#### Sympathetic inputs regulate adaptive thermogenesis in brown adipose tissue through cAMP-Salt inducible kinase axis

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#### Supplementary information:

#### Supplementary Figure 1: Gnas was specifically deleted in iBAT in Gnas<sup>BKO</sup>

mice. Immunoblots showing amounts of Gnas and Hsp90 in different tissues

from ~8-week-old male CON and Gnas<sup>BKO</sup> mice.

#### Supplementary Figure 2: CLAMS measurements of CON and Gnas<sup>BKO</sup> mice.

Average oxygen consumption calculated as per body weight (**A**) and per mouse (**B**), respiratory exchange ratio (RER) (**C**), food intake (**D**) and physical activity (**E**) during day and night in ~8-week old male CON and Gnas<sup>BKO</sup> mice. Sample size: CON (n=4) and Gnas<sup>BKO</sup> (n=3).

#### Supplementary Figure 3: Mass spectrometry analysis of iBAT

**mitochondrial proteome. A**, Frequency of mitochondrial and non-mitochondrial proteins identified by mass spectrometry from isolated mitochondria from iBAT of ~8-10-week old male CON and Gnas<sup>BKO</sup> mice housed at RT. Sample size: n=3 for each genotype. **B**, Immunoblots showing amounts of Hk1 and Hsp60 in iBAT of ~8-10-week old male CON and Gnas<sup>BKO</sup> mice housed at RT.

Supplementary Figure 4: Cold-induced beige adipocytes formation in iWAT was not affected in Gnas<sup>BKO</sup> mice. A, g-PCR analysis of *Ucp1* mRNA levels in iWAT from wild-type B6 mice at different ages and conditions. RT: room temperature; Cold: 8°C for 7 days; PBS or 6-OHDA was injected prior to 3-week of age or cold at 8-week of age. Sample sizes: 3w PBS/RT (n=9), 3w 6-OHDA/RT (n=4), 8w PBS/RT (n=3), 8w PBS/Cold (n=12) and 8w 6-OHDA/Cold (n=4). **B**, Immunoblots showing protein amounts of Ucp1, phosphor-PKA substrates and Hsp90 in iWAT from 3-week-old wild-type B6 pups with or without 6-OHDA injection. **C**, Immunoblots showing protein amounts of Ucp1, phosphor-PKA substrates and Hsp90 in iWAT from 8-week-old wild-type B6 mice under room temperature or after 7-day 8°C cold stimulation with or without 6-OHDA injection. **D**, g-PCR analysis of *Ucp1* mRNA levels in iWAT from ~8-week-old CON and βless mice at room temperature and after 7-day 8°C cold. Sample sizes: CON/RT (n=4), βless/RT (n=6), CON/Cold (n=6) and βless/Cold (n=4). E, q-PCR analysis of *Ucp1* mRNA levels in iWAT from Gnas<sup>f/f</sup> (CON), Adiponectin-Cre;Gnas<sup>f/f</sup> (Gnas<sup>AKO</sup>) and Ucp1-Cre;Gnas<sup>f/f</sup> (Gnas<sup>BKO</sup>) mice at 3 and 8-week of age, and with 7-day 8°C cold stimulations. Sample sizes: 3w CON (n=15), 3w Gnas<sup>BKO</sup> (n=4), 3w Gnas<sup>AKO</sup> (n=5), 8w RT/CON (n=6), 8w RT/Gnas<sup>BKO</sup> (n=4), 8w RT/Gnas<sup>AKO</sup> (n=3), 8w Cold/CON (n=6), 8w Cold/Gnas<sup>BKO</sup> (n=4) and 8w/Cold Gnas<sup>AKO</sup> (n=3). **F**, Representative H&E staining of iWAT from 6-8-week-old CON, Gnas<sup>AKO</sup> and Gnas<sup>BKO</sup> mice at RT or after 7-day 8°C cold stimulation. Scale bar: 50µm. Arrows showing multilocular beige adipocytes. **G**, Diagram showing

effects of sympathetic innervation and adipocyte cAMP signaling on beige adipocyte genesis and renaissance.

Supplementary Figure 5: Gnas<sup>BKO</sup> mice exhibited a fat redistribution phenotype at normal chow. **A**, Body mass, lean and fat mass (measured by DEXA) of ~10-week-old male CON and Gnas<sup>BKO</sup> mice housed at RT and 30°C. Sample size: CON/RT (n=4), Gnas<sup>BKO</sup>/RT (n=5), CON/30°C (n=4) and Gnas<sup>BKO</sup>/30°C (n=5). **B**, Tissue mass of iWAT, eWAT, BAT and liver from ~10week-old male CON and Gnas<sup>BKO</sup> mice housed at RT and 30°C. Sample size: CON/RT (n=11), Gnas<sup>BKO</sup>/RT (n=7), CON/30°C (n=7) and Gnas<sup>BKO</sup>/30°C (n=8).

