profiles. Trp $^{\rm B26}$ insulin analogs are unique in their ability to protract the PD profile of insulin by stabilizing the hexamer without significantly compromising the potency of the analog.

		B26		D26
Residue	r	Dih.A	r	Dih.A
B16	13.6	76	6.0	52
B24	7.3	85	5.9	87
B25	10.9	34	10.6	70
B26			12.0	77
D ₁₆	5.8	58	13.1	89
D ₂₄	5.8	38	7.5	78
D ₂₅	12.6	38	9.5	45
D26	12.0	77		

Table S11a. Intercentroid distances and angles between Tyr^{B26} and local aromatic residues^a

Table S11b. Intercentroid distances and angles between Tyr^{B26} and local aromatic residues^a

		B26	D26	
Residue	r	Dih.A	r	Dih.A
B16	12.8	90	5.5	20
B24	7.8	79	5.9	25
B25	9.7	37	13.0	44
B26			12.2	73
D ₁₆	5.6	51	13.7	66
D24	6.6	46	7.0	89
D ₂₅	10.9	73	11.1	10
D26	12.2	73		

^a Tables S11a and S11b (respectively calculated from WT structure 1TRZ and the present crystal structure of the TrpB26 analog) indicate the character of aromaticaromatic interactions involving Tyr or Trp^{B26} at the insulin dimer interface. Whereas some interactions are classical ETF interactions, others show some deviation in ring-to-ring dihedral angle (Dih.A).

Side Chain	ΔSA	$\Delta G_{\rm u}$ ^b (kcal/mol)	Side Chain	ASA	ΔG_{u} (kcal/mol)
Tvr^{B26}	68.9% 0.49 $^{\rm a}$		$Trp^{B2\overline{6}}$	69.3%	1.4
Tyr^{D26}	78.6% 0.56		Trp^{D26}	70.1%	1.5
Mean $\Delta\Delta G_u$: 0.9 kcal/mol (not observed)					

Table S12a. Thermodynamic stabilization associated with the B26 side chain during folding of insulin analogs a

Table S12b. Thermodynamic stabilization associated with the B26 side chain during dimerization of insulin analogs^a

Side Chain	ΔSA.	Λ G _u (kcal/mol)	Side Chain	ASA	$\Delta G_{\rm n}$ (kcal/mol)	
Tvr^{B26}	25.9% 0.18		Trp^{B26}	26.6% 0.55		
Tyr^{D26}	78.6% 0.56		Trp^{D26}	26.5% 0.55		
Mean $\Delta\Delta G_{\rm u}$: 0.37 + 0.42 = 0.8 kcal/mol/dimer interface						

^a The relative thermodynamic stabilities of Tyr^{B26} and Trp^{B26} insulin monomers calculated using hydrophobic transfer free energies contrasted with those determined by guanidinedenaturation assays. However, NMR data suggests dimerization may be favored in Trp^{B26} lispro to a greater extent than in native lispro.

^b Tabulated values of respective H₂O/octanol transfer free energies are -0.71 kcal/mol (Tyr) and -2.09 kcal/mol (Trp) as described [\(29\)](#page-34-2). This calculation pertains only to changes in exposure of residue B26 and does not consider secondary changes in exposure of neighboring side chains.

PDB ID or Reference	Modification		
1EV6	WT Human Insulin		
1ZNJ	Porcine insulin: Ala ^{B30} at chain terminus		
4E7V	Bovine Insulin: Ala ^{A8} , Val ^{A10} , Ala ^{B30} at surfaces		
3GKY	His^{A8} , Val ^{A16} at surface (A8) or in core (A16)		
4P65	Cha ^{B24} at dimer interface		
3ROV	D-Ala ^{B20} , D-Ala ^{B23} , lispro within β -turn		
5HRQ	cis-hydroxyproline ^{B28} at edge of dimer interface		
5HPU	trans-hydroxyproline ^{B28} at edge of dimer interface		
5URU	Dihydroxyproline ^{B28} at edge of dimer interface		
5UQA	4-fluoro-proline ^{B28} at edge of dimer interface		
2WS6	N-methyl-Tyr ^{B26} at dimer interface and inter-chain		
2WS7	Pro ^{B26} at dimer interface		
3ZS2	Tyr^{B25} , N-methyl-Phe ^{B26} , lispro at dimer interface		
3ZQR	N-methyl-Phe ^{B25} at dimer interface		
5EMS	3-iodotyrosine ^{B26} , Norleucine ^{B29} at dimer interface		
1QIY	Tyr ^{B5} at inter-chain crevice		
1ZEG	Asp ^{B28} at edge of dimer interface		
3ZU1	Des-B30, Ne- ω -carboxyheptadecanoyl-Lys ^{B29}		
Derewenda, 1987 (30)	$Val^{B12} \rightarrow$ Ile at dimer interface		

