Supplementary Information for

Synchronized mesenchymal cell polarization and differentiation shape the formation of murine trachea and esophagus

by Kishimoto et al.,

Supplementary Table 1 Primary antibody list and conditions for immunofluorescence.

Antibody, Dilution	Company, Catalog code	Fixative	Tissue Preparation	Antigen retrieval	Secondary antibody, Amplification
phospho-Histon H3	Santa Cruz	4% PFA	Paraffin	105°C for 15 min	Rabbit-Alexa488
(1:100)	Biotechnology, Inc,			in Histo ^{VT} One	
	sc-12927				
CDH1	Cell Signaling	4% PFA	Paraffin	105°C for 15 min	Rabbit-Alexa488
(1:800)	Technology,			in Histo ^{VT} One	
	#3195				
β-catenin	Cell Signaling	4% PFA	Paraffin	105°C for 15 min	Mouse-Alexa488
(1:200)	Technology,			in Histo ^{VT} One	
	#8847				
Phospho-MLC2	Cell Signaling	4% PFA	Paraffin	105°C for 15 min	Mouse-Alexa488
(Ser 19)	Technologies, ell			in Histo ^{VT} One	
	Signaling				
	Technologies,				
	#3675				
Phospho-MLC2	Cell Signaling	4% PFA	Paraffin	105°C for 15 min	Rabbit-Alexa488
(Thr19/Ser19)	Technology,			in Histo ^{VT} One	
(1/200)	#3674				
Cy3-SMA	Sigma-Aldrich,	4% PFA	Paraffin	105°C for 15 min	
(1:1,000)	C6198			in Histo ^{VT} One	
SM22α	Abcam,	4% PFA	Paraffin	105°C for 15 min	Rabbit-Alexa594
(1:1,000)	ab14106			in Histo ^{VT} One	
GM130	BD Bioscences,	4% PFA	Paraffin	105°C for 15 min	Mouse-Alexa488
(1:100)	610822			in Histo ^{VT} One	
γ-tubulin	Abcam,	4% PFA	Paraffin	105°C for 15 min	Rabbit-Alexa647
(1/100)	Ab11321			in Histo ^{VT} One	
Sox9	MILLIPORE,	4% PFA	Paraffin	105°C for 15 min	Rabbit-Alexa488
(1/100)	AB5535			in Histo ^{VT} One	
Foxj1	Gift from Dr.	4% PFA	Paraffin	105°C for 15 min	Mouse-Alexa488
(1/100)	Steven L. Brody			in Histo ^{VT} One	
CC10 (T-18)	Santa Cruz	4% PFA	Paraffin	105°C for 15 min	Goat-Alexa594
(1/400)	Biotechnology, Inc,			in Histo ^{VT} One	
	sc-9772				
Krt5	Abcam,	4% PFA	Paraffin	105°C for 15 min	Rabbit-Alexa647
(1/800)	Ab24647			in Histo ^{VT} One	
			-		
GFP	Thermo Fisher	4% PFA	Frozen	90°C for 5 min in	Chicken-Alexa488
(1/200)	Scientific,			Histo ^{v1} One	
(1/200)	A10262				



С

% 4

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Chr. 23. 13. 14. 14. 16. 15. 10. 16. 10. 14.

views are shown below. (**b**) Distribution of pHH3⁺ cells in developing trachea. Sections were stained for pHH3 (*Green*) and TOPRO-3 (*Blue*). (**c**) Proportion of proliferating cells determined by pHH3 expression. D, Dorsal; V, Ventral. Data represent means \pm SEM (n \geq 4). Scale bar; 200 µm (**a**, **b**), 50 µm (**a**; *Magnified view*).



Supplementary Figure 2 Orientation of cell division in epithelium during embryogenesis. (a) Representative images of cell division in SHH^{Cre} ; $R26R^{RG}$ tracheal epithelium at E16.5. Red indicates nuclei genetically labeled by H2B-mCherry. Green indicates plasma membrane labeled by EGFP-GPI. (b) Quantification of cell division orientation based on (a). 0° corresponds to clockwise circumferential direction. 90° corresponds to anterior direction. (n = 30, 37, 35, and 23 in more than three embryos each for E12.5, 14.5, 16.5 and 18.5). Scale bar; 10 µm.



Supplementary Figure 3 Ectopic epithelial cell proliferation in phase 2 inhibited epithelial rearrangement, but neither tube elongation nor expansion.

