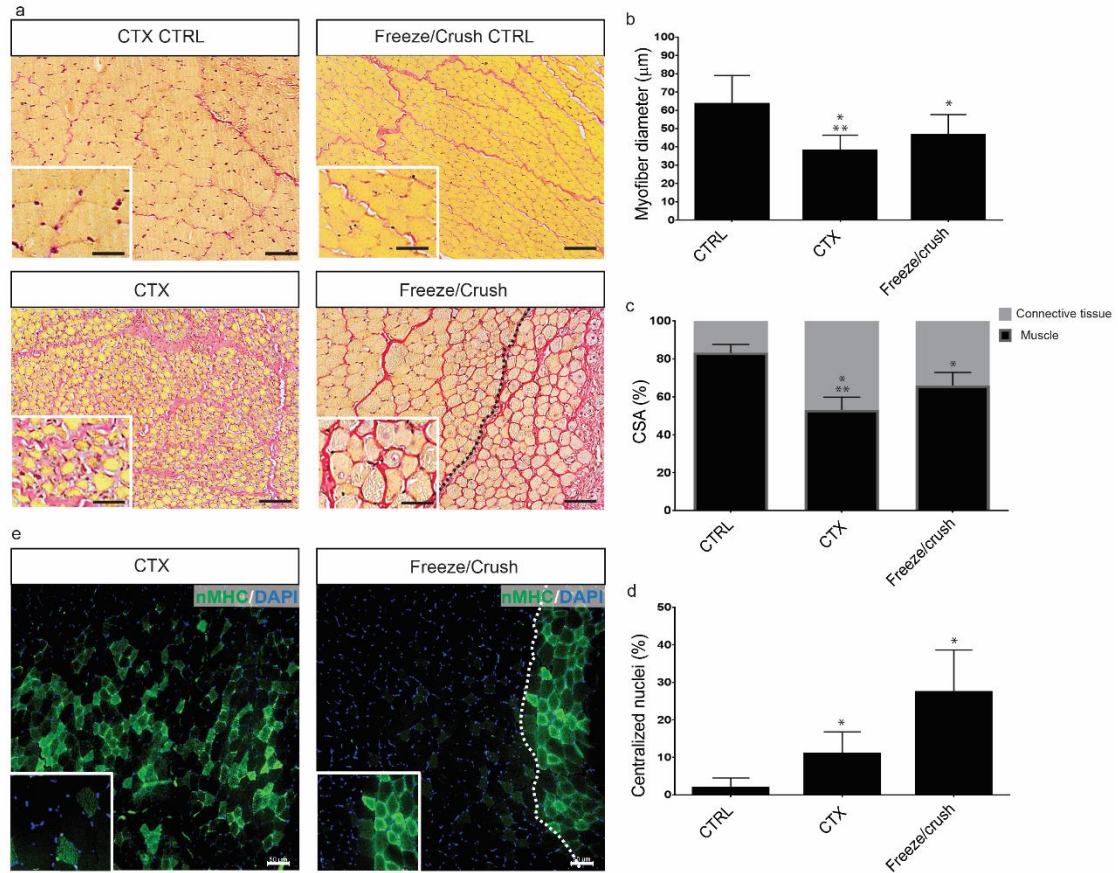


# **Transplantation of Allogeneic PW1<sup>pos</sup>/Pax7<sup>neg</sup> Interstitial Cells (PICs) Enhance Endogenous Repair of Injured Porcine Skeletal Muscle**

