

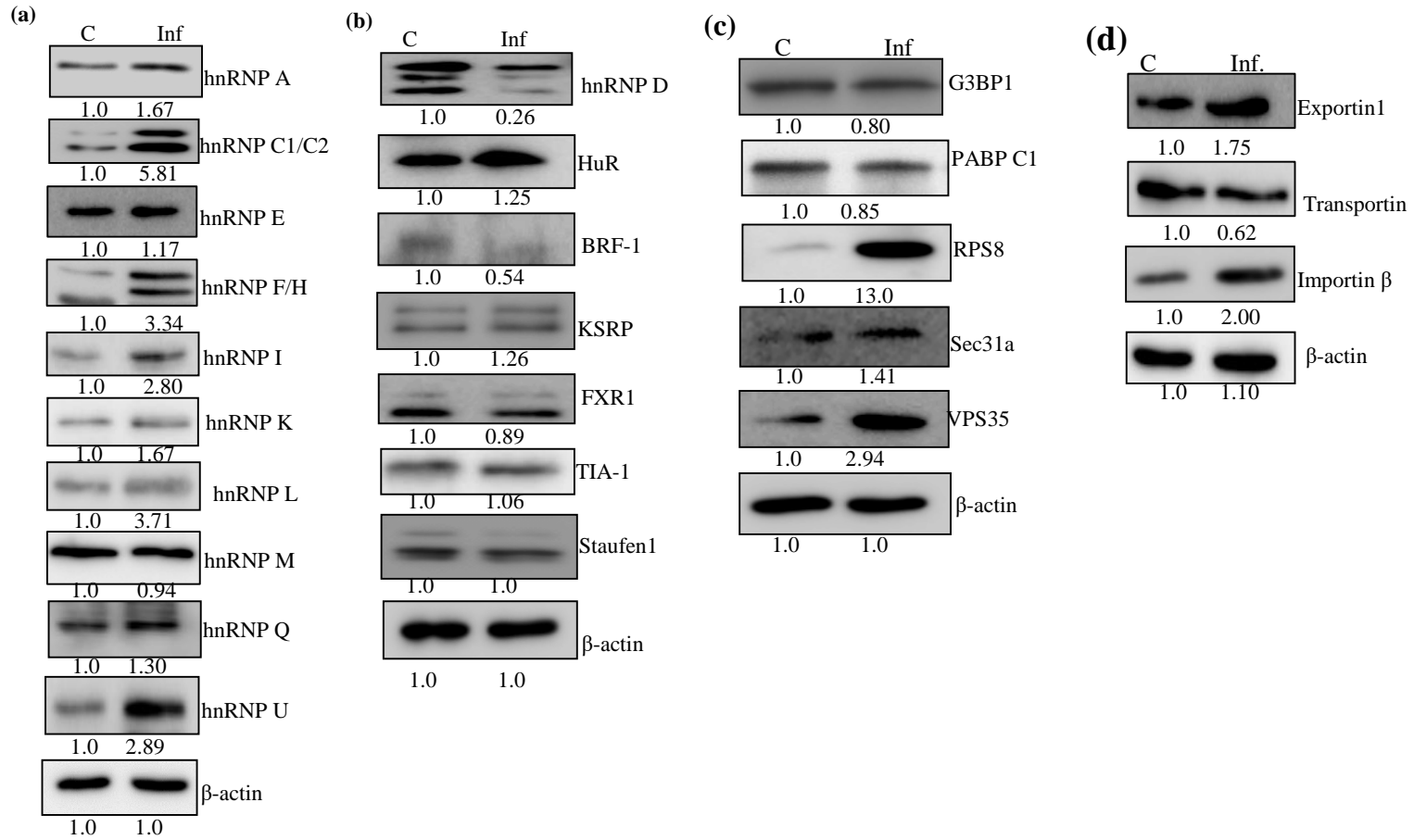
Supplemental Table S3 Time-dependent fold change in the level of cellular proteins in the cytoplasm and the nucleus during the course of rotavirus infection. The fold change in the cytoplasm or the nucleus was calculated by normalizing the values at different time points with reference to that in the respective compartments in the serum-grown control cells (Figs. 2, 3, 4 and 6). The fold changes in the cytoplasm and nucleus of host protein levels in each figure are normalized with respect to the levels of β -actin and PCNA, respectively, shown in the same figure. For proteins hnRNP A1, D, F/H, K/J, L, Q and U, HuR, KSRP and BRF1 which were undetectable in the cytoplasm of control cells, the fold change in the cytoplasm with reference to that in serum-control control cells could not be calculated. So it was calculated by normalizing the values with reference to that of their first appearance in the respective compartments during the course mock infection or virus infection. For example, the fold change for hnRNP L in cytoplasm of mock-infected cells was calculated with reference to that at 2 hr post serum starvation. Quantification of band intensities in western blots were done using Image J software. S.C., serum-grown control cells. The significant fold changes in the protein levels at 8 hpi are highlighted.