**Supplementary Figure 6: Abundance of Pdgfra<sup>+</sup>Sca1<sup>+</sup> progenitors in the eWAT of Gnas<sup>BKO</sup> mice after HFD.** Numbers of Pdgfra<sup>+</sup>Sca1<sup>+</sup> progenitors (determined by flow cytometry) in the eWAT of Gnas<sup>BKO</sup> mice after HFD. Sample size: CON (n=5) and Gnas<sup>BKO</sup> (n=8).

Supplementary Figure 7: Sik2 was a SIK isoform enriched in adipose tissue. Immunoblots of Sik2 and Hsp90 in different tissues from ~8-week-old male wild-type (WT) and Sik2 KO mice.

Supplementary Figure 8: Effects of SIK deficiency on Ucp1 expression. A, q-PCR analysis of *Ucp1*, *Dio2* and *Pgc1* $\alpha$  mRNA levels in differentiated brown adipocytes treated with control or Sik1 and Sik2 knockdown adenovirus. Effects of 4-hour FSK stimulation shown. **B**, q-PCR analysis of *Ucp1* and *Sik2* mRNA levels in iBAT from ~8-week-old male WT and Sik2 KO mice. Sample size: WT (n=3) and Sik2 KO (n=4). **C**, Immunoblots showing Ucp1, Sik2 and Hsp90 in iBAT from ~8-week-old male WT and Sik2 KO mice. **D**, q-PCR analysis of *Ucp1* and *Sik1* mRNA levels in iBAT from ~8-week-old male WT and Sik2 KO mice. Sample size: n=3 per each genotype. **E**, Immunoblots showing Ucp1 and Hsp90 in iBAT from ~8-week-old male WT and Sik1 KO mice.

**Supplementary Figure 9: Expression of SIK isoforms in iBAT at RT and thermoneutrality.** q-PCR analysis of *Sik1, Sik2* and *Sik3* in iBAT from ~10week-old male C57bl/6J mice housed at RT and thermoneutrality.

Supplementary Figure 10: CLAMS measurements, lipolysis activity and mitochondrial gene expression in iBAT of WT and Sik2 KO mice at thermoneutrality. Average oxygen consumption calculated as per body weight (A) and per mouse (B), respiratory exchange ratio (RER) (C), food intake (D) and physical activity (E) during day and night in ~12-week old male CON and Sik2 KO mice. Sample size: n=4 per genotype. F, Area under the curve (AUC) of oxygen consumption recordings in response to norepinephrine in ~10-week-old male WT and Sik2 KO mice housed 30°C. Sample size: n=3 for each genotypes. G, *In vitro* glycerol release rates (treated with 10uM FSK) of iBAT from ~10-week-old male WT and Sik2 KO mice housed 30°C. Sample size: n=4 for each genotypes. q-PCR analysis of relative mRNA levels of nuclear (H) and

mitochondrial (I) encoded ETC gene expression in ~12-week old male CON and Sik2 KO mice. Sample size: n=5 per genotypes.

Supplementary Figure 11: Brown adipocyte-specific deletion of Hdac4 did not show thermogenic defects *in vivo*. **A**, q-PCR analysis of *Ucp1* and *Glut4* in differentiated brown adipocytes. Effects of FSK and LMK235 alone or combined shown. **B**, q-PCR analysis of *Hdac4, Ucp1* and other thermogenic genes in iBAT from ~8-week-old male CON and Hdac4<sup>BKO</sup> mice. Sample size: CON (n=4) and Hdac4<sup>BKO</sup> (n=3). **C**, Immunoblots showing amounts of Ucp1, Hdac4 and Hsp90 in iBAT of from ~8-week-old male CON and Hdac4<sup>BKO</sup> mice. Oxygen consumption recordings in response to NE in ~8-week-old male CON and Hdac4<sup>BKO</sup> mice. Average oxygen consumption calculated as per body weight (**E**) and per mouse (**F**), respiratory exchange ratio (RER) (**G**), food intake (**H**) and physical activity (**I**) during day and night in ~8-week-old male CON and Hdac4<sup>BKO</sup>

CON













30 -2 25 CON/RT 1.5 Mass (g) 20 Mass (g) ■ Gnas<sup>вко</sup>/RT 15 □ CON/30°C \*\*\* 10 \*\*\* □ Gnas<sup>BKO</sup>/30°C 0.5 5 0