(a) Gross morphology of developing *Nkx2.1^{CreERt2}, LSL-Kras^{G12D}* trachea. Tube length (b). External diameter (c). Data represent means \pm SEM (n \geq 3). (d) Ki67 expression and phosphorylated ERK1/2 in *Nkx2.1^{CreERt2}, LSL-Kras^{G12D}* trachea. Sections were stained for pERK1/2 (*Green*), Ki67 (*Red*), and DAPI (*Blue*). (e) Rate of proliferating epithelial cells determined by Ki67 expression at E18.5. Data represent means \pm SEM (n=3). (f) The number of GFP⁺ epithelial cells per 100µm². Data represent means \pm SD (Control, n = 123, *Nkx2.1^{CreERt2}; LSL-Kras^{G12D}*, n =167 in 3 fields, respectively). (g) Apical surface area of GFP⁺ epithelial cells at E18.5. Centre line, box limits and whiskers represent mean, 25 and 75% confidence limits, and min and max values, respectively. (Control, n = 52, *Nkx2.1^{CreERt2}; LSL-Kras^{G12D}*, n =50) (h) Proportion of luminal cells in GFP⁺ epithelial cells. Data represent means \pm SD (Control, n = 123, *Nkx2.1^{CreERt2}; LSL-Kras^{G12D}*, n =167 in 3 fields, respectively. N = 123, *Nkx2.1^{CreERt2}; LSL-Kras^{G12D}*, n = 50) (h) Proportion of luminal cells in GFP⁺ epithelial cells. Data represent means \pm SD (Control, n = 123, *Nkx2.1^{CreERt2}; LSL-Kras^{G12D}*, n =167 in 3 fields, respectively. P values (*; P<0.05, **; P<0.01) show the significance with Student's *t*-test, NS, Not Significance



Supplementary Figure 4 Cell proliferation of epithelial cells in developing Wnt5a^{-/-} trachea.

(a) Distribution of BrdU-incorporated cells in developing trachea of $Wnt5a^{-/-}$ mice. Sections were stained for BrdU (*Green*) and TOPRO-3 (*Blue*). Magnified views are shown below. (b) Rate of proliferating epithelial cells determined by BrdU-incorporation assay. Each column indicates the mean with SEM (n=3). (c) Epithelial cell numbers per 100 μ m in $Wnt5a^{-/-}$ trachea at E12.5. (d) Cartilage morphology in $Wnt5a^{-/-}$ trachea and control at E16.5. Whole tracheas were stained for Sox9(*Green*) and SMA (*Red*). D, Dorsal; V, Ventral. Data represent means \pm SEM (n \geq 4). Scale bar; 200 μ m (a, upper panel, d), 20 μ m (a; Magnified view).



Supplementary Figure 5 Indistinguishable appearance of splanchnic mesodermal Wnt5a or Ror2 mutant embryos from control.

(a, i, I) Gross morphology of $Foxg1^{Cre}$, $Wnt5a^{flox/flox}$ (a), $Foxg1^{Cre}$, $Ror2^{flox/flox}$ (i), and $Dermo1^{Cre}$, $Wnt5a^{flox/flox}$ (I), embryos at E12.5. (b and j) Body length of $Foxg1^{Cre}$, $Wnt5a^{flox/flox}$ (b) and $Foxg1^{Cre}$, $Wnt5a^{flox/flox}$ (j) embryos at E12.5 (Control; n=5, $Foxg1^{Cre}$; $Wnt5a^{flox/flox}$ n=6; Control; n=4, $Foxg1^{Cre}$, $Ror2^{flox/flox}$; n=3). (c and e) Gross morphology of $Foxg1^{Cre/+}$, and $Foxg1^{Cre/Cre}$ embryos at E12.5 (c) and 18.5 (e). (d and f) Gross morphology of $Foxg1^{Cre/+}$, and $Foxg1^{Cre/Cre}$ tracheas at E12.5 (d) and 18.5 (f). (g and k) Gross morphology of SHH^{Cre} ; $Ror2^{flox/flox}$ trachea, and $Dermo1^{Cre}$, $Wnt5a^{flox/flox}$ (k) at E12.5. (h) Tracheal tube length (Control; n = 3, SHH^{Cre} , $Ror2^{flox/flox}$; n = 5). NS, Not Significance Scale bar, 1cm (e),1mm (c, d, I), 500 \mum (f), 200 \mum (k)



Supplementary Figure 6 Cell proliferation of SM cells in Wnt5a^{-/-} trachea during elongation process.

NS

E12.5

(a) Distribution of proliferating Ki67⁺ SM cells in Wnt5a^{-/-} trachea and control littermate at E11.5 and 12.5. Sections were stained for Ki67 (Red), SM22a (White) and DAPI (Blue). Magnified views are shown below. (b) Proliferating rate of SM cells, which is determined by Ki67 expression. Data represent means ± SEM (n=3). NS; Not Significance. Scale bar; 50 μm





b

GM130 y-tubulin SMA DAPI





Ror2 GM130 DAPI

Supplementary Figure 7 Subepithelial cell polarity of smooth muscle cells are determined by the Golgiapparatus and centrosome positions relative to the nucleus.

