

Supporting Information

Identification of novel binding sites for heparin in RPTPs: implications for proteoglycan signaling

Yasuhiro Katagiri, Ashlea A. Morgan, Panpan Yu, Nathanael J. Bangayan, Radoslaw Junka, and Herbert M. Geller

Figure S1

Schematic structure of RPTP σ recombinant proteins used in this study

Figure S2

Similar expression levels of recombinant proteins were detected in transfected 293 cells.

Figure S3

Intact phosphatase domain of RPTP σ is essential for the upregulated intracellular phosphorylation upon heparin exposure.

Figure S4

Size exclusion chromatography of GAGs used in this study

Figure S5

Heparinase treatment of transfected cells increases the sensitivity to exogenous heparin

Table S1

Disaccharide composition analysis of chondroitin / dermatan sulfate and heparin

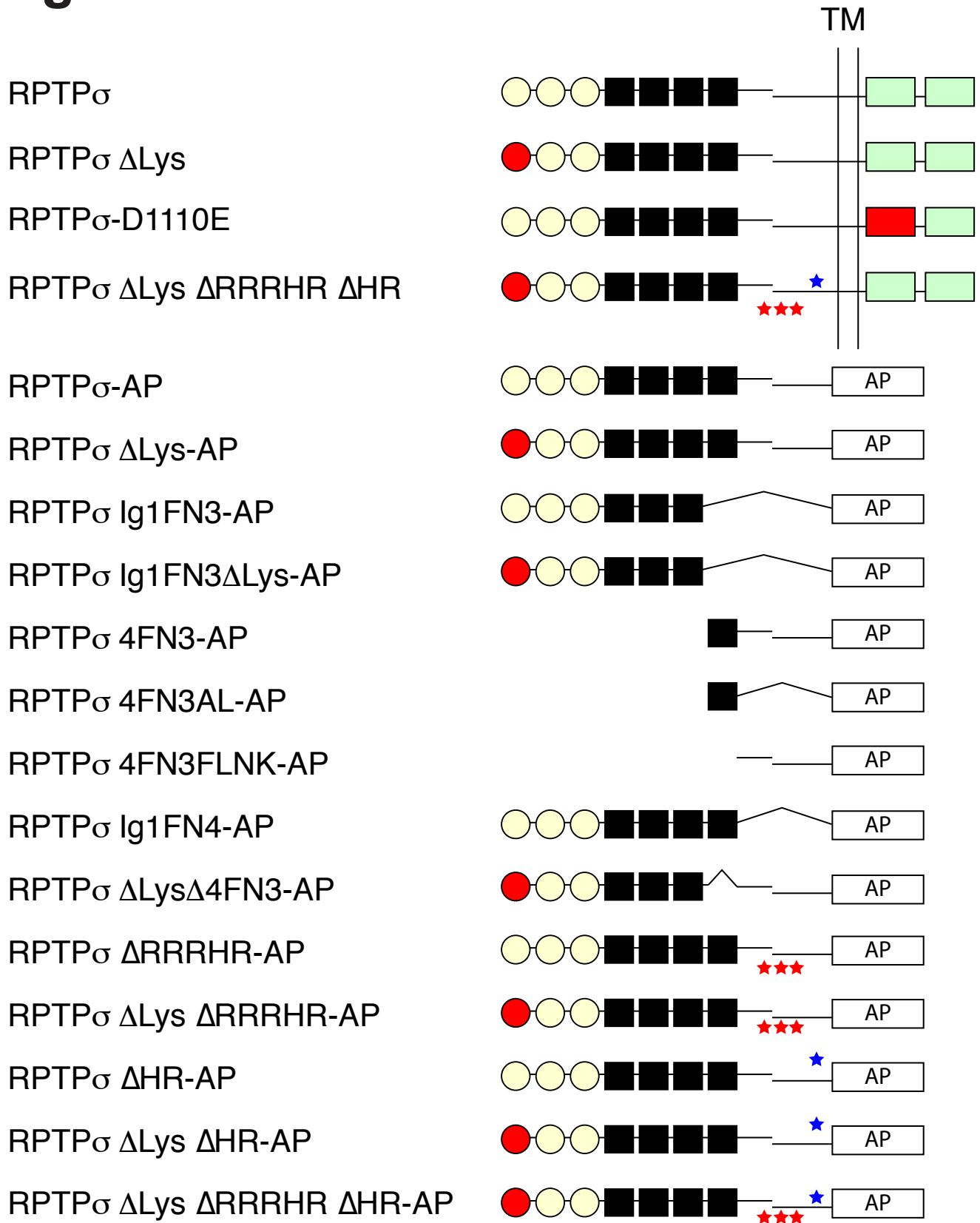
Table S2