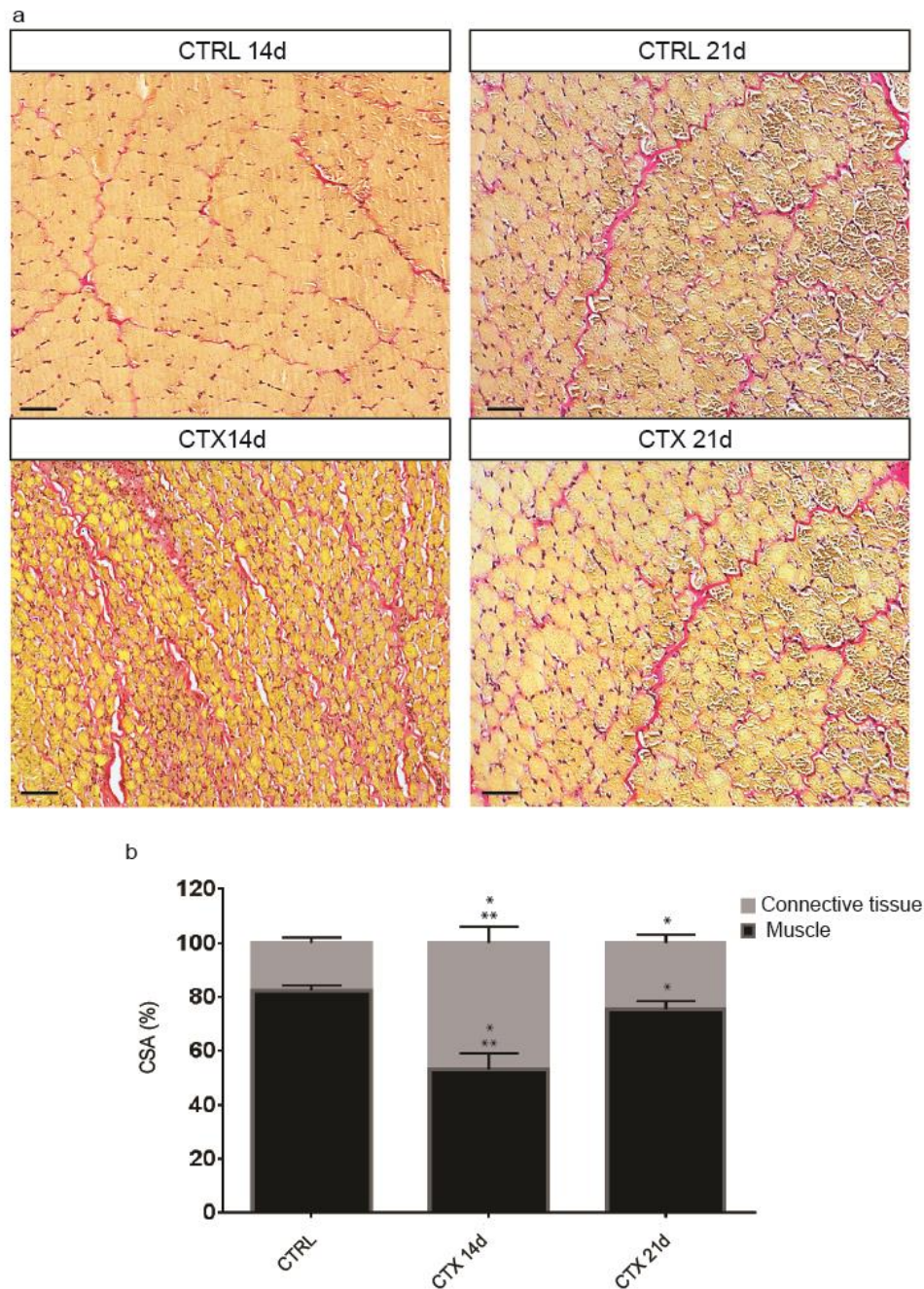
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Cheyenne C. S. Tseng<sup>3</sup>, Steven A. J. Chamuleau<sup>3</sup>, Bernardo Nadal-Ginard<sup>1†</sup>, Georgina M.  
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Figure S1



**Fig. S1. Comparison of two porcine models of skeletal muscle injury.** (a) HVG staining of CTRL, CTX and freeze/crush-injured paraffin-embedded muscle, showing connective tissue (red) and skeletal muscle (yellow), scale bar 100μm. Inset shows 40x magnification of centralized nuclei, scale bar 20μm. Dotted line demarcates the restricted injury area observed in freeze/crush-injured muscle only. (b) Myofiber diameter per 100 myofibers in CTRL, CTX and freeze/crush-injured muscle, n=2 animals per group. Data are mean ± SD, \* $P < 0.0001$  vs. CTRL, \*\*  $P < 0.0001$  vs. freeze/crush-injured muscle. (c) Quantification of the ratio of skeletal muscle to connective tissue CSA of CTRL, CTX and freeze/crush-injured muscle, determined from five fields of view per muscle, n=2 animals per group. Data are mean ± SD, \* $P < 0.0001$  vs. CTRL, \*\*  $P = 0.018$  vs. freeze/crush-injured muscle. (d) Number of centralized nuclei in CTRL, CTX and freeze/crush-injured muscle determined from five fields of view per muscle, n=2 animals per group. Data are mean ± SD, \* $P < 0.0001$  vs. CTRL (e) Immunohistochemical staining indicates a diffuse pattern of nMHC expression in CTX-injured muscle compared to restricted expression in freeze/crush-injured muscle outlined by the white dotted line, scale bar 50μm.