S.No.	Cellular protein	Cytoplasm S.C	Mock-infected Cytoplasm (hpi)				Infected Cytoplasm (hpi)				Nucleus S.C.	Mock-infected Nucleus (hpi)				Infected Nucleus (hpi)			
			2	4	8	10	2	4	8	10		2	4	8	10	2	4	8	10
1	Importin- β	1.0	1.1	1.2	0.9	1.0	1.0	1.6	2.0	2.0	1	1.4	0.9	1.4	1.4	1.3	1.3	1.4	1.4
2	Exportin1	1.0	0.8	0.9	0.4	0.4	1.2	1.9	2.8	3.0	1	1.4	1.3	1.3	1.4	1.4	1.3	1.4	1.4
3	Transportin	1.0	1.1	0.9	0.9	1.0	0.8	0.8	1.0	1.3	1	0.9	1.0	1.0	0.7	0.6	0.8	0.2	0.3
4	Ran	1.0	0.9	0.9	0.8	0.9	1.3	1.7	2.7	1.9	1	1.5	0.9	1.0	0.8	1.2	1.3	1.3	1.3
5	KSRP	1.0	1.2	0.5	1.2	0.2	1.8	1.2	3.0	6.0	1	0.6	1.5	1.4	1.8	1.3	1.9	1.7	2.2
6	TTP	1.0	0.6	0.9	1.3	1.6	1.1	2.0	1.2	1.1	1	1.3	1.4	1.2	1.2	1.0	1.1	0.9	1.2
7	FXR1	1.0	1.2	1.3	1.1	1.4	1.4	1.9	1.4	1.1	1	0.5	0.4	0.3	0.2	0.5	0.4	0.4	0.3
8	hnRNP D	--	---	---	---	---	---	1.0	1.4	1.1	1	0.8	0.9	1.0	0.7	1.0	0.1	---	---
9	HuR	--	---	---	---	---	---	1.0	11.4	7.8	1	0.7	0.8	0.3	0.5	0.8	0.4	0.2	---
10	BRF1	--	---	---	---	---	1.0	1.2	2.3	3.1	1	0.6	0.8	1.2	1.2	1.5	0.2	---	---
11	TIAL-1	1.0	2.2	3.3	2.5	3.3	2.5	3.1	2.2	1.6	1	0.8	0.9	0.8	0.8	0.8	0.5	0.4	0.2
12	TIA-1	1.0	2.0	1.3	0.4	0.2	0.3	1.8	1.4	1.6	1	2.5	0.6	2.0	2.7	1.9	0.8	1.8	1.6
13	hnRNP A1	1.0	0.6	2.9	0.6	0.8	2.2	9.5	21.0	21.1	1	0.8	0.7	0.4	0.4	0.6	0.5	0.4	0.06
14	hnRNP C1/C2	1.0	1.6	2.6	1.9	3.0	1.4	2.0	2.3	4.8	1	0.8	0.6	0.3	0.4	0.2	0.6	0.8	0.7
15	hnRNP E (PCBP1)	1.0	1.3	1.1	2.2	2.0	1.6	1.6	2.8	2.1	1	2.0	2.0	1.7	0.8	1.1	2.2	2.5	2.1
16	hnRNP F/H	---	1.0	0.5	2.5	0.9	3.1	5.8	7.9	5.6	1	2.0	0.8	2.2	2.6	2.0	2.0	2.2	2.7
17	hnRNP I (PTBP1)	1.0	1.3	0.9	0.9	1.0	1.9	2.6	5.4	9.6	1	1.4	1.1	1.4	1.4	1.1	1.0	0.8	0.4
18	hnRNP K/J	---	---	---	---	---	1.0	1.6	5.4	2.5	1	1.0	1.0	1.0	1.0	1.0	0.9	0.6	0.2
19	hnRNP L	---	1.0	2.3	6.8	7.6	4.2	9.3	9.8	28.0	1	0.9	0.9	0.4	0.6	0.5	0.7	0.5	0.1
20	hnRNP M	1.0	0.7	1.4	1.3	1.7	1.1	2.5	4.3	2.0	1	0.6	0.9	0.7	0.8	1.0	0.8	0.6	0.6
21	hnRNP Q	---	---	---	---	---	---	---	---	---	1	1.8	1.1	1.6	1.7	1.5	1.3	1.6	1.5
22	hnRNP U	---	---	---	---	---	1.0	1.2	1.8	1.6	1	2.3	1.6	2.7	2.3	2.2	2.4	2.0	1.5
23	Staufen-1	1.0	1.0	1.1	0.7	0.8	1.0	1.1	1.3	1.3	1	1.7	1.1	1.6	1.6	1.6	1.4	1.9	1.8
24	PABP C1	1.0	1.2	3.5	0.7	0.5	0.4	0.5	0.1	0.1	1.0	3.1	4.0	5.5	2.5	3.1	5.8	9.8	2.5
25	Sec31A	1.0	0.9	0.7	0.8	0.06	0.6	0.7	3.7	3.0	---	---	---	---	---	---	---	---	---
26	VPS35	1.0	2.1	3.2	4.4	2.5	2.7	5.6	6.8	7.5	---	---	---	---	---	---	---	---	---
27	RPS8	1.0	0.8	1.0	0.6	0.8	0.5	0.5	2.3	0.8	1.0	1.8	10.8	9.0	2.6	2.0	7.2	10.6	2.0
28	β -actin (for FIG. 2c, 2d)	1.0	1.1	1.2	1.0	1.1	1.1	1.0	1.0	1.2	--	---	---	---	---	---	---	---	---
29	β -actin (for FIG. 3c)	1.0	0.7	1.0	0.9	1.0	1.0	1.0	1.0	1.0	--	---	---	---	---	---	---	---	---
30	β -actin (for FIG. 6a)	1.0	1.2	1.1	1.0	1.2	1.0	1.1	1.0	0.9	--	---	---	---	---	---	---	---	---
31	β -actin (for FIG. 3d-e)	1.0	0.9	0.7	0.8	0.7	1.0	0.8	0.9	1.0	--	---	---	---	---	---	---	---	---
32	PCNA (for FIG. 2c,2d)	--	---	---	---	---	---	---	---	---	1	1.4	1.4	1.6	1.8	1.4	1.5	1.7	1.4
33	PCNA (for FIG. 3c)	--	---	---	---	---	---	---	---	---	1	0.8	1.2	0.8	0.8	1.5	1.2	0.9	1.0
34	PCNA (for FIG. 6a)	--	---	---	---	---	---	---	---	---	1	1.2	1.0	1.1	1.2	1.0	1.0	1.2	1.0
35	PCNA (for FIG. 3d-e)	--	---	---	---	---	---	---	---	---	1.0	1.0	1.0	0.9	0.9	0.8	1.0	1.0	0.7