Potential heparin binding sites within the fourth FNIII flanking region selected by “heparin protection” assays

Table S3

DNA plasmids and oligonucleotides used in this study

Figure S1



○ Ig domain ■ FNIII domain □ Phosphatase domain

● Δ Lys mutation ★★★ Δ RRRHR mutation ★ Δ HR mutation

Schematic structure of RPTP σ recombinant proteins used in this study

Figure S2

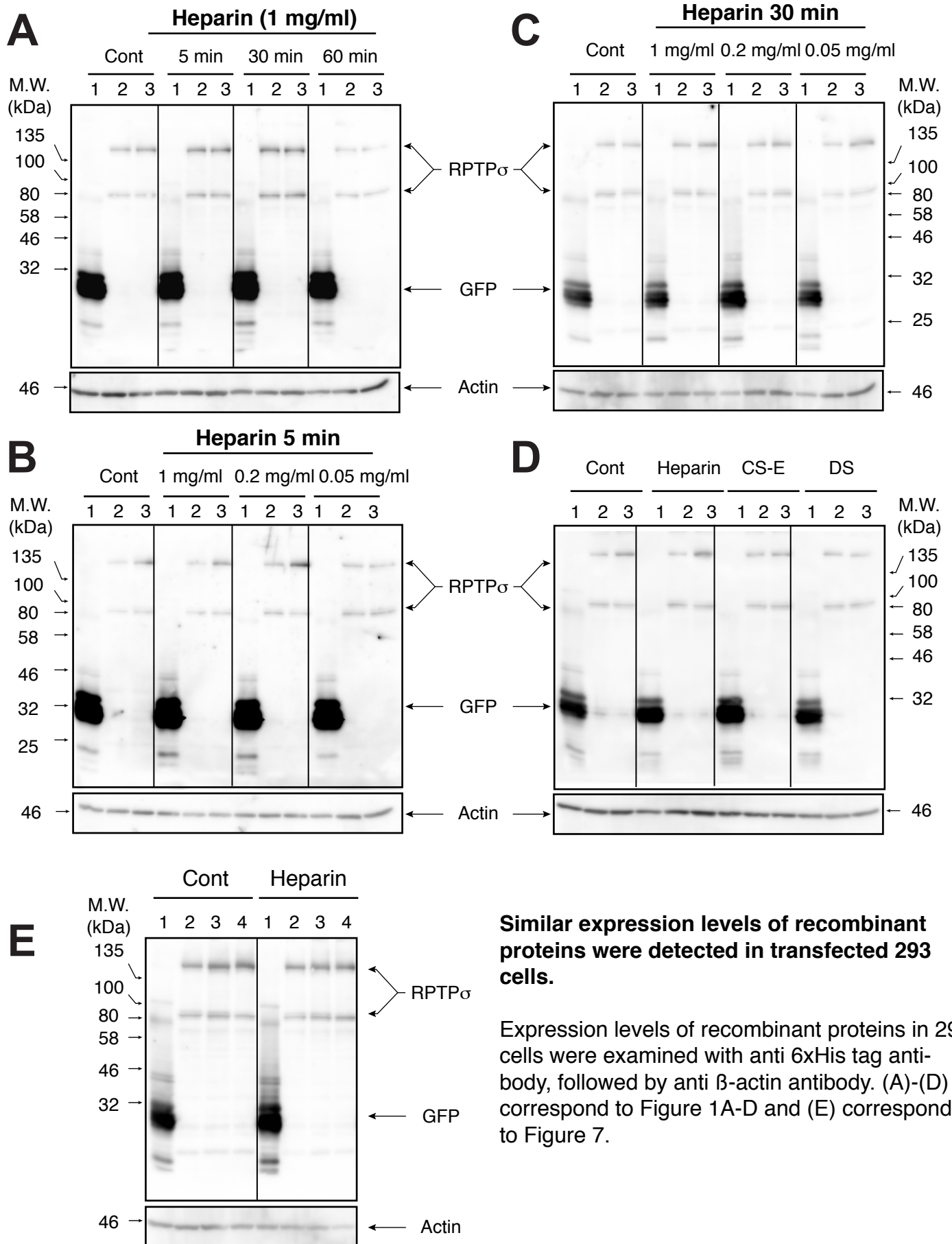
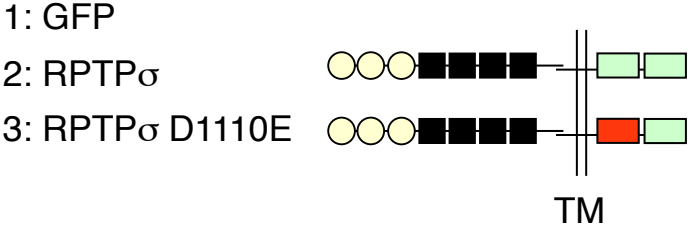
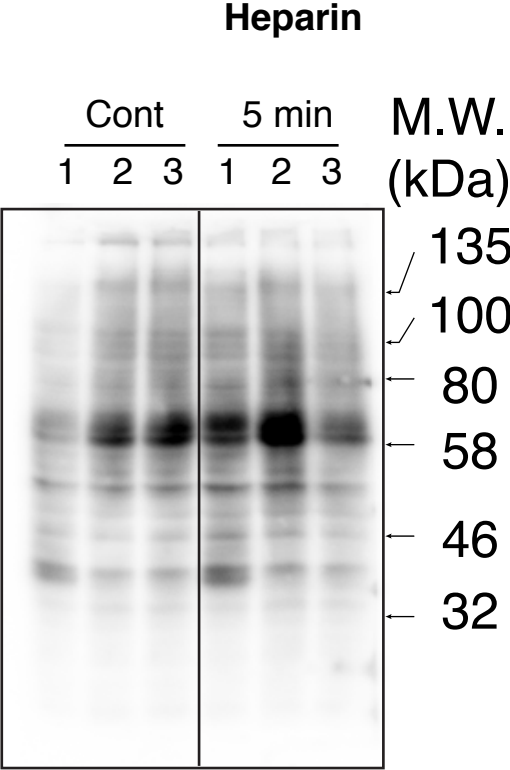