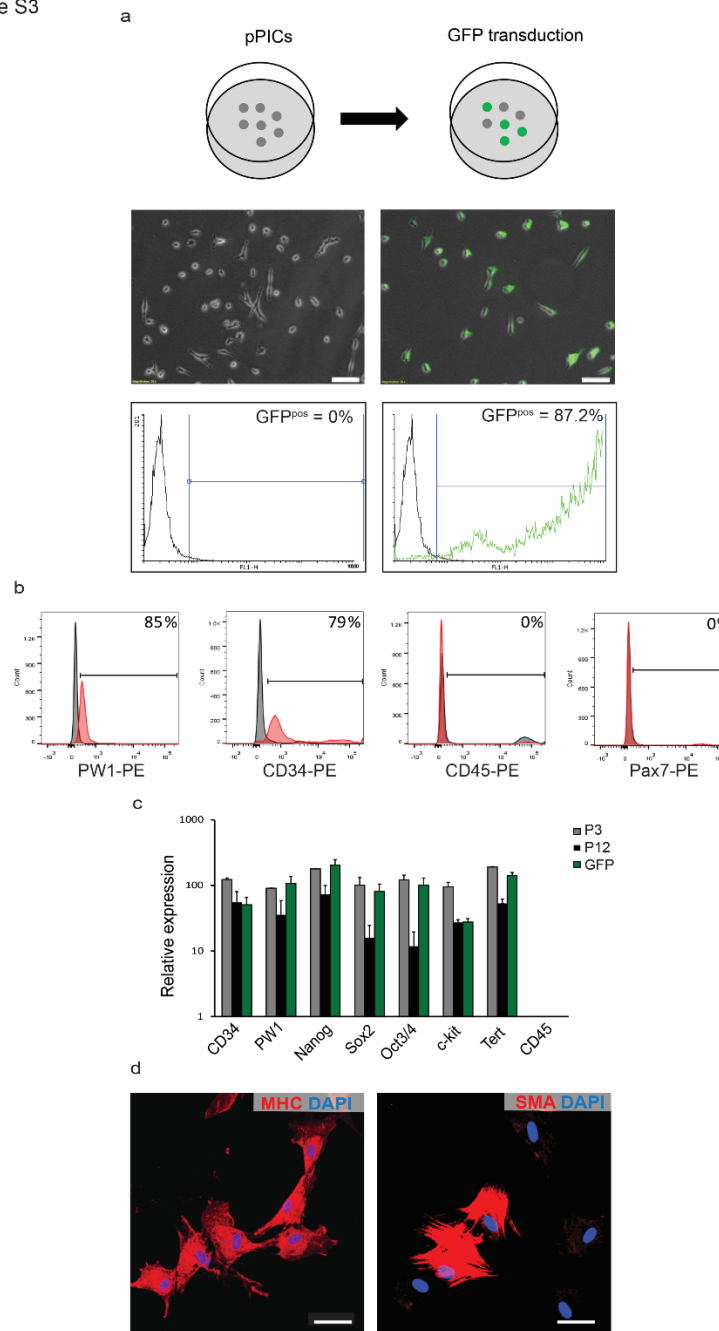
Figure S2



**Fig. S2. CTX-injured muscle undergoes significant regeneration 21 days post-injury.**

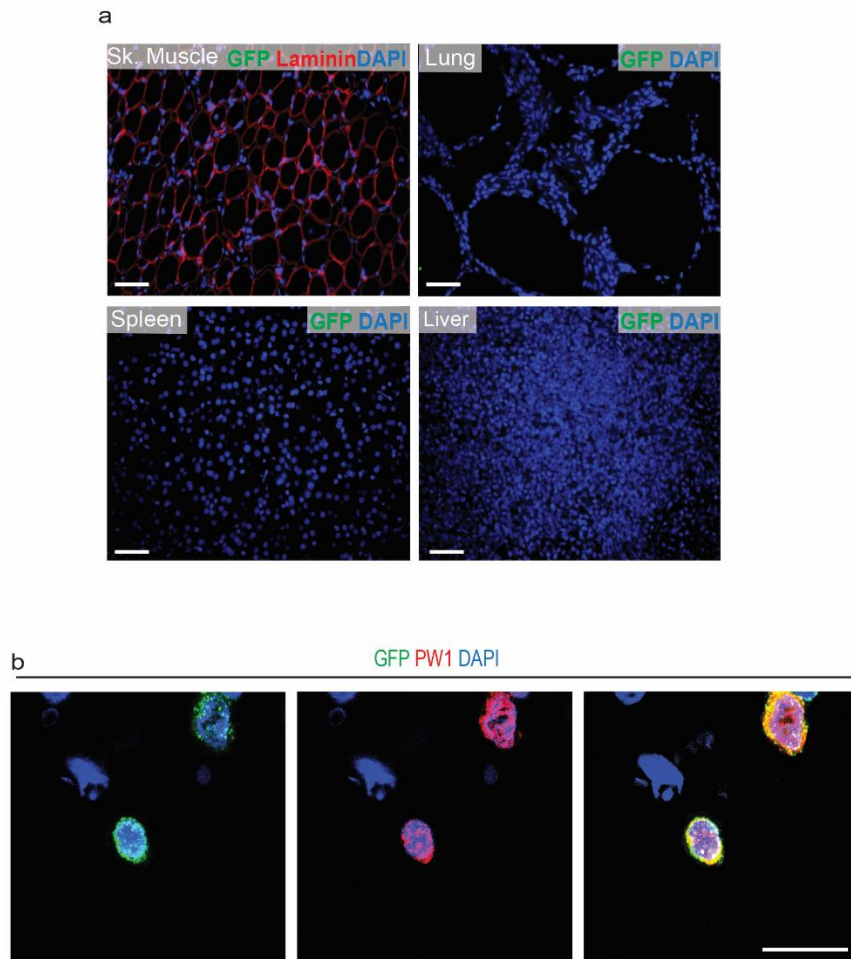
(a) Representative image of HVG stained CTX-injured paraffin-embedded muscle 21 days post-injury compared to 14 days post-CTX injury, scale bar 100 $\mu$ m. (b) Quantification of the ratio of skeletal muscle to connective tissue CSA in CTRL, CTX: 14 days and CTX: 21 days post-injury determined from five fields of view per muscle, n=2 animals per group. Data are mean  $\pm$  SD, \* $P$ <0.01 vs. CTRL, \*\* $P$ <0.0001 vs. CTX: 21 days post-injury, n=2.

