1 SUPPLEMENTARY INFORMATION

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3	The habu genome reveals accelerated evolution of venom protein
4	genes.
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41 Characterization of the habu venom genes

42 Followings are descriptions of each of the 18 families.

43

44 (a) Metalloproteinases (MP)

45Metalloproteinases (MPs) are one of the major constituents of snake venoms and key 46toxins involved in venom-induced pathogenesis such as hemorrhage, fibrinolysis and 47apoptosis. Snake venom MPs (svMPs) are classified into four groups (P-I to P-IV) 48according to their domain architecture and size: P-I MPs possess a metalloproteinase 49domain only and are largely hemorrhagic; P-II contain metalloproteinase and disintegrin 50domains and are larger; P-III have metalloproteinase, disintegrin, and cysteine-rich 51domains; and P-IV have lectin-like domain(s) linked by disulfide bonds to a P-III 52structure¹. The structural complexity of P-III MPs yields a great diversity of their 53functions. P-II, P-III, and P-IV groups belong to the 'disintegrin and metalloproteinase' 54(ADAM) family.

55A thorough examination of P. flavoviridis gene models and transcriptomes identified at 56least 11 svMP genes (svMP01 to svMP11) (Supplementary Fig. S3a; Table 1) and 55 57transcripts that are expressed in the venom gland (Supplementary Fig. S3a; Table 1). 58svMP01, svMP02, svMP03 and svMP11 (Gene model ID: habu1 s2862 g10314a/b/c/d) 59were located tandem on the same scaffold (habu1 scaffold 2862, Acc no. 60 BFFQ01002098) (Supplementary Fig. S8a). They encoded а vascular 61 apoptosis-inducing protein HV1, flavorase, a VMP III-like protease, and NaMP-like 62 protease, respectively (Supplementary Fig. S3a). Three genes, svMP06 (Gene model ID: 63 habu1 s14911 g21429), svMP07 (g21430), and svMP08 (g21431), which encode H2 64 metalloproteinase, HR2a, and flavoridin, respectively, were also located on the same 65scaffold, habu1 scaffold 14911 (Acc no. BFFQ01007560) (Supplementary Fig. S3a; 66 Supplementary Fig. S8a). Since the two scaffolds form a super-scaffold, it is highly 67 likely that svMPs (svMP01-03, 11 and svMP06-08) are located tandem in close 68 proximity to each other (Supplementary Fig. S8a). In addition, svMP04 genes (Gene 69 model ID: habu1 s3258 g11210) and svMP05 (g11211) encoded a jerdonitin-like 70protease, the major hemorrhage factor, HR1a, and a jerdonitin-like protease, 71respectively, and they were all located on habul scaffold 3258 (Acc no. 72BFFQ01002364) (Supplementary Fig. S3a). svMP09 and svMP10 (Gene model ID: 73habu1 s399953 g24864a/b) encoded HR1b and Mt-b/elegantin-like protease. 74respectively, and were located on habu1 scaffold 399953 (Acc no. BFFQ01081176) 75(Supplementary Fig. S3a). Numbers of exons have also become highly variable in this 76gene family, ranging from 13 to 17, of which exon 11 contains a catalytic zinc binding 77motif, HEXGHNLGXXHD (Supplementary Fig. S10).

78

Furthermore, among non-venom (nv) MPs, we identified 17 paralogous genes (*nvMP01* to *nvMP17*) of *ADAM* (a disintegrin and metalloproteinase) and 26 paralogous genes (*nvMP18* to *nvMP43*) of *ADAMTS* (ADAM with thrombospondin motifs) and 11 paralogous genes (*nvMP44* to *nvMP54*) of *MMP* (matrix metalloproteinase) (Supplementary Fig. S3b). ADAM and ADAMTS include trans-membrane and secreted proteins with functions in cell adhesion and proteolytic processing of the ectodomain of

85 cell-surface receptors and signaling proteins as key contributing molecules to various 86 physiological functions including cell adhesion, fertilization, migration, proteolysis and signaling²⁻⁵ in addition to other MPs such as MMP (*nvMP44* to *nvMP54*). Predicted 87 88 genes encoding nvADAMs and nvADAMTS were located on different scaffolds than svMPs, except for nvMP57 located on habu1 s2862, which contains svMP01, svMP02, 89 90 svMP03 and svMP11 (Supplementary Fig. 3b, Supplementary Fig. 8a). 91 Phylogenetic analysis shows that habu SV and NV metalloproteinase genes are 92distinctly clustered into SV and NV homologs such as ADAM, ADAMTS, and MMP 93from different venomous snakes and other reptile except for ADAMTS9 94(Supplementary Figs. S3c, S3d and S3e). Phylogenetic tree of MP also shows that 95svMPs has been derived from the ancestor of ADAM clade including ADAM8, 9, 12, 96 15, 19, 21, 32, and 33 (Supplementary Fig. S3c). Moreover, it was found that the 97 apparent clade including NaMP-like svMP (svMP11) has initially diverged from a 98 common ancestor of P. flavoviridis svMPs (Supplementary Fig. S3d).

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100 (b) Serine proteases (SP)

101 Serine proteases (SPs) are also major constituents of venom proteins. They act on 102of blood coagulation, various macromolecular substrates fibrinolytic, and kallikrein-kinin systems, and on platelets to cause hemostatic defects⁶. Although svSPs 103 104 belong to the chymotrypsin subfamily (peptidase subfamily S1A), they show distinct 105substrate specificities toward macromolecules, compared the relatively non-specific 106activity of trypsin or chymotrypsin.

107 The P. flavoviridis genome contains at least 11 svSP genes (svSP01 to svSP11) 108 (Supplementary Figs. S4a and S11) with complete sequences. svSP01 (TLf1) was the 109 most abundant svSP transcript in the venom gland corresponds to an acidic protein, known as flavoxobin^{7,8} and habutobin⁹. It encodes a weakly thrombin-like enzyme of 110 111 242 amino acids that specifically releases fibrinopeptide A from fibrinogen⁷. Although 112no information is available with regard to possible kallikrein-like activity, it was shown 113 that flavoxobin acts as a heterologous C3-convertase that selectively releases human C3b and C3a¹⁰. svSP02 (*TLf2*) is also major svSP transcript that encodes a basic 114 115thrombin-like enzyme (pI >10) (Supplementary Fig. S4a).

116More recently, we found that TLf2 enhances the myonecrotic activity of Lys49-PLA2, 117 although TLf2 was hydrolytically inactive due to replacement of its catalytic residue 118 His57 with Arg (Ogawa et al., in preparation). From the P. flavoviridis venom, a 119kinin-releasing enzyme, flavorase, and thrombin-like enzyme possessing fibrinopeptide A- and B-releasing activity, flavoviridiobin, have been reported^{11, 12}. However, since no 120121sequence data are available, we could not identify them among transcripts and genome 122sequences of our analyses. From the sequence homologies of P. flavoviridis svSPs with 123other snake svSPs using phylogenetic tree analysis (Supplementary Fig. S4c), svSP03, 124svSP04, and svSP05 correspond to beta fibrinogenase, catroxase-like kinin-releasing 125enzyme, and flavorase, respectively, which have kinin-releasing activity similar to KN-BJ from *Bothrops jararaca* venom¹³. Several of the genes were located tandem in 126127the certain scaffolds, that is, svSP06 (Gene ID: habu1 s4106 g13431a), svSP07 (g13431b) and svSP10 (g13432) were located on scaffold 4106 (Acc no. 128

BFFQ01002946) (Supplementary Fig. S8b). svSP04 (habu1_s6789_g17480), svSP05(g17481a), and svSP08 (g17481b) were located on the scaffold_4106 (Acc no. BFFQ01002946), while svSP09 (habu1_s7597_g18190c), svSP01 (g18190a), and svSP03 (g18190b) were located on the scaffold 7597 (Acc no. BFFQ01004842) (Supplementary Fig. S8b).

134Based on the similarity of enzymatic activities and the conservation of primary and 135tertiary structures of SPs, it is believed that svSPs evolved from an ancestral 136 kallikrein-like SP. svSP genes consist of 6 exons except for svSP06, which contains 137additional two exons (exons 7 and 8) including 3' non-coding region (Supplementary 138Fig. S11). svSP possess the same exon-intron junctions as coding regions of mammalian kallikrein (KLK)¹⁴ and trypsin gene rather than other SPs such as elastase 139 140 (Supplementary Fig. S11b). In addition to sv-related SP genes, we found at least 34 141SP-family genes with non-venom related functions (Supplementary Fig. S4b).

Phylogenetic analysis shows that habu SV and NV serine proteinase genes are distinctly
clustered with SV and NV homologs from different venomous snakes and other reptile.
Furthermore, phylogenetic tree of SP also shows that svSPs derived from the
trypsin-like SPs (Supplementary Fig. S4c).

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147 (c) C-type lectin-like proteins (or snaclecs) / Galactose binding lectins (CTLP)

148 C-type lectins (CTLPs) are one of the major animal lectin families, members of which

149 bind in Ca²⁺-dependent fashion to carbohydrates. In general, they adopt multidomain

150 structures and contain one or more highly conserved carbohydrate recognition domain

151 (CRD) consisting of 115–130 amino acids with a unique α/β topology. To date, the CTL 152 domain superfamily has been classified into 17 groups¹⁵⁻¹⁸ first discovered lectin 153 activity (agglutinating activity of erythrocytes and leukocytes) in snake venoms, and the 154 first isolation and characterization were reported by Gartner et al.¹⁹.

155To date, primary structures of 15 kinds of snake venom galactose-binding lectins (svGBL) have been determined, and all svGBLs are C-type lectins²⁰. Furthermore, 156157C-type lectin-like proteins (CTLP) lacking carbohydrate-binding activity have been 158isolated from snake venom and characterized as heterodimers, which were dimerized by 159unique domain swapping between alpha and beta chains, resulting in a conformational change in the central loop with a new concave surface^{21,22}. Dimeric svCTLPs showed 160161 several pharmacological activities such as anticoagulant, platelet aggregations 162(agonistic) and their inhibition (antagonistic) via specific binding to coagulant factors 163IX and X, von Willebrand factor, and integrins on the platelet such as GPIa/IIa, GPIb, 164GPVI.

165The present study identified 10 genes encoding svCTLPs (Supplementary Fig. S5a) and 166 39 genes for non-venomous (nv) CTLs (Supplementary Fig. S5b). Molecular phylogeny 167 shows that the ten svCTLP genes are subdivided into three groups, lectin-type 168(svCTLP08 and svCTLP10), B-chain type (svCTLP05, 06 and 07), and A-chain type 169(svCTLP01, 02, 03, 04 and 09) (Supplementary Fig. S5c). The genes encoding svCTLPs 170were composed of six exons with conserved exon-intron junctions except for svCTLP08, 171which lacks exons 1 - 3, resulting in four exons (Supplementary Fig. S5a). svCTLP01, 172svCTLP02, svCTLP05 and svCTLP09 encode factor XI/X binding protein alpha,

173flavocetin A alpha, factor XI/X binding protein beta, and new factor XI/X binding 174protein alpha-like with 83% identity, respectively, although they were not assigned gene 175model ID numbers (Supplementary Fig. S5a). Interestingly, four genes, svCTLP03 176 (Gene model ID: habu1 s10061 g19810 c), svCTLP04 (g19810 a), svCTLP06 177(g19809), and svCTLP07 (g19810 b), which encode stejaggregin-A like alpha, 178rhodocetin/EmEMS 16-like alpha, flavocetin A beta, and rhodocetin/EmEMS 16-like 179beta, respectively, were located tandemly on habul scaffold 10061 (Acc no. 180 BFFQ01005768) (Supplementary Fig. S8c).

181 Non-venom CTL genes encoding C-type lectins, nvCTLP01 (Gene model ID: 182habul s10061 g19804), nvCTLP02 (g19805), nvCTLP03 (g19806), nvCTLP04 183 (g19807), and nvCTLP05 (g19808) (Supplementary Fig. S5b), were located on the same 184 scaffold, habu1 scaffold 10061 (Acc no. BFFQ01005768) (Supplementary Fig. S8c). 185On the other hand, svCTLP08 (Gene model ID: habu1 s3168 g10977) and svCTLP10 186 (g10975), which encoded venom C-type lectins, were located on habu1 scaffold 3168 187 (Acc no. BFFQ01002306). Non-venom CTL genes, nvCTLP12 (Gene model ID: 188 habu1 s3168 g10976), nvCTLP13 (g10978), and nvCTLP14, and (g10979) 189 (Supplementary Fig. S5b) were also located on habul scaffold 3168 (Acc no. 190 BFFQ01002306) (Supplementary Fig. S8c).

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192 (d) Phospholipases A₂ (PLA2)

193 Phospholipase A₂ (PLA₂, EC 3.1.1.4) catalyzes hydrolysis of 2-acyl ester bonds of

194 3-sn-phosphoglycerides in the presence of Ca²⁺, to liberate 3-sn-lysophosphoglycerides

and fatty acids²³. Snake venoms PLA₂s are major toxins and are classified into groups I
and II based on disulfide bonds²⁴. Group-I PLA₂s are found in elapid venoms while
group II are in viperid venoms. Group-II PLA₂s are further divided into two subgroups,
[Asp49] PLA₂ forms and [Lys49] PLA₂ forms²⁵. It has been reported that they share the
same scaffold²⁶⁻²⁸. Numerous studies have shown the presence of multiple PLA₂ genes
in habu venoms²⁹⁻³⁴.

The P. flavoviridis genome contains ten types of group II venom PLA2 genes 201202(svPLA201a, b to svPLA209) (Supplementary Fig. 6a). In addition, we also confirmed 203that four PLA₂ transcripts, including a hemolytic [Asp49] PLA₂ (svPLA201) [Accession #: D10070.1, D10720.1, D10722.1, AB778558.1, AB072174.1, AB778559.1], 204205edema-inducing basic [Asp49]PLA2 (svPLA202), a weak neurotoxin PLA-N 206 (svPLA203) [Accession #: AB848131], and [Lys49]PLA2 myotoxins (svPLA204 to 207svPLA206)) (BPI, BPII and BPIII) [Accession #: D10718.1, D10719.1, AB470470.1], 208were expressed in the venom gland (Supplementary Fig. S6a).

209 In addition, the P. flavoviridis genome contains at least 31 genes encoding non-venom 210PLA2s (Supplementary Fig. S6b). These include type I PLA2s such as nvPLA201 and 211nvPLA202 (habu1 s9792 g19655 and habu1 s9792 g19656, respectively) and type II 212PLA2, nvPLA203 (habu1 s4562 g14470) (Supplementary Fig. S6b). Molecular 213phylogeny of PLA2 showes that svPLA2 genes have been evolved from ansestoral gene 214of GIIE gene, and then diversified into multiple gene types with several toxic activities 215such as myotoxic, hemolytic, edema-inducing and neurotoxic (Supplementary Fig. 216S6c).

217

218 (e) Three-finger toxin (3FTX)

219Three-finger toxins (3FTXs) are non-enzymatic neurotoxins composed of about 60-70 220amino acid residues. They are major components in the venoms of elapids (cobras, 221kraits and mambas), hydrophiids (sea snakes) and colubrids, but extremely minor components in viperid snakes^{42,43}. Although the gross structures of 3FTXs are 222223conserved, they have diverse pharmacologies such as L-type calcium channel inhibition, 224 acetylcholinesterase inhibition, and muscarinic and nicotinic acetylcholine receptor antagonism^{44,45}. 3FTXs were considered specific to elapid venoms until they were 225shown to be present in Sistrurus catenatus edwardsii venom (0.83% abundance)^{37,46} and 226 a venom gland transcriptome of *P. flavoviridis*²¹. 227

228We identified four genes encoding 3FTXs in the P. flavoviridis genome (3FTX-01, 2293FTX-02, 3FTX-03 and 3FTX-04; Gene models habu1 s4579 g14476, g14477, _g14478, _g14475, respectively) (Supplementary Fig. S12a). These are located 230231tandemly on the same scaffold, habu1 scaffold 4579 (Acc no. BFFQ01003259). From 232RNA-seq data of *P. flavoviridis* tissues, these *3FTX* genes generate single transcripts, 233respectively (Supplementary Fig. S12a). 3FTX-01 and 3FTX-02 were expressed in 234venom gland. Three 3FTXs, 3FTX-01, 02 and 04, have highly conserved Cys residues. 235Furthermore, two non-venom 3FTX genes, 3FTX-05 (habu1 s138 g00722) and 2363FTX-06 (habu1 s138 g00721), encoding UPAR-Ly6 and CD59, respectively, were 237identified on the same scaffold, habu1 scaffold 138 (Acc no. BFFQ01000122)

238 (Supplementary Fig. S12a). Seven transcript variants were generated from 3FTX-05,

although other *3FTX* genes generate single transcript. Molecular phylogeny shows that
toxic 3FTX proteins form a clade different from non-toxic protein clade (Supplementary
Fig. S12b).

242

243 (f) Aminopeptidase (APaseN)

244Aminopeptidases (EC 3.4.11) remove one or more specific N-terminal residues from 245target proteins or peptides. For example, aminopeptidase L (APL: leucyl 246aminopeptidase) removes an N-terminal leucine residue. Aminopeptidase A (APA) and 247aminopeptidase B (APB) remove an acidic N-terminal residue and a basic N-terminal 248residue, respectively. Aminopeptidase N (APN) removes neutral N-terminal residues, 249typically alanine. To date, several aminopeptidase activities have been detected in 250venoms from elapids and vipers, and the cDNA sequence of APA from Gloydius 251*blomhoffii brevicaudus* snake has been determined⁴⁷.

252In this study, we identified two svAPases genes and ten nvAPases genes 253(Supplementary Fig. S13a). svAPases genes include APase01 (habu1 s1390 g05001) 254and APase 03 (habu1 s3769 g12636), which encode APN and APA, respectively. They 255reside in independent scaffolds. On the other hand, ten nvAPases genes include 256aminopeptidase-like (APNPEPL) (habu1 s5205 g15384), APB (habu1 s89848 g19021), APO (habu1 s4166 g13605), APQ (habu1 s3446 g11762), 257258APD (habu1 s133 g00676), two APMs (habu1 s7599 g18195 and 259habu1 s9114 g19166), and three XPAPs (habu1 s4753 g14695, habu1 s4159 g13556, 260and habu1 s1233 g04248) (Supplementary Fig. S13a).

261 Molecular phylogeny shows that APN and APA form a clade, and this SV-related clade

- has affinity with a clade including APases O, B and Q (Supplementary Fig. S13b).
- 263
- 264 (g) Cysteine-rich secretory proteins (CRISP)

265Cysteine-rich secretory proteins (CRISP) are glycoproteins implicated in the 266mammalian male reproductive functions spanning haploid germ cell development, 267epididymal maturation, capacitation, motility, and the actual processes of fertilization⁴⁸. 268The first discovered CRISP, acidic epididymis glycoprotein (AEG, also known as 269protein D/E or CRISP-1), is an androgen-responsible secretory protein involved in 270gamete fusion. Two other mammalian CRISPs, testis-specific CRISP-2 (TPX-1) and 271salivary gland-specific CRISP-3 (28 kDa specific granule protein, SGP28) have been 272isolated and characterized.