Table S13. Native-like crystal structures of R_6 hexamers formed by mutant insulin.

Supplemental References:

- 1. Holleman, F., and Hoekstra, J. B. (1997) Insulin lispro. *N. Engl. J. Med.* **337**, 176-183
- 2. Brange, J., Ribel, U., Hansen, J. F., Dodson, G., Hansen, M. T., Havelund, S., Melberg, S. G., Norris, F., Norris, K., and Snel, L. (1988) Monomeric insulins obtained by protein engineering and their medical implications. *Nature* **333**, 679-682
- 3. Brange, J., and Vølund, A. (1999) Insulin analogs with improved pharmacokinetic profiles. *Adv. Drug Deliv. Rev.* **35**, 307-335
- 4. van Bon, A. C., Bode, B. W., Sert-Langeron, C., DeVries, J. H., and Charpentier, G. (2011) Insulin glulisine compared to insulin aspart and to insulin lispro administered by continuous subcutaneous insulin infusion in patients with type 1 diabetes: a randomized controlled trial. *Diabetes Technol. Ther.* **13**, 607-614
- 5. Markussen, J. M., Hougaard, P., Ribel, U., Sørensen, A. R., and Sørensen, E. (1987) Soluble, prolonged-acting insulin derivatives. I. Degree of protraction and crystallizability of insulins substituted in the termini of the B-chain. *Protein Eng.* **1**, 205-213
- 6. Gillies, P. S., Figgitt, D. P., and Lamb, H. M. (2000) Insulin glargine. *Drugs* **59**, 253-260
- 7. Markussen, J., Havelund, S., Kurtzhals, P., Andersen, A., Halstrøm, J., Hasselager, E., Larsen, U., Ribel, U., Schäffer, L., and Vad, K. (1996) Soluble, fatty acid acylated insulins bind to albumin and show protracted action in pigs. *Diabetologia* **39**, 281-288
- 8. Gough, S., Harris, S., Woo, V., and Davies, M. (2013) Insulin degludec: overview of a novel ultra long‐acting basal insulin. *Diab. Obes. Metab.* **15**, 301-309
- 9. Steensgaard, D. B., Schluckebier, G., Strauss, H. M., Norrman, M., Thomsen, J. K., Friderichsen, A. V., Havelund, S., and Jonassen, I. (2013) Ligand-controlled assembly of hexamers, dihexamers, and linear multihexamer structures by the engineered acylated insulin degludec. *Biochemistry* **52**, 295-309
- 10. Bentley, G. A., Brange, J., Derewenda, Z., Dodson, E. J., Dodson, G. G., Markussen, J., Wilkinson, A. J., Wollmer, A., and Xiao, B. (1992) Role of B13 Glu in insulin assembly. The hexamer structure of recombinant mutant (B13 Glu→Gln) insulin. *J. Mol. Biol.* **228**, 1163-1176
- 11. Frisch, M. J., Trucks, G. W., Schlegal, H. B., Scuseria, G. E., Robb, M. A., Cheeseman, J. R., Scalmani, G., Barone, V., Mennucci, B., Petersson, G. A., Nakatsuji, H., Caricato, M., Li, X., Hratchian, H. P., Izmaylov, A. F., Bloino, J., Zheng, G., Sonnenberg, J. L., Hada, M., Ehara, M., Toyota, K., Fukuda, R., Hasegawa, J., Ishida, M., Nakajima, T., Honda, Y., Kitao, O., Nakai, H., Vrenven, T., Montogomery, J. A., Peralta, J. E., Ogliaro, F., Bearpark, M., Heyd, J. J., Brothers, E., Kudin, K. N., Staroverov, V. N., Kobayashi, R., Normand, J., Raghavachari, K., Rendell, A., Burant, J. C., Iyengar, S. S., Tomasi, J., Cossi, M., Rega, N., Millam, J. M., Klene, M., Knox, J. E., Cross, J. B., Bakken, V., Adamo, C., Jaramillo, J., Gomperts, R., Stratmann, R. E., Yazyev, O., Austin, A. J., Cammi, R., Pomelli, C., Ochterski, J. W., Martin, R. L., Morokuma, K., Zakrzewski, V. G., Voth, G. A., Salvador, P., Dannenberg, J. J., Dapprich, S., Daniels, A. D., Farkas, Foresman, J. B., Ortiz, J. V., Cioslowski, J., and Fox, D. J. (2009) Gaussian09 Revision D.01. Gaussian Inc., Wallingford, CT