Figure S3



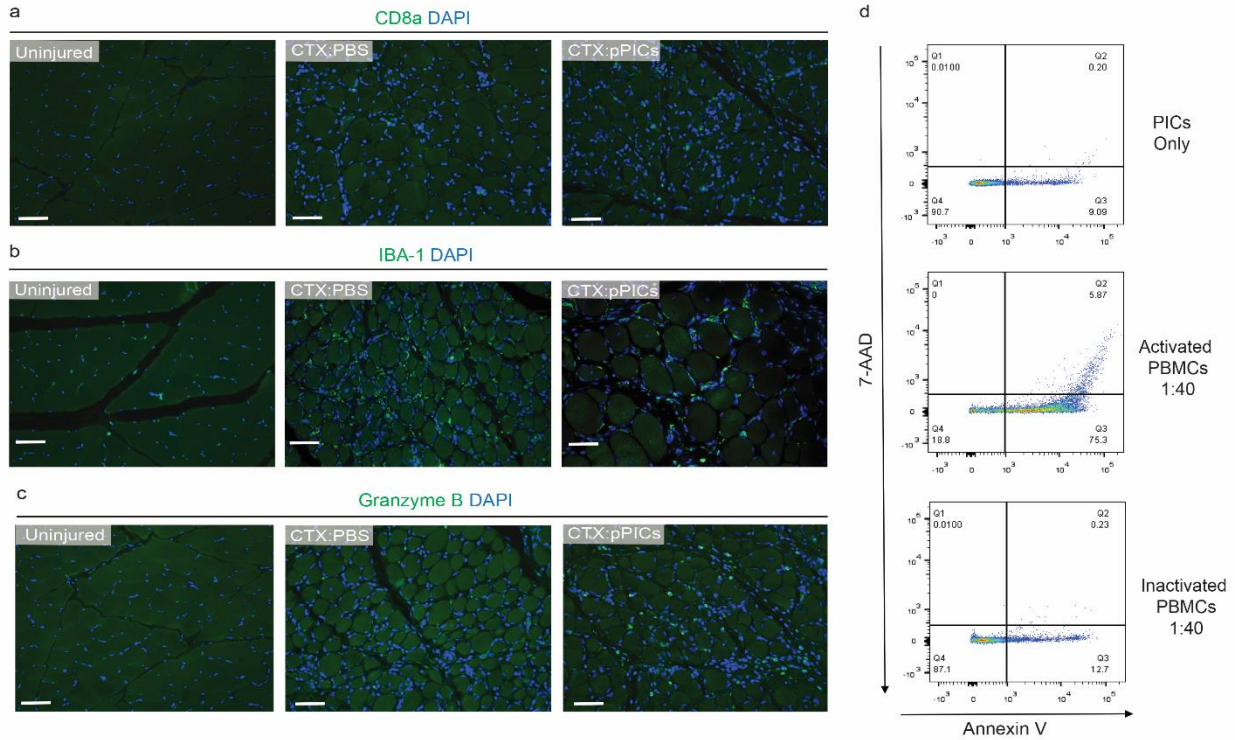
**Fig. S3. *GFP<sup>pos</sup> pPICs retain their phenotype and myogenic differentiation potential.*** (a) Porcine PICs were transduced with a GFP lentiviral construct at an efficiency of 87% as determined by flow cytometric analysis. (b) Flow cytometry confirmed  $GFP^{pos}$  pPICs expressed PW1 and CD34 while being negative for CD45 and Pax7. (c) qRT-PCR profile of  $GFP^{pos}$  pPICs compared to unlabelled pPICs confirms maintenance of pPIC phenotype. (d) Porcine PICs were subjected to differentiation media and display bi-potent muscle differentiation potential