Liver

iBAT

**iWAT** 

eWAT

■ CON/RT ■ Gnas<sup>BK0</sup>/RT ■ CON/30<sup>°</sup>C ■ Gnas<sup>BK0</sup>/30<sup>°</sup>C

## **Supplementary Figure 5**

Fat

Lean

Body





iBAT ewait to the the the set and the the set and the the set and Sik2 Hsp90

WT













#### Supplementary table 1: List of primer sequences for q-PCR

36B4-F	TTTGGGCATCACCACGAAAA
36B4-R	GGACACCCTCCAGAAAGCGA
Gnas-F	GCAGAAGGACAAGCAGGTCT
Gnas-R	CCCTCTCCGTTAAACCCATT
Ucp1-F	ACTGCCACACCTCCAGTCATT
Ucp1-R	CTTTGCCTCACTCAGGATTGG
Cox8b-F	GAACCATGAAGCCAACGACT
Cox8b-R	GCGAAGTTCACAGTGGTTCC
Cidea-F	TGCTCTTCTGTATCGCCCAGT
Cidea-R	GCCGTGTTAAGGAATCTGCTG
Pgc1α-F	AGCCGTGACCACTGACAACGAG
Pgc1α-R	GCTGCATGGTTCTGAGTGCTAAG
mt-Nd4-F	CTAATAATCGCACATGGCCTC
mt-Nd4-R	CGTAGTTGGAGTTTGCTAGG
mt-Nd5-F	CATCCTTCTCAACTTTACTGGG
mt-Nd5-R	TTTATGGGTGTAATGCGGT
mt-Cyb-F	CCATTCTACGCTCAATCCCCA
mt-Cyb-R	AGGCTTCGTTGCTTTGAGGTA
mt-Co1-F	ACACAACTTTCTTTGATCCCG
mt-Co1-R	AGAATCAGAACAGATGCTGG
mt-Co2-F	ATAATCCCAACAAACGACCT
mt-Co2-R	CTCGGTTATCAACTTCTAGCA
mt-Co3-F	GGTATAATTCTATTCATCGTCTCGG
mt-Co3-R	AGAACGCTCAGAAGAATCCT
Ndufs1-F	AGGATATGTTCGCACAACTGG
Ndufs1-R	TCATGGTAACAGAATCGAGGGA
Ndufs4-F	CTGCCGTTTCCGTCTGTAGAG
Ndufs4-R	TGTTATTGCGAGCAGGAACAAA
Ndufs8-F	AGTGGCGGCAACGTACAAG
Ndufs8-R	TCGAAAGAGGTAACTTAGGGTCA
Sdha-F	GGAACACTCCAAAAACAGACCT
Sdha-R	CCACCACTGGGTATTGAGTAGAA
Sdhb-F	AATTTGCCATTTACCGATGGGA
Sdhb-R	AGCATCCAACACCATAGGTCC
Sdhc-F	GCTGCGTTCTTGCTGAGACA
Sdhc-R	ATCTCCTCCTTAGCTGTGGTT
Sdhd-F	TGGTCAGACCCGCTTATGTG
Sdhd-R	GGTCCAGTGGAGAGATGCAG
Cyc1-F	CAGCTTCCATTGCGGACAC
Cyc1-R	GGCACTCACGGCAGAATGAA
Cox4-F	ATGTCACGATGCTGTCTGCC
Cox4-R	GTGCCCCTGTTCATCTCGGC
Cox4i1-F	ATTGGCAAGAGAGCCATTTCTAC
Cox4i1-R	CACGCCGATCAGCGTAAGT

Atp5a1-F	TCTCCATGCCTCTAACACTCG
Atp5a1-R	CCAGGTCAACAGACGTGTCAG
Atp5j2-F	TGCCGAGCTGGATAATGATGC
Atp5j2-R	ACCATGCTAATCCCCGAGATG
Atp5b-F	GCAAGGCAGGGACAGCAGA
Atp5b-R	CCCAAGGTCTCAGGACCAACA
mt-Atp8-F	GGCACCTTCACCAAAATCACT
mt-Atp8-R	GGGGTAATGAATGAGGCAAATAGA
mt-Atp6-F	CCTTCAATCCTATTCCCATCC
mt-Atp6-R	GTTGGAAAGAATGGAGACGG
Hdac4-F	CACACCTCTTGGAGGGTACAA
Hdac4-R	AGCCCATCAGCTGTTTTGTC
mtDNAF	CGACCTCGATGTTGGATCA
mtDNAR	AGAGGATTTGAACCTCTGG
B2MF	TCTCTGCTCCCCACCTCTAAGT
B2MR	TGCTGTCTCGATGTTTGATGTATCT