(**a**) Smooth muscle cells in subepithelial layer show synchronized cell polarity toward epithelium. Sagittal sections of E12.5 trachea were stained for GM130 (Golgi-apparatus; *Green*), γ-tubulin (Centrosome; *White*), SMA (Smooth muscle; *Red*), and DAPI (nucleus; *Blue*). Scale Bar; 10 µm (Low magnification), 2 µm (High magnification). (**b**) Radial cell polarity toward epithelium. Transversal sections were stained for Ror2 (*Green*), GM130 (*White*), and DAPI (*Blue*). Red arrows indicate the direction of polarity. Scale bar; 20 µm (Low magnification), 10 µm (High magnification).



Supplementary Figure 8 Directional migration of SM cells to the epithelium *in vitro.* (a) 3D co-culture system of primary SM cells and epithelial cells. (b) Quantification of approaching distance and directionality. See Materials and Methods for details.



Supplementary Fig. 9 Normal tracheal tube elongation and subepithelial cell polarity in *Foxg1^{Cre}*, *Srf^{flox/flox}* mice.

(a) SM morphology in *Foxg1^{Cre}*, *Srf^{flox/flox}* trachea and control at E12.5. Transversal sections were stained for SMA (*Red*), DAPI (*Blue*). Coronal (*Upper*) and transversal (*Lower*) sections. (b) QUantification of subepithelial cell polarity, determined by the Golgi-apparatus position relative to the nucleus. (Control; n=26, *Foxg1^{Cre}*, *Srf^{flox/flox}*; n=29) (c) Gross morphology of *Foxg1^{Cre}*, *Srf^{flox/flox}* tracheas and littermate control at E12.5. (d) Cell morphology at dorsal subepithelial tissue in *Foxg1^{Cre}*, *Srf^{flox/flox}* and *Wnt5a^{-/-}* trachea. Sections were stained for F-actin (*Green*), DAPI (*Blue*).
(e) Quantification of bundle orientation toward lumen. (Control; n=47, *Foxg1^{Cre}*, *Srf^{flox/flox}*; n=44, Wnt5a^{-/-}; n=50) Epi.; Epithelium, Scale bar, 200 µm (c), 10 µm (a, b, d)



Supplementary Figure 10 The distribution of Ror2 protein in diverse organs derived from foregut endoderm.

(**a-c**) Expression of Ror2 in the esophagus (**a**, **b**), trachea (**a**), stomach (**b**), and intestine (**c**) at E12.5. Sections were stained for Ror2 (*Green*), SMA (*Red*). Nuclei were counterstained by DAPI (*Blue*). Scale bar; 100 μ m (**b**), 50 μ m (**a**), 40 μ m (**c**).



Supplementary Figure 11 Cartilage and SM architecture of the tracheas in *Foxg1^{Cre}, Sox9^{flox/flox}* **mice.** (a) SM morphology in *Foxg1^{Cre}, Sox9^{flox/flox}* trachea and control at E16.5. Whole tracheas were stained for SMA (*Red*). Coronal sections were shown. (b) Cartilage and SM histology of *Foxg1^{Cre}, Sox9^{flox/flox}* tracheas. Sections were stained for Sox9 (*Green*), SMA (*Red*) and DAPI (*Blue*). Magnified views are shown below. Scale bar, 50 μm



Supplementary Figure 12 Cell proliferation of epithelial cells in *Foxg1^{Cre};* Sox9^{flox/flox} tracheas.

(a) Distribution of BrdU-incorporated cells in $Foxg1^{Cre}$; $Sox9^{flox/flox}$ tracheas. Sections were stained for BrdU (*Green*) and TOPRO-3 (*Blue*). (b) Proportion of proliferating cells determined by cumulative BrdU incorporation assay in $Foxg1^{Cre}$; $Sox9^{flox/flox}$ tracheas. (n = 4 or 5). Magnified views are shown below. D, Dorsal; V, Ventral. Data represent means \pm SEM (n \geq 4). Scale bar; 200 µm, 20 µm (*Magnified view*).



Supplementary Figure 13 Psuedostratification index in Foxg1^{Cre}; Sox9^{flox/flox} tracheas.

(a) Schematic model for calculating the pseudostratification index, the distance between the nuclear centroid to the basement membrane. (b) Optical image of the tracheal epithelium in SHH^{Cre} ; $R26R^{RG/+}$ mice. (c) Pseudostratification index in SHH^{Cre} ; $R26R^{RG/+}$. Centre line, box limits and whiskers represent mean, 25 and 75% confidence limits, and min and max values, respectively. Numbers represent means (n = 264, 207, 152 cells in three embryos for E14.5, E16.5, and E18.5). (d) Optical image of the tracheal epithelium in $Foxg1^{Cre}$; $Sox9^{flox/flox}$. Scale bar: 20 µm.