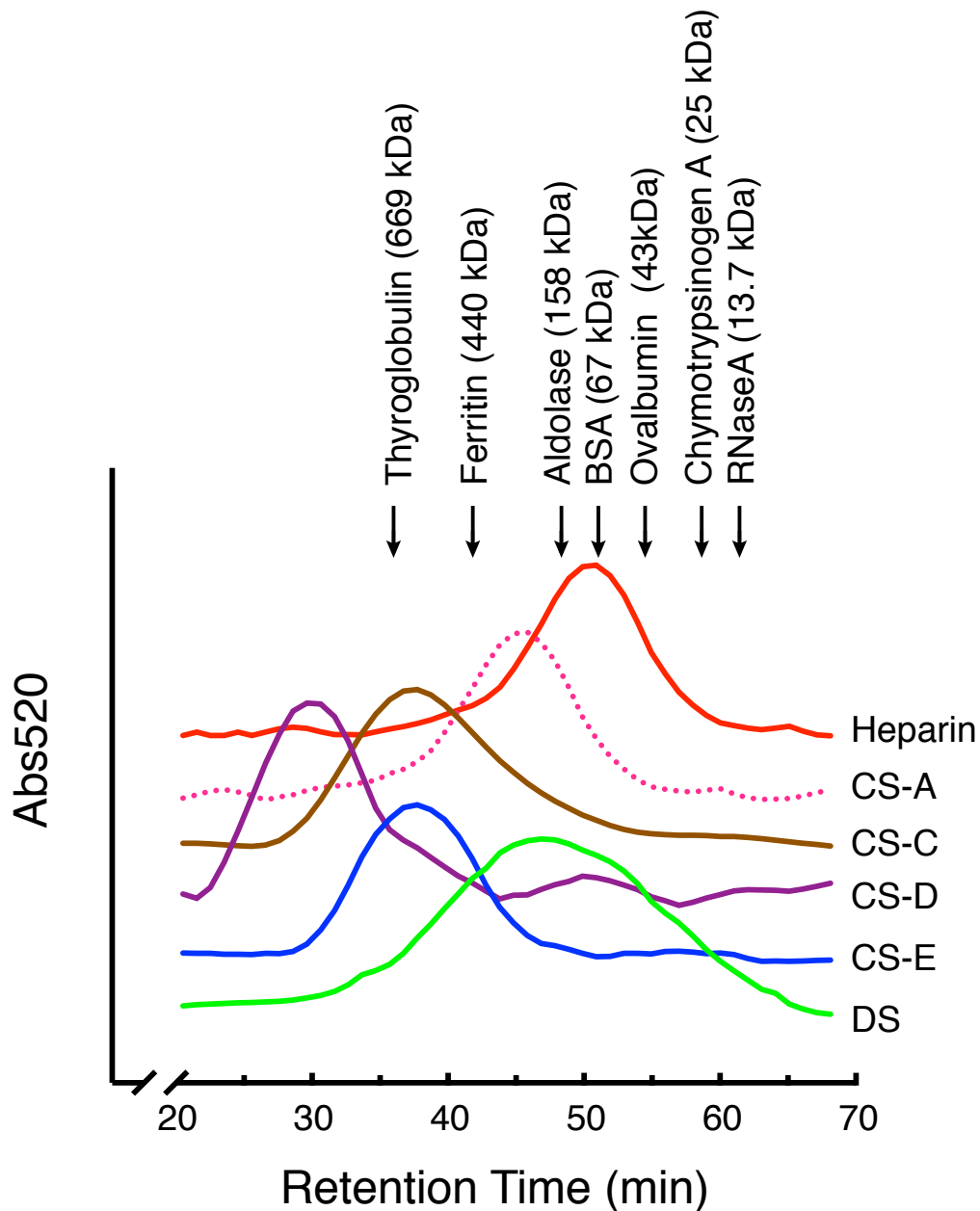
Figure S3



Intact phosphatase domain of RPTP σ is essential for the upregulated intracellular phosphorylation upon heparin exposure.

293 cells expressing (1) GFP, (2) RPTP σ , and (3) RPTP σ D1110E (catalytically inactive form) were treated with heparin (1 mg/ml) for 5 min and Tyr phosphorylation was examined by immunoblot.