273CRISP-family proteins belonging to CRISP-3 have also been found in various snake venoms⁴⁹⁻⁵² and the Mexican beaded lizard⁵³⁻⁵⁵. CRISPs contain conserved C-terminal 274275helical bundle subdomains termed ion channel regulatory (ICR) domains. ICR domains 276show homology and similar disulfide-bonding patterns to peptide toxins, BgK and ShK, which are potent inhibitors of K channels isolated from the sea anemones, Bunodosoma 277granulifera^{56, 57} and Stichodactyla helianthus^{58, 59}, respectively. For example, triflin, 278279ablomin, latisemin, ophanin and piscivorin block smooth muscle contruction caused by inhibition of L-type Ca²⁺ channels. Naturin from Naja atra acts as a blocker of BK_{Ca} 280channels⁶⁰. On the other hand, unique CRISP family proteins, pseudechetoxin (PsTx) 281282and pseudecin, which block olfactory and retinal cyclic nucleotide-gated ion (CNC) 283 channels, and crovirin, which shows anti-parasitic activity against infective 284 trypanosoma forms have been identified⁶¹. Thus, venom CRISPs show specific 285 activities against a variety of ion channels and possess various physiological activities. 286 To date, cDNAs (transcriptome data) for triflin (AF384219.1, AB848115.1, 287 AB985232.1) and CRISPs with EGF-like domains (AB851959.1) have been identified 288 in *P. flavoviridis* snake venom^{50,32}.

The present study identified four CRISP-related genes, CRISP01 to CRISP04, in the P. 289290flavoviridis genome (Supplementary Fig. S14a). The genes encoding triflin (CRISP01; 291Gene IDs: habu1 s22025 g22470a) and a novel triflin-like peptide (CRISP02; Gene IDs: 292habu1 s22025 g22470b) were located in the same scaffold, 293habul scaffold 22025 (Acc no. BFFQ01031149) (Supplementary Fig. S14a). On the 294other hand, genes encoding two types of CRISP with an EGF-like domain, CRISP03 295(Gene ID: habu1 s1243 g04322) and CRISP04 (Gene ID: habu1 s264 g01336), were 296located on habu1 scaffold 1243 (Acc no. BFFQ01000866) and habu1 scaffold 264 297 (Acc no. BFFQ01000220), respectively (Supplementary Fig. S14a). Interestingly, 298although both triflin and triflin-like genes contain eight exons (ex1 to ex8 for triflin, and 299ex1a, ex1b, ex2 to ex7 for triflin-like) and their N-terminal regions including signal 300 sequences and pathogenesis-related CRISP domains corresponding to exons 2 to 5 were 301 conserved, the number and structures of exons encoding C-terminal regions, including 302 ICR domains were quite different (Supplementary Figs. S14c and S14d). These results 303 suggest that CRISP/triflin and triflin-like genes arose through the gene duplication and exon shuffling. On the other hand, genes encoding CRISPs with EGF-like domains 304

showed the same exon-intron architecture and homologies without a C-terminal
extension of *CRISP04*. Molecular phylogeny supports these notions (Supplementary Fig.
S14b)

308

309 (h) SPRY/Vespryns (Vespryin)

310The vespryn family is a relatively recently discovered as a snake venom protein family 311 containing consensus PRY-SPRY domains. The SPRY domain is an interaction module 312 consisting of ~120 amino acids, which is implicated in the biological pathways 313 including innate and adaptive immunity. The SPRY domain was first identified in SPIA 314(spore lysis A) kinase from Dictyostelium discoideum, and in mammalian ryanodine receptors^{62, 63}. Ohanin, which was isolated from the king cobra, Ophiophagus hannah 315 venom, shows dose-dependent hypolocomotive and hyperalgesic effects in mice^{64, 65}. 316 317The present study identified a gene encoding vespryn (Ves01) and another 11 genes

318 encoding SPRY domain-containing proteins (Ves02 to Ves12) (Supplementary Fig. 319 S15a). Vespryn gene includes 5 exons and 4 introns, while butyrophilin2A1 320 (btn2A1)-encoding gene (Ves02) includes 10 exons. Butyrophilins (BTN) are members 321 of a protein family that belong to immunoglobulin superfamily that have 322 immunomodulatory functions. Both vespryn and butyrophilin genes are located on the 323 same scaffold, habu1 scaffold 402940 (Acc no. BFFQ01082524) (Supplementary Fig. 324 S15a), and molecular phylogeny shows affinity or similarity of these two gene products 325 (Supplementary Fig. S15b), indicating that vespryn is more likely to be derived from 326ancestor gene of butyrophilin.

327

328 (i) 5'-Nucleotidases (5Nase)

5'-nucleotidase (5Nase, EC: 3.1.3.5) dephosphorylates 5'-mononucleotides to release purine and pyrimidine nucleosides. Viperidae and crotalidae venoms display more 5'-nucleotidase activity than elapidae venoms⁶⁶. 5Nase inhibits platelet aggregation due to the liberation of adenosine. Venom 5Nase act synergistically *in vivo* with other toxins such as ADPases, hemorrhagic proteases, fibrinogenases and disintegrins to exert a more pronounced anti-coagulant effect⁶⁷.

335The present study identified one sv5Nase gene, sv5Nase1 (habu1 s6028 g16570) 336 (Supplementary Fig. S16a). In addition, the P. flavoviridis genome contained at least 10 337 nv5Nase (habu1 s1426 g05181 for 5Nase02 to habu1 s2849 g10145 for 5Nase11) 338 (Supplementary Fig. S16a). Molecular phylogeny shows that sv5Nase01 form a clade 339 with other SV genes while the other nv5Nase genes cluster with other reptile NV 5Nase 340genes (Supplementary Fig. S16b), indicating the affinity or similarity of the SV and NV 341 gene products in the 5Nase01 clade. An ancestral gene appears to have diverged first 342 into NV forms, and then one of them has been functionalized as a toxin (Supplementary 343Fig. S16b).

344

345 (j) Dipeptidyl Peptidase IV (DPP IV)

Dipeptidyl peptidases (DPP) (EC 3.4.14.-) cleave dipeptides from the N-termini of
polypeptides, and are classified into nine distinct types (DPP I-IV, VI-X). Mammalian
DPP-IV is a highly glycosylated serine protease and a type II membrane protein that

349 selectively cleaves N-terminal dipeptides, Xxx-Pro (or Ala) from polypeptides. DPP-IV 350 has been detected on the surface of immune cells as being CD26, a cell-surface differentiation marker in the T cell linage⁶⁸, and also detected in serum⁶⁹ and seminal 351plasma⁷⁰ as a soluble form. More recently, it has been reported that DPP-IV plays as a 352 353 processing enzyme for liberation of biologically active peptides in the skin of Xenopus *laevis*⁷¹ and snake venoms⁷². It was proposed that svDPP-IV contribute to hypotension 354 by destroying vasoconstrictive peptides such as Peptide YY, neuropeptide Y and 355 substance P, and shows synergistic effect with BPPs-ACE inhibitory peptide⁷³. 356

The present study identified 8 genes encoding dipeptidyl peptidases, *DPP01* to *DPP08* (Supplementary Fig. S17a), one of which was venom svDPP-IV (*DPP05*, habu1_s1020_g03622). Molecular phylogeny supports the evolutionary history of svDPP-IV and other NV DPPs that diversified into 8 genes (Supplementary Fig. S17b).

361

362 (k) Hyaluronidases (Hyal)

Hyaluronidases (Hyals) (EC: 3.2.1.35) are enzymes that cleave predominantly the glycosaminoglycans, hyaluronan (HA) and less efficiently, chondroitin (Ch) and chondroitin sulphate (ChS) in the extracellular matrix (ECM) of animals⁷⁴. HA participate in physiological processes such as embryogenesis, cell migration, wound healing, tissue turn over, and malignancies⁷⁵.

368 Hyaluronidase is also common component of snake venoms known as a venom 369 spreading factor to degrade HA in ECM⁷⁶. To date, the full-length cDNA sequences 370 encoding snake venom Hyal (svHyal) have been reported from *Echis ocellatus* 371 (EOC00242; DQ840249), Echis pyramidum leakeyi (EplHy-1: DQ840253), Bitis
372 arientans (BaHy-1: DQ840256, BaHy-2: DQ840257), Cerastes cerastes cerastes
373 (CccHy-1: DQ840250, CccHy-3: DQ840251, CccHy-4: DQ840257)⁷⁷ and, Bothrops
374 pauloensis⁷⁸. They showed different primary structures compared with other Hyals.

375

376In this study, we identified six genes encoding P. flavoviridis Hyals, Hyal01 to Hyal06 377(Supplementary Fig. S2a). Hyal01 (habu1 s7188 g17820) encodes a svHyal, which 378 was composed of six exons. On the other hand, other Hyal genes, Hyal02 (HYAL1), 379 Hyal03 (HYAL2), Hyal04 (HYAL2/4), Hyal05 (HYAL3) and Hyal06 (HYAL4), showed 380 quite different gene structures with different exon numbers (four to seven), although 381 core exons (exons 4 and 5 in *svHyal*) were conserved in all genes. Molecular phylogeny 382 supports such a relationship between P. flavoviridis hyaluronidase family members 383(Supplementary Fig. S2a).

384

385 (l) Nerve growth factor (NGF)

Nerve growth factor (NGF) is one of polypeptide hormones belonging to the neurotrophin family that is necessary for neuronal differentiation, survival, and maintenance. The neurotrophin family also includes brain-derived neurotrophic factor (BDNF)⁷⁹, neurotrophin-3 (NT-3)⁸⁰, and neurotrophin-4/5 (NT-4/5)⁸¹, which interact with cell surface receptors, p75 and Trk subfamily receptors, and stimulate tyrosine phosphorylation of Trk receptors⁸². NGFs are present in the venoms of elapids and viperids^{83, 84}. Cobra NGFs show lower biological activities than mouse NGF⁸⁵. More 393 recently, Sunagar et al reported that a duplication of NGF genes occurred in Elaphidae 394 snake, and discussed their putative roles in venom and their unique molecular evolution under the positive-selection⁸⁶. Furthermore, Hargreaves *et al.* reported the 395 396 transcriptomic analysis for body tissues and salivary and venom glands from five 397 species of venomous and nonvenomous reptiles, and discussed their molecular evolution based on the expression profiles⁸⁷. It suggested that snake venom does not 398399 evolve through the hypothesized process of duplication and recruitment of genes 400 encoding body proteins, and that many proposed venom toxin genes have been 401 restricted to the venom gland following duplication, not recruited. In the case for NGF, 402the nontoxic form of NGF is expressed in a diversity of tissues, including the salivary 403 glands of nonvenomous reptiles. The putatively toxic NGF has therefore also been 404 restricted to the venom gland following duplication⁸⁷.

In the *P. flavoviridis* genome, we identified a single gene encoding svNGF located on habu1_scaffold_3536 (*Neu01*, Gene ID: habu1_s3536_g11975) (Supplementary Fig. S2b), which consists of four exons (Ex1 to 4) as in chicken and mammalian NGFs. Interestingly, the expression profiles of *P. flavoviridis* NGF is specific to venom gland although several transcripts with different 5'exons were present (Supplementary Table S7). This alternative usage of 5' exons containing the 5'-untranslated region has been reported in the chicken NGF gene⁸⁸.

412 Furthermore, neurotrophin family genes encoding, *Neu02, 03 and 04*, BDNF (Gene ID:

413 habu1_s3803_g12736), NT-3 (habu1_s105_g00548), and NT-4 (habu1_s6123_g16797),

414 respectively, were identified in the genome (Supplementary Fig. S2b). Molecular

phylogeny indicates that *svNGF* and other three *BDNF*, *NT-3* and *NT-4* diverged first in
their history (Supplementary Fig. S2b), suggesting that the neurotrophin family
including NGF is a typical example of SV genes generated by 2R-WGD.

418

419 (m) Vascular endothelial growth factor-like proteins (VEGF)

420Vascular endothelial growth factors (VEGFs) are key regulators of vascular 421development during embryogenesis (vasculogenesis), blood-vessel formation (angiogenesis), skeletal growth, and reproductive functions⁸⁹. In mammals, five VEGFs 422423 (VEGF-A, -B, -C, -D, and -E (viral VEGF)) have been identified so far. These ligands 424bind to three receptor tyrosine kinases, VEGF receptor-1 (VEGFR1, Flt-1, fms-like 425tyrosine kinase-1), -2 (VEGFR2, KDR, kinase insert domain-containing receptor) and 426 -3 (VEGFR3, Flt-4), as well as to co-receptors such as heparan sulphate glycans. 427VEGFs have been found in snake venoms, and they showed strict receptor specificities 428 compared with mammalian VEGF. For example, Vammin and VR-1 from the venoms 429 of Vipera a. ammodytes and Daboia r. russelli, respectively, bind only KDR with high affinity but not to other VEGF receptors⁹⁰. Tf-svVEGF from P.flavoviridis venom have 430 been shown to bind Flt-1 in preference to KDR, unlike vammin and VR-1⁹¹. The gene 431432 structures of tissue- and venom-types VEGFs from *P. flavoviridis* have been reported⁹². 433 We identified three VEGF genes in the P. flavoviridis genome (Supplementary Fig. 434 S18a). One is svVEGF01 (habu1 s565 g02679) encoding svVEGF-F and the others are 435nvVEGF01 and nvVEGF02, encoding VEGF-A (habu1 s6836 g17529) and VEGF-C 436(habu1 s9381 g19343), respectively. Molecular phylogeny indicates that svVEGF-F is 437 within toxin-related protein group (Supplementary Fig. S18b).

438

439 (n) L-Amino acid oxidases (LAAO)

440 L-Amino acid oxidases (LAAO: EC 1.4.3.2) are flavoenzymes that catalyze the 441 oxidative deamination of L-amino acids to produce α -keto acids with the release of NH₃ 442and H₂O₂. LAAOs are abundant in some snake venoms and are cytotoxic due to the liberated H_2O_2 , which also induces or inhibits platelet aggregation⁹³. It was reported that 443444solble guanylate cyclase (sGC) is activated by H_2O_2 in the presence of superoxide 445dismutase (SOD), and that H₂O₂ activates nitric oxide synthase (NOS). Thus, LAAOs 446show the synergic effects on hypertension with other toxins such as BPPs via H₂O₂ production⁷³. 447

The *P. flavoviridis* genome contains three genes for L-amino acid oxidase, $LAAO_01$, $LAAO_02$, and $LAAO_03$ (Gene ID: habu1_s402940_g24950, g24949, and g24947, respectively), all of them being located in the same scaffold (Supplementary Fig. S2c). The results indicate that these LAAO genes diversified by gene duplication. $LAAO_01$ encodes the venom type L-amino acid oxidase composed of seven exons. Molecular phylogeny indicates that $LAAO_01$ encodes the venom enzyme, but $LAAO_02$, and $LAAO_03$ encode non-venom type (Supplementary Fig. S2c).

455

456 (o) Phosphodiesterases (PDE)

457 Phosphodiesterases (PDEs: E.C. 3.1.4.1) catalyze the hydrolysis of phosphodiester 458 bonds of cyclic adenosine monophosphate (cAMP) and cyclic guanosine 459monophosphate (cGMP). PDEs have been isolated and characterized from numerous 460 snake venoms. They are generally high molecular mass proteins (> 90 kDa) with single 461 polypeptide Р. flavoviridis venom **PDEs** chains. are reported to be metalloglycoproteins⁹⁴. More recently, Trummal et al. reported the structure and 462 characterization of PDE from Vipera lebetina venom⁹⁵. It inhibits ADP- and 463 464collagen-induced platelet aggregation in a dose-dependent manner. Mamillapalli et al 465reported that the activity of svPDE is enhanced by lysophospholipids, which are liberated by PLA2 during venomous action⁹⁶. Thus, svPDE and svPLA2 show the 466 467synegistic effect.

468In this study, we identified two PDE genes, PDE01 (Gene ID: habu1 s149 g00804) 469 and PDE02 (habu1 s149 g008059), which are tandemly located on 470habul scaffold 149 (Supplementary Fig. S19a). Molecular phylogeny shows that Pr. 471*flavoviridis PDE01* is the venom enzyme and PDE02 is the tissue form (Supplementary 472Fig. S19b). PDE01 and PDE-02 include 889 and 904 amino acids, respectively, and 473 both conserve catalytic amino acid residues. Although both genes are encoded by 25 474exons with the same exon-intron junctions, they showed different expression profiles. 475PDE01 is expressed only in the venom gland. The cAMP-specific 0', 5'-cyclic phosphodiesterase gene was also identified on the same scaffold (Gene ID: 476 habul s149 g00834) (Supplementary Fig. S19a). However, it showed no sequence 477478homology with PDE01 and PDE02.

479

480 (p) Phospholipases B (PLB)

481 Phospholipases B (PLB: EC 3.1.1.5) are enzymes with combined PLA₁ and PLA₂ 482 activities that cleaves acyl chains from both *sn*-1 and *sn*-2 positions of phospholipids. 483 PLB also acts on lysolecithin, which is formed by PLA₂, then PLBs are known as 484 lysophospholipases. PLBs are present in most snake venoms, but generally at very 485 low levels⁹⁷.

486In this study, we identified five PLB genes, PLB01 (Gene ID: habu1 s1233 g04284), 487PLB02 (habu1 s1964 g07200), PLB03, (habu1 s313 g01520), PLB04 488 (habu1 s3352 g11424), and PLB05 (habu1 s303 g01427) in the P. flavoviridis 489 genome, (Supplementary Fig. S20a). PLB01 showed high sequence identity (99-85%) 490with phospholipases B isolated from other snakes. On the other hand, PLB02 is 491 phospholipase B-like 2 and PLB03 is a membrane-associated PLB1. PLB05 and 492 PLB06 are 60 kDa lysophospholipase and lysophospholipase-like protein, respectively. 493PLB01 and PLB02 showed low but distinct sequence similarity (30.2%) each other, 494 while PLB03, PLB04 and PLB05 showed no similarity. From transcriptome data, 495PLB01 showed venom-specific expression, while PLB02, PLB04 and PLB05 are 496 ubiquitous housekeeping genes, and PLB03 showed small intestine-specific expression. 497Molecular phylogeny supports the evolutionary history of SV and other NV genes 498 (Supplementary Fig. S20b).

499

500 (q) Bradykinin-potentiating peptide (BPP) and C-type natriuretic peptide (CNP)

501 Bradykinin-potentiating peptides (BPPs) are known as painful stimulation factors502 following snakebites through the inhibition of enzymatic digestion of bradykinin. BPPs

are also well known as inhibitors of angiotensin-converting enzyme (ACE), and were the basis for development of antihypertensive drugs such as captopril and enalapril⁹⁸. BPPs activate argininosuccinate synthetase, resulting in increased nitric oxide production, which reduces arterial blood pressure⁹⁹. Typically, BPPs contain 5 to 13 amino acid residues with a pyroglutamyl residue (pGlu) at the N-terminus and a proline residue at the C-terminus a high proline content and/or the tripeptide sequence Ile–Pro– Pro¹⁰⁰.