Supplementary Figure 14 Phase 2 trachea tubulogenesis in Col2a1-/- mice.

(a) Generation of Col2a1 knockout mice with CRISPR/CAS genome editing technology. Guide RNAs were designed at 1st and 3rd intron in order to delete exon2 and exon3. (b) Genomic PCR for Col2a1 genotyping. (c) Gross appearance of Col2a1+/- and Col2a1-/- mice. (d - g) Gross morphology of Col2a1+/- and Col2a1-/- tracheas (d). Tracheal tube length (e), external diameter (f) (Control; n = 8, Col2a1^{-/-}; n = 9), and internal circumference (g) (Control; n = 3, Col2a1^{-/-}; n = 3). (h) Cartilage and SM architecture of Col2a1^{-/-} trachea at E18.5. Sections were stained for Sox9 (Green), SMA (Red) and DAPI (Blue). (i) Apical surface area of epithelial cells in Nkx2.1^{mTomato}, Col2a1^{-/} trachea at E18.5. Centre line, box limits and whiskers represent mean, 25 and 75% confidence limits, and min and max values, respectively. (Control; n = 140, Col2a1-/-; n = 60 from 3 embryos, respectively) (j) Psuedostratification index in Col2a1^{-/-}. Centre line, box limits and whiskers represent mean, 25 and 75% confidence limits, and min and max values, respectively. (Control; n = 58, Col2a1^{-/-}; n = 118 from 2 embryos, respectively). (k and I) Cell proliferation of epithelial cells in Col2a1-/- trachea at E18.5. Sections were stained for Ki67 (Magenda) and DAPI (Blue) (k). Proliferating rate of epithelial cells in Col2a1-/- trachea at E18.5 (I) (Control; n = 878, Col2a1^{-/-}; n = 672 from 3 embryos, respectively). (m and n) Cell differentiation of epithelial cells in Col2a1-/ trachea at E18.5. Sections were stained for Foxj1 (Green), SCGB1A1 (Red), Krt5 (Grey), and DAPI (Blue) (m). Cell proportion of epithelial cells in $Col2a1^{-/-}$ trachea at E18.5 (n) (Control; n = 826, $Col2a1^{-/-}$; n = 632 from 3 embryos, respectively). P values (*; P<0.05, **; P<0.01) show the significance with Student's t-test, NS, Not Significance, Scale bar, 2000 μm (c), 500 μm (d), 50 μm (h and k), 20 μm (m)



Supplementary Figure 15 Expression and distribution of adhesion-, polarity-, and cytoskeleton-related proteins.

(a) Expression and distribution of adhesion- and polarity-related proteins. Sections were stained for CDH1, CTNNB1, ZO-1, or SCRIB. (b) Expression and distribution of cytoskeletal components. Sections were stained for F-actin, pMLC^{Ser19}, ppMLC^{Thr18/Ser19}, or α -tubulin. Nuclei were counterstained by DAPI or TOPRO-3. Scale bar; 10 µm.



Supplementary Figure 16 Phase 2 tracheal tubulogenesis in Wnt5a/Ror2 mutant mice.

(**a**, **b**) External diameter of *Wnt5a^{-/-}* (**a**), *Foxg1^{Cre}*, *Ror2^{flox/flox}*, *Foxg1^{Cre}*, *Ror1^{flox/flox}*, *Ror2^{flox/flox}*, trachea (**b**) at E18.5. Data represent means ± SEM (Control; n = 3, *Wnt5a^{-/-}*; n = 3, *Foxg1^{Cre}*, *Ror2^{flox/flox}*; n = 3, *Foxg1^{Cre}*, *Ror1^{flox/flox}*; n = 4). (**c**) Gross morphology of *Foxg1^{Cre}*, *Ror1^{flox/flox+}*, *Ror2^{flox/flox}* and *Foxg1^{Cre}*, *Ror1^{flox/flox}*; n = 15. (**d**) Apical surface area of epithelial cells in *Wnt5a^{-/-}* trachea at E16.5 and 18.5. Centrol, n=262, *Wnt5a^{-/-}*, n=152). (**e**) Pseudostratification index in *Wnt5a^{-/-}* tracheas at E16.5 and 18.5. Centre line, box limits and whiskers represent mean, 25 and 75% confidence limits, and min and max values, respectively. Numbers represent means (E16.5; Control, n=125, *Wnt5a^{-/-}*, n=96, E18.5; Control, n=109, *Wnt5a^{-/-}*, n=130). P values (*; P<0.