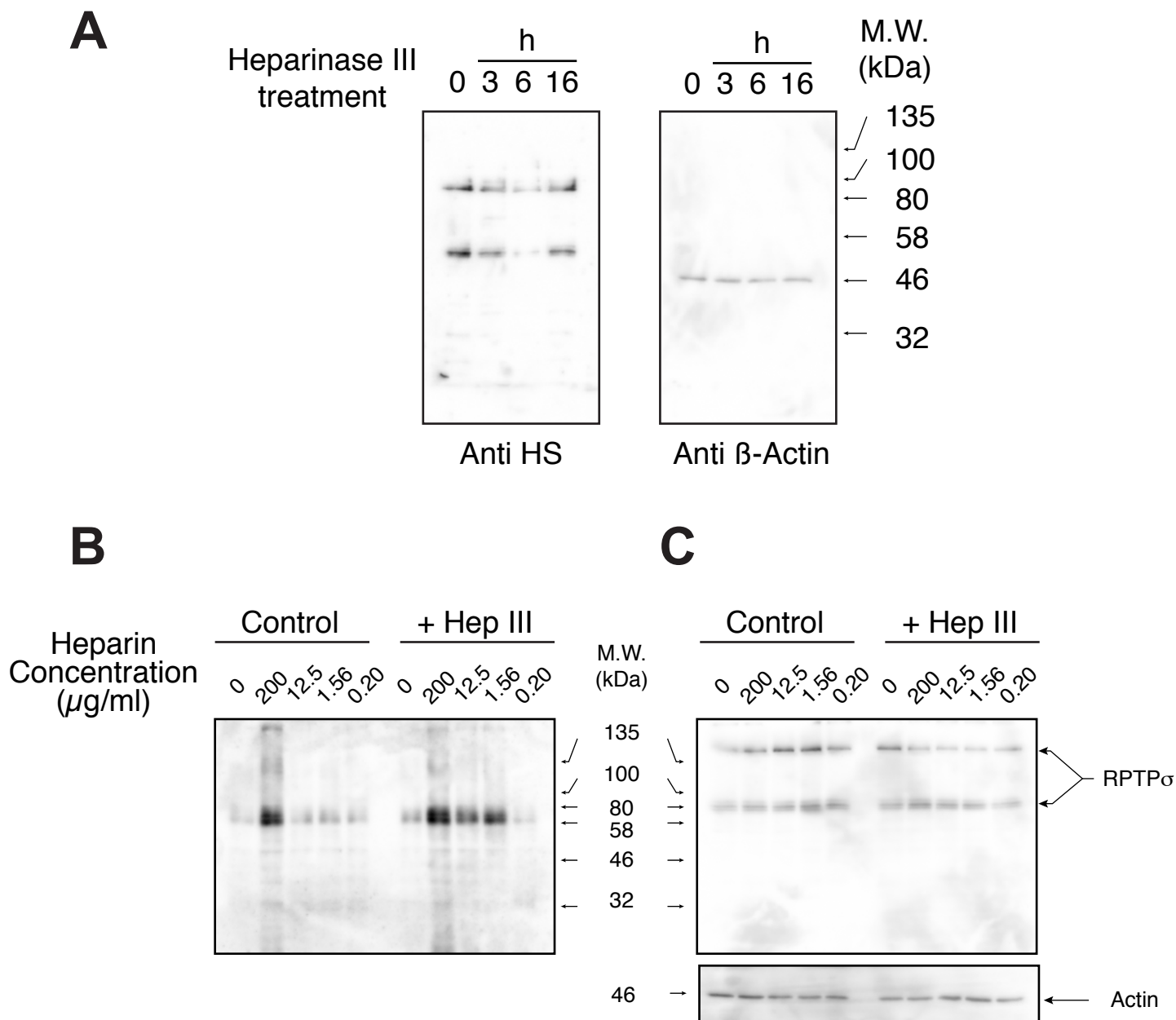
Figure S4



Size exclusion chromatography of GAGs used in this study

Each GAG was separated in 150 mM ammonium bicarbonate by size-exclusion chromatography (Superdex 200, 10 x 300 mm, GE Healthcare Life Sciences) attached to a Shimadzu UFLC system at a flow rate of 0.3 ml/min. Amount of uronic acid in each fraction was measured by the carbazole method.

Figure S5



Heparinase treatment of transfected cells increases the sensitivity to exogenous heparin

Transfected 293 cells with RPTP σ (full length) were treated with heparinase III (Sigma, H8891). (A) At indicated time, cell lysates were collected with 1% TritonX100 in PBS containing Protease Inhibitor Cocktail (Sigma #539137), followed by immunoblot with anti HS antibody (F58-10E4, AMSBIO, # 370255). After stripping, PVDF membrane was reprobbed with anti β -actin antibody. (B) After 8 h treatment with heparinase III, cells were treated with heparin for 5 min and harvested with 2 x SDS sample buffer. Tyrosine phosphorylation was detected by immunoblot with anti-phosphotyrosine antibody. (C) Similar expression level of RPTP σ was confirmed with anti-6xHis tag antibody, followed by reprobbed with anti β -actin antibody.