510Natriuretic peptide (NP) plays a fundamental role in cardiovascular homeostasis by 511modulating fluid and electrolyte balance and vascular tone, and exhibit hypotensive and 512vasodepressor activity by activating natriuretic receptors. Previously, we have cloned 513cDNAs (NCBI AB749764.1) encoding both BPPs and C-type naturetic peptide (CNP) from P. flavoviridis and other snakes^{101, 102}, suggesting that CNP coded on the same 514515gene as BPP. Recently, Aird et al. reported the transcriptomes and proteomes of P. 516flavoviridis and Ovophis okinavensis venom, showed only two BPP-related peptides, QSKPGRSPPISP, which corresponds to BPP1 3, and QGRPRSEVPP¹⁰¹. 517

In the *P. flavoviridis* genome, we identified two genes. *CNP01* encodes a BPP-CNP precursor protein, which was located at two scaffolds, habu1_scaffold_258676: 1...337 and habu1_scaffold_4348: 15402 ... 16141 (Gene model ID: habu1_s258676_g24318 and habu1_s4348_g14020, respectively) (Supplementary Fig. S21a). *CNP02* encodes B-type natriuretic peptide (BNP) (habu1_s20540_g224311) (Supplementary Fig. S21a). The former is venom peptides, while the latter is of another tissue origin. Molecular phylogeny supports the evolutionary history of SV and other NV genes (Supplementary 525 Fig. S21b).

526

527 (r) Glutaminyl peptide cyclotransferases (GPCase)

528Glutaminyl peptide cyclases (GPCase, QC; glutaminyl cyclotransferase, EC 2.3.2.5) 529 catalyze N-terminal pyroglutamate (pGlu) formation on proteins and peptides. This 530N-terminal modification confers resistance to aminopeptidase degradation. Among 531various snake venom components, glutaminyl cyclase (svQC) is one of the least 532understood protein family. Wang et al. reported the presence of vQC activity in a wide 533spectrum of venom species and their structures and characterization including cDNA sequences from seven species¹⁰³. svQC present in each venom with the low content 534535and only a single form, suggesting its house-keeping role for posttranslational 536 modification of the venom proteins. For example, svQC cyclizes the N-terminal Gln residue of snake venom bioactive peptides and proteins including BPPs^{100,104} and 537crotoxin subunits¹⁰⁵, resulting the stability and prolongation of peptides in prey plasma 538539 by protecting against the degradation.

We identified two genes, svGPCase01 (habu1_s510_g02384) and nvGPCase02(habu1_s3067_g10859), encoding glutaminyl peptide cyclases in *P. flavoviridis* genome (Supplementary Fig. S22a). Both svGPCase01 and nvGPCase02 were composed of seven exons with the conserved exon-intron junctions. Recently, Aird et al. reported four QC transcripts for two pairs of toxins (AB848133, AB848134, AB851933, AB851934)¹⁰⁶, the correspondence of those to svGPCase01 and nvGPCase02 remains to be elucidated. Molecular phylogeny shows that svGPCase01 forms a clade with other 547 SV members while *nvGPCase02* is grouped with other NV members (Supplementary 548 Fig. S22b). This indicates that the product of *svGPCase01* is sure to act as toxin or toxin 549 stabilizing factor in *P. flavoviridis* venom although QC activity has not yet been 550 demonstrated in the venom.

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552 References

- Fox, J. W, & Serrano, S. M. T. Structural considerations of the snake venom
 metalloproteinases, key members of the M12 reprolysin family of
 metalloproteinases. Toxicon 2005; 45, 969–985.
- 556 2. Worlfsberg, T. G. et al. ADAM, a widely distributed and developmentally 557 regulated gene family encoding membrane proteins with a disintegrin and 558 metalloprotease domain. Devlop Biol. 1995; 169, 378-383.
- 559 3. Edwards, D. R., Handsley, M. M., Pennington, C. J. The ADAM
 560 metalloproteinases. Mol. Asp. Med. 2009; 29, 258–289.
- 4. Reiss K, Saftig P. The "A Disintegrin and Metalloprotease" (ADAM) family of
 sheddases: Physiological and cellular functions. Semin Cell Dev Biol. 2009; 20:
 126–37.
- 564 5. Takeda S. ADAM and ADAMTS family proteins and snake venom 565 metalloproteinases: A structural overview. Toxins. 2016; 8: 155.
- 566 6. Seeger WH, Ouyang C. Snake venoms and blood coagulation, In: Lee CY editor.

567 Snake venoms. New York: Springer; 1979. p. 684-750.

568	7.	Shieh TC, Kawabata S, Kihara H, Ohno M, Iwanaga S. Purification and
569		characterization of a coagulant enzyme from Trimeresurus flavoviridis venom. J
570		Biochem. 1985; 98: 713–61.

- 571 8. Deshimaru M, Ogawa T, Nakashima K, Nobuhisa I, Chijiwa T, Shimohigashi Y,
- 572 et al. Accelerated evolution of crotalinae snake venom gland serine proteases.
 573 FEBS Lett. 1996; 397: 83-8.
- 574 9. Sunagawa M, Nakamura M, Kosugi T. Cloning of habutobin cDNA and
 575 antithrombotic activity of recombinant protein. Biochem Biophys Res Commun.
 576 2007; 362: 899-904.
- 577 10. Yamamoto C, Tsuru D, Oda-Ueda N, Ohno M, Hattori S, Kim ST. Flavoxobin, a
 578 serine protease from *Trimeresurus flavoviridis* (habu snake) venom,
 579 independently cleaves Arg726-Ser727 of human C3 and acts as a novel,
 580 heterologous C3 convertase. Immunology. 2002; 107: 111-7.
- 581 11. Komori Y, Tatematsu R, Tanida S, Nikai T. Thrombin-like enzyme, flavovilase,
 582 with kinin-releasing activity from *Trimeresurus flavoviridis* (habu) venom. J Nat
 583 Toxins. 2001; 10: 239-48.
- Tatematsu R, Komori Y, Nikai T. A new thrombin-like enzyme, flavoviridiobin
 from the venom of *Trimeresurus flavoviridis* (habu). J Nat Toxins. 2000; 9:
 327-39.
- 587 13. Serrano SM, Hagiwara Y, Murayama N, Higuchi S, Mentele R, Sampaio CA, et
 al. Purification and characterization of a kinin-releasing and fibrinogen-clotting
 serine proteinase (KN-BJ) from the venom of *Bothrops jararaca*, and molecular

cloning and sequence analysis of its cDNA. Eur J Biochem. 1998; 251: 845-53.

- 591 14. van Leeuwen BH, Evans BA, Tregear G, Richards RI. Mouse glandular kallikrein
- 592 gene identification, structure, and expression of the renal kallikrein gene. J Biol593 Chem. 1986; 261: 5529-35.
- 594 15. Drickamer K. Evolution of Ca(2+)-dependent animal lectins. Prog Nucl Acid Res
 595 Mol Biol. 1993; 45: 207–32.
- 596 16. Drickamer K, Fadden AJ. Genomic analysis of C-type lectins. Biochem Soc
 597 Symp. 2002; 69: 59–72.
- 598 17. Zelensky AN, Gready JE. The C-type lectin-like domain superfamily. FEBS J.
 599 2005; 272: 6179–217.
- Flexner S, Noguchi H. The constitution of snake venom and snake sera. J Pathol.
 1903; 8: 379-410.
- 602 19. Gartner TK, Stocker K, Williams DC. Thrombolectin: a lectin isolated from
 603 *Bothrops atrox* venom. FEBS Lett. 1980; 117: 13–6.
- 604 20. Sartim MA, Sampaio SV. Snake venom galactoside-binding lectins: a structural
 605 and functional overview. J Venom Anim Toxins Incl Trop Dis. 2015; 21: 35.
- 607 evolution of C-type lectin-like proteins from snake venom. Toxicon. 2005; 45:608 1-14.

Ogawa T, Chijiwa T, Oda-Ueda N, Ohno M. Molecular diversity and accelerated

606

21.

609 22. Morita T. Structures and functions of snake venom CLPs (C-type lectin-like
610 proteins) with anticoagulant-, procoagulant-, and platelet-modulating activities.
611 Toxicon 2005; 45: 1099-114.

- 612 23. Dijkstra <u>BW</u>, <u>Drenth J</u>, <u>Kalk KH</u>. Active site and catalytic mechanism of
 613 phospholipase A2. Nature 1981; 289: 604-6.
- 614 24. Dufton MJ, Hider RC. Classification of phospholipases A₂ according to sequence.
- Evolutionary and pharmacological implications. Eur J Biochem. 1983; 137:545-51.
- 617 25. Maraganore JM, Merutka G, Cho W, Welches W, Kézdy FJ, Heinrikson RL. A
 618 new class of phospholipases A2 with lysine in place of aspartate 49. Functional
 619 consequences for calcium and substrate binding. J Biol Chem. 1984; 259:
 620 13839-43.
- 621 26. Renetseder R, Brunie S, Dijkstra BW, Drenth J, Sigler PB. A comparison of the
 622 crystal structures of phospholipase A₂ from bovine pancreas and *Crotalus atrox*623 venom. J Biol Chem. 1985; 260: 11627-34.
- 624 27. Holland DR, Clancy LL, Muchmore SW, Ryde TJ, Einspahr HM, Finzel BC, et
 625 al. The crystal structure of a lysine 49 phospholipase A2 from the venom of the
 626 cottonmouth snake at 2.0-Å resolution. J Biol Chem. 1990; 265: 17649-56.
- 627 28. Suzuki A, Matsueda E, Yamane T, Ashida T, Kihara H, Ohno M. Crystal
 628 structure analysis of phospholipase A2 from *Trimeresurus flavoviridis* (Habu
 629 snake) venom at 1.5 Å resolution. J Biochem. 1995; 117: 730-40.
- 630 29. Ikeda N, Chijiwa T, Matsubara K, Oda-Ueda N, Hattori S. <u>Matsuda Y, Ohno M.</u>
 631 Unique structural characteristics and evolution of a cluster of venom
 632 phospholipase A₂ isozyme genes of *Protobothrops flavoviridis* snake. Gene. 2010;
 633 461: 15-25.

- 634 30. Nakashima K, Ogawa T, Oda N, Hattori M, Sakaki Y, Kihara H, Ohno M.
- 635 Accelerated evolution of *Trimeresurus flavoviridis* venom gland phospholipase
- 636 A₂ isozymes. Proc Natl Acad Sci U S A. 1993; 90: 5964-8.
- 637 31. Nakashima, K. et al. Accelerated evolution in the protein-coding regions is
- 638 universal in crotalinae snake venom gland phospholipase A₂ isozyme genes. *Proc.*639 *Natl. Acad. Sci. USA.* 92, 5605-5609 (1995)
- 640 32. Ohno, M. et al. Molecular evolution of snake toxins: Is the functional diversity of
- snake toxins associated with a mechanism of accelerated evolution? *Prog. Nucleic Acid Res. Mol. Biol.* 59, 307-364 (1998).
- 643 33. Ogawa, T. et al. Unusually high conservation of untranslated sequences in cDNAs
- 644 for *Trimeresurus flavoviridis* phospholipase-A2 isozymes. *Proc. Natl. Acad. Sci.*645 USA. 89, 8557-8561 (1992).
- 646 34. Ogawa, T., Kitajima, M., Nakashima, K., Sakaki, Y. & Ohno, M. Molecular
 647 evolution of group II phospholipases A2. J. Mol. Evol. 41, 867-877 (1995).
- 648 35. Nair DG, Fry BG, Alewood P, Kumar PP, Kini RM. Antimicrobial activity of649 omwaprin, a new member of the waprin family of snake venom proteins.
- 650 Biochem J. 2007; 402: 93-104.
- 651 36. Fry, B. G. et al. Evolution of an arsenal: structural and functional diversification
- of the venom system in the advanced snakes (Caenophidia). *Mol. Cell Proteomics.* 7, 215-46 (2008).
- 654 37. St Pierre Let al. Common evolution of waprin and kunitz-like toxin families in
 655 Australian venomous snakes. Cell Mol Life Sci. 2008; 65: 4039-54.

Bahari S, Mackessy SP, Kini MR. The venom gland transcriptome of the Desert
Massasauga Rattlesnake (Sistrurus catenatus edwardsii): towards an
understanding of venom composition among advanced snakes (Superfamily
Colubroidea). BMC Mol Biol. 2007; 8: 115.

- Boley R, Pahari S, Reza M. A., Mackessy SP, Kini RM. The gene structure and
 evolution of ku-wap-fusin (Kunitz Waprin Fusion Protein), a novel evolutionary
 intermediate of the Kunitz serine protease inhibitors and Waprins from *Sistrurus catenatus* (Massasauga Rattlesnake) venom glands. The Open Evolution J. 2010;
 4: 31-41.
- 40. Mourao CBF, Schwartz EF. Protease inhibitors from marine venomous animals
 and their counterparts in terrestrial venomous animals. Mar Drugs. 2013; 11:
 2069-112.
- Bohlen CJ, Chesler AT, Sharif-Naeini R, Medzihradszky KF, Zhou S, King D, et
 al. A heteromeric Texas coral snake toxin targets acid-sensing ion channels to
 produce pain. Nature. 2011; 479: 410–4.
- 42. Utkin YN. Three-finger toxins, a deadly weapon of elapid venom--milestones ofdiscovery. Toxicon. 2013; 62: 50-5.
- 673 43. Sunagar K, Jackson TN, Undheim EA, Ali SA, Antunes A, Fry BG.
- 674 Three-fingered RAVERs: Rapid accumulation of variations in exposed residues of
 675 snake venom toxins. Toxins. 2013; 5: 2172-208.
- 676 44. Ménez A. Functional architectures of animal toxins: a clue to drug design?
 677 Toxicon. 1998; 36: 1557–72.

678 45. Tsetlin V. Snake venom α-neurotoxins and other 'three-finger' proteins. Eur J
679 Biochem. 1999; 264: 281–6.

- 46. Junqueira-de-Azevedo IL, Ching AT, Carvalho E, Faria F, Nishiyama MY Jr, Ho
 PL, Diniz MR. *Lachesis muta* (Viperidae) cDNAs reveal diverging pit viper
 molecules and scaffolds typical of cobra (Elapidae) venoms: implications for
 snake toxin repertoire evolution. Genetics. 2006; 173: 877-89.
- 684 47. Ogawa Y, Murayama N, Fujita Y, Yanoshita R. Characterization and cDNA
 685 cloning of aminopeptidase A from the venom of *Gloydius blomhoffi brevicaudus*.
 686 Toxicon. 2007; 49: 1172–81.
- Burnett LA, Washburn CA, Sugiyama H, Xiang X, Olson JH, Al-Anzi B, Bieber
 AL, Chandler DE. Allurin, an amphibian sperm chemoattractant having
 implications for mammalian sperm physiology. Int Rev Cell Mol Biol. 2012; 295:
 1–61.
- 49. Yamazaki Y, Brown RL, Morita T. Purification and cloning of toxins from elapid
 venoms that target cyclic nucleotide-gated ion channels. Biochemistry. 2002a; 41:
 11331–7.
- 50. Yamazaki Y, Koike H, Sugiyama Y, Motoyoshi K, Wada T, Hishinuma S, et al.
 Cloning and characterization of novel snake venom proteins that block smooth
 muscle contraction. Eur J Biochem. 2002b; 269: 2708–15.
- 51. Yamazaki Y, Hyodo F, Morita T. Wide distribution of cysteine-rich secretory
 proteins in snake venoms: isolation and cloning of novel snake venom
 cysteine-rich secretory proteins. Arch Biochem Biophys. 2003; 412: 133–41.

- 700 52. Yamazaki Y, Morita T. Structure and function of snake venom cysteine-rich
 701 secretory proteins. Toxicon. 2004; 44: 227-31.
- 53. Mochca-Morales J, Martin BM, Possani LD. Isolation and characterization of
 helothermine, a novel toxin from *Heloderma horridum horridum* (Mexican
 beaded lizard) venom. Toxicon. 1990; 28: 299–309.
- 54. Morrissette J, Kratzschmar J, Haendler B, El-Hayek R, Mochca-Morales J, Martin
 BM, et al. Primary structure and properties of helothermine, a peptide toxin that
 blocks ryanodine receptors. Biophys J. 1995; 68: 2280–8.
- 55. Nobile M, Noceti F, Prestipino G, Possani LD. Helothermine, a lizard venom
 toxin, inhibits calcium current in cerebellar granules. Exp Brain Res. 1996; 110:
 15–20.
- 56. Cotton J, Crest M, Bouet F, Alessandri N, Gola M, Forest E, et al. A
 potassium-channel toxin from the sea anemone *Bunodosoma granulifer*a, an
 inhibitor for Kv1 channels. Revision of the amino acid sequence, disulfide-bridge
 assignment, chemical synthesis, and biological activity. Eur J Biochem. 1997;
 244: 192–202.
- 57. Dauplais M, Lecoq A, Song J, Cotton J, Jamin N, Gilquin B, et al. On the
 Convergent Evolution of Animal Toxins Conservation of a diad of functional
 residues in potassium channel-blocking toxins with unrelated structures. J Biol
 Chem. 1997; 272: 4302–9.

- 58. Castañeda O, Sotolongo V, Amor AM, Stöcklin R, Anderson AJ, Harvey AL, et
 al. Characterization of a potassium channel toxin from the Caribbean Sea
 anemone *Stichodactyla helianthus*. Toxicon 1995; 33: 603–13.
- 59. Tudor JE, Pallaghy PK, Pennington MW, Norton RS. Solution structure of ShK
 toxin, a novel potassium channel inhibitor from a sea anemone. Nat Struct Biol.
 1996; 3: 317–20.
- Wang F, Li H, Liu M, Song H, Han H, Wang Q, et al. Structural and functional
 analysis of natrin, a venom protein that targets various ion channels. Biochem
 Biophys Res Commun. 2006; 315: 443–8.
- 729 61. Adade CM, Carvalho ALO, Tomaz MA, Costa TFR, Godinho JL, Melo PA, et al.
- Crovirin. A snake venom cysteine-rich secretory protein (CRISP) with promising
 activity against tryptozoma. PLoS Negl Trop Dis. 2014; 8: e3252.
- 732 62. Ponting C, Schultz J, Bork P. SPRY domains in ryanodine receptors (Ca²⁺-release
 733 channels). Trends Biochem Sci. 1997; 22: 193–4.
- 63. Woo JS, Imm JH, Min CK, Kim KJ, Cha SS, Oh BH. Structural and functional
 insights into the B30.2/SPRY domain. EMBO J. 2006; 25: 1353-63.
- 64. Pung YF, Wong PTH, Kumar PP, Hodgson WC, Kini RM. Ohanin, a novel
 protein from king cobra venom, induces hypolocomotion and hyperalgesia in
 mice. J Biol Chem. 2005; 280: 13137–47.
- Pung YF, Kumar SV, Rajagopalan N, Fry BG, Kumar PP, Kini RM. Ohanin, a
 novel protein from king cobra venom: its cDNA and genomic organization. Gene
 2006; 371: 246-56.