- 12. Willighagen, E., and Howard, M. (2007) Fast and Scriptable Molecular Graphics in Web Browsers without Java3D. in *Nature Precedings*
- 13. Brooks, B. R., Brooks, C. L., Mackerell, A. D., Nilsson, L., Petrella, R. J., Roux, B., Won, Y., Archontis, G., Bartels, C., Boresch, S., Caflisch, A., Caves, L., Cui, Q., Dinner, A. R., Feig, M., Fischer, S., Gao, J., Hodoscek, M., Im, W., Kuczera, K., Lazaridis, T., Ma, J., Ovchinnikov, V., Paci, E., Pastor, R. W., Post, C. B., Pu, J. Z., Schaefer, M., Tidor, B., Venable, R. M., Woodcock, H. L., Wu, X., Yang, W., York, D. M., and Karplus, M. (2009) CHARMM: The biomolecular simulation program. *J. Comput. Chem.* **30**, 1545-1614
- 14. MacKerell, A. D., Brooks, B., Brooks, C. L., Nilsson, L., Roux, B., Won, Y., and Karplus, M. (2002) CHARMM: The Energy Function and Its Parameterization. in *Encyclopedia of Computational Chemistry* (Schleyer, P. v. R. C., N. L. A. T. , Gasteiger, J., Kollman, P. A., Schaefer, H. F., III, Schreiner, P. R. S. ed.), John Wiley & Sons, Ltd, Chichester. pp 271-277
- 15. Frisch, M. J., Trucks, G. W., Schlegel, H. B., Scuseria, G. E., Robb, M. A., Cheeseman, J. R., Scalmani, G., Barone, V., Mennucci, B., Petersson, G. A., Nakatsuji, H., Caricato, M., Li, X., Hratchian, H. P., Izmaylov, A. F., Bloino, J., Zheng, G., Sonnenberg, J. L., Hada, M., Ehara, M., Toyota, K., Fukuda, R., Hasegawa, J., Ishida, M., Nakajima, T., Honda, Y., Kitao, O., Nakai, H., Vreven, T., Montgomery, J., J. A. , Peralta, J. E., Ogliaro, F., Bearpark, M., Heyd, J. J., Brothers, E., Kudin, K. N., Staroverov, V. N., Kobayashi, R., Normand, J., Raghavachari, K., Rendell, A., Burant, J. C., Iyengar, S. S., Tomasi, J., Cossi, M., Rega, N., Millam, J. M., Klene, M., Knox, J. E., Cross, J. B., Bakken, V., Adamo, C., Jaramillo, J., Gomperts, R., Stratmann, R. E., Yazyev, O., Austin, A. J., Cammi, R., Pomelli, C., Ochterski, J. W., Martin, R. L., Morokuma, K., Zakrzewski, V. G., Voth, G. A., Salvador, P., Dannenberg, J. J., Dapprich, S., Daniels, A. D., Farkas, O., Foresman, J. B., Ortiz, J. V., Cioslowski, J., and Fox, D. J. (2009) Gaussian 09, Revision A.02. in *Gaussian, Inc., Wallingford CT*, Revision A.02 Ed.
- 16. Baker, N. A., Sept, D., Joseph, S., Holst, M. J., and McCammon, J. A. (2001) Electrostatics of nanosystems: application to microtubules and the ribosome. *Proc. Nat. Acad. Sci.* **98**, 10037- 10041
- 17. Galloway, J. A., Hooper, S. A., Spradlin, C. T., Howey, D. C., Frank, B. H., Bowsher, R. R., and Anderson, J. H. (1992) Biosynthetic human proinsulin. Review of chemistry, *in vitro* and *in vivo* receptor binding, animal and human pharmacology studies, and clinical trial experience. *Diab. Care* **15**, 666-692