Table S1**Disaccharide composition analysis of chondroitin and dermatan sulfate**

Mol %	Δ Di-0S ^a	Δ Di-4S	Δ Di-6S	Δ Di-2,6S	Δ Di-4,6S	Δ Di-2,4,6S
CS-A	6.3	54.2	39.5	0	0	0
CS-C	13.9	26.9	58.3	1.0	0	0
CS-D	2.2	12.8	42.3	42.7	0	0
CS-E	3.4	26.6	25.5	0	44.6	0
DS	7.7	74.6	16.0	0.9	0.9	0

Disaccharide composition analysis of heparin

Mol %	D0H0 ^b	D0A0	D0H6	D2H0	D0S0	D0A6	D2A0	D2H6	D0S6	D2S0	D2A6	D2S6
Heparin	0	12.4	0	0	7.7	7.4	6.8	0	20.7	23.1	2.9	19.0

Unsaturated chondroitin disaccharides generated by digestion with chondroitinase ABC were analyzed by anion-exchange HPLC after labeling with the fluorophore 2AB as described in Experimental Procedures section. Analysis of heparin composition was performed by the UCSD Glycotechnology Core.

^a Δ Di-0S= Δ HexUA-GalNAc; Δ Di-6S= Δ HexUA-GalNAc(6-*O*-sulfate); Δ Di-4S= Δ HexUA-GalNAc(4-*O*-sulfate); Δ Di-2,6S= Δ HexUA(2-*O*-sulfate)-GalNAc(6-*O*-sulfate); Δ Di-4,6S= Δ HexUA-GalNAc(4,6-*O*-disulfate); Δ Di-2,4,6S= Δ HexUA(2-*O*-sulfate)-GalNAc(4,6-*O*-sulfate).

^bD0H0, Δ HexUA-GlcNH₂; D0A0, Δ HexUA-GlcNAc; D0H6, Δ HexUA-GlcNH₂-6S; D2H0, Δ HexUA2S-GlcNH₂; D0S0, Δ HexUA-GlcNS; D0A6, Δ HexUA-GlcNAc6S; D2A0, Δ HexUA2S-GlcNAc; D2H6, Δ HexUA2S-GlcNH₂6S; D0S6, Δ HexUA-GlcNS6S; D2S0, Δ HexUA2S-GlcNS; D2A6, Δ HexUA2S-GlcNAc6S; D2S6, Δ HexUA2S-GlcNS6S.

Table S2**Potential heparin binding sites within the fourth FNIII flanking region selected by “heparin protection” assays**

Peptides are sorted according to the position in RPTP σ . Modified amino acids are shown as the addition of numbers; [+226.07760] indicates a modification by biotin and [+15.99492] indicates Met oxidation.

Position	Sequence	Observed m/z	Calculated m/z	ppm error
667	F.VLTNR[+226.07760]GSSL.G	586.8114	586.8082	5.39
669	L.TNRGSSLGGLQQTVTAR[+226.07760]TAF.N	764.3949	764.3902	6.10
669	L.TNRGSSLGGLQQTVTAR[+226.07760].T	658.0101	658.0058	6.49
669	L.TNR[+226.07760]GSSLGGLQQTVTAR.T	658.0094	658.0058	5.41
669	L.TNR[+226.07760]GSSLGGLQQT.VT.A	872.9407	872.936	5.47
669	L.TNR[+226.07760]GSSLGGLQQT.VT	822.4167	822.4121	5.55
671	N.R[+226.07760]GSSLGGLQQT.VT.A	765.3954	765.3907	6.26
676	L.GGLQQTVTAR[+226.07760]TAF.N	788.4061	788.401	6.48
676	L.GGLQQTVTAR[+226.07760].T	628.8281	628.8244	5.89
684	T.AR[+226.07760]TAF.N	396.1991	396.1971	5.13
689	F.NM[+15.99492]LSGK[+226.07760]PSVAPKPDNDGF.I	706.0024	705.9994	4.22
689	F.NM[+15.99492]LSGK[+226.07760]PS.V	538.2523	538.2492	5.86
689	F.NMLSGK[+226.07760]PS.V	530.2546	530.2517	5.41
692	L.SGK[+226.07760]PSVAPKPDNDGFIVVY.L	739.378	739.3734	6.21
692	L.SGKPSVAPK[+226.07760]PDNDGF.I	871.42	871.4143	6.57
697	S.VAPK[+226.07760]PDNDGF.I	643.3	643.2977	3.56
724	Y.FIVM[+15.99492]VPL.R	834.4849	834.4793	6.68
731	L.RKSR[+226.07760]GGQFPVLL.G	528.636	528.6328	6.03
732	R.KSR[+226.07760]GGQFPVLL.G	714.3995	714.395	6.26
732	R.KSR[+226.07760]GGQFPVLL.G	714.3994	714.395	6.09
733	K.SR[+226.07760]GGQFPVLL.G	650.3516	650.3475	6.30
733	K.SR[+226.07760]GGQFPVLL.G	650.3512	650.3475	5.64
754	L.IQDISR[+226.07760]LQ.R	599.8191	599.8161	5.05
754	L.IQDISR[+226.07760]L.Q	535.7903	535.7868	6.55
783	F.SILPAVFHPGNQK[+226.07760]QY.G	962.9961	962.9905	5.76
788	A.VFHPGNQK[+226.07760]QY.G	722.3497	722.3455	5.75
790	F.HPGNQK[+226.07760]QYGGF.D	729.8372	729.8328	6.01
790	F.HPGNQK[+226.07760]QY.G	599.2809	599.2771	6.40
790	F.HPGNQK[+226.07760]Q.Y	517.7482	517.7454	5.26
801	F.DNRGLEPGHR[+226.07760]YVLF.V	633.6529	633.6492	5.82
801	F.DNRGLEPGHR[+226.07760]Y.V	513.9119	513.9089	5.93
815	F.VLAVLQK[+226.07760]NEPTF.A	792.9311	792.9263	6.00
817	L.AVLQK[+226.07760]NEPTF.A	686.8535	686.8501	4.97
820	L.QK[+226.07760]NEPTFAASPF.S	781.8738	781.869	6.17
820	L.QK[+226.07760]NEPTF.A	545.2583	545.2553	5.48