Supplemental Table S4 Details including source, host, dilution, and catalogue numbers of different antibodies. MABs against the cellular proteins were primarily used in confocal imaging experiments. PABs were used only when MABs were not available. Lower and higher dilutions were used in confocal imaging and immunoblotting experiments, respectively.

Antibody	Source	Dilution	Antibody	Source	Dilution
HuR Millipore: 07-468	Rabbit	1:5000	hnRNP E1/PCBP1	MAB	1:2000
Santacruz: Sc-374285 Mouse	MAB	1:500	Santacruz Biotech:sc-137249- mouse		1:100
hnRNP D Abnova: H00003184-BO1P Mouse	MAB	1:1000 1:100	Importin-β Millipore: 05-1530 - Mouse	MAB	1:2000 1:200
BRF-1 Abnova: H00000677-MO2 Mouse	MAB	1:1000 1:200	Exportin-1 BD: 611832 - Mouse	Mab	1:2000 1:200
TIAR Santacruz Biotech:sc-398373 Mouse	MAB	1:500 1:200	Transportin-1 Millipore: 0501515- Mouse	MAB	1:2000 1:200
TIA-1 Santacruz Biotech:sc-365349 Mouse	MAB	1:500 1:100	Ran Upstate: 07-517	Rabbit	1:2000 1:200
hnRNP F Snataacruz Biotech: sc32309 Mouse & Thermo-pierce:MA5-18024	MAB	1:500 1:100	CPSF6 Santacruz: sc-376228 -Mouse	MAB	1:2000 1:200
hnRNP I (PTB1) Santacruz Biotech: 73391 – Mouse	MAB	1:500 1:100	SG I VP6 (Dr. H. B. Greenberg) SGII VP6 (Dr. H. B. Greenberg)	MAB Mab	1:200 1:200
FXR-1 Millipore: 05-1529 Mouse	MAB	1:2000	NSP5 Lab generated NSP5: (Dr. O. Burrone)	Rabbit Guinea pig	1:2000 1:200
hnRNP A1 Millipore: 05-1521 Mouse	MAB	1:3000 1:200	NSP2 Lab generated NSP2: (Dr. O. Burrone)	Rabbit Guinea pig	1:2000 1:1000
hnRNP C1/C2 Millipore: 05-1520 Mouse	MAB	1:3000 1:200	DLP-VP6 Lab generated	Rabbit	1:1000 1:1000
hnRNP K/J Millipore: 05-1519 Snataacruz Biotech: sc-28380 - mouse	MAB	1:2000 1:200	SC-35 Millipore: 04-1550 - Mouse	MAB	1:1000
hnRNP L Millipore: 05-1518- Mouse ThermPierce: PA5-19599 - Rabbit	MAB	1:3000 1:200	β-tubulin Thermo-Pierce: MA16308 - Mouse	MAB	1:2000
hnRNP M Thermo-Pierce: PA5-30247 & MA1-91607 - Mouse	Rabbit	1:3000 1:200	PCNA Millipore: 1742353 - Mouse	MAB	1:1000
hnRNP Q Millipore: 05-1517 - Mouse	MAB	1:2000	TTP Millipore: ABE-285 & MABE65- Mouse	MAB	1:2000 1:200
KSRP Millipore: 611286 - Mouse	MAB	1:3000 1:200	Staufen1-Thremo-Pierce: sc-390992-Mouse Thermo-Pierce: PA5-28479 -Rabbit	MAB	1:2000 1:200
G3BP1 BD: 611126 - Mouse	Mab	1:2000 1:200	Sec31A BD: 612351- Mouse	MAB	1:2000 1:200
hnRNP U Millipore: 05-1516 – mouse	Mab	1:2000 1:200	VPS35 Thermo-Pierce: PA5-21898- Rabbit Santacruz: sc-374372- Mouse	MAB	1:2000 1:100
β-Actin Thermo-Pierce: MA5-15739- Mouse	MAB	1:2000	RPS8 Thermo-Pierce: PA5-31676	Rabbit	1:1000 1:100
ZBP1 Thermo-Pierce: PA5-20455	Rabbit	1:200	HSP70 Thermo=Pierce: PA5-28003	Rabbit	1:2000 1:200
mCherry Clontech: 632543- Mouse	MAB	1:200	Hsc70 BD: 51-6892GR- Mouse	MAB	1:2000 1:200
Anti-6X-His Clontech:894-1 – Mouse	MAB	1:2000 1:200	anti- Myc Clontech: 51826 –Mouse	Mab	1:2000 1:200
Anti-GFP Chemicon: A33080	Rabbit	1:2000 1:200	Anti-rabbit secondary IgG- Alexa488- Invitr –ogen); -Cy5 –GEHealthcare	goat anti-rabbit	1:200
Anti-guinea pig secondary IgG-Alex633 Invitrogen		1:200	Ati-mouse secondary IgG-Cy3: GE Healthcare	Goat anti-mouse	1:200

Fig. S1 Comparative analysis of the total steady-state levels of cellular proteins in virus-infected and uninfected MA104 cells. (a) hnRNPs, (b) ARE-BPs (c) cytoplasmic proteins and (d) Nuclear transport proteins. 50 μ g of the control uninfected and infected cell extract (8 hpi) was analysed by immunoblotting using specific antibodies. C, uninfected serum-grown control cells; Inf, RRV-infected cells. Fold change in the level of each protein in virus-infected cells compared to that in the control cells is indicated.