- Aird SD. Taxonomic distribution and quantitative analysis of free purine and
 pyrimidine nucleosides in snake venoms. Comp Biochem Physiol B Biochem Mol
 Biol. 2005; 140: 109–26.
- 745 67. Dhananjaya BL, D'Souza CJM. The pharmacological role of nucleotidases in
 746 snake venoms. Cell Biochem Funct 2010; 28: 171–7.
- 68. Gorrell MD, Gysbers V, McCaughan GW. CD26: a multifunctional integral
 membrane and secreted protein of activated lymphocytes. Scand J Immunol.
 2001; 54: 249–64.
- 750Maes M, Scharpé S, Demeester I, Goossens P, Wauters A, Neels H, et al. 69. 751Components biological variation in endopeptidase of prolyl and 752dipeptidyl-peptidase-iv activity in plasma of healthy subjects. Clin Chem. 1994; 40: 1686-91. 753
- 754 70. Ohkubo I, Huang K, Ochiai Y, Takagaki M, Kani K. Dipeptidyl peptidase IV
 755 from porcine seminal plasma: purification, characterization, and N-terminal amino
 756 acid sequence. J Biochem. 1994; 116: 1182-6.
- 757 71. Mollay C, Vilas U, Hutticher A, Kreil G. Isolation of a dipeptidyl aminopeptidase,
 758 a putative processing enzyme, from skin secretion of *Xenopus laevis*. Eur J
 759 Biochem. 1986; 160: 31–5.
- 760 72. Ogawa Y, Mamura Y, Murayama N, Yanoshita R. Characterization and cDNA
 761 cloning of dipeptidyl peptidase IV from the venom of *Gloydius blomhoffi*762 *brevicaudus*. Comp Biochem Physiol B Biochem Mol Biol. 2006; 145:
 763 35-42.
- 764 73. Aird SD. Ophidian envenomation strategies and the role of purines. Toxicon
 765 2002; 40: 335-393.
- 766 74. Stern R, Jedrzejas MJ. Hyaluronidases: their genomics, structures, and
 767 mechanisms of action. Chem Rev. 2006; 106: 818–39.
- 768 75. Toole BP. Hyaluronan: from extracellular glue to pericellular cue. Nat Rev769 Cancer. 2004; 4: 528-39.
- 770 76. Kemparaju K, Girish KS. Snake venom hyaluronidase: a therapeutic target. Cell
 771 Biochem Function. 2006; 24: 7–12.
- 772 77. Harrison RA, Ibison F, Wilbraham D, Wagstaff SC. Identification of cDNAs
 773 encoding viper venom hyaluronidases: cross-generic sequence conservation of
 774 full-length and unusually short variant transcripts. Gene 2007; 392: 22-33.
- 775 78. Castanheira LE, Rodrigues RS, Boldrini-França J, Fonseca FPP, Henrique-Silva
 776 F, Homsi-Brandeburgo MI, Rodrigues VM. Molecular cloning of a hyaluronidase
 777 from *Bothrops pauloensis* venom gland. J Venom Anim Toxins Incl Trop Dis.
 778 2014; 20: 25.
- 779 79. Hohn A, Leibrock J, Bailey K, Barde Y-A. Identification and characterization of a
 780 novel member of the nerve growth factor/brain-derived neurotrophic factor
 781 family. Nature. 1990; 344: 339-41.
- 80. Maisonpierre PC, Belluscio L, Squinto S, Ip NY, Furth ME, Lindsay RM,
 Yancopoulos GD. Neurotrophin-3: a neurotrophic factor related to NGF and
 BDNF. Science. 1990; 247: 1446-51.
- 785 81. Ibanez CF, Ernfors P, Timmusk T, Ip NY, Arenas E, Yancopoulos GD, Persson

- H. Neurotrophin-4 is a target-derived neurotrophic factor for neurons of the
 trigeminal ganglion. Development 1993; 117: 1345-53.
- Wong V, Arriaga R, Ip NY, Lindsay RM. The neurotrophins BDNF, NT-3 and
 NT-4/5, but not NGF, up-regulate the cholinergic phenotype of developing motor
 neurons, Eur J Neurosci, 1993; 5: 466-74.
- 791 83. Kostiza T, Meier J. Nerve growth factors from snake venoms: Chemical
 792 properties, mode of action and biological significance. Toxicon 1996; 34: 787–
 793 806.
- 794 84. Trummal K, Tõnismägi K, Paalme V, Järvekülg L, Siigur J, Siigur E. Molecular
 795 diversity of snake venom nerve growth factors. Toxicon 2011; 58: 363–8.
- Hayashi K, Inoue S, Ikeda K. Purification and characterization of nerve growth
 factors (NGFs) from the snake venoms. In: Singh BR and Tu AT editors. Natural
 Toxins 2: Structure, Mechanism of Action, and Detection. 1996; 391: p. 403-16.
- 799 86. Sunagar K, Fry BG, Jackson TNW, Casewell NR, Undheim EAB, Vidal N, Ali
- SA, King GF, Vasudevan K, Vasconcelos V, Antunes A. Molecular Evolution of
 Vertebrate Neurotrophins: Co-Option of the Highly Conserved Nerve Growth
 Factor Gene into the Advanced Snake Venom Arsenalf. Plos ONE 2013; 8:
 e81827
- 804 87. Hargreaves AD, Swain MT, Hegarty MJ, Logan DW, Mulley JF. Restriction and
 805 Recruitment—Gene Duplication and the Origin and Evolution of Snake Venom
 806 Toxins. Genome Biol. Evol. 2014; 6: 2088–2095. doi:10.1093/gbe/evu166

- 807 88. Bertaux O, Toselli-Mollereau E, Auffray C, Devignes M-D. Alternative usage of
 5' exons in the chicken nerve growth factor gene: refined characterization of a
 809 weakly expressed gene. Gene 2004; 334: 83-97.
- 810 89. Ferrara N, Gerber HP, LeCouter J. The biology of VEGF and its receptors. Nat
 811 Med. 2003; 9: 669-76.
- 90. Yamazaki Y, Takani K, Atoda H, Morita T. Snake venom vascular endothelial
 growth factors (VEGFs) exhibit potent activity through their specific recognition
 of KDR (VEGF receptor 2). J Biol Chem. 2003; 278: 51985-8.
- 815 91. Takahashi H, Hattori S, Iwamatsu A, Takizawa H, Shibuya M. A novel snake
 816 venom vascular endothelial growth factor (VEGF) predominantly induces
 817 vascular permeability through preferential signaling via VEGF receptor-1. J Biol
 818 Chem. 2005; 279: 46304-14.
- 92. Yamazaki Y, Matsunaga Y, Tokunaga Y, Obayashi S, Saito M, Morita T. Snake
 venom vascular endothelial growth factors (VEGF-Fs) exclusively vary their
 structures and functions among species. *J Biol Chem.* 2009; 284: 9885–91.
- 822 93. Guo C, Liu S, Yao Y, Zhang Q, Sun MZ. Past decade study of snake venom
 823 L-amino acid oxidase. Toxicon 2012; 60: 302-11.
- 824 94. Kini RM, Gowda TV. Rapid method for separation and purification of four
 825 isoenzymes of phosphodiesterase from *Trimeresurus flavoviridis* (Habu snake)
 826 venom. J Chromatogr. 1984; 291: 299-305.

- 827 95. Trummal K, Aaspõllu A, Tõnismägi K, Samel M, Subbi J, Siigur J, Siigur E
 828 Phosphodiesterase from *Vipera lebetina* venom structure and characterization.
 829 Biochimie. 2014; 106, 48-55.
- 830 96. Mamillapalli R, Haimovitz R, Ohad M, Shinitzky M. Enhancement and inhibition
- of snake venom phosphodiesterase activity by lysophospholipids. FEBS Letters
 1998; 436: 256-258.
- 833 97. Doery HM, Pearson JE. Phospholipase B in snake venoms and bee venom.
 834 Biochem J. 1964; 92: 599-602.
- 835 98. Smith CG, Vane JR. The discovery of captopril. FASEB J. 2003; 17: 788–9.
- 836 99. Camargo AC, Ianzer D, Guerreiro JR, Serrano SM. Bradykinin-potentiating
 837 peptides: beyond captopril. Toxicon 2012; 59: 516-23.
- 838 100. Cotton J, Hayashi MA, Cuniasse P, Vazeux G, Ianzer D, De Camargo AC, Dive
- 839 V. Selective inhibition of the C-domain of angiotensin I converting enzyme by
 840 bradykinin potentiating peptides. Biochemistry 2002; 41: 6065–71.
- 841 101. Higuchi S, Murayama N, Saguchi K, Ohi H, Fujita Y, Camargo AC, et al.
 842 Bradykinin-potentiating peptides and C-type natriuretic peptides from snake
 843 venom. Immunopharmacol. 1999; 44: 129-35.
- 844 102. Murayama N, Hayashi MA, Ohi H, Ferreira LA, Fernandes BL, Yamane T,
- Camargo ACM. Cloning and sequence analysis of a *Bothrops jararaca* cDNA
 encoding a precursor of seven bradykinin-potentiating peptides and a C-type
 natriuretic peptide. Proc Natl Acad Sci U S A. 1997; 94: 1189–93.
- 848 103. Wang YM, Huang KF, Tsai IH. Snake venom glutaminyl cyclases: Purification,

- 849 cloning, kinetic study, recombinant expression, and comparison with the human850 enzyme. Toxicon 2014; 86: 40-50.
- 104. Calvete JJ, Fasoli E, Sanz L, Boschetti E, Righetti PG. Exploring the venom
 proteome of the western diamondback rattlesnake, *Crotalus atrox*, via snake
 venomics and combinatorial peptide ligand library approaches. J Proteome Res
 2009; 8: 3055–3067.
- 855 105. Aird SD, Yates JR III, Martino PA, Shabanowitz J, Hunt DF, Kaiser II: The
 856 amino acid sequence of the acidic subunit B-chain of crotoxin. Biochim Biophys
 857 Acta 1990; 1040: 217–224.
- 858 106. Aird SD, Watanabe Y, Villar-Briones A, Roy MC, Terada K, Mikheyev AS.
 859 Quantitative high-throughput profiling of snake venom gland transcriptomes and
 860 proteomes (*Ovophis okinavensis* and *Protobothrops flavoviridis*). BMC Genomics.
 861 2013; 14: 790.

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864	Supplementary Table S1. Summary of Roche 454 reads and Miseq and Hiseq
865	reads of the Protobothrops flavoviridis genome.
866	
867	Supplementary Table S2. Summary statistics of the Habu genome assembly,
868	HabAm1.
869	
870	Supplementary Table S3. Comparison of genome sequence assemblies among four
871	snakes.
872	
873	Supplementary Table S4. RNA-seq analysis of tissues and organs of Protobothrops
874	flavoviridis.
875	
876	Supplementary Table S5. Keywords used to identify venom-related proteins.
877	
878	Supplementary Table S6. Best-fit model for estimating the phylogenetic trees.
879	
880	Supplementary Table S7. Expression profiles of neutrophin family genes in
881	Protobothrops flavoviridis.

883 Supplementary Figures

884 Supplementary Figure S1. Genome size estimation based on flow cytometry and 885*k*-mer analysis. a, Flow cytometry analysis of *Gallus gallus* (blue), *Protobothrops* 886 flavoviridis (green), and Eublepharis macularius (red). The vertical axis indicates the 887 number of nuclei counted. The horizontal axis indicates fluorescence intensity using an 888 FL2-H filter. Simple linear regression using the two known genome sizes gave a rough 889 estimate of the *P. flavoviridis* genome size at 1.8 Gb. b, k-mer distribution of whole 890 genome shotgun sequencing reads of *P. flavoviridis* (k = 27). The vertical axis indicates 891 the number of k-mers. The horizontal axis indicates k-mer coverage. Total number of 892 k-mers (k = 27) was 65,661,771,084. Peak coverage was 43x. The left peak at low 893 frequency (< 10x depth) likely results from mismatches due to heterozygous SNPs. The 894 genome size was estimated to be 1.41 Gb (total length of used reads / peak coverage). 895 896 Supplementary Figure S2. Summary and molecular phylogeny of SV and NV 897 genes of the hyaluronidase (Hyal) family (a), the Nerve growth factor (NGF) 898 family (b), and the L-amino acid oxidase (LAAO) family (c). Multiple sequence 899 alignments were performed using Clustalw. Phylogenetic trees were constructed with

900 the maximum likelihood method using IQ-TREE (<u>http://www.iqtree.org</u>) based on 901 aligned amino acid sequences. Numbers on branches are bootstrap values with 1000x 902 resampling.

903 Ac: Anolis carolinensis, Acc: Agkistrodon contortrix contortrix, Af: Azemiops feae,
904 Am: Alligator mississippiensis, Ap: Agkistrodon piscivorus, Ar: Aspidites ramsayi, As:

905 Alligator sinensis, Ba: Bitis arietans, Bb: Brachymeles bonitae, Bcc: Boa constrictor 906 constrictor, Bco: Boa constrictor occidentalis, Ben: Boa constrictor nebulosa, Bem: 907 Bellatorias major, Bf: Bungarus fasciatus, Bia: Bitis arietans, Bj: Bothrops jararacussu, 908 Bm: Bungarus multicinctus, Boa: Bothrops asper, Bp: Bothrops pauloensis, Bt: 909 Brachymeles tiboliorum, Ca: Crotalus adamanteus, Car: Calabaria reinhardtii, Cc: 910 Cerastes cerastes, Cdc: Candoia superciliosa crombiei, Cdt: Crotalus durissus 911 terrificus, Ce: Chilabothrus exsul, Cg: Chilabothrus granti, Ch: Crotalus horridus, Cha: 912 Chilabothrus angulifer, Chm: Chelonia mydas, Chss: Chilabothrus strigilatus 913 strigilatus, Cm: Chelonia mydas, Cn: Cryptophis nigrescens, Coa: Corallus annulatus, 914Coc: Corallus cookii, Coh: Corallus hortulanus, Cor: Corallus ruschenbergerii, Cr: 915 Calloselasma rhodostoma, Css: Candoia superciliosa superciliosa, Ct: Cophosaurus 916 texanus, Cu: Charina umbratica, Cym: Cyclophiops major, Dc: Drysdalia coronoides, 917 Dd: Dipsosaurus dorsalis, Dip: Ditypophis sp. AC-2011, Dr: Daboia russelii, Dv: 918 Demansia vestigiata, Ea: Epicrates alvarezi, Ec: Epicrates cenchria, Em: Eryx muelleri, 919 En: Eunectes notaeus, Eo: Echis ocellatus, Epl: Echis pyramidum leakeyi, Et: Eryx 920 tataricus, Fp: Feylinia polylepis, Gb: Gloydius blomhoffii, Gg: Gallus gallus, Gh: 921 Gloydius halys, Gi: Gloydius intermedius, Gj: Gekko japonicus, Gw: Gambelia 922 wislizenii, Hs: Hoplocephalus stephensii, Lc: Lepidophyma cuicateca, Lm: Lachesis 923 muta, Lv: Lacerta viridis, Mg: Meleagris gallopavo, Ml: Macrovipera lebetina, Nk: 924 Naja kaouthia, Ns: Naja sputatrix, Nss: Notechis scutatus scutatus, Oh: Ophiophagus 925hannah, Om: Oxyuranus microlepidotus, Oo: Ovophis okinavensis, Oss: Oxyuranus 926 scutellatus scutellatus, Pa: Pseudechis australis, Pb: Python bivittatus, Pc: Phrynosoma

927 coronatum, Pe: Protobothrops elegans, Pf: Protobothrops flavoviridis, Ph: Phrynosoma 928 hernandesi, Phm: Phalotris mertensi, Phrm: Phrynosoma mcallii, Pm: Protobothrops 929 mucrosquamatus, Pn: Pseudonaja textilis, Pp: Pseudechis porphyriacus, Psa: 930 Psammodromus algirus, Pv: Pogona vitticeps, Pyc: Python curtus, Sb: Sceloporus 931 bicanthalis, Sca: Struthio camelus australis, Sct: Sistrurus tergeminus, See: Sistrurus 932catenatus edwardsi, Sib: Simalia boeleni, Sjb: Sinomicrurus japonicus boettgeri, Sjj: 933 Sinomicrurus japonicus japonicus, Sjt: Sinomicrurus japonicus takarai, Smi: 934 Sinomicrurus macclellandi iwasakii, Tc: Tropidechis carinatus, Tcu: Tropidophis 935 curtus, Tel: Tropidophis greenwayi lanthanus, Tg: Trimeresurus gracilis, Th: 936 Takydromus hsuehshanensis, Ts: Thamnophis sirtalis, Tt: Tropidophis taczanowskyi, 937 Uo: Urosaurus ornatus, Vu: Vipera ursinii, Wa: Walterinnesia aegyptia, Xv: Xantusia 938 vigilis.

939

940 Supplementary Figure S3. Summary and molecular phylogeny of the 941 Metalloproteinase (MP) family. SV genes (a), NV genes (b) and a molecular 942 phylogeny of SV and NV genes from the metalloproteinase family (c-e). Phylogenetic 943 trees were constructed with the maximum likelihood method using IQ-TREE 944 (<u>http://www.iqtree.org</u>) based on aligned amino acid sequences. Numbers on branches 945 are bootstrap values with 1000x resampling.

946 Aam: Apteryx australis mantelli, Ac: Anolis carolinensis, Acc: Aquila chrysaetos
947 canadensis, Acl: Agkistrodon contortrix laticinctus, Af: Aptenodytes forsteri, Am:
948 Alligator mississippiensis, Ap: Agkistrodon piscivorus, Apl: Agkistrodon piscivorus

949 leucostoma, As: Alligator sinensis, Ap: Anas platyrhynchos, Ba: Bothrops atrox, Bj: 950 Bothrops jararaca, Brs: Buceros rhinoceros silvestris, Ca: Crotalus adamanteus, Cad: 951Crotalus adamanteus, Cdd: Crotalus durissus durissus, Cdc: Crotalus durissus 952 collilineatus, Cg: Cricetulus griseus, Ch: Crotalus horridus, Cj: Coturnix japonica, Cm: 953 Chelonia mydas, Cp: Crocodvlus porosus, Cpb: Chrysemys picta, Cvv: Crotalus viridis 954viridis, Da: Deinagkistrodon acutus, Ec: Echis coloratus, Gh: Gloydius halys, Gi: 955Gloydius intermedius, Gj: Gekko japonicus, Gs: Gloydius saxatilis, MI: Macrovipera 956 lebetina, Oh: Ophiophagus hannah, Oo: Ovophis okinavensis, Pb: Python bivittatus, Pc: 957Phalacrocorax carbo, Pf: Protobothrops flavoviridis, Pj: Protobothrops jerdonii, Pm: 958Protobothrops mucrosquamatus, Ps: Pelodiscus sinensis, Sct: Sistrurus tergeminus, 959 Sce: Sistrurus catenatus edwardsi, Smb: Sistrurus miliarius barbouri, Ta: Trimeresurus 960 albolabris, Tg: Trimeresurus gracilis, Ths: Thamnophis sirtalis, Ts: Thamnophis 961 sirtalis, Vaa: Vipera ammodytes ammodytes.

962

963

964 Supplementary Figure S4. Summary and molecular phylogeny of the Serine
965 protease (SP) family.

966 SV genes (a), NV genes (b) and a molecular phylogeny of SV and NV genes of the 967 serine protease family (c-d). Multiple sequence alignments were performed with 968 ClustalW. Phylogenetic trees were constructed with the maximum likelihood method 969 using IQ-TREE (<u>http://www.iqtree.org</u>) based on the aligned amino acid sequences. 970 Numbers on branches are bootstrap values with 1000x resampling. 971 Ac: Anolis carolinensis, As: Alligator sinensis, Ap: Anas platyrhynchos, Bf: Bungarus fasciatus, Ca: Crotalus adamanteus, Ch: Crotalus horridus, Cm: Chelonia mydas, Ec: 972973 Echis coloratus, Gj: Gekko japonicus, Mf: Micrurus fulvius, Ml: Macrovipera lebetina, 974 Mr: Macropisthodon rudis, Mt: Micrurus tener, Ns: Notechis scutatus, Oh: 975 Ophiophagus hannah, Oo: Ovophis okinavensis, Pa: Pseudechis australis, Pb: Python 976 bivittatus, Pe: Protobothrops elegans, Pf: Protobothrops flavoviridis, Pm: 977 Protobothrops mucrosquamatus, Pp: Pseudechis porphyriacus, Smb: Sistrurus 978 miliarius barbouri, Ts: Thamnophis sirtalis.