Table S3

DNA plasmids and oligonucleotides used in this study

DNA Plamid Name	Gene of Interest	Protein Region	Vector Backbone	Cloning site(s)	Mutations	Primers used	Accession Number
pTAG5-RPRP σ	RPTP σ	1-1502	pAPTAG5	NheI(5') and XhoI(3')	None	1,2	NM_001252456
pTAG5-RPRP σ Δ Lys	RPTP σ	1-1502	pAPTAG5	NheI(5') and XhoI(3')	K68A, K69A, K71A, K72A	1,2,4,5	NM_001252456
pTAG5-RPRP σ D1110E	RPTP σ	1-1502	pAPTAG5	NheI(5') and XhoI(3')	D1110E	1,2,16,17	NM_001252456
pTAG5-RPRP σ Δ Lys Δ RRRHR Δ HR	RPTP σ	1-1502	pAPTAG5	NheI(5') and XhoI(3')	K68A, K69A, K71A, K72A	1,2,4,5,10-13,18,19	NM_001252456
pAPTAG5-RPRP σ	RPTP σ	1-847	pAPTAG5	NheI(5') and HindIII (3')	None	1,3	NM_001252456
pAPTAG5-RPRP σ Δ Lys	RPTP σ	1-847	pAPTAG5	NheI(5') and HindIII (3')	K68A, K69A, K71A, K72A	1,3-5	NM_001252456
pAPTAG5-RPRP σ Ig1FN3	RPTP σ	1-602	pAPTAG5	NheI(5') and HindIII (3')	None	1,6	NM_001252456
pAPTAG5-RPRP σ Ig1FN3 Δ Lys	RPTP σ	1-602	pAPTAG5	NheI(5') and HindIII (3')	K68A, K69A, K71A, K72A	1,4-6	NM_001252456
pAPTAG5-RPRP σ 4FN3	RPTP σ	601-847	pAPTAG5	HindIII (5' and 3')	None	3,8	NM_001252456
pAPTAG5-RPTP σ Ig1FN4	RPTP σ	1-682	pAPTAG5	NheI(5') and HindIII (3')	None	1,7	NM_001252456
pAPTAG5-RPTP σ 4FN3 FLNK	RPTP σ	673-847	pAPTAG5	HindIII (5' and 3')	None	3,9	NM_001252456
pAPTAG5-RPTP σ 4FN3AL	RPTP σ	601-682	pAPTAG5	HindIII (5' and 3')	None	8,9	NM_001252456
pAPTAG5-RPTP σ Δ 4FN3	RPTP σ	1-602, 673-847	pAPTAG5	NheI(5') and HindIII (3')	None	1,3,6,9	NM_001252456
pAPTAG5-RPTP σ Δ 4FN3 Δ Lys	RPTP σ	1-602, 673-847	pAPTAG5	NheI(5') and HindIII (3')	K68A, K69A, K71A, K72A	1,3-6,9	NM_001252456
pAPTAG5-RPTP σ Δ RRRHR	RPTP σ	1-847	pAPTAG5	NheI(5') and HindIII (3')	R762A, R763A, R766A, H767A, R769A	1,3,10-13	NM_001252456
pAPTAG5-RPTP σ Δ Lys Δ RRRHR	RPTP σ	1-847	pAPTAG5	NheI(5') and HindIII (3')	K68A, K69A, K71A, K72A, R762A, R763A, R766A, H767A, R769A	1,3-5,10-13	NM_001252456
pAPTAG5-RPTP σ Δ HR	RPTP σ	1-847	pAPTAG5	NheI(5') and HindIII (3')	H809A, R810A	1,3,18,19	NM_001252456
pAPTAG5-RPTP σ Δ Lys Δ HR	RPTP σ	1-847	pAPTAG5	NheI(5') and HindIII (3')	K69A, K71A, K72A, H809, R810A	1,3-5,18,19	NM_001252456
pAPTAG5-RPTP σ Δ Lys Δ RRRHR Δ HR	RPTP σ	1-847	pAPTAG5	NheI(5') and HindIII (3')	K68A, K69A, K71A, K72A, R762A, R763A, R766A, H767A, R769A, H809A, R810A	1,3-5,10-13,18,19	NM_001252456
pFUSE-RPTP σ	RPTP σ	1-847	pFUSE hIgl	EcoRI(5') and EcoRV(3')	None	14,15	NM_001252456
pGFPTAG5	EGFP	1-239	pAPTAG5	BspEI(5') and XhoI (3')	None	20,21	U55763
pGFPTAG5-RPTP σ	RPTP σ	1-847	pAPTAG5	NheI(5') and HindIII (3')	None	1,3,20,21	U55763, NM_001252456
pLNCX chick v-src	v-src		pLNCX				NM_205457