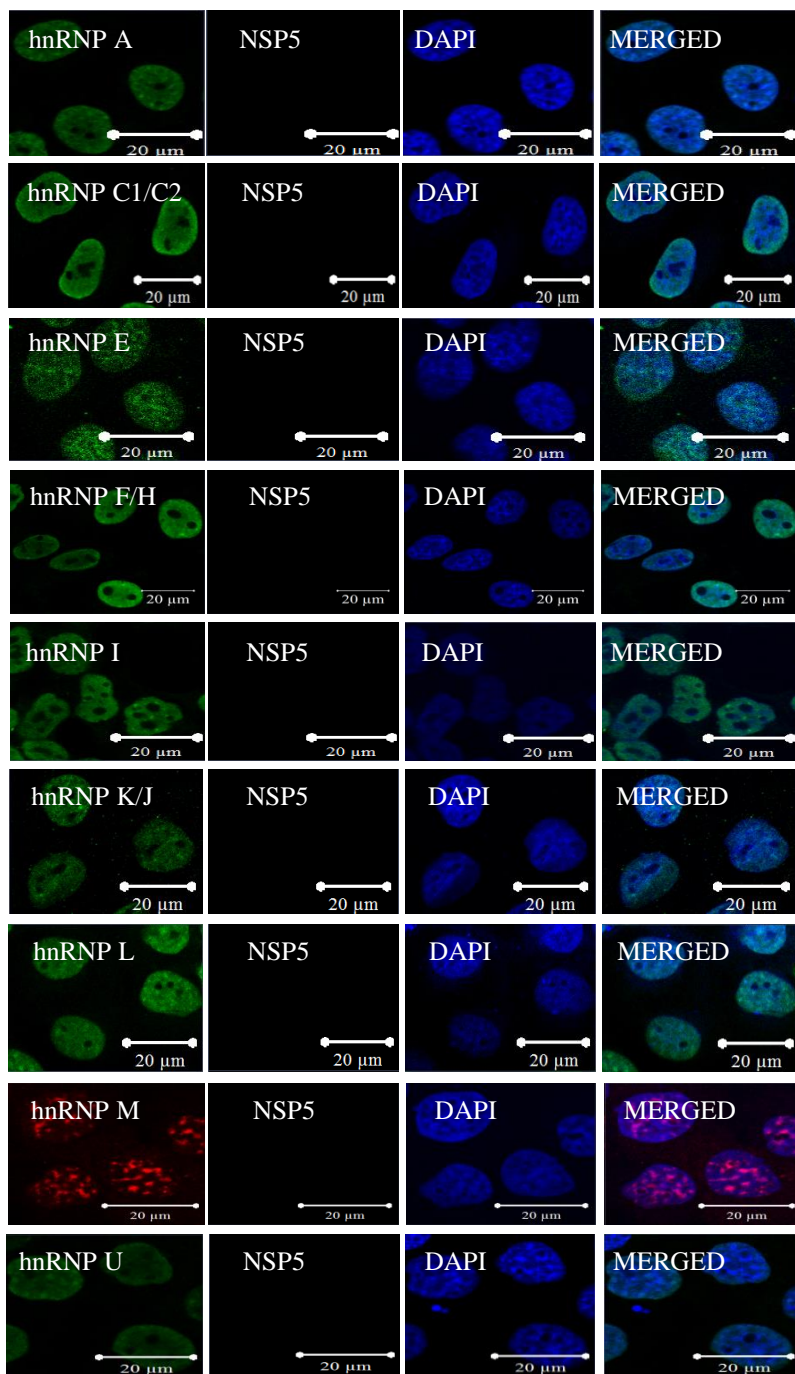
(a)

Fig. S2 Predominant nuclear localization of hnRNPs and ARE-BPs in serum-grown control MA104 cells. MA104 cells grown in the presence of 10% FBS were processed for confocal imaging as described in Methods and Material section. The host proteins were detected using specific primary antibodies (MAbs) and fluorescent Cy3-tagged secondary antibodies. NSP5 and VP6 were detected using