979

980 Supplementary Figure S5. Summary and molecular phylogeny of the C-type 981 lection-like protein (CTLP) family. SV genes (a), NV genes (b) and a molecular 982 phylogeny of ten SV genes from the C-type lectin-like protein family (c). Multiple 983 sequence alignments were performed with ClustalW. Phylogenetic trees were 984constructed with the maximum likelihood method using **IQ-TREE** 985 (http://www.iqtree.org) based on the aligned amino acid sequences. Numbers on 986 branches are bootstrap values with 1000 resampling.

Ac: Anolis carolinensis, Acc: Agkistrodon contortrix contortrix, Ap: Agkistrodon
piscivorus, Ba: Bitis arietans, Bf: Bungarus flaviceps, Bj: Bothrops jararaca, Ca:
Crotalus adamanteus, Ch: Crotalus horridus, Cn: Cryptophis nigrescens, Coh: Crotalus
oreganus helleri, Cr: Calloselasma rhodostoma, Da: Deinagkistrodon acutus, Dv:
Demansia vestigiata, Ecs: Echis carinatus sochureki, Gb: Gloydius brevicaudus, Gh:
Gloydius halys, Lm: Lachesis muta, Mc: Micrurus corallinus, MI: Macrovipera

993 lebetina, Mt: Micrurus tener, Ns: Notechis scutatus, Oh: Ophiophagus hannah, Pa:
994 Pseudechis australis, Pb: Python bivittatus, Pf: Protobothrops flavoviridis, Pj:
995 Protobothrops jerdonii, Pm: Protobothrops mucrosquamatus, Pn: Parasuta nigriceps,

996 Pp: Pseudechis porphyriacus, Ps: Pelodiscus sinensis, Pt: Pseudonaja textilis, Sce:

997 Sistrurus catenatus edwardsi, Ta: Trimeresurus albolabris, Ts: Thamnophis sirtalis

998

999 Supplementary Figure S6. Summary and molecular phylogeny of the 1000 Phospholipase A_2 (PLA2) family. SV genes (a), NV genes (b) and a molecular 1001 phylogeny of SV and NV genes of the phospholipase A_2 family (c). Multiple sequence 1002 alignments were performed using ClustalW. Phylogenetic trees were constructed by 1003 maximum likelihood method using IQ-TREE (<u>http://www.iqtree.org</u>) based on aligned 1004 amino acid sequences. Numbers on branches are bootstrap values with 1000x 1005 resampling.

1006 Ac: Anolis carolinensis, Acc: Agkistrodon contortrix contortrix, Ap: Agkistrodon 1007 piscivorus, Ba: Bitis arietans, Bas: Bothrops asper, Bd: Bothrops diporus, Bj: Bothrops 1008 jararaca, Bl: Bothrops leucurus, Bm: Bothrops moojeni, Bs: Bothriechis schlegelii, Ca: 1009 Crotalus adamanteus, Cc: Crotalus cerberus, Cdt: Crotalus durissus terrificus, Cg: 1010 Cerrophidion godmani, Cmn: Crotalus molossus nigrescens, Coa:Crotalus oreganus 1011 abyssus, Coc: Crotalus oreganus concolor, Cpb: Chrysemys picta bellii, Cr: 1012 Calloselasma rhodostoma, Css: Crotalus scutulatus scutulatus, Gb: Gloydius 1013 brevicaudus, Gh: Gloydius halys, Gi: Gloydius intermedius, Gu: Gloydius ussuriensis, 1014Lm: Lachesis muta, Oh: Ophiophagus hannah, Om: Ovophis makazayazaya, Oo: 1015 Ovophis okinavensis, Pb: Python bivittatus, Pe: Protobothrops elegans, Pf:
1016 Protobothrops flavoviridis, Pm: Protobothrops mucrosquamatus, Scc: Sistrurus
1017 catenatus catenatus, Sct: Sistrurus tergeminus, Sm: Sistrurus miliarius, Smb: Sistrurus
1018 miliarius barbouri, Sms: Sistrurus miliarius streckeri, Ta: Trimeresurus albolabris, Tg:
1019 Trimeresurus gracilis, Ts: Thamnophis sirtalis.

1020

1021Supplementary Figure S7. Three possible evolutionary scenarios. Thee possible 1022 evolutionary scenarios to explain the relationship between 2R-WGD and duplication of 1023 four ohnologs (A, B, C and D) into genes that encode venom proteins (red bars) and 1024those that encode non-venom proteins (blue bars). 1R and 2R indicates first and second 1025round WGD, respectively. (ABCD), (AB) and (CD) represent ancestral genes. (a) a 1026 molecular phylogeny showing the diversification occurred randomly in all ohnologs. (b) 1027a molecular phylogeny showing the diversification occurred in one of the two ohnologs 1028 that were produced after the second round of WGD. (c) a molecular phylogeny showing 1029 only one ohnolog (in this case, ohnolog A) was duplicated to give rise to venom protein 1030 genes while in the three others, duplication did not involve the development of venom 1031protein genes.

1032

Supplementary Figure S8. Tandem duplications of SV genes. Seven examples of
tandem duplications of MP (a), SP (b), and CTLP (c) genes. SV copies and NV copies
are shown in red and grey arrows, respectively. Sizes of gene clusters are shown on the
right side.

1037

Supplementary Figure S9. Five examples of FISH mapping of cDNA clones to 1038 1039 metaphase spreads of Protobothrops flavoviridis and Elaphe quadrivirgata. 1040 Arrowheads indicate hybridization signals. TEX14 (b, h), CBX2 (c, i) and AR (d, j) 1041 genes were mapped to chromosomes 1p, 2q, and a pair of microchromosomes, 1042respectively, in P. flavoviridis (b, c, d) and E. quadrivirgata (h, i, j). Hoechst-stained 1043 G-banded pattern of the same metaphase chromosomes as in b and h is shown in a and g, 1044 respectively. While GAD2 genes mapped to Zq in P. flavoviridis (e) and Zp in E. 1045quadrivirgata (k), EIF1 genes to Zp in P. flavoviridis (f) and Zq in E. quadrivirgata (l). 1046Arm ratios of Z chromosomes were different between the two species due to a small 1047pericentric inversion that occurred in Z chromosomes in the *Elaphe* lineage. 1048 1049 Supplementary Figure S10. Schematic diagram of the gene structures of the snake 1050venom metalloproteinases (svMP), HV1, flavorase, VMP-III-like, jerdonitin-like, 1051 HR1a, H2-protease, HR2a, flavoviridin, HR1b, elegantin and NaMP-like. Red box

shows the eight core exons (4 to 11) encoding metalloproteinase domains. HHHindicate catalytic sites for MP.

1054

1055 Supplementary Figure S11. Schematic structure of the snake venom serine 1056 protease (svSP) family genes. Flavoxobin (svSP01, TLf1), TLf2 (svSP02), TLf3 1057 (svSP03), and svSP04 to svSP11 (a), and a gene structure comparison among 1058 representative serine proteases (b) are shown. These svSP genes essentially consist of 6 exons, the first of which contains only the 5'non-coding region, except for svSP06
having additional two exons Ex7 and Ex8, and svSP05 possessing a sub-exon, Ex3b. H,
D, and S in the boxes indicate the catalytic triad. Arrowheads show the initiation
codons.

1063

Supplementary Figure S12. Summary and a phylogenetic tree of the three-finger toxin (3FTX) family. SV and NV genes (a) and a molecular phylogeny (b) of the 3FTX family. Multiple sequence alignments were performed using ClustalW. Phylogenetic trees were constructed with the maximum likelihood method using IQ-TREE (<u>http://www.iqtree.org</u>) based on aligned amino acid sequences. Numbers on branches are bootstrap values with 1000x resampling.

1070 Af: Azemiops feae, Ahp: Ahaetulla prasina, Am: Alligator mississippiensis, Bc: 1071Bungarus candidus, Bf: Bungarus flaviceps, Bi: Boiga irregularis, Bm: Bungarus 1072multicinctus, Cm: Chelonia mydas, Cpb: Chrysemys picta bellii, Dt: Dispholidus typus, 1073Lm: Leioheterodon madagascariensis, Mt: Micrurus tener, Nn: Naja naja, Oh: 1074 Ophiophagus hannah, Oo: Ovophis okinavensis, Pe: Protobothrops elegans, Pf: Protobothrops flavoviridis, Pm: Protobothrops mucrosquamatus, Psm: Psammophis 10751076mossambicus, Sce: Sistrurus catenatus edwardsi, Tb: Trimorphodon biscutatus, Td: 1077Telescopus dhara, Tj: Thrasops jacksonii, Ths: Thamnophis sirtalis.

1078

1079 Supplementary Figure S13. Summary and a phylogenetic tree of the
1080 aminopeptidase family. SV and NV genes (a) and a molecular phylogeny (b) of the

aminopeptidase family. Multiple sequence alignments were performed using ClustalW.
Phylogenetic trees were constructed with the maximum likelihood method using
IQ-TREE (<u>http://www.iqtree.org</u>) based on aligned amino acid sequences. Numbers on
branches are bootstrap values with 1000x resampling.

1085 Ac: Anolis carolinensis, Br: Bitis rhinoceros, Gb: Gloydius brevicaudus, Oh:
1086 Ophiophagus hannah, Oo: Ovophis okinavensis, Pb: Python bivittatus, Pf:
1087 Protobothrops flavoviridis, Pm: Protobothrops mucrosquamatus, Ts: Thamnophis
1088 sirtalis.

1089

1090 Supplementary Figure S14. Summary and a phylogenetic tree and structure of the 1091 Cysteine-rich secretory protein (CRISP) family. SV and NV genes (a) and a molecular phylogeny (b) of the CRISP family. Comparison of amino-acid sequences 1092 1093 between CRISP01 and 02 (c), and CRISP03 and 04 (d). Multiple sequence alignments 1094 were performed using ClustalW. Phylogenetic trees were constructed with the 1095 maximum likelihood method using IQ-TREE (http://www.iqtree.org) based on aligned 1096 amino acid sequences. Numbers on branches are bootstrap values with 1000x 1097resampling.

Ab: Agkistrodon blomhoffi, Ac: Anolis carolinensis, Af: Azemiops feae, Am: Alligator
mississippiensis, An: Atheris nitschei, App: Agkistrodon piscivorus piscivorus, As:
Austrelaps superbus, Bc: Bungarus candidus, Bs: Bothriechis schlegelii, Ca: Crotalus
atrox, Cg: Cerrophidion godmani, Ch: Crotalus horridus, Cr: Causus rhombeatus, Clr:
Calloselasma rhodostoma, Cv: Crotalus viridis, Da: Deinagkistrodon acutus, Dr:

1103 Daboia russellii, Dt: Dispholidus typus, Dv: Demansia vestigiata, Ep: Enhydris 1104 polylepis, Gi: Gloydius intermedius, Hs: Hoplocephalus stephensii, Lh: Lapemis 1105hardwickii, Lm: Leioheterodon madagascariensis, Lp: Liophis poecilogyrus, Ls: 1106 Laticauda semifasciata, Mi: Micropechis ikaheca, Na: Naja atra, Nk: Naja kaouthia, 1107 Ns: Notechis scutatus, Oh: Ophiophagus hannah, Om: Oxyuranus microlepidotus, Oo: 1108 Ovophis okinavensis, Pa: Pseudechis australis, Pb: Python bivittatus, Pf: Protobothrops 1109 flavoviridis, Pe: Protobothrops elegans, Pm: Protobothrops mucrosquamatus, Po: 1110 Philodryas olfersii, Pp: Pseudechis porphyriacus, Pt: Pseudonaja textilis, Rn: 1111 Rhinoplocephalus nigrescens, Rtt: Rhabdophis tigrinus tigrinus, Sce: Sistrurus 1112catenatus edwardsi, Sct: Sistrurus catenatus tergeminuss, Tb: Trimorphodon biscutatus, 1113 Tc: Tropidechis carinatus, Td: Telescopus dhara, Tg: Trimeresurus gracilis, Tj: 1114 Trimeresurus jerdonii, Ts: Thamnophis sirtalis., Vb: Vipera berus, Vn: Vipera nikolskii, 1115Vs: Viridovipera stejnegeri.

1116

1117 Supplementary Figure S15. Summary and a phylogenetic tree of the vespryn 1118 family. SV and NV genes (a) and a molecular phylogeny (b) of the vespryn family. Multiple sequence alignments were performed using ClustalW. Phylogenetic trees were 1119 1120constructed with the maximum likelihood method using **IQ-TREE** 1121(http://www.iqtree.org) based on aligned amino acid sequences. Numbers on branches 1122are bootstrap values with 1000x resampling.

1123 Ac: Anolis carolinensis, Ca: Crotalus atrox, Ch: Crotalus horridus, Lm: Leioheterodon

1124 madagascariensis, Mi: Micropechis ikaheka, Oh: Ophiophagus hannah, Pb: Python

1125 *bivittatus*, Pf: *Protobothrops flavoviridis*, Pm: *Protobothrops mucrosquamatus*.

1126

1127 Supplementary Figure S16. Summary and a phylogenetic tree of the
1128 5'-nucleotidase family. SV and NV genes (a) and a molecular phylogeny (b) of the
1129 5'-nucleaotidase family. Multiple sequence alignments were performed using ClustalW.
1130 Phylogenetic trees were constructed with the maximum likelihood method using
1131 IQ-TREE (<u>http://www.iqtree.org</u>) based on aligned amino acid sequences. Numbers on
1132 branches are bootstrap values with 1000x resampling.
1133 Ac: *Anolis carolinensis*, Oh: *Ophiophagus hannah*, Oo: *Ovophis okinavensis*, Pb:

1134 Python bivittatus, Pe: Protobothrops elegans, Pf: Protobothrops flavoviridis, Pm:
1135 Protobothrops mucrosquamatus.

1136

Supplementary Figure S17. Summary and a phylogenetic tree of the dipeptidyl peptidase (DPP) family. SV and NV genes (a) and a molecular phylogeny (b) of the dipeptidyl peptidase family. Multiple sequence alignments were performed using ClustalW. Phylogenetic trees were constructed with the maximum likelihood method using IQ-TREE (http://www.iqtree.org) based on aligned amino acid sequences. Numbers on branches are bootstrap values with 1000x resampling.

1143 Ac: Anolis carolinensis, Am: Alligator mississippiensis, As: Austrelaps superbus, Gj:

1144 Gekko japonicus, Ns: Notechis scutatus, Oh: Ophiophagus hannah, Oo: Ovophis

1145 okinavensis, Pa: Pseudechis australis, Pb: Python bivittatus, Pe: Protobothrops elegans,

1146 Pf: Protobothrops flavoviridis, Pm: Protobothrops mucrosquamatus, Ts: Thamnophis

1147 sirtalis.

1148

1149Supplementary Figure S18. Summary and a phylogenetic tree of the vascular 1150endothelial growth factor (VEGF)-like protein family. SV and NV genes (a) and a molecular phylogeny (b) of the VEGF family. Multiple sequence alignments were 11511152performed using ClustalW. Phylogenetic trees were constructed with the maximum 1153likelihood method using IQ-TREE (http://www.iqtree.org) based on aligned amino acid 1154sequences. Numbers on branches are bootstrap values with 1000x resampling. 1155Ac: Anolis carolinensis, Ap: Agkistrodon piscivorus, App: Agkistrodon piscivorus 1156piscivorus, Ba: Bitis arietans, Be: Bothrops erythromelas, Bg: Bitis gabonica, Bj:

Bothrops jararaca, Ca: Crotalus adamanteus, Ch: Crotalus horridus, Gt: Gloydius
tsushimaensis, Mf: Micrurus fulvius, Mt: Micrurus tener, Oh: Ophiophagus hannah,
Oo: Ovophis okinavensis, Pb: Python bivittatus, Pe: Protobothrops elegans, Pf:
Protobothrops flavoviridis, Pm: Protobothrops mucrosquamatus, Sce: Sistrurus
catenatus edwardsi, Sct: Sistrurus catenatus tergeminuss, Smb: Sistrurus miliarius
barbouri, Ts: Thamnophis sirtalis, Vaa: Vipera ammodytes ammodytes.

1163

Supplementary Figure S19. Summary and a phylogenetic tree of the phosphodiesterase (PDE) family. SV and NV genes (a) and a molecular phylogeny (b) of the PDE family. Multiple sequence alignments were performed using ClustalW. Phylogenetic trees were constructed with the maximum likelihood method using IQ-TREE (http://www.iqtree.org) based on aligned amino acid sequences. Numbers on

branches are bootstrap values with 1000x resampling. A red dot shows the branch pointbetween toxic and non-toxic proteins.

- 1171 Ac: Anolis carolinensis, Am: Alligator mississippiensis, Ca: Crotalus adamanteus, Ch:
- 1172 Crotalus horridus, Mf: Micrurus fulvius, Mr: Macropisthodon rudis, Mt: Micrurus
- 1173 tener, Oo: Ovophis okinavensis, Pb: Python bivittatus, Pe: Protobothrops elegans, Pf:
- 1174 Protobothrops flavoviridis, Pm: Protobothrops mucrosquamatus, Smb: Sistrurus
 1175 miliarius barbouri.
- 1176

1177 Supplementary Figure S20. Summary and a phylogenetic tree of the phospholipase

B (**PLB**) **family.** SV and NV genes (a) and a molecular phylogeny (b) of the PLB family. Multiple sequence alignments were performed using ClustalW. Phylogenetic trees were constructed with the maximum likelihood method using IQ-TREE (<u>http://www.iqtree.org</u>) based on aligned amino acid sequences. Numbers on branches are bootstrap values with 1000x resampling.

1183 Ac: Anolis carolinensis, Dc: Drysdalia coronoides, Oh: Ophiophagus hannah, Oo:

1184 Ovophis okinavensis, Pb: Python bivittatus, Pe: Protobothrops elegans, Pf:

1185 Protobothrops flavoviridis, Pm: Protobothrops mucrosquamatus.

1186

Supplementary Figure S21. Summary and a phylogenetic tree of the C-type natriuretic peptide (CNP) family. SV and NV genes (a) and a molecular phylogeny (b) of the CNP family. Multiple sequence alignments were performed using ClustalW. Phylogenetic trees were constructed with the maximum likelihood method using

1191 IQ-TREE (<u>http://www.iqtree.org</u>) based on aligned amino acid sequences. Numbers on
1192 branches are bootstrap values with 1000x resampling.

1193 Ac: Anolis carolinensis, Acc: Agkistrodon contortrix contortrix, Am: Alligator 1194 mississippiensis, Ap: Agkistrodon piscivorus, As: Alligator sinensis, Ba: Bitis arietans, 1195Bi: Boiga irregularis, Bj: Bothrops jararaca, Bp: Bothrops pauloensis, Ca: Crotalus 1196 adamanteus, Cc: Cerastes cerastes, Ch: Crotalus horridus, Cm: Chelonia mydas, Cr: 1197Calloselasma rhodostoma, Dr: Daboia russelii, Eo: Echis ocellatus, Epl: Echis 1198 pyramidum leakevi, Gb: Gloydius blomhoffii, Gg: Gallus gallus, Gh: Gloydius halys, Gi: Gloydius intermedius, Gj: Gekko japonicus, Lm: Lachesis muta, Mg: Meleagris 1199 1200 gallopavo, MI: Macrovipera lebetina, Oh: Ophiophagus hannah, Oo: Ovophis 1201okinavensis, Pb: Python bivittatus, Pc: Protobothrops elegans, Pf: Protobothrops flavoviridis, Phm: Phalotris mertensi, Pm: Protobothrops mucrosquamatus, Sca: 12021203 Struthio camelus australis, Sct: Sistrurus tergeminus, Smb: Sistrurus miliarius barbouri, 1204Tg: Trimeresurus gracilis, Ts: Thamnophis sirtalis.