as demonstrated by expression of MHC and SMA (both red). DAPI stain nuclei in blue. Scale bar 50 $\mu$ m.

Figure S4



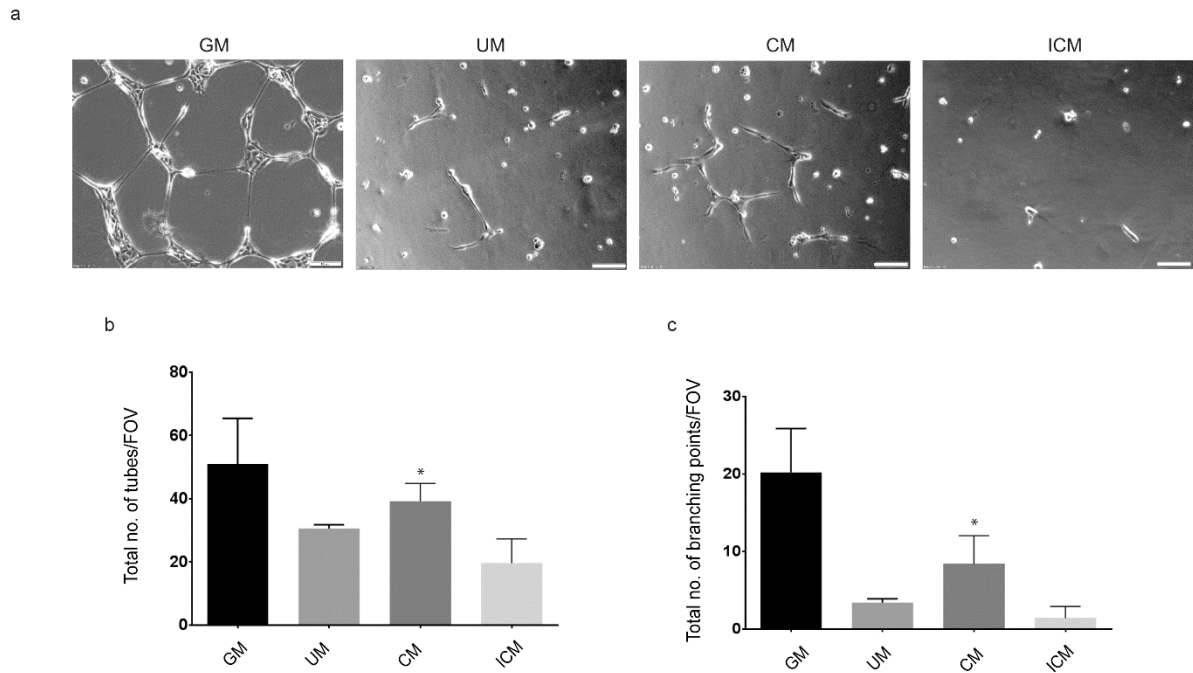
**Fig. S4. Evaluation of  $GFP^{pos}$  pPIC distribution in host tissues.** (a) No donor  $GFP^{pos}$  PICs were identified in PBS-CTRL skeletal muscle, or the lung, spleen or liver examined from animals transplanted with allogeneic  $GFP^{pos}$  pPICs 14 days post-transplantation, scale bar  $50\mu m$ . (b) Co-expression of GFP and PW1 confirmed that the interstitial nuclei were donor  $GFP^{pos}$  pPICs, scale bar  $10\mu m$ . DAPI stain nuclei in blue.