rabbit PABs and Cy5-tagged rabbit secondary antibody. Since the control cells do not express viral proteins, NSP5 is undetectable, further indicating that the rabbit polyclonal antibodies are specific to NSP5 and do not cross react with cellular proteins. Note the significant presence of TTP and TIAL-1 in both compartments, correlating with the results in Fig. 2b. Scale bar: 20 μm.

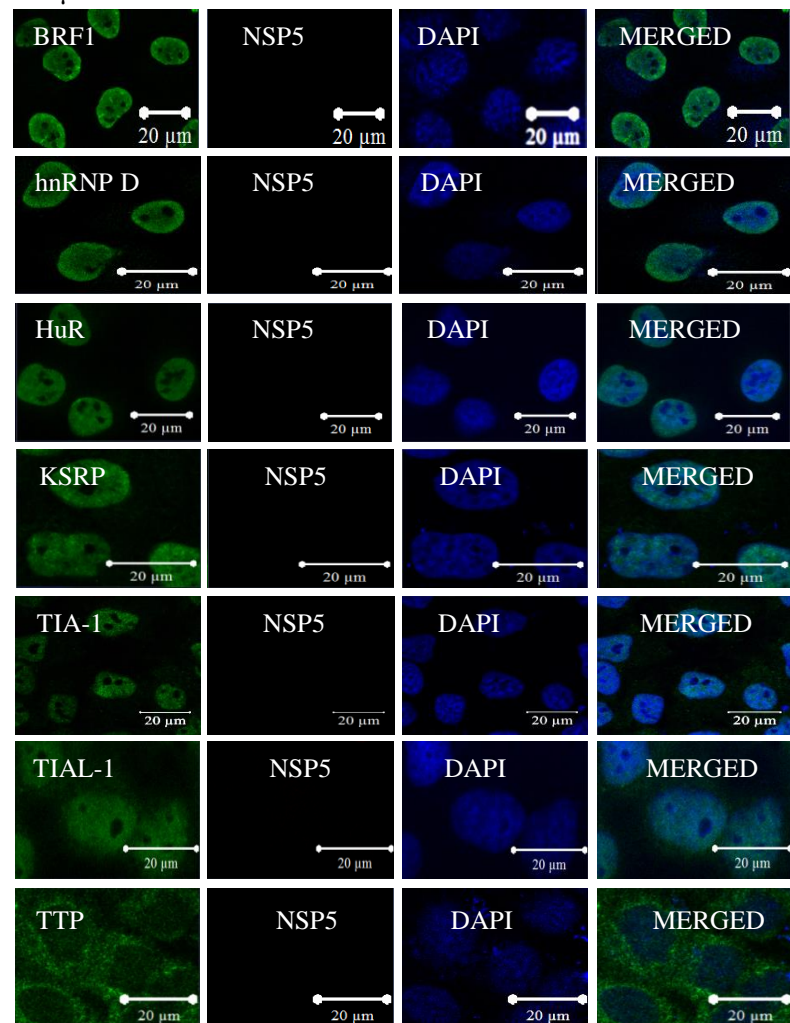
(b)