1205

Supplementary Figure S22. Summary and a phylogenetic tree of the glutaminyl peptide cyclotransferase (GPCase) family. SV and NV genes (a) and a molecular phylogeny (b) of the GPCase family. Multiple sequence alignments were performed using ClustalW. Phylogenetic trees were constructed with the maximum likelihood method using IQ-TREE (http://www.iqtree.org) based on aligned amino acid sequences. Numbers on branches are bootstrap values with 1000x resampling.

1212 Ac: Anolis carolinensis, Bd: Bothrops diporus, Bi: Boiga irregularis, Bj: Bothrops

- 1213 jararaca, Cg: Cerrophidion godmani, Cpb: Chrysemys picta bellii, Dr: Daboia russelii,
- 1214 Gj: Gekko japonicus, Mf: Micrurus fulvius, Oo: Ovophis okinavensis, Pb: Python
- 1215 bivittatus, Pe: Protobothrops elegans, Pf: Protobothrops flavoviridis, Pm:
- 1216 Protobothrops mucrosquamatus. Sct: Sistrurus catenatus tergeminus, Tg: Trimeresurus
- 1217 gracilis, Ts: Thamnophis sirtalis.
- 1218
- 1219

Supplementary Figure S1.



Supplementary Fig. S2.

(a) Hyaluronidase (Hyal) Member of the family

Wieniber of the	Weinder of the failing				
Gene	Name	Locus	Gene model ID	Transcripts	
Hyal01 (svHyal)	svHyaluronidase (svHYAL)	habu1_scaffold7188 : 185477 226495 : -	habu1_s7188_g17820	pb016481_c260499_f1p0_1519	
Hyal02 (nvHyal)	hyaluronidase-1 (HYAL1)	habu1_scaffold2564 : 518042 530062 : +	habu1_s2564_g09254	habu1_s2564_g09254.t1	
Hyal03 (nvHyal)	hyaluronidase-2 (HYAL2)	habu1_scaffold2564 : 478562 496616 : +	habu1_s2564_g09252	habu1_s2564_g09252.t1	
Hyal04 (nvHyal)	hyaluronidase-2 (HYAL2)	habu1_scaffold6849 : 149616 194647 : +	habu1_s6849_g17560	habu1_s6849_g17560.t1	
Hyal05 (nvHyal)	hyaluronidase-3 (HYAL3)	habu1_scaffold2564 : 546642 569966 : +	habu1_s2564_g09256	habu1_s2564_g09256.t1	
Hyal06 (nvHyal)	hyaluronidase-4 (HYAL4)	habu1_scaffold7188 : 99577 184945 : -	habu1_s7188_g17819	habu1_s7188_g17819.t1	

Molecular phylogeny of members



Fig. S2 contineued.

(b) Nerve growth factor (NGF) Member of the family

Withingth of the	vicinity of the family				
Gene	Name	Locus	Gene model ID	Transcripts	
Neu01 (svNeu)	svNGF	habu1_scaffold3536 : 227336 309758 : +	habu1_s3536_g11975	pb016481_c100488_f2p5_1112	
Neu02 (nvNeu)	BDNF	habu1_scaffold3803 : 208016 266737 : +	habu1_s3803_g12736	habu1_s3803_g12736.t1	
Neu03 (nvNeu)	neurotrophin-3 isoform	habu1_scaffold105 : 263936 278118 : +	habu1_s105_g00548	habu1_s105_g00548.t1	
Neu04 (nvNeu)	neurotrophin-4 isoform	habu1_scaffold6123 : 46517 70395 : -	habu1_s6123_g16797	habu1_s6123_g16797.t1	

Molecular phylogeny of members



Supplementary Figure S2 contineued.

(c) L-amino acid oxidase (LAAO) Member of the family

Michiger of the	Archiber of the family				
Gene	Name	Locus	Gene Model ID	Transcripts	
LAAO01 (svLAAO)	LAAO01 (svLAAO)	habu1_scaffold402940 : 463276 513813 : -	habu1_s402940_g24950	pb016481_c420538_f1p26_2821	
LAAO02 (nvLAAO)	LAAO02 (nvLAAO)	habu1_scaffold402940 : 432924 460003 : -	habu1_s402940_g24949	habu1_s402940_g24949.t1	
LAAO03 (nvLAAO)	LAAO03 (nvLAAO)	habu1_scaffold402940 : 288786 347897 : +	habu1_s402940_g24947b	habu1_s402940_g24947.t1	

Molecular phylogeny of members



(a) SV genes

Gene	Name	Locus	Gene model ID	Transcripts
svMP01	HV1, Vascular apoptosis-inducing protein	habu1_scaffold2862: 139192-161910: +	habu1_s2862_g10314a	pb016480_c42639_f26p8_2346
svMP02	flavorase	habu1_scaffold2862: 165955-186529: +	habu1_s2862_g10314b	pb016480_c42674_f12p2_2354
svMP03	VMP-III -like	habu1_scaffold2862: 188915-209119: +, habu1_scaffold14911:15918-22474: +	habu1_s2862_g10314c	pb016481_c551930_f88p36_2337
svMP04	jerdonitin-like	habu1_scaffold3258:115093: +, habu1_scaffold415864:1818: -, habu1_scaffold191139: 1797: -, habu1_scaffold403873:1701: -	habu1_s3258_g11210	pb016481_c222387_f3p5_2004
svMP05	HR1a	habu1_scaffold3258: 15841-52645: +	habu1_s3258_g11211	pb016480_c115296_f1p1_2355
svMP06	H2 metalloproteinase (metalloprotease P-lia)	habu1_scaffold14911:20225557: +	habu1_s14911_g21429	pb016481_c225249_f1p9_1815
svMP07	HR2a	habu1_scaffold14911: 6078976673: + habu1_scaffold14911: 121550: +	habu1_s14911_g21430a	pb016480_c226_f26p9_2010
svMP08	flavoridin	habu1_scaffold14911:373353141: +	habu1_s14911_g21430b	pb016480_c222_f46p26_1997
svMP09	HR1b	habu1_scaffold399953:2078536711: -	habu1_s399953_g24864a	pb016481_c917376_f5p9_2037
svMP10	Mt-b/ elegantin-like	habu1_scaffold399953:138225: - habu1_scaffold410279:1747: - habu1_scaffold408733:151: - habu1_scaffold412967:1795: -	habu1_s399953_g24864b	pb016481_c16287_f13p13_2020
svMP11	NaMP	habu1_scaffold2862: 106532130000+	habu1_s2862_g10314d	habu1_s2862_g10314.t2

(b) NV genes

(D) IN V	genes			
Gene	Name	Locus	Gene model ID	Transcripts
nvMP01	ADAM8/9-like	habu1 scaffold2 : 71286 110080 : +	habu1 s2 q00003	habu1 s2 g00003.t1
nvMP02	ADAM9		habu1 s179838 g24144	habu1 s179838 g24144.t1
nvMP03	ADAM 9		habu1 s2858 g10274	habu1 s2858 g10274.t1
nvMP04	ADAM 10-like	habu1_scaffold2336; 11126231112538; -	habu1 s4524 g14387/	habu1 s2934 g10526.t1
		habu1 scaffold4524 : 1 13872 : +,	habu1 s333519 g24419/	
		habu1_scaffold333519 : 1 395 : -,	habu1_s2934_g10526	
		habu1_scaffold2934 : 1 18140 : +,		
		habu1_scaffold244794:1917-		
nvMP05	ADAM10	habu1_scaffold878 : 135568 161715 : -	habu1_s878_g03107	habu1_s878_g03107.t1
nvMP06	ADAM11	habu1_scaffold2140 : 97648 117201 : -	habu1_s2140_g07747	habu1_s2140_g07747.t1
nvMP07	ADAM11	habu1_scaffold5075 : 1 61735 : -	habu1_s5075_g15185	habu1_s5075_g15185.t1
nvMP08	ADAM 12	habu1_scaffold2690 : 97936 361842 : +	habu1_s2690_g09595	habu1_s2690_g09595.t1
nvMP09	ADAM 15	habu1_scaffold5687 : 27897 62375 : -	habu1_s5687_g16173	habu1_s5687_g16173.t1
nvMP10	ADAM 17	habu1_scaffold602 : 44893 82435 : -	habu1_s602_g02843	habu1_s602_g02843.t1
nvMP11	ADAM 19	habu1_scaffold5506 : 105086 193255 : -	habu1_s5506_g15874	habu1_s5506_g15874.t1
nvMP12	ADAM 20	(habu1_scaffold8405:494795504794: +)		
nvMP13	ADAM21	habu1_scaffold399842 : 646178 651147 : +	habu1_s399842_g24578	habu1_s399842_g24578.t1
nvMP14	ADAM 22	habu1_scaffold4012 : 359513 383057 : -	habu1_s4012_g13214	habu1_s4012_g13214.t1
nvMP15	ADAM 32	habu1 scaffold2858 : 1837688 1857057 : +	habu1 s2858 g10275	habu1 s2858 g10275.t1
nvMP16	ADAM 33	habu1_scaffold3839 : 5573 17043 : -	habu1 s3839 g12793	habu1 s3839 g12793.t1
nvMP17	ADAM 33		habu1 s44599 g23329	habu1 s44599 g23329.t1
nvMP18	ADAMTS-like	habu1_scaffold5048 · 540792 774172 · -	habu1 s5048 g15167	habu1_s5048_g15167 t1
nvMP19	ADAMTS1	habu1_scaffold2614 · 247016 269192 · +	habu1 s2614 g09476	habu1_s2614_g09476.t1
nvMP20	ADAMTS 2	habu1_ccaffold5378 : 27996 91121 : +	habu1_5378_d15691	habu1_55378_d15691_t1
nvMP21	ADAMTS2	habu1_scaffold4909:1 71717:+	habu1_s4909_g14946	habu1_cccro_g1ccc1.t1
nvMD22	ADAMTS2	habu1_scaffold4909 : 72026 141808 : +	habu1_s4909_g14940	habu1_34303_g14340.01
nvMD23	ADAMTS 2	habu1_scaffold3496 : 12525 141606 : 1	habu1_34909_g14947	habu1_34909_g14947.t1
nvMD24	ADAMTS 3	habu1_scallold3490 : 103170 243933 : -	habu1_33490_g11075	habu1_33490_g11073.t1
nvMD25	ADAMTS 4 like	habu1_scallold2091.29300 221749.+	habu1_22091_g07045	habu1_s2091_g07045.t1
nvMD26		habu1_scallolu2309 . 17219 36126	habu1_s2309_g08591	habu1_s2309_g08391.t1
	ADAMTS5 like	habu1_scallolu121.92410110766.+	habu1_\$121_g00004	habu1_s121_g00004.t1
NVIVIP27	ADAM 155-like	habu1_scaffold6970 : 1 21977 : +	habu1_s6970_g17669	habu1_s6970_g17669.t1
nvMP28	ADAMTS5	habu1_scaffold2614 : 187286 229869 : +	habu1 s2614 g09475	habu1 s2614 g09475.t1
nvMP29	ADAMTS6	habu1_scaffold16578 : 176146 270945 : -	habu1 s16578 g21764	habu1 s16578 g21764.t1
nvMP30	ADAMTS7	habu1_scaffold1390 1630741 1657463 -	habu1 s1390 g05010	habu1 s1390 g05010 t1
nvMP31	ADAMTS 8	habu1_ccaffold2647 240949 299435 -	habu1_2647_009516	habu1_c1647_d09516.t1
nvMP32	ADAMTS9	habu1_ccaffold523 : 106186 238219 : +	habu1 s523 d02405	habu1_523_002405.t1
nvMP33	ADAMTS9-like	habu1_scaffold524 : 46586 55632 : +	habu1_524_002406	habu1_5524_002406.t1
nvMP34		habu1_scaffold2275:129686195770:+	habu1_0024_002400	habu1_0024_902400.01
nvMP35		habu1_scaffold2255:1235000 135770: 1	habu1_32275_g00107	habu1_32275_g00157.t1
nvMD36		habu1_scaffold2256 : 1 12075 :	habu1_32256_g00000	habu1_32255_g00000.11
nvMD37		habu1_scaffold1017:171206 107845:	habu1_s2230_g08001	habu1_s2230_g00001.t1
DVMD20	ADAMTS13	habu1_scalloid1917 : 171200 197040 : -	habu1_51917_g07045	habu1_51317_g07045.t1
nvMD20	ADAMIS14	habu1_scallolu5729.515490367470.+	habu1_32647_c00517	habu1_s3729_g10218.t1
nvMD40	ADAMISIS	habu1_scallolu2047 . 327310 300932 . +	habu1_52047_909517	habu1_52047_909517.t1
		habu	habu1_317734_921874	$habu1_517734_921374.11$
	ADAMTS17	habu1_scallolu215.17775521900665.+	habu1_s215_g01205	habu1_s215_g01205.t1
	ADAMTO10	habu1_scallolu490 . 205045 029934	habu1_5490_902284	habu1_\$490_902284.01
IIVIVIP43	ADAMIS 19	habu1_scallold65 : 129465 215465 : -	habu1_\$83_g00469	habu1_\$65_000469.01
111111111111111111111111111111111111111		habu1_scallold2519:40667 63655	habu1_s2519_g09165	habu1_s2519_g09165.t1
nviviP45	MMP-15	nabu1_scaffold2277 : 991963 999434 : +	habu1_s22/7_g08190	nabu1_s2277_g08190.t1
nviviP46	MMP-16	nabu1_scaffold2447 : 249336 445542 : +	habu1_s2447_g08959	nabu1_s2447_g08959.t1
nvMP47	MMP-17	habu1_scaffold2313 : 1036 67938 : +	habu1_s2313_g08399	habu1_s2313_g08399.t1
nvMP48	MMP-17	nabu1_scattold1466 : 578466 598489 : -	nabu1_s1466_g05412	nabu1_s1466_g05412.t1
nvMP49	MMP-19	habu1_scaffold402967 : 95446 134769 : +	habu1_s402967_g25016	habu1_s402967_g25016.t1
nvMP50	MMP-20	habu1_scaffold532 : 778467 796669 : -	habu1_s532_g02441	habu1_s532_g02441.t1
nvMP51	MMP-24	habu1_scaffold4809 : 306319 347795 : -	habu1_s4809_g14788	habu1_s4809_g14788.t1
nvMP52	MMP-25	habu1_scaffold9747 : 1 14152 : +	habu1_s9747_g19608	habu1_s9747_g19608.t1
nvMP53	MMP-25	habu1_scaffold21648 : 40170 53088 : -	habu1_s21648_g22429	habu1_s21648_g22429.t1
nvMP54	MMP-28	habu1_scaffold1501 : 177931 192935 : -	habu1_s1501_g05556	habu1_s1501_g05556.t1
nvMP55	ATP-dependent zinc metalloprotease YME1L1	habu1_scaffold403007 : 90596 121556 : +	habu1_s403007_g25105	habu1_s403007_g25105.t1
nvMP56	Metalloprotease TIKI2	habu1_scaffold13164 : 180021 242825 : -	habu1_s13164_g20985	habu1_s13164_g20985.t1
nvMP57	MPtypeIII-1	habu1_scaffold2862: 2690690317:+	habu1_s2862_g10314e	habu1_s2862_g10314.t1



MMP (see Fig. S3e)





3.0

(a) SV genes

<u> </u>	8			
Gene	Name	Locus	Gene model ID	Transcripts
svSP01	TLf1/flavoxobin	habu1_scaffold7597 : 107550 117068 : -	habu1_s7597_g18190a	pb016481_c628_f19p22_1897
3001 02		habu1_scaffold189200:12644: + habu1_scaffold142871:1912: +		pb010401_000000_1140p00_0720
svSP03	TLf3	habu1_scaffold7597 : 146701 152624 : -	habu1_s7597_g18190b	pb016481_c663_f11p12_1578
svSP04		habu1_scaffold6789 : 134126 143039 : +	habu1_s6789_g17480	pb016480_c50972_f1p2_1888
svSP05		habu1_scaffold6789: 168298179481: +	habu1_s6789_g17481a	pb016481_c237569_f1p19_1959
svSP06		habu1_scaffold4106 : 1768 14932 : +, habu1_scaffold375860:1711: +, habu1_scaffold50513:13838: +, habu1_scaffold73303:1843: +, habu1_scaffold270900:1692: +	habu1_s4106_g13431a	pb016481_c782298_f2p3_6239
svSP07		habu1_scaffold4106 : 20911 28186: +	habu1_s4106_g13431b	pb016481_c395211_f1p12_2348
svSP08		habu1_scaffold6789 : 187000 207128 : +	habu1_s6789_g17481b	pb016481_c1327_f9p6_1578
svSP09		habu1_scaffold7597 : 96885 155905 : -	habu1_s7597_g18190c	pb016481_c243922_f1p5_2691
svSP10		habu1_scaffold4106 : 28886 46059 : +	habu1_s4106_g13432	pb016481_c944_f4p2_1299
svSP11	Jerdonobin-II like, GPV-PA	habu1_scaffold22023 : 12371 34705 : -	habu1_s22023_g22469	pb016481_c591372_f1p3_1420