- 18. Hagedorn, H., Jensen, B. N., Krarup, N., and Wodstrup, I. (1936) Protamine insulinate. *JAMA* **106**, 177-180
- 19. Lauritzen, T., Pramming, S., Gale, E., Deckert, T., and Binder, C. (1982) Absorption of isophane (NPH) insulin and its clinical implications. *Brit. Med. J.* **285**, 159-162
- 20. Owens, D. R., and Bolli, G. B. (2008) Beyond the era of NPH insulin—long-acting insulin analogs: chemistry, comparative pharmacology, and clinical application. *Diabetes Technol. Ther.* **10**, 333-349
- 21. Stewart, W. J., Mcsweeney, S. M., Kellett, M. A., Faxon, D. P., and Ryan, T. J. (1984) Increased risk of severe protamine reactions in NPH insulin-dependent diabetics undergoing cardiac catheterization. *Circulation* **70**, 788-792
- 22. Markussen, J., Diers, I., Engesgaard, A., Hansen, M. T., Hougaard, P., Langkjaer, L., Norris, K., Ribel, U., Sorensen, A. R., Sorensen, E., and et al. (1987) Soluble, prolonged-acting insulin derivatives. II. Degree of protraction and crystallizability of insulins substituted in positions A17, B8, B13, B27 and B30. *Protein Eng.* **1**, 215-223
- 23. Markussen, J., Diers, I., Hougaard, P., Langkjaer, L., Norris, K., Snel, L., Sørensen, A. R., Sørensen, E., and Voight, H. O. (1988) Soluble, prolonged-acting insulin derivatives. III. Degree of protraction, crystallizability and chemical stability of insulins substituted in positions A21, B13, B23, B27 and B30. *Protein Eng.* **2**, 157-166
- 24. Kurtzhals, P., Havelund, S., Jonassen, I., Kiehr, B., Larsen, U., Ribel, U., and Markussen, J. (1995) Albumin binding of insulins acylated with fatty acids: characterization of the ligandprotein interaction and correlation between binding affinity and timing of the insulin effect in vivo. *Biochem. J.* **312**, 725-731
- 25. Suissa, S., Azoulay, L., Dell'Aniello, S., Evans, M., Vora, J., and Pollak, M. (2011) Long-term effects of insulin glargine on the risk of breast cancer. *Diabetologia* **54**, 2254-2262
- 26. Yamamoto, C., Miyoshi, H., Fujiwara, Y., Kameda, R., Ichiyama, M., Nomoto, H., Kameda, H., Nakamura, A., and Atsumi, T. (2016) Degludec is superior to glargine in terms of daily glycemic variability in people with type 1 diabetes mellitus. *Endocr. J.* **63**, 53-60
- 27. Sommerfeld, M. R., Muller, G., Tschank, G., Seipke, G., Habermann, P., Kurrle, R., and Tennagels, N. (2010) *In vitro* metabolic and mitogenic signaling of insulin glargine and its metabolites. *PLoS One* **5**, e9540
- 28. Jonassen, I., Havelund, S., Hoeg-Jensen, T., Steensgaard, D. B., Wahlund, P. O., and Ribel, U. (2012) Design of the novel protraction mechanism of insulin degludec, an ultra-long-acting basal insulin. *Pharm. Res.* **29**, 2104-2114
- 29. Wimley, W. C., and White, S. H. (1996) Experimentally determined hydrophobicity scale for proteins at membrane interfaces. *Nat. Struct. Molec. Biol.* **3**, 842
- 30. Derewenda, U., Derewenda, Z., Dodson, G., and Brange, J. (1987) The Crystal-Structure of the B-12 Ile Human Insulin Prepared by Site-Directed Mutatgenesis. *Protein Eng. 222, 425-433*