Fig. S3 Time-dependent detection of viroplasm formation after infection of MA104 cell with RRV. Note the initiation of formation of punctate structures of NSP5 and TIA1, and those of VP6 and RPS8 and their colocalization by 4 hpi, suggesting that the colocalization of the nuclear proteins as well as the abundant cytoplasmic proteins with VS is specific, but is not due to cell lysis or abundance of the host protein. See Fig. 2 legend for experimental details. Scale bar: 10µm.

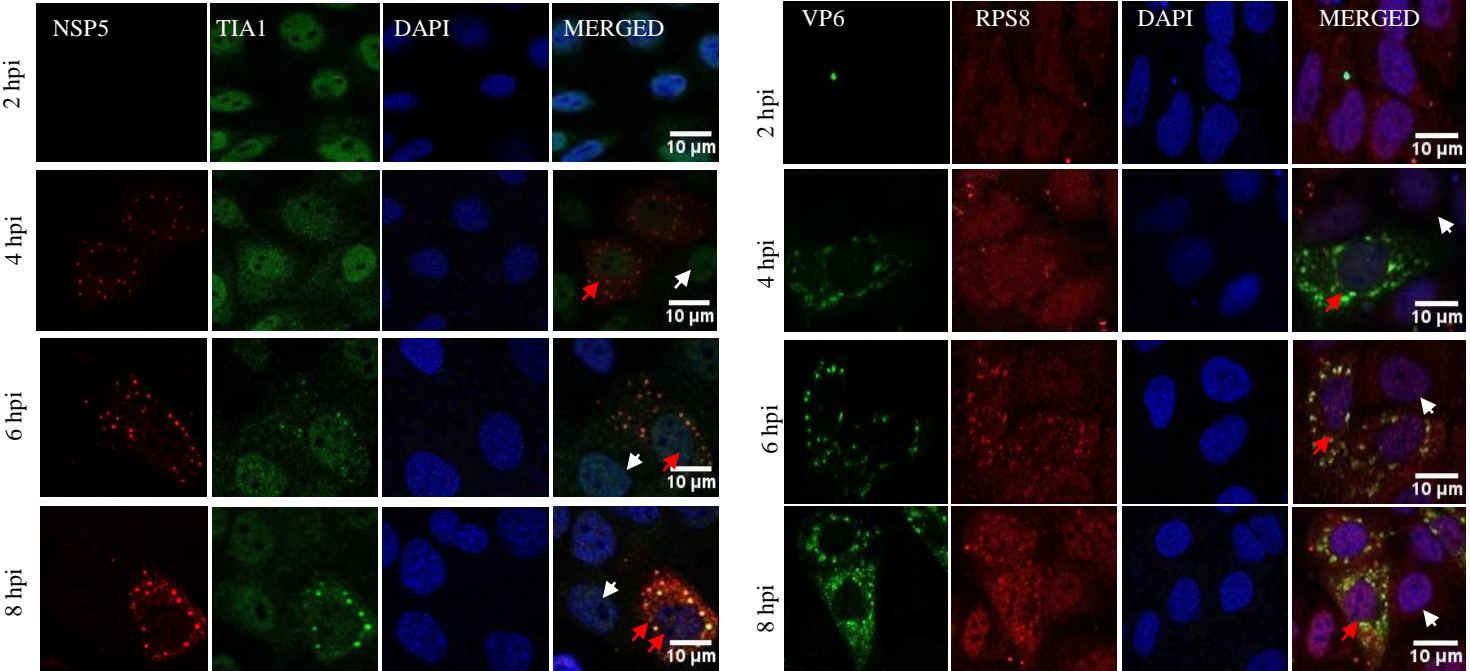


Fig. S4 Z-stack analysis of colocalization of viral and nuclear proteins in the VS. **(a)** The viroplasmic viral protein NSP2 and the host protein hnRNP D, and **(b)** NSP5 and the host protein hnRNP L in the punctate VSs in RRV-infected MA104 cells (at 6 hpi) were visualized by immunofluorescence using rabbit anti-NSP2 and anti-NSP5, and mouse anti-hnRNP D and hnRNP L primary antibodies, and Cy5-tagged (Red) anti-rabbit and Cy3-tagged (Green) anti-mouse secondary antibodies. Z-stacks of images, collected at 0.35 μm intervals ranging from 0 to 2.10 μm at 4x zoom using 63x objective under Zeiss LSM 880 microscope, indicate that both the viral and host proteins are enriched in the same punctate structures in the cytoplasm and that the colocalization is not due to nonspecific fluorescence or due to overlapping diffuse distribution of the two proteins. Z-stack analyses for several host proteins also revealed similar results. Note the perfect localization of the punctate structures of the viral and host proteins at the same site in different sections in the cytoplasm of the infected MA104 cells. Scale bar: 20 μm .