Figure S5

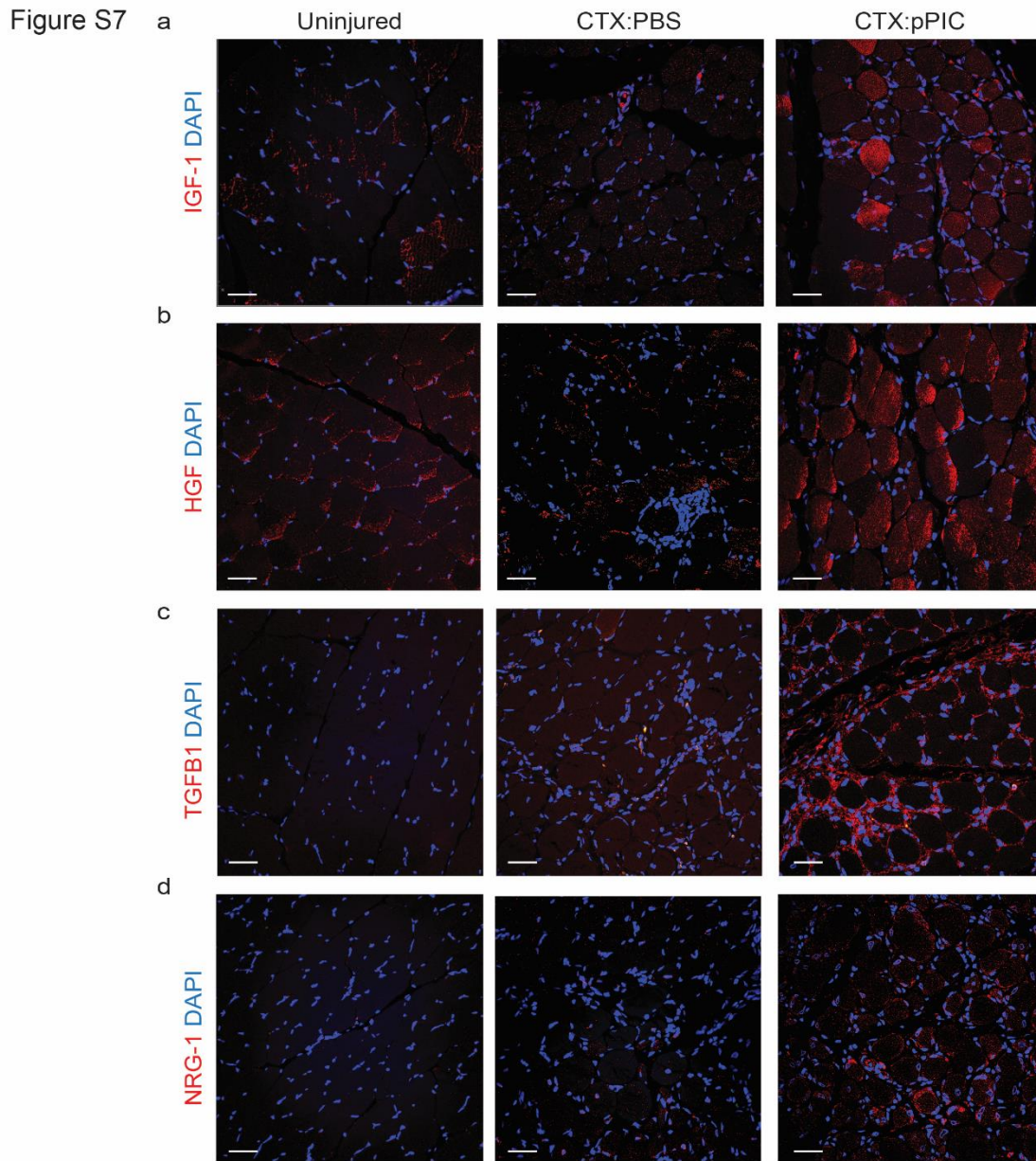


**Fig. S5. Allogeneic pPICs are cleared by immune cells** (a) Representative micrographs of skeletal muscle sections stained for (a) the cytotoxic T-lymphocyte marker, CD8a, (b) the macrophage marker, IBA-1, (c) the NK cell and lymphocyte marker, Granzyme B at 14 days post-injury, scale bar 50 $\mu$ m. (d) Representative Annexin V/7-AAD flow cytometry dot plots of pPICs only, co-culture of pPICs with activated porcine PBMCs, and co-culture of pPICs with inactivated porcine PBMCs. Apoptosis was measured by 7-AAD and Annexin V.

Figure S6



**Fig. S6. *pPIC* conditioned media promotes *in vitro* angiogenesis.** (a) Representative micrographs of HUVECs subjected to a 24h angiogenesis assay supplemented with endothelial growth media (GM), unconditioned media (UM), 24h *pPIC* conditioned media (CM) or heat inactivated conditioned media (ICM), scale bar 100 $\mu$ m. Endothelial network formation was quantified based on parameters including number of capillaries per field of view (b), and total number of branching points per field of view (c), which were found to be significantly increased ( $p < 0.05$ ) where HUVECs were exposed to 24h *pPIC* conditioned media compared to heat-inactivated conditioned media.