(b) NV genes

Gene	Name	Locus	Gene Model ID	Transcripts
nvSP01	anionic trypsin-like	habu1_scaffold2510 : 115346 122903 : +	habu1_s2510_g09123	habu1_s2510_g09123.t1
nvSP02	anionic trypsin-2-like	habu1_scaffold2510 : 78736 98860 : +	habu1_s2510_g09122	habu1_s2510_g09122.t1
nvSP03	trypsin I-P38-like	habu1_scaffold2510 : 68775 74705 : -	habu1_s2510_g09121	habu1_s2510_g09121.t1
nvSP04	kallikrein-14-like	habu1_scaffold9123 : 219136 242356 : +	habu1_s9123_g19184	habu1_s9123_g19184.t1
nvSP05	trypsin-like	habu1_scaffold9123 : 194396 219087 : +	habu1_s9123_g19183	habu1_s9123_g19183.t1
nvSP06	chymotrypsin-C	habu1_scaffold9572 : 83215 86125 : -	habu1_s9572_g19445	habu1_s9572_g19445.t1
nvSP07	chymotrypsin-like elastase family member 3B	habu1_scaffold3296 : 1 6460 : +	habu1_s3296_g11282	habu1_s3296_g11282.t1
nvSP08	chymotrypsin-like protease CTRL-1	habu1_scaffold493 : 678049 684597 : -	habu1_s493_g02255	habu1_s493_g02255.t1
nvSP09	Neurotrypsin	habu1_scaffold1537 : 430486 473512 : +	habu1_s1537_g05780	habu1_s1537_g05780.t1
nvSP10	neurotrypsin-like	habu1_scaffold2696 : 320034 343955 : -	habu1_s2696_g09609	habu1_s2696_g09609.t1
nvSP11	thrombin	habu1_scaffold10983 : 253158 268223 : -	habu1_s10983_g20317	habu1_s10983_g20317.t1
		/habu1_scaffold10995 : 15124 44505 : -		
nvSP12	tissue-type plasminogen activator (tPA)	habu1_scaffold2858 : 1135301 1175837 : -	habu1_s2858_g10253	habu1_s2858_g10253.t1
nvSP13	plasma kallikrein	habu1_scaffold11004 : 283304 308535 : -	habu1_s11004_g20335	habu1_s11004_g20335.t1
nvSP14	Inactive serine protease 35	habu1_scaffold1630 : 31548 63325 : -	habu1_s1630_g06161	habu1_s1630_g06161.t1
nvSP15	inactive serine protease PAMR1-like	habu1_scaffold3643 : 56659 132295 : -	habu1_s3643_g12251	habu1_s3643_g12251.t1
nvSP16	Serine protease 23, partial	habu1_scaffold6061 : 41736 53088 : +	habu1_s6061_g16640	habu1_s6061_g16640.t1
nvSP17	serine protease 27-like	habu1_scaffold13321 : 1 17475 : -	habu1_s13321_g21080	habu1_s13321_g21080.t1
nvSP18	serine protease 27-like	habu1_scaffold2928 : 8492 20060 : +	habu1_s2928_g10510	habu1_s2928_g10510.t1
nvSP19	serine protease 27-like	habu1_scaffold661 : 20981 65935 : -	habu1_s661_g03024	habu1_s661_g03024.t1
nvSP20	serine protease 27-like	habu1_scaffold14979 : 1 65141 : -	habu1_s14979_g21447	habu1_s14979_g21447.t1
nvSP21	serine protease 33-like	habu1_scaffold661 : 80056 108788 : +	habu1_s661_g03025	habu1_s661_g03025.t1
nvSP22	transmembrane protease serine 12-like: partial	habu1_scaffold5131 : 399731 416735 : -	habu1_s5131_g15292	habu1_s5131_g15292.t1
nvSP23	serine protease 53	habu1_scaffold3327 : 161929 190072 : -	habu1_s3327_g11330	habu1_s3327_g11330.t1
nvSP24	serine protease 55-like	habu1_scaffold16606 : 34165 36715 : -	habu1_s16606_g21794	habu1_s16606_g21794.t1
nvSP25	serine protease 56	habu1_scaffold2472 : 86176 122259 : +	habu1_s2472_g09051	habu1_s2472_g09051.t1
nvSP26	serine protease HTRA4	habu1_scaffold2861 : 833394 841595 : +	habu1_s2861_g10303	habu1_s2861_g10303.t1
nvSP27	serine protease HTRA1-like	habu1_scaffold1135 : 72756 111731 : +	habu1_s1135_g03965	habu1_s1135_g03965.t1
nvSP28	serine protease HTRA2, mitochondrial-like	habu1_scaffold2921 : 104599 106495 : -	habu1_s2921_g10480	habu1_s2921_g10480.t1
nvSP29	thymus-specific serine protease-like	habu1_scaffold1205 : 38676 53312 : +	habu1_s1205_g04155	habu1_s1205_g04155.t1
nvSP30	mannan-binding lectin serine protease 1-like: partial	habu1_scaffold76496 : 1 797 : +	habu1_s76496_g23722	habu1_s76496_g23722.t1
nvSP31	mannan-binding lectin serine protease 1 :partial	habu1_scaffold120144 : 1 3501 : +	habu1_s120144_g23985	habu1_s120144_g23985.t1
nvSP32	mannan-binding lectin serine protease 2	habu1_scaffold1702 : 13930 24737 : -	habu1_s1702_g06326	habu1_s1702_g06326.t1
nvSP33	mannan-binding lectin serine protease 2	habu1_scaffold1703 : 1 17225 : -	habu1_s1703_g06327	habu1_s1703_g06327.t1
nvSP34	serine protease 27-like	habu1_scaffold14979 : 1 65141 : -	habu1_s14979_g21447	habu1_s14979_g21447.t1







(a) SV genes

Gene	Name	Locus	Gene Model ID	Transcripts
svCTLP01	IX/X-bp alpha	habu1_scaffold11033: 1290919459: +		pb016481_c1025238_f1p0_769
svCTLP02	flavocetin-A alpha	habu1_scaffold375321: 193344: + habu1_scaffold81761: 50263746: -		pb016481_c1024339_f1p0_729
svCTLP03	stejaggregin-A-like alpha	habu1_scaffold10061: 161378161319, 200154203895: +	habu1_s10061_g19810c	pb016481_c1014958_f1p2_701
svCTLP04	rhodocetin/EmEMS16-like alpha homologue/ stejaggregin-B-like alpha	habu1_scaffold10061 : 161378178891: -	habu1_s10061_g19810a	pb016481_c1017202_f1p0_682
svCTLP05	IX/X-bp beta_M	habu1_scaffold117853: 386430: - habu1_scaffold241669: 62151: - habu1_scaffold399881: 745837: - habu1_scaffold415837: 179: + habu1_scaffold4161573: 496: +		pb016480_c398342_f1p1_706
svCTLP06	flavocetin-A beta	habu1_scaffold10061 : 155982 167335 : -	habu1_s10061_g19809	pb016481_c1018124_f1p2_702
svCTLP07	rhodocetinEMS16-like-beta	habu1_scaffold10061: 184382193870: -	habu1_s10061_g19810b	pb016481_c1013095_f1p6_704
svCTLP08	C-lectin02	habu1_scaffold3168 : 129063 132925 : -	habu1_s3168_g10977	habu1_s3168_g10977.t1
svCTLP09	Pf_CTLP_FIX/X BP-A_like	habu1_scaffold399941:17140:+ habu1_scaffold75126_1926:+		pb016481_c1017356_f1p0_692
svCTLP10	pb016481 c1008289 f1p0 724	habu1_scaffold3168 : 97112 103333 : +	habu1_s3168_g10975	pb016481_c1008289_f1p0_724

(b) NV genes

Gene	Name	Locus	Gene Model ID	Transcripts
nvCTLP01	C-type lectin	habu1_scaffold10061 : 25086 32544 : +	habu1_s10061_g19804	habu1_s10061_g19804.t1
nvCTLP02	C-type lectin-like	habu1_scaffold10061 : 44806 51663 : +	habu1_s10061_g19805	habu1_s10061_g19805.t1
nvCTLP03	C-type Lectin CRL-like	habu1_scaffold10061 : 52556 89545 : +	habu1_s10061_g19806	habu1_s10061_g19806.t1
nvCTLP04	thymidine phosphorylase	habu1_scaffold10061 : 93712 132005 : -	habu1_s10061_g19807	habu1_s10061_g19807.t1
nvCTLP05	C-type lectin: regenerating islet-derived protein 4-like	habu1_scaffold10061 : 139236 146528 : +	habu1_s10061_g19808	habu1_s10061_g19808.t1
nvCTLP06	C-type lectin BiL-like	habu1_scaffold10589 : 1 10687 : +	habu1_s10589_g20136	habu1_s10589_g20136.t1
nvCTLP07		habu1_scaffold10590 : 1 60935 : -	habu1_s10590_g20137	habu1_s10590_g20137.t1
nvCTLP08		habu1_scaffold10590 : 62389 78515 : -	habu1_s10590_g20138	habu1_s10590_g20138.t1
nvCTLP-09	C-type lectin BiL-like	habu1_scaffold20885 : 1 6175 : -	habu1_s20885_g22359	habu1_s20885_g22359.t1
nvCTLP-10	C-type lectin Cal-like	habu1_scaffold20885 : 9716 23508 : +	habu1_s20885_g22360	habu1_s20885_g22360.t1
nvCTLP-11		habu1_scaffold3162 : 1 44155 : -	habu1_s3162_g10961	habu1_s3162_g10961.t1
nvCTLP-12	C-lectin01	habu1_scaffold3168 : 105257 110525 : -	habu1_s3168_g10976	habu1_s3168_g10976.t1
nvCTLP-13	C-lectin03 (olfactory receptor 5V1-like [Python bivittatus)	habu1_scaffold3168 : 132988 153875 : -	habu1_s3168_g10978	habu1_s3168_g10978.t1
nvCTLP-14	C-lectin04	habu1_scaffold3168 : 158476 201437 : +	habu1_s3168_g10979	habu1_s3168_g10979.t1
nvCTLP-15	C-type lectin	habu1_scaffold5624 : 612654 645778 : -	habu1_s5624_g16096	habu1_s5624_g16096.t1
nvCTLP-16	C-type lectin domain family 2 member A-like	habu1_scaffold2729 : 242646 255780 : +	habu1_s2729_g09708	habu1_s2729_g09708.t1
nvCTLP-17	C-type lectin domain family 2 member B-like	habu1_scaffold4914 : 32203 44645 : -	habu1_s4914_g14978	habu1_s4914_g14978.t1
nvCTLP-18	C-type lectin domain family 2 member D	habu1_scaffold16096 : 1593 16985 : -	habu1_s16096_g21607	habu1_s16096_g21607.t1
nvCTLP-19	C-type lectin domain family 2 member D	habu1_scaffold16095 : 1 33634 : +	habu1_s16095_g21604	habu1_s16095_g21604.t1
nvCTLP-20	C-type lectin domain family 2 member D-like isoform	habu1_scaffold7470 : 7148 64124 : +	habu1_s7470_g18072	habu1_s7470_g18072.t1
nvCTLP-21	C-type lectin domain family 2 member D-like	habu1_scaffold2729 : 287626 301914 : +	habu1_s2729_g09710	habu1_s2729_g09710.t1
nvCTLP-22	C-type lectin domain family 2 member D-like	habu1_scaffold2729 : 269596 280053 : +	habu1_s2729_g09709	habu1_s2729_g09709.t1
nvCTLP-23	C-type lectin domain family 2 member F	habu1_scaffold650 : 139753 147435 : -	habu1_s650_g02983	habu1_s650_g02983.t1
nvCTLP-24	C-type lectin domain family 2 member F	habu1_scaffold16095 : 20706 30572 : +	habu1_s16095_g21605	habu1_s16095_g21605.t1
nvCTLP-25	C-type lectin domain family 4 member F	habu1_scaffold5437 : 1 163815 : -	habu1_s5437_g15753	habu1_s5437_g15753.t1
nvCTLP-26	C-type lectin domain family 4 member F	habu1_scaffold5437 : 175001 184325 : -	habu1_s5437_g15754	habu1_s5437_g15754.t1
nvCTLP-27	C-type lectin domain family 4 member G	habu1_scaffold9852 : 245036 264242 : +	habu1_s9852_g19732	habu1_s9852_g19732.t1
nvCTLP-28	C-type lectin domain family 5 member A-like	habu1_scaffold356 : 6432 15155 : -	habu1_s356_g01674	habu1_s356_g01674.t1
nvCTLP-29	C-type lectin domain family 10 member A	habu1_scaffold399929 : 2366 26492 : +	habu1_s399929_g24822	habu1_s399929_g24822.t1
nvCTLP-30	C-type lectin domain family 10 member A	habu1_scaffold399929 : 28656 34341 : +	habu1_s399929_g24823	habu1_s399929_g24823.t1
nvCTLP-31	C-type lectin domain family 10 member A	habu1_scaffold399929 : 35256 46873 : +	habu1_s399929_g24824	habu1_s399929_g24824.t1
nvCTLP-32	C-type lectin domain family 11 member A	habu1_scaffold6124 : 381916 408060 : +	habu1_s6124_g16816	habu1_s6124_g16816.t1
nvCTLP-33	C-type lectin domain family 17, member A: ow affinity immunoglobulin epsilon Fc receptor-like	habu1_scaffold9852 : 223468 243595 : -	habu1_s9852_g19731	habu1_s9852_g19731.t1
nvCTLP-34	C-type lectin domain family 17, member A	habu1_scaffold2388 : 1078710 1129431 : -	habu1_s2388_g08701	habu1_s2388_g08701.t1
nvCTLP-35	C-type lectin domain family 17, member A	habu1_scaffold4489 : 1161650 1171504 : -	habu1_s4489_g14309	habu1_s4489_g14309.t1
nvCTLP-36	collectin-10	habu1_scaffold24 : 236156 286149 : +	habu1_s24_g00208	habu1_s24_g00208.t1
nvCTLP-37	collectin-11	habu1_scaffold1294 : 282428 290611 : -	habu1_s1294_g04540	habu1_s1294_g04540.t1
nvCTLP-38	collectin-12	habu1_scaffold2067 : 4679 91925 : -	habu1_s2067_g07546	habu1_s2067_g07546.t1
nvCTLP-39	E-selectin isoform X1	habu1_scaffold20406 : 1 19656 : -	habu1_s20406_g22285	habu1_s20406_g22285.t1
nvCTLP-40	olfactory receptor 5V1-like	habu1_scaffold3168 : 205916 208854 : +	habu1_s3168_g10980	habu1_s3168_g10980.t1

Supplementary Figure S5 continued

(c)



Supplementary Figure S6.

(a) SV genes

Gene	Name	Locus	Gene Model ID	Transcripts
svPLA2-01	Asp49PLA2/PL1a/PL2a	habu1_scaffold9571 : 7266 11848 : +	habu1_s9571_g19434	pb016481_c1004773_f1p3_727
svPLA2-02	PL-Y/PL-X'/PL-B	habu1_scaffold47459 : 1268 2998 : -	habu1_s47459_g23397a	a pb016481_c1023909_f1p0_734
svPLA2-03	PLA-N	habu1_scaffold47459 : 1264 3188 : -	habu1_s47459_g23397b	pb016481_c1016263_f1p3_727
svPLA2-04	BPI	habu1_scaffold50376:26421454: -		pb016481_c1030113_f1p0_731
svPLA2-05	BPII	habu1_scaffold9571 : 1 1869 : -		pb016481_c1028655_f1p0_728
svPLA2-06	BPIII	habu1_scaffold401404: 1365: -		
svPLA2-07	PL1b/PL2b	habu1_scaffold9570:1742: +	habu1_s9570_g19432	pb016481_c1024179_f1p5_745
svPLA2-08	PfPLA6	habu1_scaffold9571 : 1 8417 : -	habu1_s9571_g19433	habu1_s9571_g19433.t2
svPLA2-09	pgPLA2b	habu1_scaffold9571:1621617662: +		pb016481_c1027830_f1p0_729

(b) NV genes

Gene	Name	Locus	Gene Model ID	Transcripts
nvPLA2-01	PLA2_G1_1	habu1_scaffold9792:202026204267: +	habu1_s9792_g19655	habu1_s9792_g19655.t1
nvPLA2-02	PLA2_G1_2	habu1_scaffold9792:206686208528: +	habu1_s9792_g19656	habu1_s9792_g19656.t1
nvPLA2-03	PLA2_G2E_1	habu1_scaffold4562 : 336326 340919: -	habu1_s4562_g14470	habu1_s4562_g14470.t1
nvPLA2-04	PLA2_G3_2	habu1_scaffold2696 : 770786 776844: +	habu1_s2696_g09640	habu1_s2696_g09640.t1
nvPLA2-05	PLA2_G3_4	habu1_scaffold3088 : 458 5355: -	habu1_s3088_g10883	habu1_s3088_g10883.t1
nvPLA2-06	PLA2_G10_1	habu1_scaffold194 : 345256 364398: +	habu1_s194_g01103	habu1_s194_g01103.t1
nvPLA2-07	PLA2_G10_2	habu1_scaffold194 : 366490 382195: -	habu1_s194_g01104	habu1_s194_g01104.t1
nvPLA2-08	PLA2_G10_3	habu1_scaffold2789 : 707889 711217: +	habu1_s2789_g09989	habu1_s2789_g09989.t1
nvPLA2-09	PLA2_G12A_1	habu1_scaffold3769 : 1122983 1127025: +	habu1_s3769_g12642	habu1_s3769_g12642.t1
nvPLA2-10	PLA2_G12B_1	habu1_scaffold2074 : 358299 399065: -	habu1_s2074_g07581	habu1_s2074_g07581.t1
nvPLA2-11	PLA2_G15_1	habu1_scaffold499 : 736999 788724: +	habu1_s499_g02314	habu1_s499_g02314.t1
nvPLA2-12	PLA2_G15_3	habu1_scaffold53923 : 12382 17295: -	habu1_s53923_g23499	habu1_s53923_g23499.t1
nvPLA2-13	PLA2_G15_4	habu1_scaffold5562 : 106910 108039: -		pb016480_c10250_f1p0_1130
nvPLA2-14	PLA2	habu1_scaffold2192 : 72907 74929: -	habu1_s2192_g07871	habu1_s2192_g07871.t1
nvPLA2-15	PLA2G4A	habu1_scaffold8317 : 5635 95005: -	habu1_s8317_g18615	habu1_s8317_g18615.t1
nvPLA2-16	PLA2G4B_1	habu1_scaffold3646:16155: -	habu1_s3646_g12259	habu1_s3646_g12259.t1
nvPLA2-17	PLA2G4B_2	habu1_scaffold3645 : 202517 283015: -	habu1_s3645_g12258	habu1_s3645_g12258.t1
nvPLA2-18	PLA2G4C_3	habu1_scaffold11920:77736127822: +	habu1_s11920_g20651	habu1_s11920_g20651.t1
nvPLA2-19	PLA2G4C_4	habu1_scaffold32464 : 1 60356: +	habu1_s32464_g23033	habu1_s32464_g23033.t1
nvPLA2-20	PLA2_B	habu1_scaffold2320 : 573149 601069 : +	habu1_s2320_g08440	habu1_s2320_g08440.t1
nvPLA2-21	PLA2_B	habu1_scaffold2321 : 2106 33861 : +	habu1_s2321_g08441	habu1_s2321_g08441.t1
nvPLA2-22	PLA2G4F	habu1_scaffold2323 : 1 38699 : +	habu1_s2323_g08442	habu1_s2323_g08442.t1
nvPLA2-23	PLA2G4E_2	habu1_scaffold3645 : 9056 58091: +	habu1_s3645_g12255	habu1_s3645_g12255.t1
nvPLA2-24	PLA2G4E_4	habu1_scaffold2324 : 1 70129: +	habu1_s2324_g08443	habu1_s2324_g08443.t1
nvPLA2-25	PLA2G4E_5	habu1_scaffold2324 : 74886 119324: +	habu1_s2324_g08444	habu1_s2324_g08444.t1
nvPLA2-26	PLA2G4E_6	habu1_scaffold2324 : 122456 211367: +	habu1_s2324_g08445	habu1_s2324_g08445.t1
nvPLA2-27	PLA2G6	habu1_scaffold7565 : 780358 836117: +	habu1_s7565_g18177	habu1_s7565_g18177.t1
nvPLA2-28	PNPLA8	habu1_scaffold5091 : 1 61165: -	habu1_s5091_g15231	habu1_s5091_g15231.t1
nvPLA2-29	PLA1	habu1_scaffold5830 : 1665 118015 : -	habu1_s5830_g16316	habu1_s5830_g16316.t1
nvPLA2-30	PLA2_G16	habu1_scaffold1542 : 1 34425 : +	habu1_s1542_g05787	habu1_s1542_g05787.t1
nvPLA2-31	PLA1mA	habu1_scaffold19666 : 1 11400 : +	habu1_s19666_g22161	habu1_s19666_g22161.t1
Supplementary Figure S6 continued