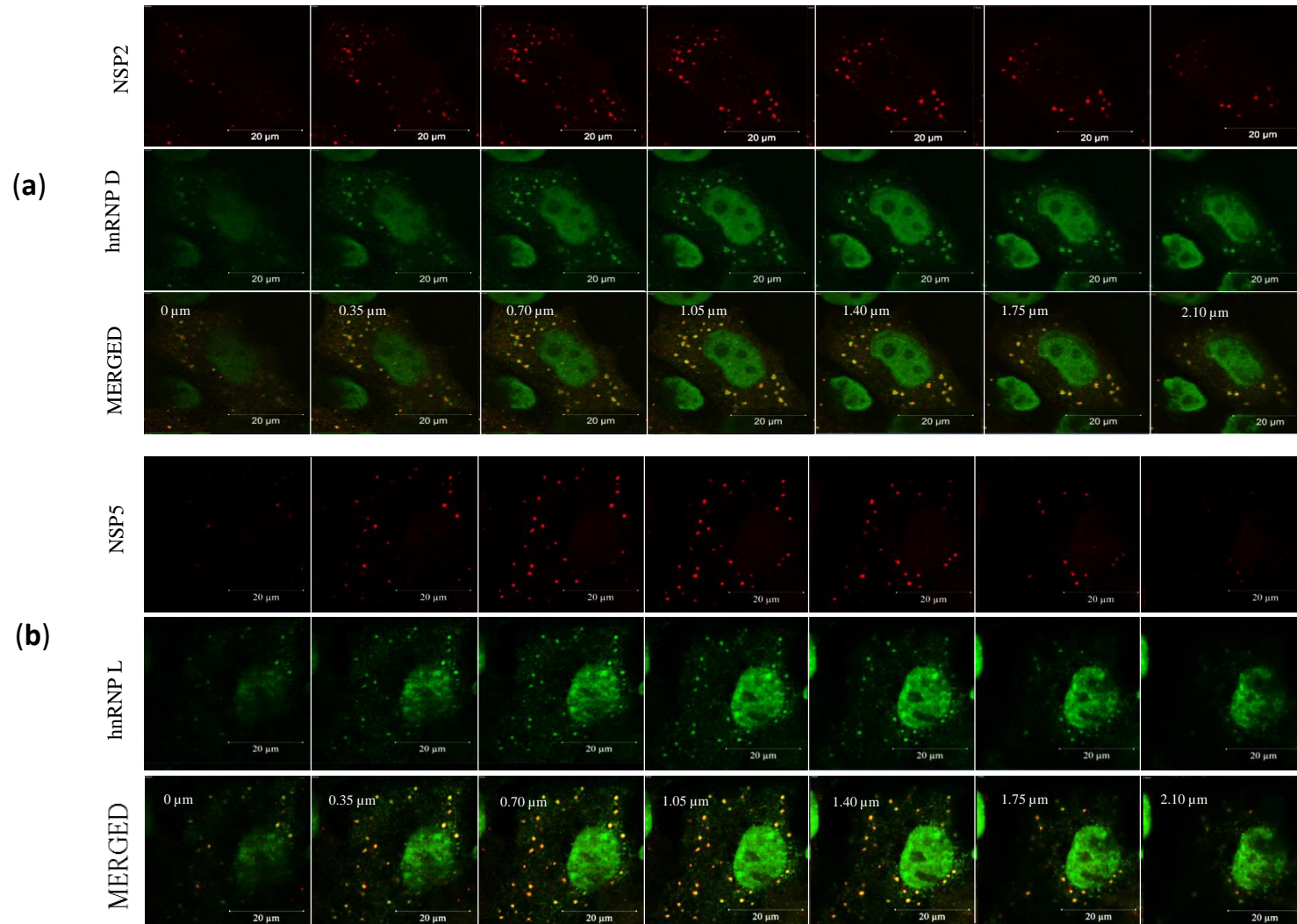


Fig. S5 Colocalization of fluorescent protein-tagged host proteins with viroplasm. HEK293T cells were transfected with plasmid vectors expressing ECFP-NSP5, mCH-VPS35, ECFP-hnRNP1, ECFP-hnRNP D, ECFP-hnRNP K or ECFP. 36 hrs posttransfection, the cells were infected with RRV. At 8 hpi, NSP2 and NSP5 expressed from the virus were detected by specific PABs and fluorescent dye-tagged secondary antibodies. The proteins expressed from the vectors were detected by their fluorescence. Note the colocalization of the ectopically-expressed tagged NSP5 and host proteins with viroplasm. The transfected and infected cells are indicated by orange arrow heads. The sites of quantification of colocalization are indicated by white broken line. Scale bar: 20 μ m.