**Fig. S7. Evaluation of growth factor expression in pPIC-treated muscle.** The growth factors IGF-1, HGF, TGF $\beta$ 1, NRG-1, were screened in uninjured, CTX:PBS and CTX:pPICs-treated muscle at 14 days post-injury, n=5 animals per group. Scale bar 50 $\mu$ m.



**Table S1.** Volcano plot dataset listing all genes that were significantly upregulated or downregulated by pPICs, which had undergone 24h of myogenic differentiation compared to undifferentiated pPICs.

<b>Genes Upregulated in Differentiated vs. Undifferentiated pPICs</b>		
<b>Gene Symbol</b>	<b>Fold Regulation</b>	<b>p-value</b>
INHBA	22.284	0.000488
SPP1	8.8333	0.0123
IL8	5.3279	0.001386
BMP4	5.2978	0.02984
IGF1	4.9029	0.120844
POSTN	3.8978	0.001753
TGFB2	3.7774	0.007102
BDNF	3.3156	0.007734
CCL2	3.0488	0.081075
FGF9	2.7595	0.092373
VEGFd	2.7526	0.010973
GDF15	2.7265	0.036191
AFGF	2.6136	0.11545
NTF3	2.3613	0.155942
IGF2	2.3254	0.141795
IL1A	2.3105	0.138098
PDGFRC	2.134	0.020948
INHBB	2.0393	0.155071
Neuregulin 1	2.0185	0.045671
<b>Genes Downregulated in Differentiated vs. Undifferentiated pPICs</b>		
<b>Gene Symbol</b>	<b>Fold Regulation</b>	<b>p-value</b>
IL6	-10.5331	0.013034
PGF	-3.9006	0.067643
GPI	-3.45	0.026743
HGF	-3.1783	0.077588
NOGGIN	-3.0586	0.140538
BMP1	-2.7257	0.35925
LIF	-2.2785	0.277737
SCF	-2.0049	0.067705