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svSP01_TLf1_flavoxobin habu1_scaffold7597:103760..119614 -Ex1 Ex2 Ex3 Ex4 Ex5 Ex6 -----_ ſ svSP02_TLf2 $\begin{array}{ccc} {}^{habu1_scaffold409039:1..737:} & {}^{habu1_scaffold189200:1..2644:} + & {}^{habu1_scaffold142871:1..912:} + \\ & Ex1_Ex2_Ex3_Ex4_Ex5_Ex6 \end{array}$ ÷C svSP03_TLf3 habu1_scaffold7597:146701...152624:-Ex1 Ex2 Ex3 Ex4 Ex5 Ex6 -----svSP04 habu1_scaffold6789:134126 ... 143039:+ Ex1 Ex2 Ex3 Ex4 Ex5 Ex6 -0-0------**_**___ -0 svSP05 habu1_scaffold6789:149856 ... 207128:+ Ex1 Ex2 Ex3 Ex4 Ex5 Ex6 svSP06 habu1_scaffold4106 : 1768 ... 14932 : +, habu1_scaffold375860:1...711: +, habu1_scaffold50513:1..3838: +, habu1_scaffold73303:1..843: +, habu1_scaffold270900:1..692: +

	Ex1 Ex2	Ex3	Ex4	Ex5	Ex6	Ex7	Ex8
svSP07	habu1_scaffold4106 Ex1 Ex2	Ex3	28186:+ Ex4	Ex5	Ex6	-	
svSP08	habu1_scaffold6789 Ex1 Ex2	Ex3	207128 Ex4	:+ Ex5	Ex6	—	
svSP09	habu1_scaffold7597 Ex1 Ex2	Ex3	155905 : Ex4	- Ex5	Ex6	-	
svSP10	habu1_scaffold4106	; 28886 .	46059 : +	÷			
	Ex1 Ex2	Ex3	Ex4	Ex5	Ex6	-	
svSP11	habu1_scaffold2202 Ex1b Ex1 E	x2 Ex	34705 : 3 Ex4	- Ex5	Ex	6	

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(a)

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Gene	Name	Locus	Gene Model ID	Transcripts
3FTX01 (sv3FTX)	Pf_3FTX-01	habu1_scaffold4579 : 59513 76175 : -	habu1_s4579_g14476	pb016480_c396810_f1p0_507
3FTX02 (sv3FTX)	Pf_3FTX-02	habu1_scaffold4579 : 119426 122076 : +	habu1_s4579_g14477	habu1_s4579_g14477.t1
3FTX03 (sv3FTX)	Pf_3FTX-03	habu1_scaffold4579 : 142940 144965 : -	habu1_s4579_g14478	habu1_s4579_g14478.t1
3FTX04 (sv3FTX)	Pf_3FTX-04	habu1_scaffold4579 : 45965 48865 : -	habu1_s4579_g14475	habu1_s4579_g14475.t1
3FTX05 (nv3FTX)	Pf_3FTX-05 (UPAR_LY6, CD59A)	habu1_scaffold138 : 73600 130505 : -	habu1_s138_g00722	habu1_s138_g00722.t1
3FTX06 (nv3FTX)	Pf_3FTX-06 (CD59B)	habu1_scaffold138 : 62681 68945 : -	habu1_s138_g00721	habu1_s138_g00721.t1



(a)

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Gene		Name	Locus	Gene model ID	Transcripts
APase01	APaseN (svAPase)	aminopeptidase N	habu1_scaffold1390 : 1416444 1437202 : +	habu1_s1390_g05001	pb016481_c405527_f2p3_3459
APase02	APaseNPEPL	aminopeptidase NPEPL	habu1_scaffold5205 : 360816 379669 : +	habu1_s5205_g15384	habu1_s5205_g15384.t1
APase03	APaseA (svAPase)	aminopeptidase A	habu1_scaffold3769 : 814477 861472 : -	habu1_s3769_g12636	pb016481_c564840_f1p8_3942
APase04	APaseB	aminopeptidase B	habu1_scaffold8948 : 297629 311470 : +	habu1_s8948_g19021	habu1_s8948_g19021.t1
APase05	APaseO	aminopeptidase O	habu1_scaffold4166 : 161346 345243 : +	habu1_s4166_g13605	habu1_s4166_g13605.t1
APase06	APaseQ	aminopeptidase Q	habu1_scaffold3446 : 147415 149419 : -	habu1_s3446_g11762	habu1_s3446_g11762.t1
APase07	APaseD	aspartyl aminopeptidase	habu1_scaffold133 : 520486 540509 : +	habu1_s133_g00676	habu1_s133_g00676.t1
APase08	APaseM1	methionine aminopeptidase 1	habu1_scaffold7599 : 75984 96985 : -	habu1_s7599_g18195	habu1_s7599_g18195.t1
APase09	APaseM2	methionine aminopeptidase 2	habu1_scaffold9114 : 242443 254705 : -	habu1_s9114_g19166	habu1_s9114_g19166.t1
APase10	XP APase1	xaa-pro aminopeptidase 1	habu1_scaffold4753 : 86146 168417 : +	habu1_s4753_g14695	habu1_s4753_g14695.t1
APase11	XP APase2	xaa-pro aminopeptidase 2	habu1_scaffold4159 : 322646 384475 : +	habu1_s4159_g13556	habu1_s4159_g13556.t1
APase12	XP APase3	xaa-pro aminopeptidase 3	habu1_scaffold1233 · 167726 _ 197694 · +	habu1 s1233 g04248	habu1 s1233 d04248 t1



<u>(a)</u>

Gene	Name	Locus	Gene model ID	Transcripts
CRISP01 (svCRISP)	CRISP/triflin	habu1_scaffold22025: 114800: -	habu1_s22025_g22470a	pb016480_c42550_f61p14_1314
CRISP02 (svCRISP)	CRISP_like	habu1_scaffold22025:3000967537: +	habu1_s22025_g22470b	habu1_s22025_g22470.t1
CRISP03 (nvCRISP)	CRISP w egf-like domain	habu1_scaffold1243 : 310296 329704 : +	habu1_s1243_g04322	habu1_s1243_g04322.t1
CRISP04 (nvCRISP)	CRISP w egf-like domain	habu1_scaffold264 : 3584 27775 : -	habu1_s264_g01336	habu1_s264_g01336.t1



 \leftarrow Signal sequence \Rightarrow α_1 α_2 β_1 β_2 ex2 ex3 CRISP01 MIAFIVLPILAAVLQQSSGNVDFDSESPRKPEIQNEIIDLHNSLRRSVNPTASNMLKMEW MIAFIALLSFAAVLQQSSGTVDFASESANERETQKEILDKHNALRRSVRPTARNMLQMEW CRISP02 ex2 || ex3 || -<u>a3</u><u>a4</u><u>b3</u><u>b4</u><u>a5</u><u>b5</u> Pathogenesis-related domain ex4 ex5 CRISP01 YPEAAANAERWAYRCIESHSSRDSRVIGGIKCGENIYMATYPAKWTDIIHAWHGEYKDFK NFNAAQNATRWADRCSFAHSPQHLRTVGELKCGENLFMSSHPFPWTRVIQSWYDENKNFK CRISP02 ex4 || ex5 —<u>β7</u>___ ex6 CRISP01 YGVGAVPSNAVVGHYTQI-----YWYKSYRAGCAAAYCPSSKYS-----Y YGVGANPPNAVIGHYTQACLLTFLQNETSGRYLFHNSENSHVAIPFRNTSSKTLMECLDY CRISP02 ex6 T ***** * ********* *
 β9
 β10
 β11
 α7

 ex7
 Hinge domain
 ex8
____β8__) ex8 FYVCQYCPAGNIIGKTATPYKSGPPCGDCPSDCDNGLCTNPCTRENEFTNCDSLVQKSSC CRISP01 CRISP02 RYFTCYGFIQHLLSISSPP---APSCFWKRQSAGALPVPGASAEPRQFCSCLSVRLGSLL ex7 *. * :::. ::.* .*.*:. .:* .* *: * -<u>α8</u>-<u>α9</u>-<u>β12</u>> ICR domain QDNYMKSKCPASCFCQNKII-----CRISP01 CRISP02 L---IPLPCYTWLAIVYHLSCGKVPQVVHQLKSPDLDYR * .*: *:

(d)

CRISP03	ex1 MKAGRAAWRWLEAFAFLVLLAVSLPPASPALGSSEERLRQACNTCRGIADRFTQGLTDTAKKNF	GG
CRISP04	MAFSRMKVPPALFSFLLLFQWMPNSGSSQQDTCQTCRGLVDNFNKGLERTQRENF ex2	GG
	* .* ::. *:**:*: * ***:: ::*:***:.*.*.**	**
	DUF34556(TLR4 regulator)	
	ex2 ex3 ex4	
CRISP03	GNTAWEEKTLSKYESSEIRLVEIIENLCDSSNFECNNMVEEHEEHIENWWFKWKKKYPDLFKWLCIET	IE
CRISP04	GNTAWEEEKLAKYANSETRLLEVLESVCSTSDFACHQLLERSEDHVEHWWFHEQQQHPDFFQWLCMDT	LK
	ex3 ex4 ex5	
	*******:.*:** .** **:*::*.:*:* *::::*. *:*:*:*:	::
	ex5	
CRISP03	VCCPACTHGPCCVCCPGCSEPPCHGNGNCDGDGTPAGDGSCKCOKEVOGEECLDCSDGVVNEVKNDTH	sv
CRISP04	LCCPSGTYGPDCOTCPGGAEKPCSGYGOCDGEGTSGGTGLCMCOTGYGGPEGSECGDGYYEAARNDSH	1.11
CRIDING		114
	100010010000 10 0010100 0 01000100 10 0 0 001 0 0 0 10100001 10010	^
	Our wish (Dunis like) warset	
	Cys-rich (Furin-like) repeat	
	ext ex7	
CRISP03	CTACHDSCKTCTGATNKDCKDCKEGWLRNEEACVDEDECAVEESPCNSDQYCLNTDGSFSCKACDLSC	LG
CRISP04	CAECYRACGRCSGPEDTSCLRCKRGWMLHNQRCIDIDECGTDMAHCRSNQFCVNTEGSYECRDCAKPC	IG
	ex7 ex8	
	*: *: :* *:*. :* **.**: ::: *:* ***: : *.*!*:*:*:*:*:*:*: *: *	:*
	Cys-rich (Furin-like) repeat Ca_binding EGF_like domain	
	ex8 ex9	
CRISP03	CTGEGPNKCKSCVTGYEMKEETCTDVDECSQTEEVCTRENTNCINTPGGYKCICSEGFEDKDDICV	PS
CRISP04	CMGAGPSRCKKCNKGYQRDGVKCLDVDECAGEVEEPVCTGANEVCENTDGSYRCVCAEGHLRKEGICV	-Е
	ex9 ex10	
	* * **.:**.* .**:* *****: * *** * * *	
	ex10	
CRISP03	IKAEEKTSANISSPDTHEDL	
CRISP04	DKPPDAPEKGFFDDTTDDEVVVI.00MFFGATTCALATLAAKGDMVFTATFTGAVAAMAGVWMSERSDR	VT.
0.1.01 0 1	ex11	

CRISPOS		
CRISPUS		
CRISPU4	DGFPINGR	

(c)

(4)				
Gene	Name	Locus	Gene model ID	Transcripts
Ves01 (svVesp)	Vespryn	habu1_scaffold402940 : 157656 165646 : +	habu1_s402940_g24938	habu1_s402940_g24938.t1
Ves02	butyrophilin subfamily	habu1_scaffold402940 : 136268 173215 : -	habu1_s402940_g24937	habu1_s402940_g24937.t1
Ves03	spry domain-containing socs box protein 3 isoform x1	habu1_scaffold2038 : 674725 686992 : +	habu1_s2038_g07466	habu1_s2038_g07466.t1
Ves04	spry domain-containing protein	habu1_scaffold1489 : 27447 30825 : -	habu1_s1489_g05514	habu1_s1489_g05514.t1
Ves05	spry domain-containing socs box protein 4 isoform x1	habu1_scaffold2998 : 118432 298625 : -	habu1_s2998_g10640	habu1_s2998_g10640.t1
Ves06	F-box/SPRY domain-containing protein 1	habu1_scaffold12804 : 318189 339575 : -	habu1_s12804_g20892	habu1_s12804_g20892.t1
Ves07	SPRY domain-containing protein 7	habu1_scaffold13689 : 124686 130582 : +	habu1_s13689_g21202	habu1_s13689_g21202.t1
Ves08	SPRY domain-containing protein 3	habu1_scaffold1516 : 380747 424339 : +	habu1_s1516_g05661	habu1_s1516_g05661.t1
Ves09	fibronectin type III and SPRY domain-containing protein 2-like	habu1_scaffold3496 : 592235 607031 : +	habu1_s3496_g11883	habu1_s3496_g11883.t1
Ves10	spry domain-containing socs box protein 1	habu1_scaffold3776 : 8796 59035 : +	habu1_s3776_g12675	habu1_s3776_g12675.t1
Ves11	fibronectin type iii and spry domain-containing protein 1	habu1_scaffold2525 : 6768 32830 : -	habu1_s2525_g09169	habu1_s2525_g09169.t1
Ves12	ring finger and spry domain-containing protein 1	habu1_scaffold503 : 116026 143018 : +	habu1 s503 g02344	habu1_s503_g02344.t1

(b)



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(a)

<u>(a)</u>				
Gene	Name	Locus	Gene Model ID	Transcripts
5Nase01 (sv5Nase)	sv5Nase	habu1_scaffold6028 : 169856 203427 : +	habu1_s6028_g16570	pb016481_c551957_f58p63_2527
5Nase02 (nv5Nase)	5 -nucleotidase domain-containing protein 2- like	habu1_scaffold1426 : 2901530 2936315 : +	habu1_s1426_g05181	habu1_s1426_g05181.t1
5Nase03 (nv5Nase)	5 -nucleotidase domain-containing protein 2	habu1_scaffold6846 : 55556 83885 : +	habu1_s6846_g17547	habu1_s6846_g17547.t1
5Nase04 (nv5Nase)	5 -nucleotidase domain-containing protein 3	habu1_scaffold428 : 301964 331735 : -	habu1_s428_g02066	habu1_s428_g02066.t1
5Nase05 (nv5Nase)	cytosolic purine 5 -nucleotidase isoform	habu1_scaffold1344 : 510687 545803 : -	habu1_s1344_g04804	habu1_s1344_g04804.t1
5Nase06 (nv5Nase)	5_nucleotid	habu1_scaffold1548 : 105056 126204 : +	habu1_s1548_g05833	habu1_s1548_g05833.t1
5Nase07 (nv5Nase)	5_nucleotid	habu1_scaffold8872 : 26926 110043 : +	habu1_s8872_g18961	habu1_s8872_g18961.t1
5Nase08 (nv5Nase)	5 (3)- cytosolic type	habu1_scaffold3440 : 270746 276672 : +	habu1_s3440_g11638	habu1_s3440_g11638.t1
5Nase09 (nv5Nase)	3 (2) -bisphosphate nucleotidase	habu1_scaffold304 : 182426 196985 : -	habu1_s304_g01433	habu1_s304_g01433.t1
5Nase10 (nv5Nase)	cytosolic 5 -nucleotidase	habu1_scaffold18 : 2268164 2279432 : -	habu1_s18_g00174	habu1_s18_g00174.t1
5Nase11 (nv5Nase)	cytosolic 5 -nucleotidase 1b	habu1 scaffold2849 : 462585 469949 : +	habu1 s2849 g10145	habu1 s2849 g10145.t1





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Gene	Name	Locus	Gene Model ID	Transcripts
DPP01 (nvDPP)	DPP-I	habu1_scaffold1219 : 174906 207609 : +	habu1_s1219_g04195	habu1_s1219_g04195.t1
DPP02 (nvDPP)	DPP-II	habu1_scaffold998 : 329636 331823 : +	habu1_s998_g03532	habu1_s998_g03532.t1
DPP03 (nvDPP)	DPP-II	habu1_scaffold998 : 322306 326986 : +	habu1_s998_g03530	habu1_s998_g03530.t1
DPP04 (nvDPP)	DPP-III	habu1_scaffold7877 : 9596 37295 : +	habu1_s7877_g18328	habu1_s7877_g18328.t1
DPP05 (svDPP)	svDPP-IV	habu1_scaffold1020 : 1996805 2061742 : +	habu1_s1020_g03622	pb016480_c171896_f1p3_4438
DPP06 (nvDPP)	DPP-VIII	habu1_scaffold1390 : 679600 693217 : -	habu1_s1390_g04966	habu1_s1390_g04966.t1
DPP07 (nvDPP)	DPP-IX	habu1_scaffold5905 : 38286 91928 : +	habu1_s5905_g16391	habu1_s5905_g16391.t1
DPP08 (nvDPP)	DPP-X	habu1_scaffold13003 : 203276 412685 : -	habu1_s13003_g20945	habu1_s13003_g20945.t1



Supplementary Figure S18 (a)

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Gene	Name	Locus	Gene model ID	Transcripts
svVEGF01	svVEGF(svVEGF_F)	habu1_scaffold565 : 505831 554056 : -	habu1_s565_g02679	pb016481_c76319_f71p22_1316
nvVEGF01	VEGF-A	habu1_scaffold6836 : 129006 390238 : +	habu1_s6836_g17529	habu1_s6836_g17529.t1
nvVEGF02	VEGF-C	habu1_scaffold9381 : 115236 156444 : +	habu1_s9381_g19343	habu1_s9381_g19343.t1



(a)

<u> </u>				
Gene	Name	Locus	Gene Model ID/Annotation Link	Transcripts
PDE_01	PDE_01(svPDE)	habu1_scaffold149 : 1524169 1594790 : +	habu1_s149_g00804	pb016481_c221783_f35p18_2633
PDE_02	PDE_02 (nvPDE)	habu1_scaffold149 : 1600729 1649852 : +	habu1_s149_g00805	habu1_s149_g00805.t1
PDE_03	PDE_03(nvPDE/cAMP)	habu1_scaffold149 : 3362511 3486620 : +	habu1_s149_g00834	habu1_s149_g00834.t1



Supplementary Figure S20 (a)

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Gene	Name	Locus	Gene Model ID	Transcripts
PLB01 (svPLB)	phospholipase b	habu1_scaffold1233 : 1498267 1531920 : +	habu1_s1233_g04284	pb016481_c662_f13p9_1819
PLB02	phospholipase b-like 2	habu1_scaffold1964 : 35979 45965 : -	habu1_s1964_g07200	habu1_s1964_g07200.t1
PLB03	phospholipase b1(membrane-associated)	habu1_scaffold313 : 1861453 1973632 : -	habu1_s313_g01520	habu1_s313_g01520.t1
PLB04	60 kDa lysophospholipase	habu1_scaffold3352 : 459066 534431 : -	habu1_s3352_g11424	habu1_s3352_g11424.t1
PLB05	lysophospholipase-like	habu1_scaffold303 : 156916 176572 : +	habu1_s303_g01427	habu1_s303_g01427.t1



(a)

Gene	Name	Locus	Gene model ID	Transcripts
CNP01	BPP_CNP (svCNP)	habu1_scaffold258676: 1337: - habu1_scaffold4348 : 15402 16141 : -	habu1_s258676_g24318	habu1_s258676_g24318.t1
CNP02	B-type natriuretic peptides A-like	habu1_scaffold20540 : 7387 10895 : -	habu1_s20540_g22311	habu1_s20540_g22311.t1



(a)

Gene	Name	Locus	Gene model ID	Transcripts
GPCase01 (svGPCase)	glutaminyl-peptide cyclotransferases	habu1_scaffold510 : 291600 305425 : -	habu1_s510_g02384	pb016481_c398311_f1p1_2071
GPCase02 (nvGPCase)	glutaminyl-peptide cyclotransferases	habu1_scaffold3067 : 69888 75755 : -	habu1_s3067_g10859	habu1_s3067_g10859.t1