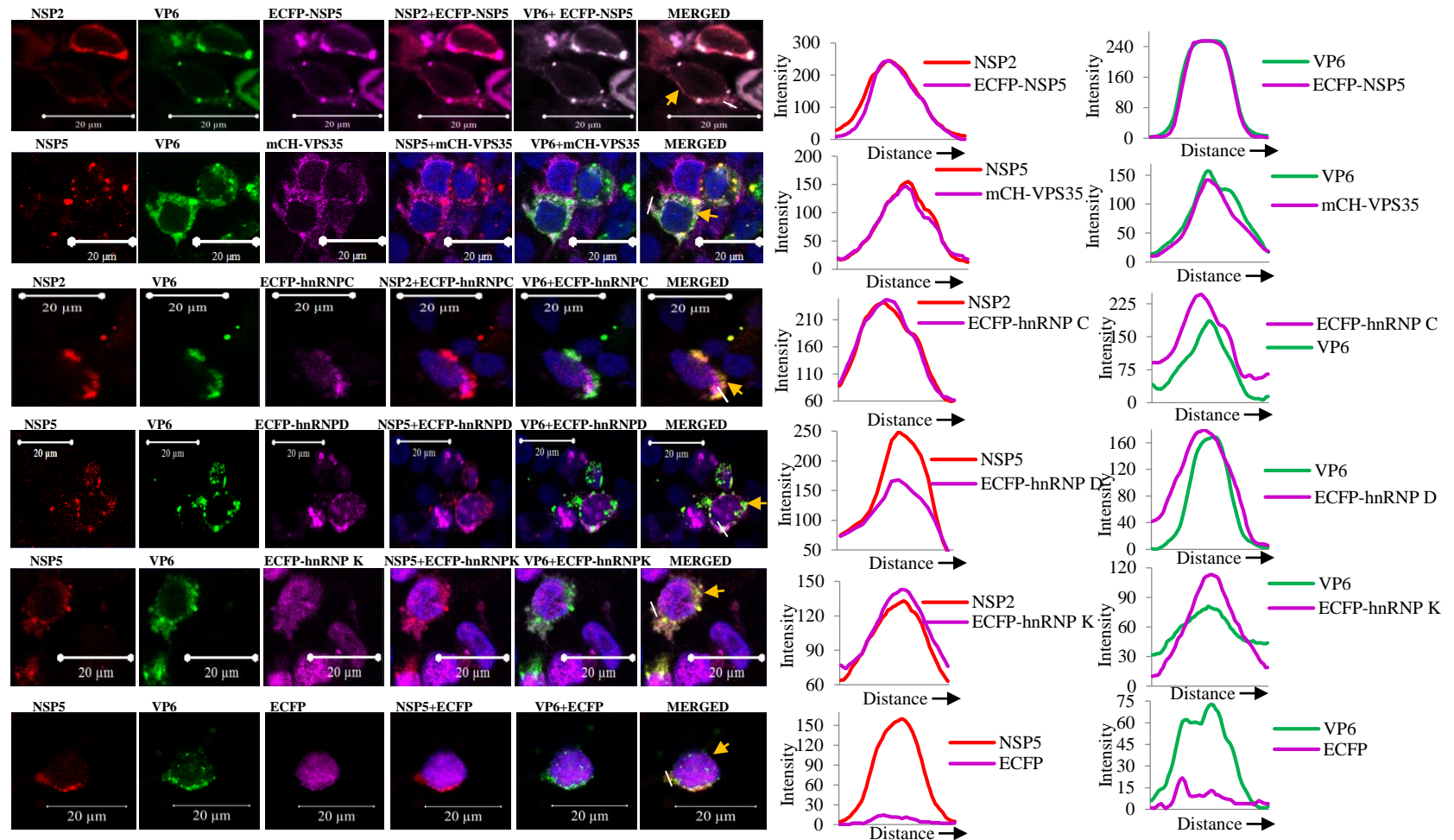


Fig. S6 Analysis of the influence of ectopic expression of (a) IS2-NSP5, (b) SA11-NSP2, and (c) IS2-VP6 in HEK293T cells on the cytoplasmic relocation of hnRNPs and ARE-BPs. Transfected and untransfected cells are indicated by orange and white arrows, respectively. Scale bar: 10 μ m.