Table S3 (continued)

Oligonucleotides	
Name	Sequence
1	TGGCTAGCGCCGCCATGGCGCCACCTGGAGTCCCA
2	GAGCGCTTCCTCGAGTCCTGTTGCATAATGATCAAACTGC
3	GGACCCTCAGCCATTGTGGAAGCTTAC
4	CCTAAGCCACGGGTGACCTGGAACgcccgcGGGagcGgcAGTGAACTCACAGCGCTTCGAGACC
5	GGTCTCGAAGCGCTGTGAGTTCAGTgcCgctCCCgcggeGTTCCAGGTCACCCGTGGCTTAGG
6	gtaagcttCCTGCAGCGTGCCTGGCGCAC
7	ttaagcttCAGTGACCGTCTGCTGCAGG
8	atgaagcttacCTGCAGGCCATCTCCCCAAAG
9	cgaagettaCAGCAGCCTGGGGGGCCTGC
10	GACATCTCCCGGCTGCAGgcGgctAGCCTGCGCCACTCCAG
11	CTGGAGTGGCGCAGGCTagcCgcCTGCAGCCGGGAGATGTC
12	CAGgcGgctAGCCTGgcCgcCTCtgcgCAGCTGGAGGTGCCTC
13	GAGGCACCTCCAGCTGcgcaGAGgcGgcCAGGCTagcCgcCTG
14	AAGAATTCAAGCTGGCTAGCGCCGCCATGG
15	TTGATATCCACAATGGGCTGAGGGTCC
16	TGGGGTACTCAGGTACtCCGTGtTCGGGCCATGCCGT
17	ACGGCATGGCCCGAaCACGGaGTACCTGAGTACCCCA
18	GCACAAAGAGGACATAtgCgCCTGGCTCCAAGCC
19	GGCTTGGAGCCAGGcCgcTATGTCCTCTTTGTGC
20	aatccggaATGGTGAGCAAGGGCGAGGAGCTG
21	aactcgagCTTGTACAGCTCGTCCATGC